




Disorders of the Nervous System

Selective Effects of PDE10A Inhibitors on Striatopallidal Neurons Require Phosphatase Inhibition by DARPP-32^{1,2,3}

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Abstract

Type 10A phosphodiesterase (PDE10A) is highly expressed in the striatum, in striatonigral and striatopallidal medium-sized spiny neurons (MSNs), which express D₁ and D₂ dopamine receptors, respectively. PDE10A inhibitors have pharmacological and behavioral effects suggesting an antipsychotic profile, but the cellular bases of these effects are unclear. We analyzed the effects of PDE10A inhibition *in vivo* by immunohistochemistry, and imaged cAMP, cAMP-dependent protein kinase A (PKA), and cGMP signals with biosensors in mouse brain slices. PDE10A inhibition in mouse striatal slices produced a steady-state increase in intracellular cAMP concentration in D₁ and D₂ MSNs, demonstrating that PDE10A regulates basal cAMP levels. Surprisingly, the PKA-dependent AKAR3 phosphorylation signal was strong in D₂ MSNs, whereas D₁ MSNs remained unresponsive. This effect was also observed in adult mice *in vivo* since PDE10A inhibition increased phospho-histone H3 immunoreactivity selectively in D₂ MSNs in the dorsomedial striatum. The PKA-dependent effects in D₂ MSNs were prevented in brain slices and *in vivo* by mutation of the PKA-regulated phosphorylation site of 32 kDa dopamine- and cAMP-regulated phosphoprotein (DARPP-32), which is required for protein phosphatase-1 inhibition. These data highlight differences in the integration of the cAMP signal in D₁ and D₂ MSNs, resulting from stronger inhibition of protein phosphatase-1 by DARPP-32 in D₂ MSNs than in D₁ MSNs. This study shows that PDE10A inhibitors share with antipsychotic medications the property of activating preferentially PKA-dependent signaling in D₂ MSNs.

Key words: biosensor imaging; cAMP; phosphodiesterase; protein kinase; schizophrenia; striatum

Significance Statement

The striatum is mainly composed of medium-sized spiny neurons that express either dopamine D₁ receptors or dopamine D₂ receptors. Their activity is associated with either the initiation of movement or action suppression, respectively. Biosensor imaging revealed that pharmacological inhibition of type 10A phosphodiesterase increased cAMP levels in D₁ and D₂ neurons in the same manner, but only D₂ neurons exhibited an increase in the protein kinase A-mediated phosphorylation level. This effect resulted from an asymmetrical regulation of phosphatases by DARPP-32. D₂ neurons are thus more prone to respond to a tonic cAMP signal than D₁ neurons, a property that may explain how phosphodiesterase 10A inhibitors produced antipsychotic-like behavioral effects. This D₁/D₂ imbalance may also be critical for reward-mediated learning and action selection.

Introduction

Schizophrenia is a devastating psychiatric disease, which results in persistent cognitive and emotional impairments. Type 10A phosphodiesterase (PDE10A) inhibitors were recently proposed as a treatment for schizophrenia (Kehler and Nielsen, 2011; Chappie et al., 2012); however, their cellular mechanisms of action remain unclear with respect to their putative therapeutic effects. PDE10A is highly and almost exclusively expressed in medium-sized spiny neurons (MSNs) of the striatum (Seeger et al., 2003; Coskran et al., 2006; Heiman et al., 2008; Lakics et al., 2010; Kelly et al., 2014). MSNs are divided into two categories based on their expression of dopamine receptors and their sites of projection, as follows: MSNs projecting to the substantia nigra highly express dopamine D₁ receptors (hereafter termed D₁ MSNs); whereas, MSNs projecting to the external globus pallidus highly express adenosine A_{2A} and dopamine D₂ receptors (hereafter termed D₂ MSNs; Gerfen et al., 1990; Le Moine and Bloch, 1995; Bertran-Gonzalez et al., 2008; Matamalas et al., 2009). The high expression of PDE10A in MSNs, and its interaction with the scaffold protein AKAP150 (A-kinase anchoring protein 150), protein kinase A (PKA), PSD-95, and NMDA receptor suggests an important role in modulating the spread of the synaptic cAMP signals into the cell body (Russwurm et al., 2015).

Besides PDE10A, striatal neurons express a number of specific signaling proteins that markedly differ from those in other brain regions (Girault, 2012), and that determine the characteristics of the cAMP/PKA signaling pathway (Castro et al., 2013). Among these specific proteins, DARPP-32 (32-kDa dopamine and cAMP-regulated phosphoprotein) is a multifunctional protein regulating phosphatase and kinase activities: for example, when DARPP-32 is phosphorylated at threonine 34 residue (Thr34) by PKA, it becomes a potent inhibitor of serine/threonine protein phosphatase-1 (PP-1; Hemmings et al., 1984; Svenningsson et al., 2004), increasing the duration of PKA-dependent signals (Castro et al., 2013).

Classical and atypical antipsychotic agents share the property of inhibiting D₂ receptors and thus, in the striatum, increase PKA-dependent phosphorylation selectively in D₂ MSNs (Bateup et al., 2008; Bertran-Gonzalez et al., 2008, 2009). In contrast, psychostimulants, which are psychotomimetic, activate many signaling responses in D₁ MSNs (Bateup et al., 2008; Bertran-Gonzalez et al., 2008, 2009). PDE10A inhibitors were shown to increase cAMP levels in the striatum (Schmidt et al., 2008) and could be expected to mimic the effects of both antipsychotic and psychotomimetic compounds. We used biosensor imaging to precisely analyze the effects of PDE10A inhibitors on cAMP/PKA signaling at the level of individual D₁ and D₂ MSNs. Our work revealed that although PDE10A inhibition increased intracellular cAMP levels in both D₁ and D₂ MSNs, the downstream consequences at the level of PKA targets were profoundly different: the cAMP signal resulting from PDE10A inhibition strongly increased PKA-dependent phosphorylation in D₂ MSNs, whereas D₁ MSNs remained mostly unaffected. Further analyses showed that the difference required DARPP-32-dependent regulation of phosphatase activity in D₁ and D₂ MSNs.

Materials and Methods

Animals

Animals were housed under standardized conditions with a 12 h light/dark cycle, stable temperature (22 ± 1°C), controlled humidity (55 ± 10%), and food and water available ad libitum. Homozygous mice expressing DARPP-32 with the T34A or T75A mutation (Svenningsson et al., 2003) were obtained by crossing heterozygous mice, on a mixed C57BL6/J-Sv129 background (a gift of Dr. P. Greengard, The Rockefeller University, New York). Male Drd2-EGFP heterozygous mice (C57Bl6/J) were generated as described previously (Gong et al., 2003). Experiments were performed in accordance with the regulations under the control of the local ethic committee Charles Darwin C2EA - 05.

Live brain slice preparation

Brain slices were prepared from male mice that were 8–12 days of age. Coronal brain slices were cut with a VT1200S microtome. Slices were prepared in an ice-cold solution of the following composition (in mM): 125 NaCl, 0.4 CaCl₂, 1 MgCl₂, 1.25 NaH₂PO₄, 26 NaHCO₃, 25 glucose, and 1 kynurenic acid, saturated with 5% CO₂ and 95% O₂. The slices were incubated in this solution for 30 min and then placed on a Millicell-CM membrane (Millipore) in culture medium (50% Minimum Essential Medium, 50% HBSS, 6.5 g/L glucose, penicillin-streptomycin; Invitrogen). We used the Sindbis virus as a vector to induce expression of the various biosensors after overnight incubation (Ehrengruber et al., 1999). The coding sequences of Epac-S^{H150} (Polito et al., 2013), AKAR2-NLS (Zhang et al., 2005), AKAR3 (Allen and Zhang, 2006), and cygnet2 (Honda et al., 2001) were inserted into the viral vector pSinRep5 (Invitrogen). The viral vector (~5 × 10⁵ particles per slice) was added, and slices were incubated overnight at 35°C under an atmosphere containing 5% CO₂. Before the experiment, slices were

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incubated for 30 min in the recording solution (125 mM NaCl, 2 mM CaCl₂, 1 mM MgCl₂, 1.25 mM NaH₂PO₄, 26 mM NaHCO₃, and 25 mM glucose, saturated with 5% CO₂ and 95% O₂). Recordings were performed with a continuous perfusion of the same solution at 32°C. MSNs constitute 95% of neurons in the striatum. Large neurons (smallest soma diameter, >14 μm), presumably cholinergic interneurons, were excluded.

Live brain slice imaging

For two-photon imaging, excitation was obtained using a Ti:sapphire laser (MaiTai HP; Spectra Physics) tuned at 850 nm for CFP excitation. Galvanometric scanners (model 6210; Cambridge Technology) were used for raster scanning, and a piezo-driven objective scanner (P-721 PIFOC; Physik Instrumente GmbH) was used for z-stack image acquisition. The system was controlled by MPscope software (Nguyen et al., 2006). The microscope was based on an Olympus BX51WI upright microscope with a 40× 0.8 numerical aperture (NA) or 60× 0.9 NA water-immersion objective. A two-photon emission filter was used to reject residual excitation light (E700 SP; Chroma Technology). A fluorescence cube containing 479/40 and 542/50 emission filters and a 506 nm dichroic beamsplitter (FF01-479/40, FF01-542/50 and FF506-Di02-25x36 Brightline Filters; Semrock) was used for the orthogonal separation of the two fluorescence signals. Two imaging channels (H9305 photomultipliers; Hamamatsu) were used for simultaneous detection of the two types of fluorescence emission. For each data point, an image stack of 30–40 images with a 0.5 μm interval was acquired.

