

Since January 2020 Elsevier has created a COVID-19 resource centre with free information in English and Mandarin on the novel coronavirus COVID-19. The COVID-19 resource centre is hosted on Elsevier Connect, the company's public news and information website.

Elsevier hereby grants permission to make all its COVID-19-related research that is available on the COVID-19 resource centre - including this research content - immediately available in PubMed Central and other publicly funded repositories, such as the WHO COVID database with rights for unrestricted research re-use and analyses in any form or by any means with acknowledgement of the original source. These permissions are granted for free by Elsevier for as long as the COVID-19 resource centre remains active. Contents lists available at ScienceDirect



International Journal of Infectious Diseases



journal homepage: www.elsevier.com/locate/ijid

Performance and acceptability of self-collected human papillomavirus testing among women living with HIV



Runzhi Wang^{a,1,*}, Kristen Lee^{b,1}, Charlotte A. Gaydos^c, Jean Anderson^a, Jean Keller^a, Jenell Coleman^a

^a Department of Gynecology and Obstetrics, Johns Hopkins University School of Medicine, Baltimore, MD, USA

^b Department of Internal Medicine, Northwestern University Feinberg School of Medicine, Chicago, IL, USA

^c Department of Medicine, Infectious Disease Division, Johns Hopkins University School of Medicine, Baltimore, MD, USA

ARTICLE INFO

Article history: Received 11 May 2020 Received in revised form 23 July 2020 Accepted 26 July 2020

Keywords: Self-collection HPV testing Cervical cancer screening Women living with HIV

ABSTRACT

Objective: To assess the validity, reliability, and acceptability of self-collected human papillomavirus (HPV) tests in women living with HIV (WLHIV) in the United States. *Methods:* WLHIV \geq 30 years of age underwent self-collected (clinic and home) and clinician-collected HPV tests. Sensitivity and specificity analyses were performed using the clinician-collected HPV tests as the comparator. The unweighted kappa statistic was used to evaluate the validity and reliability of self-collected HPV testing, and the level of agreement between the clinician-collected mRNA test and a DNA test that was used for routine care. A 13-question survey was used to assess acceptability.

Results: Among the 70 participants, the median age was 50 years, 75% had an undetectable HIV RNA, and 11% had a CD4 count of <200 cells/ μ l. Nearly 63% had at least one positive HPV test. The sensitivity and specificity of the self-collected HPV test were 84.6% (95% confidence interval (CI) 65.1–95.6%) and 62.9% (95% CI 44.9–78.5%), respectively (κ = 0.5, 95% CI 0.2–0.7). The agreement between the two self-collected tests was good (κ = 0.8, 95% CI 0.5–1.0). There was good agreement between clinician-collected mRNA tests and DNA tests (κ = 0.8, 95% CI 0.7–1.0). Self-collection was highly acceptable.

Conclusions: Among WLHIV, self-collected HPV tests had good sensitivity and moderate specificity compared to clinician-collected HPV tests. The reliability between self-collected testing locations was good. Self-collected HPV testing had high acceptability.

© 2020 Published by Elsevier Ltd on behalf of International Society for Infectious Diseases. This is an open access article under the CC BY-NC-ND license (http://creativecommons.org/licenses/by-nc-nd/4.0/).

Introduction

High-risk human papillomavirus (HPV) causes almost all cervical cancers (Walboomers et al., 1999). The American Cancer Society estimated that there were 13 170 new cervical cancer cases and 4250 cervical cancer deaths in the United States in 2019 (Siegel et al., 2019). Most HPV infections are asymptomatic and can be cleared, but if the infection persists and is left untreated, over time, it can lead to precancerous changes that may develop into cancer. One of the risk factors for persistence and progression of HPVrelated disease is immunodeficiency (IARC Working Group on the Evaluation of Carcinogenic Risks to Humans, 2012). Women living with HIV/AIDS (WLHIV) have a four-fold higher risk of developing cervical cancer than the general population but are under-screened

E-mail address: rzwang@jhmi.edu (R. Wang).

