GENOME SEQUENCES





Draft Genome Sequence of *Bradyrhizobium elkanii* BR 2003, an Efficient Rhizobium Strain for *Cajanus, Canavalia, Crotalaria*, and *Indigofera*

Jerri Édson Zilli,ª Jean Luiz Simoes-Araujo,ª Luc Felicianus Marie Rouws,ª Luis Henrique de Barros Soaresª

^aEmbrapa Agrobiologia, Seropédica, Rio de Janeiro, Brazil

ABSTRACT We report here the annotated draft genome sequence of the rhizobium strain BR 2003. This strain is able to establish symbiosis and to fix nitrogen with a broad range of leguminous species. The estimation of the average nucleotide identity confirmed the strain as a member of *Bradyrhizobium elkanii*.

The genus *Bradyrhizobium* (alphaproteobacteria) comprises some efficient nitrogenfixing bacteria that are able to nodulate leguminous species. Strains of this genus are frequently present in nodules of tropical and subtropical leguminous species, and strains that are well adapted to diverse edaphoclimatic conditions can be found in different biomes (1, 2).

For instance, strain BR 2003 was isolated in 1982 from nodules of *Crotalaria juncea* grown in Brazilian Cerrado soil (3). Field experiments have indicated that this strain is efficient in symbiotic nitrogen fixation with green manure species *C. juncea*, *Crotalaria spectabilis*, *Cajanus cajan*, *Canavalia ensiformis*, and *Indigofera hirsuta* (4). Here, we present the draft genome sequence of strain BR 2003, which is used in commercial inoculants for green manure in Brazil. The phylogeny constructed using 16S rRNA gene sequences showed that this strain is a member of the *Bradyrhizobium elkanii* clade, with high similarity to different related type strains.

Strain BR 2003 was originally isolated and purified on yeast extract-mannitol (YM) agar plates and was kept lyophilized for long-term storage after five passages on culture medium for purification. A single colony from a fresh plate was inoculated in liquid YM medium and grown for 4 days at 28°C with agitation (5). Genomic DNA was isolated from the stationary-phase bacterial culture using the Wizard DNA clean-up kit (Promega). A whole-genome paired-end library was constructed using the TruSeq PCR-free kit, and samples were run on a HiSeq 2500 sequencer (Illumina) at Macrogen (South Korea).

The Illumina sequencer generated a total of 18,045,344 paired-end reads (101 bp long), with 99.51% of the reads having a Phred quality score higher than 20. The reads were trimmed using the FASTX-Toolkit (http://hannonlab.cshl.edu/fastx_toolkit), and only bases with a quality score above 20 were used for *de novo* genome assembly using ABySS software version 2 (6). The kmer length parameter was tested from 70 to 98, and a genome with 124 contigs and 8,946,006 base pairs was the best result from the ABySS assembly, using a kmer value of 97. GFinisher software version 1.2 (7) was used to refine the BR 2003 genome ABySS assembly. GFinisher generates fuzzy GC content skew graphs for all contigs and compares them to a draft genome assembly using BLAST to find sequences that overlap gaps. The reassembly and contig/scaffold organization were carried out using *Bradyrhizobium japonicum* strain SEMIA 5079 (GenBank accession number NZ_CP007569.1) as a reference genome. Annotation and identification of metabolic pathways for the draft genome were performed using the RAST version 2.0 server (8). The genome sequence available at NCBI was annotated using the Prokaryotic

Citation Zilli JÉ, Simoes-Araujo JL, Rouws LFM, de Barros Soares LH. 2020. Draft genome sequence of *Bradyrhizobium elkanii* BR 2003, an efficient rhizobium strain for *Cajanus*, *Canavalia, Crotalaria,* and *Indigofera*. Microbiol Resour Announc 9:e01565-19. https://doi.org/ 10.1128/MRA.01565-19.

Editor Catherine Putonti, Loyola University Chicago

Copyright © 2020 Zilli et al. This is an openaccess article distributed under the terms of the Creative Commons Attribution 4.0 International license.

Address correspondence to Jerri Édson Zilli, jerri.zilli@embrapa.br.

Received 20 December 2019 Accepted 23 February 2020 Published 12 March 2020 Genome Annotation Pipeline (PGAP) version 3.1 (9). Default parameters were used for all software unless otherwise specified. The reassembly GFinisher summed 8.20 megabases, distributed in 52 contigs, with a contig N_{so} value of 298.6 kb. The size of the longest contig was 506.2 kb. The coverage reported by the assembler was $150\times$, and the genome GC content is 64.4%, which is within the range for the *Bradyrhizobium* genus. The RAST annotation identified 8,093 coding DNA sequences, distributed across 371 subsystems, and 48 RNAs. The largest number of genes was observed in the subsystem of amino acids and derivatives (629 genes), followed by carbohydrates; 51 genes were involved in nitrogen metabolism, such as nitrogen fixation (20 genes), nitrate and nitrite ammonification (11 genes), and ammonia assimilation (14 genes). However, no denitrification-related genes were found.