Wide-field images were obtained with an Olympus BX50WI or BX51WI upright microscope with a 40× 0.8 NA water-immersion objective and an ORCA-AG Camera (Hamamatsu). Images were acquired with iVision (Biovision). The excitation and dichroic filters were D436/20 and 455dcxt. Signals were acquired by alternating the emission filters, HQ480/40 for CFP, and D535/40 for yellow fluorescent protein, with a filter wheel (Sutter Instruments). These filters were obtained from Chroma Technology.

No correction for bleed-through or direct excitation of the acceptor was applied, since this correction, while increasing the absolute amplitude of ratio changes, also increases the noise in the measurement (Ducros et al., 2009).

The biosensor chromophores are sensitive to nonspecific environmental disturbances. We used a mutated version of AKAR3 in which the threonine residue of the PKA phosphorylation site was replaced with an alanine residue (T391A) as a control. This AKAR3 (T391A) control sensor reported no ratio change in response to PDE10A inhibition in MSNs.

Data analysis

Images were analyzed with custom routines written in the IGOR Pro environment (Wavemetrics). The emission ratio was calculated for each pixel, as follows: F535/F480 for AKAR2-NLS and AKAR3, and F480/F535 for cygnet2 and Epac-S^{H150} sensors. The pseudocolor images display

the ratio value coded in hue and the fluorescence of the preparation coded in intensity.

Two-photon imaging was used to separate individual neurons for a precise quantification of the amplitude of the response (Figs. 1, 2). Ratio measurements were performed on a series of 5–10 consecutive image from the image stack, centered on the cell body. With cytosolic biosensors, when visible, the nucleus was excluded from the measurement. Wide-field imaging (Figs. 3A–E) also allowed the unambiguous identification of D₁ and D₂ MSNs, provided that the infection level was kept low and no fluorescence overlap between neighboring neurons was observed. The optical cross-contamination resulting from out-of-focus light was evaluated by the final response to CGS 21680 and SKF-38393, applied sequentially: cells were rejected from analysis if the cross-contamination was >30%. For cGMP imaging (Fig. 3F,G), the data were quantified as relative ratio change.

Quantifications of cAMP signals

The amplitudes of responses were quantified for each neuron as the fractional change in ratio from its own baseline and maximal final ratio response. Responses obtained from MSNs of the same type were averaged for each experiment (i.e., brain slice).

The free cAMP concentrations were estimated with the Epac-S^{H150} biosensor from the ratio measurement using the Hill equation, with a K_d of 4.4 μM and a Hill coefficient of 0.77, as determined from Polito et al. (2013). The maximal response corresponding to biosensor saturation (R_{max}) was determined for each neuron at the end of the recording. This level was obtained by applying 13 μM forskolin (FSK), a dose known to be sufficient to maximally phosphorylate the highly sensitive probe AKAR3 in MSNs. For cAMP biosensors, this R_{max} value was obtained with 200 μM IBMX and 13 μM forskolin.

The baseline cAMP level in control conditions was evaluated by inhibiting adenylyl cyclases with 200 μM SQ22536, which resulted in a ratio decrease, measured in wide-field microscopy, of –4.0% of the maximal ratio change. This decrease in baseline ratio was -4.9 ± 0.7 and -3.3 ± 0.7 ($n = 6$, $p < 0.05$ with paired Student's t test), respectively, in D₁ and D₂ MSNs. Assuming that adenylyl cyclase inhibition effectively decreased cAMP levels down to a level sufficient to reach the minimal ratio level (R_{min}), these values suggest a baseline cAMP concentration in a range of ~100 nM.

Tissue preparation and immunofluorescence

Mice, 8–10 weeks old, were treated with the drug for 60 min and then rapidly anesthetized with pentobarbital (500 mg/kg, i.p.; Sanofi-Aventis) and were transcardially perfused with 4% (w/v) paraformaldehyde in 0.1 M PBS, pH 7.5. Brains were post-fixed overnight in the same solution and stored at 4°C. The 30-μm-thick sections were cut with a vibratome and stored at –20°C in a solution containing 30% (v/v) ethylene glycol, 30% (v/v) glycerol and 0.1 M sodium phosphate buffer, until they were processed for immunofluorescence. Sections were processed as described in Bertran-Gonzalez et al. (2009). Sodium fluoride 0.1 mM was included in all buffers and incubation solu-

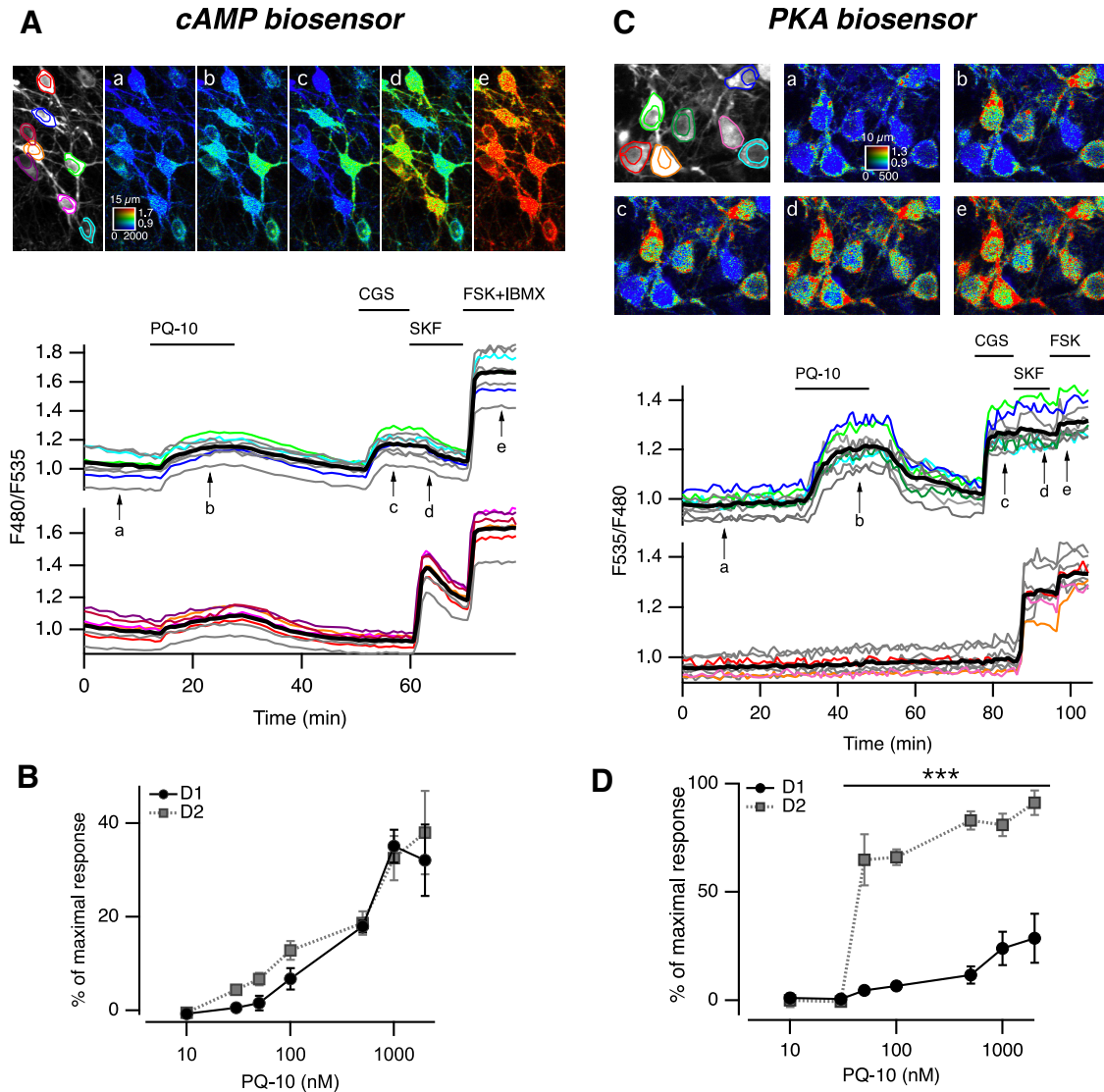


Figure 1. PDE10A inhibition increases cAMP levels in both in D₁ and D₂ MSNs, and PKA-dependent phosphorylation only in D₂ MSNs. **A**, MSNs in **a** neostriatal mouse brain slice expressing the cAMP biosensor Epac-S^{H150} were imaged with two-photon microscopy during the application of PQ-10 (100 nM). Images (vertical projection of the image stack) show the raw fluorescence at 535 nm (left, in grayscale) and the ratio (in pseudocolor) indicating intracellular cAMP concentrations, at the times indicated by the arrows on the graph below. The calibration square in **A** indicates the spatial scale (the size of the square is indicated in micrometers), and shows the ranges of intensity (horizontally) and ratio (vertically). Each trace on the graph indicates the F480/F535 emission ratio measured in regions indicated by the color contour drawn on the raw image. Traces in gray correspond to regions that are not visible on these images. Traces are plotted in two groups according to their response to either CGS 21680, an adenosine A_{2A} receptor agonist (CGS, 1 μM), or SKF-38393, a D₁-like receptor agonist (SKF, 1 μM). The thick black line represents the average of all the traces in a group. FSK (13 μM) and IBMX (200 μM) were applied at the end of the recording to determine the maximal response. **B**, The same experiment was repeated for every PQ-10 concentration tested. No significant difference was found between D₁ and D₂ MSNs (two-way ANOVA: dose effect, $F_{(6,54)} = 40.91, p < 10^{-4}$; D₁/D₂ effect, $F_{(1,54)} = 2.56, p = 0.115$; dose × D₁/D₂ interaction, $F_{(6,54)} = 0.625, p = 0.709$). Error bars indicate the SEM. **C**, Same as **A**, except that the AKAR3 biosensor was used to monitor PKA-dependent phosphorylation, and the ratio was calculated as F535/F480. **D**, Same as **B** for AKAR3 measurements. Data were analyzed with two-way ANOVA: dose effect, $F_{(6,38)} = 28.31, p < 10^{-4}$; D₁/D₂ effect, $F_{(1,38)} = 143.73, p < 10^{-4}$; dose × D₁/D₂ interaction, $F_{(6,38)} = 9.23, p < 10^{-4}$ Bonferroni's *post hoc* test, *** $p < 0.001$.