(Engels et al., 2006; Chaturvedi et al., 2009; Oster et al., 2009; Peprah et al., 2018). In addition, infection with multiple HPV types is seen more often among WLHIV compared to women without HIV (Levi et al., 2002; Moscicki et al., 2004; Menon et al., 2016), and faster HPV-related disease progression has been observed (Denslow et al., 2014).

HPV testing is a notable advancement in cervical cancer screening, given its ability to offer greater reassurance of low cancer risk compared to cytology alone. A randomized controlled trial (Ronco et al., 2010) comparing conventional liquid-based cytology versus HPV-based testing (either alone or combined with cytology) showed that HPV-based screening was more effective in preventing cervical cancer by detecting persistent high-grade lesions earlier. Additionally, HPV testing has a high sensitivity for detecting precancerous lesions (Bulkmans et al., 2007; Naucler et al., 2007; Clad et al., 2011). A negative HPV test provides greater reassurance of low cervical precancer risk compared with a negative Pap test (Cuzick et al., 2006; Gyllensten et al., 2012; Leinonen et al., 2012; Ogilvie et al., 2012; Malila et al., 2013; Ronco et al., 2014; Wright et al., 2015). Furthermore, compared to

https://doi.org/10.1016/j.ijid.2020.07.047

^{*} Corresponding author at: Johns Hopkins School of Medicine, 600 North Wolfe Street, Phipps Building Room 248, Baltimore, MD 21287, USA.

¹ These authors contributed equally to this work.

^{1201-9712/© 2020} Published by Elsevier Ltd on behalf of International Society for Infectious Diseases. This is an open access article under the CC BY-NC-ND license (http://creativecommons.org/licenses/by-nc-nd/4.0/).

cytology, HPV testing is less subjective, more reproducible, and needs less training and expertise for users (Cuzick et al., 2006). The American Society for Colposcopy and Cervical Pathology (ASCCP) and the United States Public Service Task Force (USPSTF) support primary HPV screening as one of the first-line cervical cancer prevention strategies among women older than 25 years or 30 years, respectively (Huh et al., 2015; US Preventive Services Task Force et al., 2018).

Studies have shown that HPV self-collection increases cervical cancer screening participation in healthy women who do not routinely attend traditional cervical cancer screening programs, and the acceptability has been favorable (De Alba et al., 2008; Racey et al., 2013; Nelson et al., 2015; Winer et al., 2016). Therefore, self-collected samples potentially can increase the uptake of cervical cancer screening in WLHIV who are under-screened by offering screening at primary care sites, HIV specialty clinics, or non-clinical sites (e.g., home) that do not routinely perform pelvic examinations; however, there are very few studies to support this assertion. Therefore, the aim of this study was to assess the validity, reliability, and acceptability of HPV self-collected tests in a clinical setting and at home compared to conventional clinician-collected HPV tests in US WLHIV.

Methods

Study design and participants

WLHIV \geq 30 years of age who were undergoing a Pap test with HPV DNA testing as part of routine care were eligible for inclusion. Women who were pregnant, had a hysterectomy, and had genital tract cancer were excluded. Recruitment took place at a large academic multidisciplinary clinic in Baltimore, Maryland. Institutional review board approval was obtained.