A high-resolution species tree generated by Automated Multi-Locus Species Tree software (10), using 83 marker genes, showed that BR 2003 was most closely related to Bradyrhizobium elkanii type strain USDA76 (GenBank accession number NZ_ARAG00000000.1). In fact, the estimated average nucleotide identity between BR 2003 and USDA76 was 96.12%, confirming that the strains belong to the same species. However, when compared with that of USDA76, the BR 2003 genome has 106 exclusive genes, including 12 related to the type VI secretion system (T6SS). The T6SS allows bacteria to translocate effector proteins to other bacteria or to eukaryotic cells and can act as a contact-dependent interbacterial "weapon" to keep competing strains from co-occupying sites in the host (11). Interestingly, in Azorhizobium caulinodans (strain ORS571), the T6SS seems to participate specifically in symbiosis by increasing the symbiotic competitiveness (12, 13). Other membrane transporter genes exclusive to BR 2003, including six genes from the ABC transporter family and five genes related to resistance to antibiotics and toxic compounds, were also detected; this suggests that BR 2003 has distinct capabilities with respect to the transport of specific molecules.

Data availability. The assembled genome sequence was deposited in GenBank under BioProject accession number PRJNA526863 (BioSample accession number SAMN11116018). The raw data accession number is SRX7288786. The version described in this paper is the first version under accession number SMYJ00000000, and the genome sequence available at NCBI was annotated using PGAP version 3.1 (9).

ACKNOWLEDGMENTS

We thank the Brazilian National Council for Scientific and Technological Development for productivity grants awarded to the authors and financial support of projects, especially INCT Plant Growth-Promoting Microorganisms for Agricultural Sustainability and Environmental Responsibility (grant 465133/2014-2).

REFERENCES

- Mendes I, Hungria M, Vargas M. 2004. Establishment of *Bradyrhizobium japonicum* and *B. elkanii* strains in a Brazilian Cerrado oxisol. Biol Fertil Soils 40:28–35. https://doi.org/10.1007/s00374-004-0739-1.
- Azarias Guimarães A, Florentino LA, Alves Almeida K, Lebbe L, Barroso Silva K, Willems A, de Souza Moreira FM. 2015. High diversity of *Bradyrhizobium* strains isolated from several legume species and land uses in Brazilian tropical ecosystems. Syst Appl Microbiol 38:433–441. https:// doi.org/10.1016/j.syapm.2015.06.006.
- Franco AA. 1984. Contribution of biologically-fixed nitrogen to food crop production in Brazil, p 147–166. *In* Kang BT, van der Heide J (ed), Nitrogen management in farming systems in humid and subhumid tropics. International Institute of Tropical Agriculture, Ibadan, Nigeria.
- 4. Brasil Ministério da Agricultura, Pecuária, e Abastecimento. 2011. Instrução normativa nº 13, de 24 de março de 2011: aprova as normas sobre especificações, garantias, registro, embalagem e rotulagem dos inoculantes destinados à agricultura, bem como as relações dos microorganismos autorizados e recomendados para produção de inoculantes no Brasil. DOU 58:1–24.

- Fred EB, Waksman SA. 1928. Laboratory manual of general microbiology. McGraw-Hill, New York, NY.
- Simpson JT, Wong K, Jackman SD, Schein JE, Jones SJ, Birol I. 2009. ABySS: a parallel assembler for short read sequence data. Genome Res 19:1117–1123. https://doi.org/10.1101/gr.089532.108.
- Guizelini D, Raittz RT, Cruz LM, Souza EM, Steffens MBR, Pedrosa FO. 2016. GFinisher: a new strategy to refine and finish bacterial genome assemblies. Sci Rep 6:34963. https://doi.org/10.1038/srep34963.
- Overbeek R, Olson R, Pusch GD, Olsen GJ, Davis JJ, Disz T, Edwards RA, Gerdes S, Parrello B, Shukla M, Vonstein V, Wattam AR, Xia F, Stevens R. 2014. The SEED and the Rapid Annotation of microbial genomes using Subsystems Technology (RAST). Nucleic Acids Res 42:D206–D214. https:// doi.org/10.1093/nar/gkt1226.
- Tatusova T, DiCuccio M, Badretdin A, Chetvernin V, Nawrocki EP, Zaslavsky L, Lomsadze A, Pruitt KD, Borodovsky M, Ostell J. 2016. NCBI Prokaryotic Genome Annotation Pipeline. Nucleic Acids Res 44:6614–6624. https://doi .org/10.1093/nar/gkw569.
- Alanjary M, Steinke K, Ziemert N. 2019. AutoMLST: an automated Web server for generating multi-locus species trees highlighting natural

product potential. Nucleic Acids Res 47:W276-W282. https://doi.org/10 .1093/nar/gkz282.

- Speare L, Cecere AG, Guckes KR, Smith S, Wollenberg MS, Mandel MJ, Miyashiro T, Septer AN. 2018. Bacterial symbionts use a type VI secretion system to eliminate competitors in their natural host. Proc Natl Acad Sci U S A 115:E8528–E8537. https://doi.org/10.1073/pnas.1808302115.
- 12. Lin HH, Huang HM, Yu M, Lai EM, Chien HL, Liu CT. 2018. Functional exploration of the bacterial type VI secretion system in mutualism:

Azorhizobium caulinodans ORS571–Sesbania rostrata as a research model. Mol Plant Microbe Interact 31:856–867. https://doi.org/10.1094/ MPMI-01-18-0026-R.

 Salinero-Lanzarote A, Pacheco-Moreno A, Domingo-Serrano L, Durán D, Ormeño-Orrillo E, Martínez-Romero E, Albareda M, Palacios JM, Rey L. 2019. The type VI secretion system of *Rhizobium etli* Mim1 has a positive effect in symbiosis. FEMS Microbiol Ecol 95:fiz054. https://doi.org/10 .1093/femsec/fiz054.