tions. Histone H3 phosphorylation was revealed with a rabbit polyclonal antibody against phospho-Ser10-H3 (1:1000; catalog #06570; Millipore) the specificity of which was confirmed in a previous study (Jordi et al., 2013). GFP was detected using chicken antibody against GFP (1:500;

catalog #A10262; Life Technologies). Following incubation with primary antibodies, sections were rinsed three times for 10 min in TBS and incubated for 45–60 min with goat Cy3-coupled (1:500; Jackson ImmunoResearch; Fig. 5) and goat A488 (1:500; Life Technologies; Fig. 6). Sec-

tions were rinsed for 10 min twice in Tris-buffered saline and twice in Tris buffer (0.25 M Tris) before mounting in 1,4-diazabicyclo-[2. 2. 2]-octane (Sigma-Aldrich).

Single-labeled images (Fig. 6) were obtained with a Zeiss LSM780 Confocal Microscope. Double-labeled images (Fig. 5) were obtained with a Leica TCS SPE Confocal Microscope with laser lines at 496 and 561 nm, acquiring in the 501–539 nm and 570–660 nm bands. All parameters were held constant for all sections from the same experiment.

Statistics

Data were analyzed with SPSS statistical software version 22.0. Normality in variable distributions and homogeneity of variances across groups were assessed with the Shapiro–Wilk and Levene tests, respectively. Variables that failed any of these tests were analyzed with nonparametric statistics using the Kruskal–Wallis ANOVA on ranks followed by Mann–Whitney rank sum test with a Dunn–Sidak adjustment test for pairwise multiple comparisons. Variables that passed the normality test were analyzed with ANOVA followed by Bonferroni post hoc test for multiple comparisons or by Student's *t* test for comparing two groups. Paired data were analyzed with a Student's *t* test. A *p* value of <0.05 was used as a cutoff for statistical significance. All error bars represent the SEM; *n* indicates the number of experiments (i.e., the number of brain slices tested), with at least four neurons of each type in each experiment. All experiments were performed on at least three different brain slices from at least two animals.

Drugs

SKF-38393 hydrobromide, CGS 21680 hydrochloride, 1-methyl-3-isobutylxanthine (IBMX), rolipram, papaverine, roscovitine, okadaic acid, cantharidin, gabazine, CNQX, APV, and forskolin were obtained from Tocris Cookson. TTX was from Latoxan. PQ-10, MP-10, and roflumilast were a gift from Janssen Pharmaceuticals. TP-10 was provided by Pfizer through the Compound Transfer Program.

Results

PDE10A inhibition reveals a tonic cAMP production in both D₁ and D₂ MSNs

Since both D₁ and D₂ MSNs express high levels of PDE10A protein (Nishi et al., 2008), we used biosensor-imaging approaches to compare the effects of PDE10A inhibition on the cAMP/PKA signaling cascade in D₁ and D₂ MSNs. First, we monitored changes in intracellular cAMP concentrations with two-photon microscopy in striatal brain slices expressing Epac-S^{H150}. At the end of every experiment, an agonist of adenosine A_{2A} receptors (CGS 21680, 1 μM) and an agonist of dopamine D₁ receptors (SKF-38393, 1 μM) were applied sequentially, triggering a positive cAMP response in D₂ and D₁ MSNs, respectively, thereby functionally identifying MSN subtypes. The final application of the general adenylyl cyclase activator FSK (13 μM) together with the nonselective phosphodiesterase inhibitor IBMX (200 μM) produced the maximal ratio response used for normalization.

PDE10A inhibition with PQ-10 (100 nM) increased cAMP levels in all MSNs (Fig. 1A) in a dose-dependent manner. This dose dependency was not statistically different between D₁ and D₂ MSNs (Fig. 1B). At the highest doses (1 and 2 μM), cAMP responses reached ~35% of the maximal response to FSK plus IBMX, which corresponds to a concentration of free cAMP of ~2 μM (see Materials and Methods for details on the estimation of cAMP concentrations). These experiments showed that, in the basal condition, cAMP is tonically produced in striatal slices and that PDE10A contributes significantly to its degradation.

PDE10A inhibition increases the phosphorylation of a PKA target exclusively in D₂ MSNs

We then analyzed the effects of PQ-10 on PKA-dependent phosphorylation levels using the PKA biosensor AKAR3. As for cAMP imaging, MSNs were identified at the end of each experiment by their response to either A_{2A} or D₁ receptor agonist. The maximal AKAR3 response was elicited by FSK. Although the increase in free cAMP concentration was similar in D₁ and D₂ MSNs (Fig. 1A,B), the resulting PKA-dependent phosphorylation levels were completely different (Fig. 1C): PQ-10 (100 nM) strongly increased the emission ratio of AKAR3 in the D₂ MSNs (66 ± 4% of the maximal response to FSK, *n* = 4); whereas, in D₁ MSNs, the ratio remained at a much lower level (6 ± 2%). These results indicated a significantly higher phosphorylation of AKAR3 in D₂ than in D₁ MSNs in response to PQ-10. The effect of PQ-10 on AKAR3 ratio in D₂ MSNs was steeply dose dependent with a maximal effect reached at <100 nM (Fig. 1D). In stark contrast to D₂ MSNs, even high doses of PQ-10, which increased cAMP to the same levels in D₁ and D₂ MSNs (Fig. 1B), only produced a very small effect on AKAR3 phosphorylation in D₁ MSNs (Fig. 1D). These experiments thus revealed a much stronger effect of cAMP on PKA-dependent phosphorylation in D₂ than in D₁ MSNs.

PDE10A may be addressed differentially in the cytoplasm and membranes (Kotera et al., 2004; Charych et al., 2010), and cAMP dynamics could differ in subcellular domains of different geometry, like dendrites (Castro et al., 2010). An increase in AKAR3 ratio was observed exclusively in the dendritic branches that responded to the A_{2A} agonist, whereas dendrites, which responded to the D₁ agonist, showed no response to 100 nM PQ-10 (Fig. 2A). This is consistent with other biosensor recordings in which dendrites of D₁ MSNs also exhibited no baseline response to PDE10A inhibition (Yagishita et al., 2014).

Once activated, PKA can translocate to the nucleus and phosphorylate a number of nuclear proteins. We examined whether the differential response to PQ-10 also existed in the nucleus. Using the nuclear AKAR2-NLS biosensor, we found that PDE10A inhibition induced a strong ratio increase in D₂ MSNs, while D₁ MSNs remained unresponsive (Fig. 2B). These results showed that PQ-10 efficiently increased AKAR3 phosphorylation in the cytoplasm and nucleus of D₂ but not D₁ MSNs, whereas they had a similar and selective effect on cAMP production in the two cell types.