Procedures and data collection

Participants were instructed on how to perform an unsupervised vaginal HPV mRNA cytobrush collection (Aptima; Hologic, San Diego, CA, USA). At the time of the clinic visit, participants were instructed to insert a cytobrush into the vagina as far as possible, turn the brush five full rotations, and then place the brush into a vial of transport medium. Next, participants underwent a pelvic examination, during which clinicians collected a research cervical HPV mRNA cytobrush and a routine cervical sample for HPV and Pap co-test. The clinical laboratory used an assay that detected HPV DNA (Qiagen, Hilden, Germany), although both types of HPV test are commercially available. Participants were given a home collection kit that contained instructions on self-collection, one collection brush, storage containers, and a pre-addressed, postage-paid return cardboard envelope. Two weeks after the clinic visit, participants were reminded by a text message or phone call to self-collect at home and to mail the cytobrush back. Women were given a \$20 gift card for their participation. Women with a positive result from any HPV test were phoned and advised to discuss the results with their clinicians for further guidance. AIDSinfo Guidelines for prevention and treatment of opportunistic infections in HIV-infected adults and adolescents (Panel on Opportunistic Infections in Adults and Adolescents with HIV) were used to identify women who required Colposcopy. Electronic medical records were abstracted to collect age, race, most recent HIV RNA concentration, CD4 T-cell count, antiretroviral therapy use, and smoking status. Due to the large proportion of women of Black race in the HIV clinic, race was classified to Black versus non-Black. In addition, we classified the most recent HIV RNA concentration into two groups: detectable $(\geq 20 \text{ copies/ml})$ and undetectable (<20 copies/ml).

HPV DNA and mRNA assays

The clinician-collected (CC) cervical sample was tested for HPV DNA using a hybrid capture II (HC2) DNA nucleic acid hybridization assay that detected HPV DNA types 16, 18, 31, 33, 35, 39, 45, 51, 52, 56, 58, 59, and 68. Positive samples had a relative light unit \geq 1 pg/ml. Individual genotyping results were not available. Equivocal HPV DNA results were reclassified as positive (Knoepp et al., 2007). For research purposes, the self-collected (SC) and CC samples were tested for HPV using the Aptima transcription-mediated amplification RNA test that detected E6/E7 messenger RNA of the same HPV types as the HC2 assay plus type 66. Positive samples had signal-to-cutoff values of \geq 0.5. Invalid HPV mRNA was not reclassified as there are no data available to inform the reclassification.

Survey

A 13-question survey was administered after self-collection at the clinic to assess the acceptability of self-collection. These questions assessed whether the patients (1) understood the instructions; (2) felt comfortable; (3) felt relaxed; (4) felt in control; (5) felt they were taking care of their health; (6) felt convenient; (7) felt anxious; (8) felt embarrassed; (9) felt it was painful; (10) worried that they might do the test wrong; (11) would use the cytobrush again; (12) would recommend the cytobrush to family and friends; and (13) would use the cytobrush in a clinic. A five-point Likert scale from strongly agree to strongly disagree was used. The survey also queried, "How easy or hard was it for you to collect the vaginal specimen using the brush?" with responses ranging from very easy, easy, and OK, to hard, or very hard.

Statistical analysis

To evaluate the ability of SC HPV mRNA tests to correctly identify HPV infection, sensitivity and specificity analyses with 95% confidence intervals (CI) were performed over 2×2 contingency tables using the CC HPV mRNA test as the comparator. The unweighted Cohen's kappa statistic was used to evaluate the level of agreement between SC and CC HPV mRNA tests. The McNemar test with Fisher's exact test was used to assess the significance of discordance. To examine the test-retest performance of the SC HPV mRNA tests, first (clinic) and repeated (home) test results were compared using the unweighted kappa statistic. The same method was used to compare the two CC HPV tests (mRNA test and DNA test). Kappa of 0.81-1.00 was interpreted as very good, 0.61-0.80 as good, 0.41-0.60 as moderate, 0.21-0.40 as fair, and <0.20 as poor (Altman, 1991). Based on a one-sided significance level of 0.05 and 80% power, 45 participants were needed to detect a kappa value >0.4, given that the expected kappa was 0.7. Participants with missing data or invalid test results were excluded from individual analyses. Statistics were performed using Stata version 14.1 (Stata Corp, College Station, TX, USA).