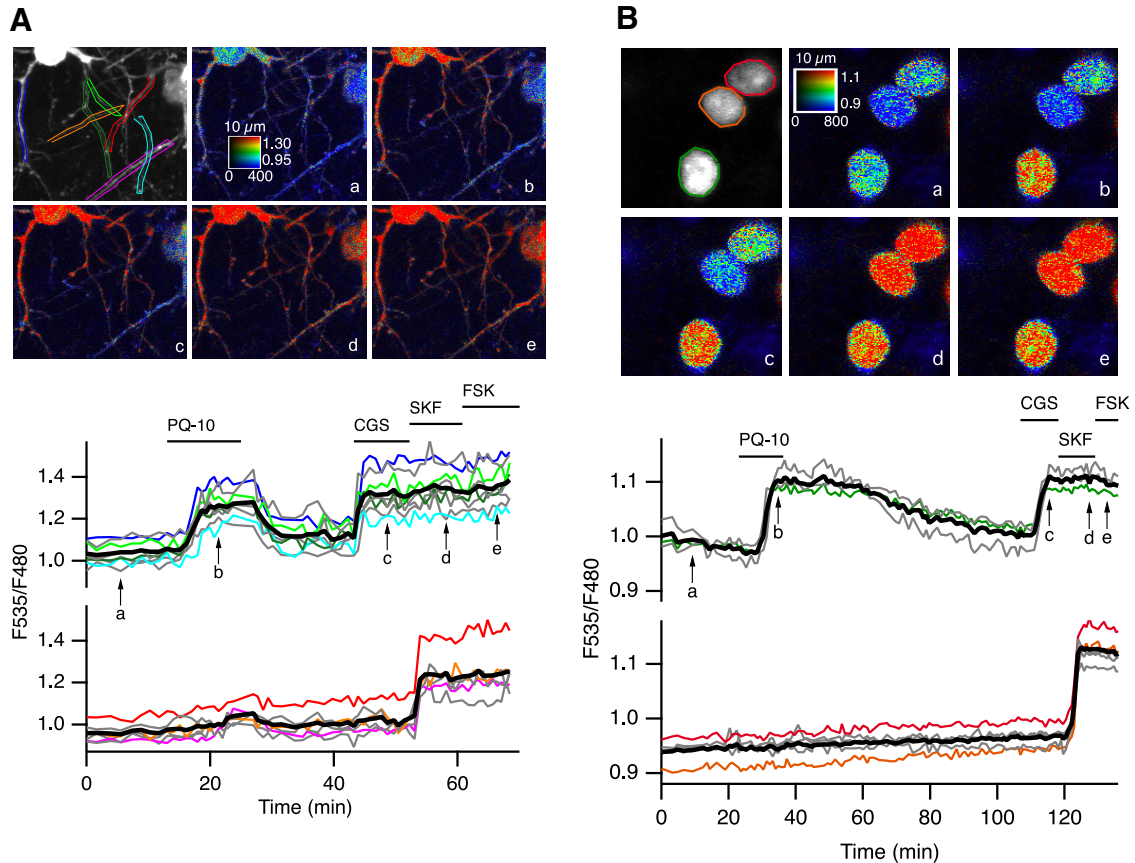


Figure 2. PDE10A inhibition triggers positive PKA responses in dendrites and nuclei preferentially in D_2 MSNs. **A**, **B**, Brain slices expressed the PKA sensor AKAR3 (**A**) or AKAR2-NLS (**B**) and were imaged by two-photon microscopy during the application of PQ-10 (100 nM). Images show the raw fluorescence at 535 nm (left in grayscale) and the ratio (in pseudocolor) indicating the PKA-dependent phosphorylation level of the biosensor, at the times indicated by the arrows on the graph below. The calibration square in **A** indicates the spatial scale (above, in micrometers), and shows the ranges of intensity (horizontally) and ratio (vertically). Each trace on the graph indicates the F535/F480 emission ratio measured on regions indicated by the color contour drawn on the raw image. Traces are plotted in two groups according to their response to either CGS 21680 (CGS, 1 μ M) or SKF-38393 (SKF, 1 μ M). The thick black line represents the average of all the traces in a group. FSK (13 μ M) was applied at the end of the recording to determine the maximal response.

PDE10A inhibition effects are antagonized by D_2 receptors and independent of A_{2A} receptors

One feature of striatopallidal MSNs is the coexpression of D_2 dopamine and A_{2A} adenosine receptors, negatively and positively coupled to adenylyl cyclase, respectively (Schiffmann et al., 1991; Schiffmann and Vanderhaeghen, 1993; Ferré et al., 1997; Svenningsson et al., 1999; Bateup et al., 2008; Bertran-Gonzalez et al., 2009). Application of the D_2 receptor agonist quinpirole (1 μ M) completely reversed the PQ-10-induced AKAR3 response in D_2 MSNs, monitored with wide-field microscopy (Fig. 3A). Application of quinpirole alone had no effect on the basal AKAR3 ratio but prevented positive responses to PQ-10 in D_2 MSNs with $11 \pm 3\%$ ($n = 5$) of the maximal FSK response in D_2 MSNs compared with $9 \pm 2\%$ in D_1 MSNs (Fig. 3B). These results showed that the activation of D_2 receptors opposed the effect of PQ-10 in D_2 MSNs, most likely via Gi-mediated inhibition of the tonic adenylyl cyclase activity. These experiments also further confirmed

that the positive AKAR3 response to PQ-10 was specific to MSNs expressing D_2 receptors.

We then determined whether the effect selectivity for D_2 MSNs was a particular property of PQ-10 or was also observed with other PDE10A inhibitors. The effects of MP-10 (100 nM) and papaverine (1 μ M) on AKAR3 ratio were similar (Fig. 3C,D), inducing an AKAR3 ratio increase selectively in D_2 MSNs. TP-10 also produced the same response profile (see below). In contrast, PDE4 inhibitors (rolipram, 100 nM, $n = 4$; and roflumilast, 1 μ M, $n = 4$) had no effect on basal AKAR3 ratio (data not shown).

As D_2 MSNs express adenosine A_{2A} receptors, we examined whether the tonic presence of extracellular adenosine in our brain slice preparation activated adenylyl cyclase and might be responsible for the positive response to PQ-10 recorded specifically in these MSNs. When A_{2A} receptors were blocked with SCH 58261 (100 nM), PQ-10 still elicited positive responses in D_2 MSNs (Fig. 3E). In contrast, SCH 58261 blocked the responses

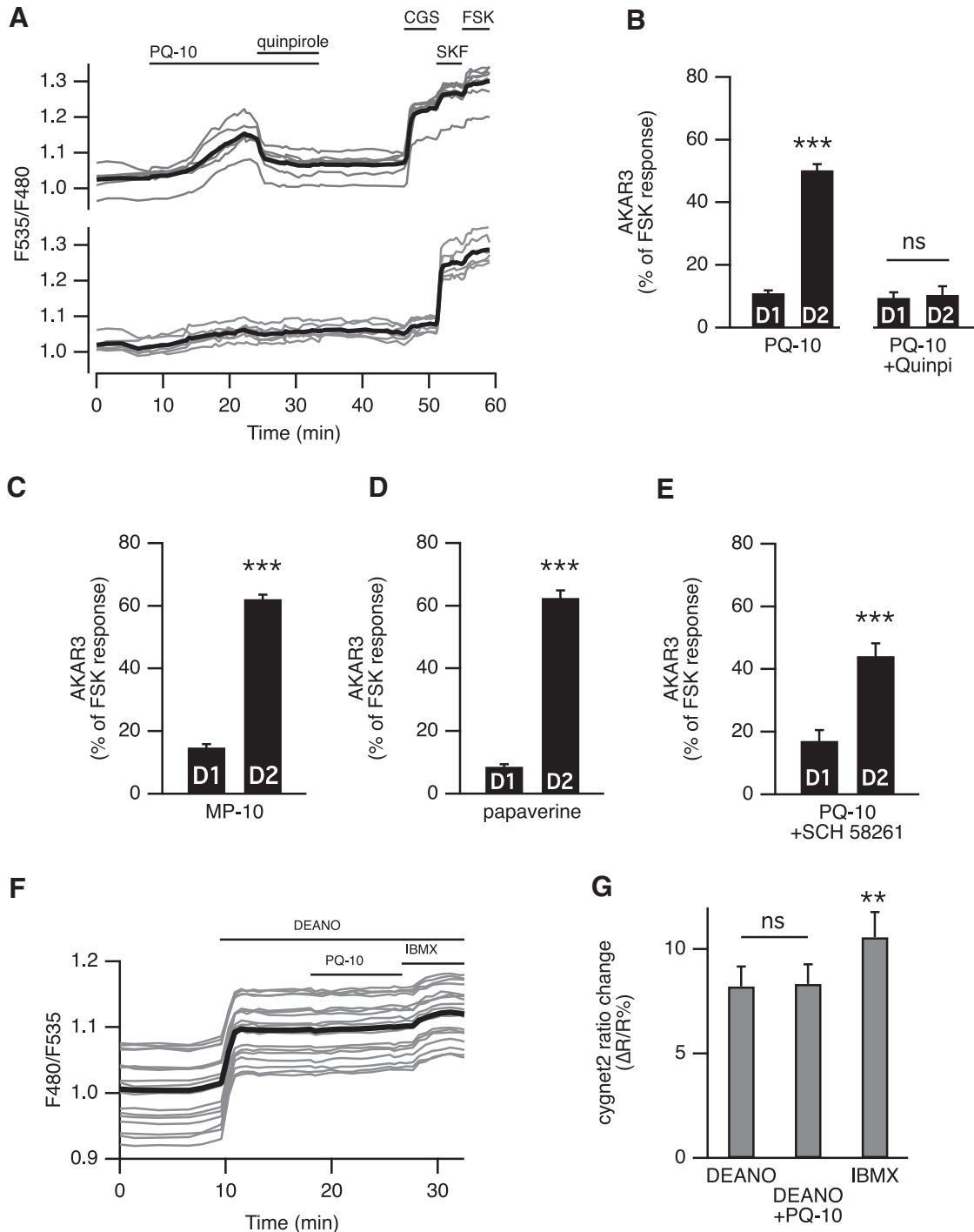


Figure 3. **A**, Activation of D₂ dopamine receptors suppressed the effect of PDE10A inhibition on AKAR3 ratio. Each trace on the graph indicates the ratio measurement on MSNs expressing AKAR3 and identified as D₁ or D₂ according to their response to either SKF-38393 (SKF, 1 μ M) or CGS 21680 (CGS, 1 μ M), respectively. The thick black line represents the average of all the traces in a group. Bath application of the agonist of dopamine D₂ receptors quinpirole (1 μ M) reversed the response to PQ-10 (100 nM). **B**, D₂ receptor activation prevented the response to PDE10A inhibition: the effect of PQ-10 was measured in the presence of the D₂ agonist quinpirole (1 μ M). No statistically significant difference ($p > 0.05$) was found between D₁ and D₂ MSNs ($n = 5$). The effect of PQ-10 is displayed for comparison on the left (same data as in Fig. 4E). **C**, **D**, Other PDE10A inhibitors also increased the AKAR3 ratio preferentially in D₂ MSNs: MP-10 (**C**; 100 nM, $n = 9$) and papaverine (**D**; 1 μ M, $n = 5$) both increased the AKAR3 ratio selectively in D₂ MSNs. **E**, PQ-10 increased AKAR3 ratio selectively in D₂ MSNs even when adenosine A_{2A} receptors were inhibited with 100 μ M SCH 58261 ($n = 4$). **B–E**, Statistical differences were tested with paired Student's *t* test. *** $p < 0.001$. **F**, PDE10A inhibition had no effect on cGMP levels measured with the cGMP sensor cygnet2. The NO donor DEANO (100 μ M) increased the ratio; after reaching a steady-state level, PQ-10 (1 μ M) was added; at the end of the recording, the maximal ratio response was elicited by DEANO plus

continued

IBMX (200 μM). **G**, No difference was measured when comparing the response with DEANO alone and DEANO with PQ-10, while IBMX produced a significant increase. The data expressed as the mean \pm SEM were analyzed by repeated-measures one-way ANOVA $F_{(1,5)} = 11,224$, $p < 0.001$, $n = 6$, followed by Bonferroni's *post hoc* test: $**p < 0.01$). **A–G**, Brain slices were imaged with wide-field microscopy. All data are expressed as the mean \pm SEM.

to the A_{2A} agonist CGS 21680 (data not shown). These results showed that tonic activation of A_{2A} receptors was not required for the phosphorylation response to PQ-10 in D_2 MSNs. We then examined whether the phosphorylation signal involved endogenous neuronal activity, or required the glutamate and GABA which may be present in the brain slice. This was not the case since, in the presence of blockers of voltage-gated sodium channels (TTX, 100 nM), calcium channels (CdCl_2 , 200 μM), non-NMDA receptors (CNQX, 10 μM), NMDA receptors (APV, 10 μM), and GABA $_A$ receptors (SR 95531, 1 μM), the selective effect of PQ-10 on D_2 MSNs was still present (25 ± 3 vs 71 ± 10 of the maximal FSK response in D_1 and D_2 MSNs, respectively; $n = 3$, paired Student's *t* test; data not shown).

Since PDE10A also degrades cGMP and PDE10A inhibition was shown to increase cGMP levels and affect synaptic transmission *in vivo* (Siuciak et al., 2006; Schmidt et al., 2008; Grauer et al., 2009; Padovan-Neto et al., 2015), we used wide-field imaging of the cGMP biosensor cygnet2 (Honda et al., 2001) to determine whether PDE10A also regulated cGMP in MSNs. PQ-10 (1 μM) had no effect on the baseline cGMP levels ($n = 4$, $p = 0.44$, one-sample Student's *t* test). In addition, PQ-10 (1 μM) had no effect on the cGMP steady-state level obtained with the nitric oxide (NO) donor DEANO (Diethylamine nitric oxide, 100 μM ; $n = 6$; Fig. 3F), while the nonspecific phosphodiesterase inhibitor IBMX produced a significant ratio increase. These results indicated that in our conditions PDE10A inhibition does not significantly affect cGMP levels in MSNs.

Different responsiveness of D_1 and D_2 MSNs is abolished by protein phosphatase inhibition

The difference between D_1 and D_2 MSNs in the phosphorylation level of AKAR3 could result from differences in the rate of phosphorylation by PKA, dephosphorylation by phosphatases, or both. A difference in PKA levels is unlikely because immunostaining of catalytic subunits in the striatum did not reveal major differences between cells (Yang et al., 2014). Since AKAR3 biosensor responses to PKA activation are reversed by the action of endogenous protein phosphatases (Gervasi et al., 2007), we investigated the role of protein phosphatases in the different responsiveness of D_1 and D_2 MSNs to PDE10A inhibitors.

Cantharidin (30 μM), a nonselective inhibitor of PP-1 and PP-2A, had no effect by itself on the AKAR3 emission ratio (Fig. 4A). However, when PQ-10 (100 nM) was applied in the bath (Fig. 4A), the AKAR3 ratio increased in virtually all D_1 and D_2 MSNs ($79 \pm 10\%$ of the FSK response, $n = 5$). Since these responses did not return to the baseline after drug washout, it was impossible to distinguish between D_1 and D_2 MSNs, and all MSNs were

pooled (Fig. 4F, gray color bar). To identify which phosphatase subtype was involved in the response, we used fostriecin (200 nM), a selective inhibitor of PP-2A (Swingle et al., 2009). We observed no effect of this inhibitor alone on basal AKAR3 ratio, and it did not alter the selective response to PQ-10 in D_2 MSNs (Fig. 4B). These results showed that PP-2A was not involved in the different responsiveness of D_1/D_2 and rather suggested the involvement of PP-1.

PP-1 regulation by DARPP-32 is necessary for the selective responsiveness of D_2 MSNs to PDE10A inhibition

DARPP-32 is expressed at high levels in both types of MSNs and constitutes a powerful and specific inhibitor of PP-1 when it is phosphorylated at Thr34 (Hemmings et al., 1984). We used a knock-in mutant mouse line in which Thr34 is replaced by an alanine (T34A; Svenningsson et al., 2003). In T34A mice, the effect of PQ-10 on AKAR3 was strongly reduced in D_2 MSNs ($14 \pm 2\%$, $n = 13$, 6 mice), whereas, as in wild-type mice, no effect was observed in D_1 MSNs ($8 \pm 1\%$; Fig. 4C,E). Normal responses to D_1 or A_{2A} stimulations were observed at the end of the recording. The phosphatase inhibitor cantharidin unmasked the response to PQ-10 in all MSNs of the DARPP-32 T34A mice (Fig. 4F), confirming that, upon inhibition of PP-1, PQ-10 was still capable of increasing the AKAR3 response in these mutant mice. Together, these results show that the inhibition of PP-1 by DARPP-32 is necessary for the difference in responsiveness of D_1 and D_2 MSNs.

DARPP-32 also inhibits PKA activity when it is phosphorylated at Thr75 by Cdk5 (Bibb et al., 1999; Nishi et al., 2000). A higher phosphorylation level of this residue in D_1 MSNs could be responsible for a weaker PKA activity in these neurons. However, in a knock-in mutant mouse line with a Thr75-to-alanine mutation (DARPP-32 T75A; Svenningsson et al., 2003), the profile of the AKAR3 response to PQ-10 was the same as that in wild-type mice (Fig. 4D,E). Moreover, the Cdk5 inhibitor roscovitine (10 μM) had no effect on the D_1/D_2 imbalance in the response to PQ-10 (Fig. 4G), ruling out the involvement of Thr75 of DARPP-32 as a critical determinant for the lack of PQ-10-dependent AKAR3 responses in D_1 MSNs.

In vivo PDE10A inhibition selectively induces histone H3 phosphorylation in D_2 MSNs of the dorsomedial striatum

Our results showed a marked difference in the responsiveness of D_1 and D_2 MSNs to PDE10A inhibitor in slices. We then investigated whether the D_1/D_2 imbalance could also be observed *in vivo* by monitoring phospho-histone H3 at Ser10 residue (PH3), a substrate for several protein