Results

Characteristics of participants

Seventy-five were women enrolled and four were excluded based on age or incomplete data. We also excluded one participant who had invalid results on CC HPV mRNA testing, leaving 70 evaluable participants. All of the 70 participants also underwent SC HPV mRNA tests at the clinic, with nine invalid samples (13%). Fifty-five participants performed home collection, with seven invalid samples (13%). Any HPV test positivity prevalence was 63% (95% CI 51–74%).

R. Wang, K. Lee, C.A. Gaydos et al. / International	Journal of Infectious Diseases 99 (2020) 452–457
---	--

Table 1 Characteristics of the study population

Characteristics	Total (N=70)		
Continuous	Range	Median (IQR)	
Age (years)	30-66	50 (41-56)	
Categorical	п	%	
Race			
Black	65	92.9	
Non-Black	5	7.1	
Smoking status			
Current	36	51.4	
Former	13	18.6	
Never	21	30.0	
On antiretroviral treatment			
Yes	66	94.3	
No	4	5.7	
CD4 T cell count (cells/µl)			
<200	8	11.4	
≥200	62	88.6	
HIV RNA			
Detectable (>20 copies/ml)	17	24.3	

IQR, interguartile range; HPV, human papillomavirus.

Undetectable (<20 copies/ml)

Any HPV test positive

The median age of the participants was 50 years (interquartile range 41–56 years), and over 92% of participants were of Black race. Over half were current smokers (51%), the majority were taking antiretroviral therapy (94%), and 76% had an undetectable HIV RNA concentration. About 11% of the participants' most recent CD4 T-cell count was <200 cells/ μ l (Table 1).

53

44

Validity and reliability of HPV sample collection strategies

Using the CC HPV mRNA test as the comparator, the sensitivity of the SC HPV mRNA test was 84.6% (95% CI 65.1–95.6%) and specificity was 62.9% (95% CI 44.9%–78.5%) (Table 2). There was a difference in the proportion of positive versus negative results between these two HPV mRNA tests, with marginal significance under the McNemar test (p = 0.049). The agreement between the CC HPV mRNA test and the SC HPV mRNA test was moderate, with a kappa value of 0.5 (95% CI 0.2–0.7). In addition, the reliability of the SC HPV mRNA tests (clinic versus home) was good, with a kappa value of 0.8 (95% CI 0.5–1.0). We performed a sensitivity analysis that included invalid test results as negative or positive. The

Table 2

Validity and reliability of self-collected HPV testing.	
---	--

Clinic self-collected	Clinician-collected HPV mRNA tests			Kappa (95% CI)
	Positive	Negative	Total	
Positive	22	13	35	0.5 (0.2-0.7)
Negative	4	22	26	
Total	26	35	61	
Sensitivity (95% CI), %	84.6 (65.1-95.6)			
Specificity (95% CI), %	62.9 (45.0-78.5)			
PPV (95% CI), %	62.9 (45.0-78.5)			
NPV (95% CI), %	84.6 (65.1-	-95.6)		

Panel B: Reliability of self-collected HPV mRNA tests

Home self-collected	Clinic self-collected			Kappa (95% CI)
	Positive	Negative	Total	
Positive	23	2	25	0.8 (0.5-1.0)
Negative	3	14	17	
Total	26	16	42	

HPV, human papillomavirus; CI, confidence interval; PPV, positive predictive value; NPV, negative predictive value.

reliability of SC HPV tests (clinic versus home) showed a good level of agreement if invalid results were included as negative results ($\kappa = 0.7$, 95% CI 0.5–0.9), and a moderate level of agreement if invalid results were included as positive results ($\kappa = 0.5$, 95% CI 0.3–0.7). The agreement between CC HPV mRNA tests and CC HPV DNA tests was good ($\kappa = 0.8$, 95% CI 0.7–1.0).

Self-collection acceptability survey

Sixty-one participants (87%) completed the survey. About 79% of women thought self-collection was easy. No participants described self-collection as hard or very hard. Almost all of the women (97%) agreed or strongly agreed that they understood the self-sampling instructions. Less than a third (27%) of participants felt anxious about self-sampling and felt it was painful (24%). Overall, 79.0% agreed that they would use the cytobrush again. An overwhelming majority (90%) agreed that self-sampling was convenient and comfortable. Additionally, 86% of participants would recommend the cytobrush to family and friends (Figure 1).

Discussion

75.7

62.9

This study found that 63% of WLHIV had an HPV infection, which is similar to other studies among WLHIV (Kojic et al., 2011) and much higher than the rate in women without HIV (2.5–4.2%) (Sargent et al., 2008). HIV-induced immunodeficiency is thought to impede HPV clearance, resulting in the persistence of HPV infection, along with an increased risk of cervical cancer and precancer (Denny et al., 2012). Thus, developing a widely acceptable and accessible screening strategy and guidelines to optimize cervical cancer prevention among WLHIV is crucial.

Compared to the clinician-collected HPV test, it was found that self-collected tests had good sensitivity, moderate specificity, and moderate agreement. This finding is in agreement with one previous study that showed moderate concordance of selfcollected vaginal samples compared with clinician-collected cervical samples (Cho et al., 2019). In the present study, the test-retest performance of the self-collected strategy was good and the overall performance of self-collected tests was acceptable, although it was still inferior to tests using clinician-collected cervical samples. Findings from several studies have also revealed similar concerns. A large population-based cervical cancer screening study in China showed self-collected HPV testing sensitivity and specificity for detecting precancerous lesions was 86.2% and 80.7%, respectively, whereas clinician-collected HPV testing sensitivity and specificity were 97% and 82.7%, respectively (Zhao et al., 2012). Results from a meta-analysis of 36 studies examining self-collected versus clinician-collected samples (Arbyn et al., 2014) also reported lower pooled sensitivity and specificity of HPV testing on self-collected than clinician-collected samples.

The present study had a higher than expected rate of invalid testing in the self-collected samples compared to cliniciancollected samples. However, the level of agreement between the first (clinic) and repeated (home) test results of self-collection was good, despite the invalid test results. These invalid test results may have been due to insufficient sample material (Engesæter et al., 2016). It is possible that participants did not adhere to the instructions to insert the brush into the vagina deep enough or did not rotate the brush enough times, which could have translated to lower sample yield. Additionally, a cytobrush was used for collection, which may have been uncomfortable to use and may have led to hesitancy with sample collection. A higher number of invalid tests were not reflected in other self-collection studies that have been done on average-risk populations (Saville et al., 2018; Smith et al., 2018). However, these studies used HPV DNA tests and different collection tools. Also, compared to self-collected samples,



Fig. 1. Survey results.

clinician-collected samples likely had greater yield since clinicians are experienced with genital sample collection and performed it under speculum examination. Lower sample material could be a drawback with self-collected methods, but it is potentially associated with the type of collection tools or assay used. Future studies should investigate the impact of different collection devices and techniques on assay test performance similar to studies that have been performed for other female genital tract analyses (Dezzutti et al., 2011).

Another important finding of this study was the high acceptance of self-collected HPV testing among WLHIV. Women had positive attitudes toward this testing strategy and thought testing was easy. Also, they felt they were taking care of themselves and would use it again. This indicates that the self-collected HPV testing strategy may be able to increase cervical cancer screening, if the purpose of HPV testing is made clear to patients. For example, one of the largest randomized clinical trials in the US of home HPV testing reported only 12% of participants returned the home HPV test kit, and there was no difference in detection of precancerous lesions compared to usual care (Winer et al., 2019). The poor kit return in this trial might be explained by the patients' lack of knowledge regarding the superior performance of HPV testing, the requirement to still attend the clinic for usual care, or the awareness that the results could not replace usual care screening methods (Moss et al., 2019).