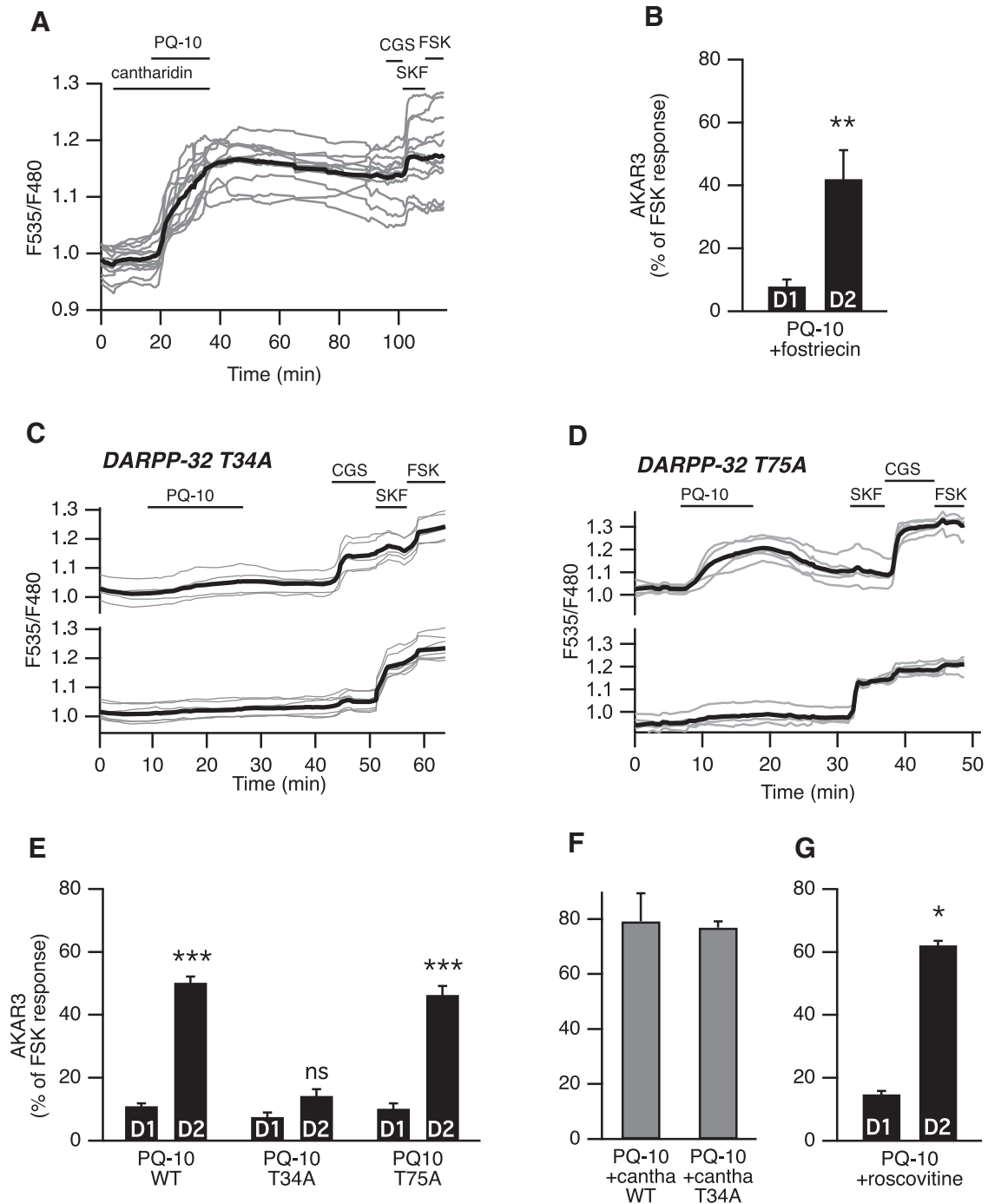


Figure 4. DARPP-32-mediated phosphatase inhibition favors PKA signaling in D₂ MSNs. **A**, PP-1 and PP-2A were inhibited with cantharidin. Cantharidin (30 μ M) alone did not change the basal ratio but strongly increased the AKAR3 response to PQ-10 (100 nM) in all MSNs. These responses were not reversible, making the final identification of D₁ and D₂ MSNs impossible (gray bars in **F**, which represent the responses of all MSNs). **B**, D₂ MSNs responded selectively to PQ-10 (100 nM) even when the PP-2A inhibitor fostriecin (200 nM) was applied ($n = 4$, paired Student's t test; $^{***}p < 0.01$). **C–E**, Mutation of the Thr34 to Ala in DARPP-32 (DARPP-32 T34A) strongly reduced the effect of PQ-10 (100 nM) in D₂ MSNs, whereas the selective effect of PQ-10 on D₂ MSNs remained in brain slices from animals bearing the Thr75 to Ala mutation in DARPP-32 (DARPP-32 T75A). **C, D**, Representative experiments performed with DARPP-32 T34A (**C**) and DARPP-32 T75A (**D**) knock-in mice. Each trace on the graph indicates the ratio measurement on MSNs expressing AKAR3 and is identified as D₁ or D₂ according to their response to either SKF-38393 (SKF, 1 μ M) or CGS 21680 (CGS, 1 μ M), respectively. The thick black line represents the average of all the traces in each group. **E**, The data expressed as the mean \pm SEM were analyzed by two-way ANOVA: genotype effect, $F_{(2,72)} = 71.12$, $p < 10^{-4}$; D₁/D₂ effect, $F_{(1,72)} = 333.07$, $p < 10^{-4}$; genotype \times D₁/D₂ interaction, $F_{(2, 72)} = 49.53$, $p < 10^{-4}$. Bonferroni's *post hoc* test: $^{***}p < 0.001$. **F**, In wild-type (WT) mice and DARPP-32 T34A mutants, and in the presence of cantharidin (30 μ M), all MSNs responded to PQ-10 (100 nM) with an increase in AKAR3 ratio such that D₁ and D₂ MSNs could not be distinguished ($n = 5$ for both). No significant difference was obtained between

continued

wild-type and DARPP-32 T34A mutant (unpaired Student's *t* test, $p > 0.05$). **G**, D₂ MSNs responded selectively to PQ-10 (100 nM) even when the Cdk5 inhibitor roscovitine (10 μM) was applied ($n = 4$, paired Student's *t* test; * $p < 0.05$).

kinases including PKA (Nowak and Corces, 2004). We used transgenic mice in which D₂ MSNs are identified by GFP fluorescence (drd2-EGFP mice; Gong et al., 2003; Bertran-Gonzalez et al., 2008) and monitored PH3 by immunofluorescence. In these *in vivo* experiments, we studied the effects of TP-10 (3 mg/kg, i.p.), a PDE10A inhibitor known to produce clear behavioral effects (Schmidt et al., 2008). At this dose, TP-10 induced a large increase in the number of PH3-positive neurons in the striatum 60 min after treatment, compared with vehicle treatment. Quantification showed that 93% of the PH3-immunoreactive neurons were GFP positive (D₂ MSNs) in the dorsomedial striatum, whereas in lateral parts of the striatum both GFP-negative and GFP-positive D₂ MSNs exhibited PH3 immunoreactivity (Fig. 5A–C). In the *nucleus accumbens*, a sparse and irregular labeling was observed in the shell region, and no immunoreactivity was detected in the core. In all striatal regions, no PH3 immunoreactivity was observed in large neurons expressing low levels of GFP, presumably corresponding to cholinergic interneurons.

Since *in vivo* the selective effects of TP-10 on D₂ MSNs was observed in the dorsomedial, but not the dorsolateral striatum, we performed a set of experiments in brain slices to compare medial and lateral dorsal striatum using the same inhibitor. The AKAR3 responses to TP-10 (100 nM) were similar, with a strong effect of TP-10 in D₂ MSNs but not in D₁ MSNs (Fig. 5D,E). Altogether, these experiments showed that PDE10A exerts selective effects on D₂ MSNs in striatal slices and that this selectivity is maintained *in vivo* in the dorsomedial striatum.

DARPP-32 is required for the *in vivo* effects of TP-10

Our experiments in striatal slices showed that the inhibition of PP-1 by DARPP-32 phosphorylated on Thr34 was necessary for the selective responsiveness to PDE10A inhibition on AKAR phosphorylation. We then investigated whether the effects of PDE10A inhibition *in vivo* also depended on the phosphorylation of DARPP-32 at Thr34. T34A knock-in and wild-type littermates were treated with TP-10 (3 mg/kg, i.p.) or vehicle (four animals for each condition) and brain sections analyzed by immunofluorescence for PH3 60 min after injection. Whereas wild-type littermates strongly responded to TP-10, the effect of TP-10 was completely abolished in the DARPP-32 T34A mutant mice in all regions of the striatum (Fig. 6A,B). These experiments clearly showed that Thr34 in DARPP-32 was required for the effects of PDE10A inhibitor on D₁ and D₂ MSNs in both the dorsomedial and dorsolateral striatum *in vivo*.

Discussion

Our study shows that although PDE10A is expressed and functional in all types of MSNs, its inhibition in striatal slices produces a higher PKA-dependent protein phos-

phorylation response in D₂ MSNs than in D₁ MSNs. We provide evidence that this difference is present in all striatal regions in brain slices and in the dorsomedial striatum of adult mice *in vivo*. Moreover, we show that the regulation of PP-1 activity by DARPP-32 is required for these specific effects of PDE10A inhibitors. These observations provide novel insights into the regulation of the cAMP/PKA pathway in the two populations of MSNs and the possible antipsychotic action of PDE10A inhibitors.

PDE10A is one of the enzymes specifically enriched in the striatum (Seeger et al., 2003; Coskran et al., 2006; Heiman et al., 2008; Lakics et al., 2010; Kelly et al., 2014), and the data reported here show that PDE10A plays an important role in degrading basally produced cAMP in both D₁ and D₂ MSNs. MSNs thus contrast with pyramidal neurons of the prefrontal cortex in which basal cAMP is predominantly controlled by PDE4 (Castro et al., 2010). Although PDE4B is also expressed in D₂ MSNs (Nishi et al., 2008), biosensor imaging of the somatic cytoplasm did not reveal significant effects of its inhibition in MSNs in our conditions.

PDE10A inhibition showed no effect on basal or stimulated cGMP production. A similar lack of effect of PDE10A inhibitors on cGMP has already been reported in brain slice preparations (Nishi et al., 2008). PDE10A thus differs from PDE1 and PDE2, other dual-specificity PDEs expressed in MSNs, which were shown in striatal homogenates to be the major PDEs involved in the control of cGMP levels (Russwurm et al., 2015). Using biosensor-imaging techniques, PDE2 was also shown to be the main PDE that regulated stimulated cGMP, while also regulating cAMP in a cGMP-dependent manner (Polito et al., 2013). *In vivo*, PDE10A inhibition was shown to increase cGMP levels and to affect synaptic transmission (Siuciak et al., 2006; Schmidt et al., 2008; Grauer et al., 2009; Padovan-Neto et al., 2015), and why this effect was not observed in brain slices remains to be determined. One hypothesis to explain this discrepancy might be that, *in vivo*, PDE10A inhibitors recruit nitric oxide synthase-positive striatal interneurons through a global increase in network activity.

While D₁ and D₂ MSNs share a number of cellular features, more detailed studies revealed subtle differences (Valjent et al., 2009), such as different excitability profiles (Gertler et al., 2008; Threlfell et al., 2009). Differences were also reported at the level of PKA-dependent phosphorylation of GABA_A receptors and DARPP-32, which were higher in D₂ MSNs than in D₁ MSNs (Janssen et al., 2009; Nishi et al., 2008). Our work reveals a possible basis for these D₁/D₂ differences, which lies at the level of DARPP-32-mediated PP-1 regulation. In D₁ MSNs, the Thr34 of DARPP-32 is in a lower phosphorylation state than in D₂ MSNs and, thus, the PP-1 activity reverts PKA target sites to the dephosphorylated state (Fig. 7). This

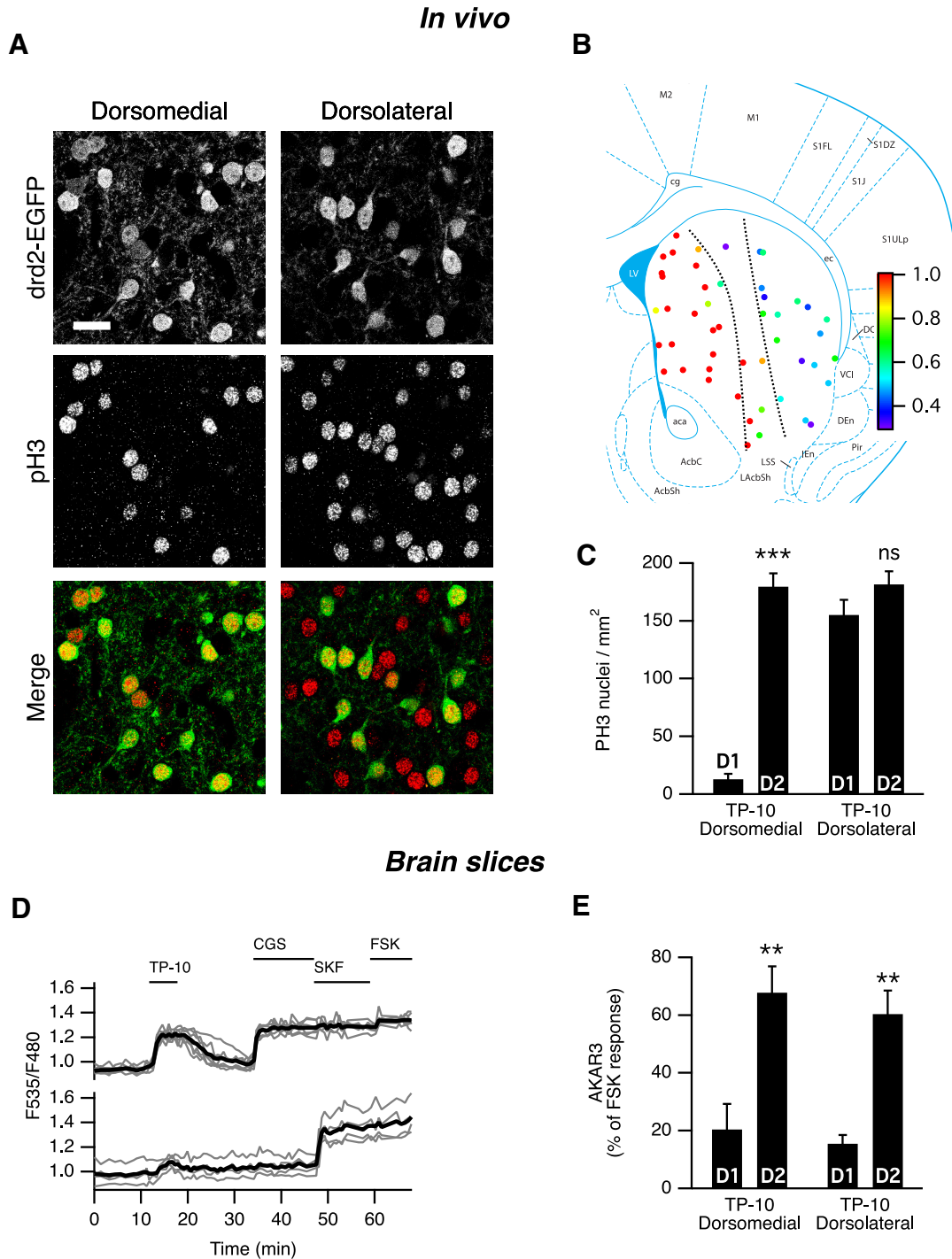


Figure 5. In vivo effects of PDE10A inhibition by TP-10. **A**, In the medial part of the dorsal striatum of *drd2-EGFP* adult mice treated with TP-10 (3 mg/kg), PH3 was selectively observed in D_2 MSNs. In the lateral part of the dorsal striatum, PH3 immunoreactivity was observed in both EGFP-positive and EGFP-negative MSNs. EGFP and PH3 are shown in grayscale, and are overlaid with EGFP in green and PH3 in red (Merge). Scale bar, 20 μ m. **B**, Each color spot represents a position where the relative distribution of $D_2/(D_1 + D_2)$ PH3-positive MSNs is indicated in pseudocolor, over a schematic of coronal mouse brain (Franklin and Paxinos, 2007). **C**, PH3-positive nuclei were quantified in medial and lateral parts of the dorsal striatum as defined by the dotted line in **B**. The effect of localization was significant (Kruskal–Wallis test followed by a Mann–Whitney test with a Dunn–Sidak adjustment test for pairwise multiple comparisons tests, $p < 10^{-4}$), with PH3-positive nuclei being preferentially D_2 MSNs in the medial striatum. ***indicates a difference between EGFP-positive (D_2) and EGFP-negative (D_1) MSNs with $p < 10^{-4}$. **D**, The preferential AKAR3 response is also observed in the lateral striatum in brain slices from neonate mice. MSNs were transduced for the expression of the AKAR3 biosensor and imaged with wide-field microscope in the lateral striatum. Each trace on the graph indicates the ratio measurement on MSNs expressing AKAR3 and was identified as D_1 or D_2 according to their response to either SKF-38393 (SKF, 1 μ M) or CGS 21680 (CGS,

continued

1 μM), respectively. The thick black line represents the average of all the traces in each group. TP-10 (100 nM) increased AKAR3 ratio selectively in D_2 MSNs. **E**, The same experiment was repeated: there was no effect of localization, and TP-10 increased the AKAR3 ratio selectively in D_2 MSNs in both the dorsolateral and dorsomedial striatum (two-way ANOVA: localization effect, $F_{(1,12)} = 0.374$, $p = 0.374$; D_1/D_2 effect, $F_{(1,12)} = 44.01$, $p < 10^{-4}$; localization \times D_1/D_2 interaction, $F_{(1,12)} = 0.042$, $p = 0.804$. Bonferroni's *post hoc* test: $**p < 0.01$). **C**, **E**, Error bars indicate the SEM.

hypothesis implies that a powerful mechanism prevents DARPP-32 from remaining phosphorylated on the Thr34 position selectively in D_1 MSNs. Thr34 residue is efficiently dephosphorylated by both PP-2A and PP-2B (but not by PP-1; Nishi et al., 1999), and further work is needed to analyze the possible differences in PP2A and PP2B activities between D_1 and D_2 MSNs. In contrast to tonic cAMP levels induced by PDE10A inhibition, cAMP signals elicited by D_1 receptor stimulation lead to a phosphorylation of T34 and inhibition of PP-1 (Bateup et al., 2008), an effect that is also clearly visible on transient responses to dopamine stimulations (Castro et al., 2013). This is consistent with the observation that, in D_1 MSNs, PDE10A inhibition only affects PKA-dependent modulation of synaptic transmission when cAMP production is stimulated (Mango et al., 2014). This nonlinearity in D_1 MSNs may improve the detection of powerful but brief events such as the phasic dopamine signal associated with reward and novelty (Schultz, 2010), while filtering out smaller fluctuations in basal cAMP level.

In contrast, in D_2 MSNs DARPP-32 is phosphorylated on the Thr34 residue, as previously demonstrated (Nishi et al., 2008). In this situation, a moderate cAMP signal,

such as that produced by PDE10A inhibition, activates PKA, and, because PP-1 is inhibited, PKA targets remain phosphorylated. Indeed, when DARPP-32 bears the T34A mutation and can no longer inhibit PP-1, D_2 MSNs fail to respond to PDE10A inhibition (Fig. 7).

Further work is needed to analyze the possible differences in PP-2A and PP-2B activities between D_1 and D_2 MSNs that may contribute to the higher level of DARPP-32 phosphorylation on Thr34.

Another potential player in the D_1/D_2 differences is the phosphorylation of DARPP-32 on Thr75, which is catalyzed by Cdk5 and is responsible for PKA inhibition (Bibb et al., 1999). Since Thr75 is dephosphorylated by a PKA-activated form of PP-2A containing the B56 subunit (Ahn et al., 2007), it could contribute to a hypersensitive feed-forward loop. However, this mechanism did not appear to be critical for PDE10A responses in D_2 MSNs, since we found no alteration of these responses in DARPP-32 T75A knock-in mutant mice.

The increased responsiveness of D_2 MSNs at the level of PKA signaling is opposed *in vivo* by the activity of D_2 receptors: the simple blockade of these receptors by D_2 antagonists strongly activates cAMP-dependent phos-

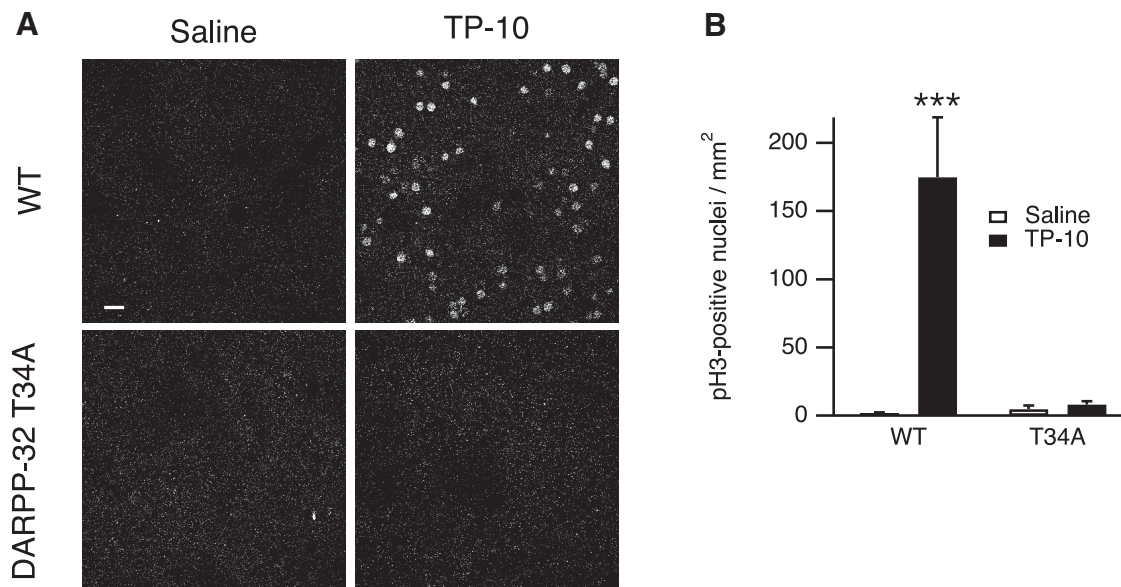


Figure 6. The DARPP-32 T34 residue is required for a TP-10-induced increase of histone H3 phosphorylation in the striatum in adult mice *in vivo*. Wild type (WT) and DARPP-32 T34A mutant mice were treated with TP-10 (3 mg/kg) or vehicle. **A**, Examples of PH3 immunofluorescence, showing the dramatic reduction of TP-10 effects in the DARPP-32 T34A mutant mice. Scale bar, 20 μm . **B**, Quantification of the number of PH3-positive neurons in striatal coronal sections. Error bars indicate the SEM. Data were analyzed by a two-way ANOVA: genotype effect, $F_{(1,12)} = 13.7$, $p < 0.01$; TP-10 effect, $F_{(1,12)} = 16.1$, $p < 0.01$; genotype \times TP-10 interaction, $F_{(1,12)} = 14.8$, $p < 0.01$. Bonferroni's *post hoc* test, $***p < 10^{-3}$.

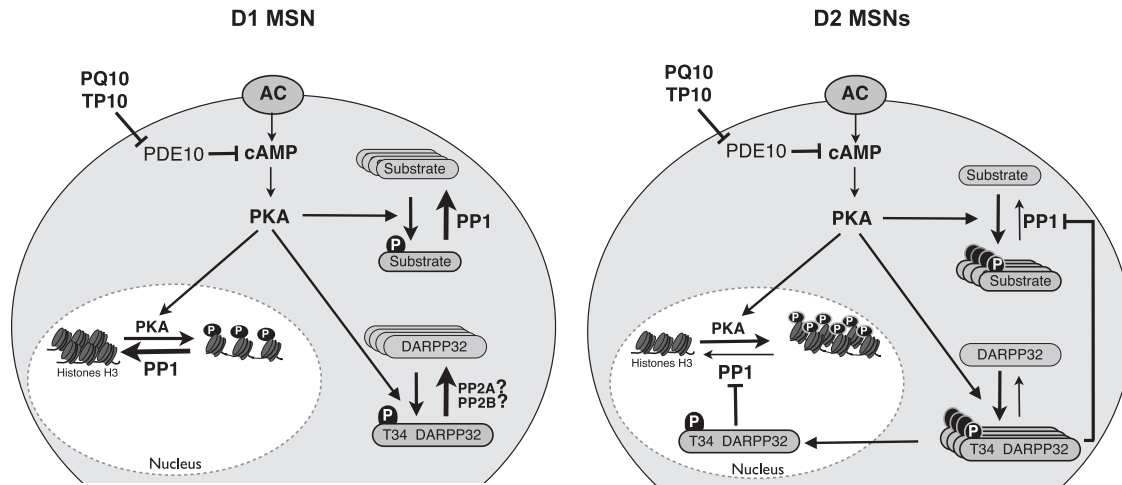


Figure 7. Diagram depicting the D₁/D₂ differential response to PDE10A inhibition. PDE10A inhibition increases cAMP and activates PKA to similar levels in D₁ and D₂ MSNs. In D₂ MSNs, DARPP-32 is phosphorylated and inhibits PP-1: PKA substrates thus remain in the phosphorylated state, both in the cytosol and in the nucleus. In D₁ MSNs, DARPP-32 is in a dephosphorylated state: PP-1 is fully active and dephosphorylates PKA substrates. Differences in PP2A/B activities between D₁ and D₂ MSNs may explain this imbalance.

phorylation (Håkansson et al., 2006; Bertran-Gonzalez et al., 2008, 2009; Bonito-Oliva et al., 2011; Valjent et al., 2011). Accordingly, haloperidol or clozapine selectively increases phospho-Thr34-DARPP-32 in D₂ MSNs, and not in D₁ MSNs (Bateup et al., 2008). This role of DARPP-32 in D₂ MSNs is functionally important for some effects of antipsychotic drugs since conditional knockout of DARPP-32 in D₂ MSNs leads to an increased locomotor activity and a strongly reduced catalepsy upon administration of D₂ receptor inhibitors (Bateup et al., 2010). Our data show that the inhibition of PDE10A has functional effects that are similar to the blockade of D₂ receptors since both potentiate the PKA pathway selectively in D₂ MSNs. The particular sensitivity of D₂ MSNs includes DARPP-32 Thr34 phosphorylation, which is more increased by PDE10A inhibitors in this population than in D₁ MSNs (Nishi et al., 2008). Thus, hypersensitivity of DARPP-32 Thr34 phosphorylation could be a critical factor to account for the responsiveness of D₂ MSNs to the blockade of either D₂ receptors or PDE10A activity.

The cellular effects of PDE10A inhibition affect MSNs neuronal properties, which are integrated through the basal ganglia network. For example, PDE10A inhibition was shown to potentiate D-amphetamine-dependent dopaminergic neuromodulation *in vivo* (Sotty et al., 2009). In addition, PDE10A inhibition was shown to massively increase cGMP levels *in vivo* (Siuciak et al., 2006; Schmidt et al., 2008; Grauer et al., 2009), whereas, no effect was observed in brain slice preparations (Nishi et al., 2008; this study). This discrepancy possibly results from the recruitment of striatal NOergic interneurons through network activity resulting indirectly from PDE10A inhibition.

In vivo, the inhibition of PDE10A also selectively activates D₂ MSNs in the medial striatum. Interestingly, this

striatal subregion is innervated by prefrontal and cingulate cortices, which are involved in the limbic system, and dopamine neurons originating from the ventral tegmental area. D₁ and D₂ receptors exert contrasting roles selectively in the dorsomedial striatum during behavioral inhibition in the stop-signal task in rats (Eagle et al., 2011), and lesions of the dorsomedial striatum disrupt prepulse inhibition (Baldan Ramsey et al., 2011). This region is also involved in early motor learning and is required for the cataleptic effects of haloperidol and for amphetamine motor response sensitization (Durieux et al., 2012). Medial striatum thus appears as a limbic system-related region that could be affected in schizophrenia.

The differences observed between medial and lateral striatum likely involve network effects and differences in synaptic plasticity reported between these two striatal subregions (Lovinger, 2010), and further work should precisely show how these spatial differences are related to behavior. Our work nonetheless shows that the D₁/D₂ difference is present in brain slices in both medial and lateral striatum, and remains when network activity is blocked, showing that the D₁/D₂ difference is an intrinsic property of D₁ and D₂ MSNs. The *in vivo* effects of PDE10A inhibitors were totally abolished in T34-DARPP-32 mutant mice, confirming the role of the DARPP-32/PP-1 loop as the initial determinant of the positive PKA response obtained during PDE10A inhibition.

Further work is needed to determine whether the imbalance in PKA signal integration between D₁ and D₂ MSNs might be of interest to understand the pathophysiology of other diseases that affect neuromodulatory processes in basal ganglia such as Parkinson's disease or Huntington's disease (Threlfall and West, 2013).

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