The present study builds on this literature by describing HPV self-collection with a specific focus on an underserved urban population with a higher risk of cervical cancer. Moreover, unlike a large-scale study testing its hypothesis under ideal highly controlled conditions, this study examined the 'real-world' experience of HPV self-collection among WLHIV. An earlier study from our clinic found that WLHIV were less likely to come to gynecology visits as compared to primary HIV care visits, and one of several reasons was fear or discomfort associated with the pelvic examination (Tello et al., 2010). Given that data are insufficient to

recommend pelvic examinations in the absence of symptoms (US Preventive Services Task Force et al., 2017) and the trend of making pelvic examinations contingent on medical history or symptoms (ACOG, 2018), the rationale for HPV self-collection at home or at non-gynecology clinic sites is strengthened. HPV self-collection could result in more WLHIV being screened for cervical cancer, which may justify its slightly inferior performance compared to clinician-collected sampling methods. Self-collected HPV testing could be a particularly effective strategy to reach underserved women, especially those living in health professional shortage areas (HPSAs) such as rural areas or those with barriers to attending medical appointments like lack of transportation or work/child care responsibilities.

This study has some limitations. First, the study population was relatively small, which limited the certainty of interpreting the results. However, the study was adequately powered and provided important evidence for future study of primary HPV cervical cancer screening strategies in WLHIV. Second, this study was limited by a higher than expected number of invalid results, although they did not appear to greatly impact the reliability of the tests. Last, the study population was predominately Black WLHIV enrolled at one urban site, which may not represent all WLHIV. However, Black patients accounted for 42% of the new HIV diagnoses in the US (CDC, 2019) and the population is a reflection of the nation's racial disparity.

In conclusion, the examination of self-collection highlights the specific needs of this population of US WLHIV and the opportunity to improve HPV screening in high-resource settings. Compared to the clinician-collected HPV test, the self-collected HPV test had good sensitivity and moderate specificity among WLHIV. Due to the possible inadequate HPV sample material resulting from self-collection, future studies should focus on how to improve self-collection techniques (including urine-based testing), since it is highly accepted by women and may improve cervical cancer screening in WLHIV who are at higher risk of cervical cancer.

Ethical approval

This study was approved by Johns Hopkins Institutional Review Boards.

Funding

This study was funded by grants from the Robert Wood Johnson Foundation (grant number 90042344).

Conflict of interest

Hologic Company donated HPV mRNA test kits for this study.

Acknowledgements

We would like to acknowledge the patients who participated in the project and Perry Barnes, Nicole Quinn, and Rosemary Ramroop for their assistance with the laboratory assay and recruitment.

References

- ACOG. ACOG Committee Opinion No. 754: the utility of and indications for routine pelvic examination. Obstet Gynecol 2018;132(4):e174–80.
- Altman DG. Some common problems in medical research. Practical statistics for medical research. London: Chapman and Hall; 1991. p. 403–9.
- Arbyn M, Verdoodt F, Snijders PJF, Verhoef VMJ, Suonio E, Dillner L, et al. Accuracy of human papillomavirus testing on self-collected versus clinician-collected samples: a meta-analysis. Lancet Oncol 2014;15(Feb (2)):172–83.
- Bulkmans N, Berkhof J, Rozendaal L, van Kemenade F, Boeke A, Bulk S, et al. Human papillomavirus DNA testing for the detection of cervical intraepithelial neoplasia grade 3 and cancer: 5-year follow-up of a randomised controlled implementation trial. Lancet 2007;370(Nov (9601)):1764–72.
- CDC. Centers for Disease Control and Prevention. HIV Surveillance Report, 2018 (preliminary) 2019;vol 30: [Internet]. Available from: https://www.cdc.gov/hiv/ library/reports/hiv-surveillance.html.
- Chaturvedi AK, Madeleine MM, Biggar RJ, Engels EA. Risk of human papillomavirusassociated cancers among persons with AIDS. J Natl Cancer Inst 2009;101(Aug (160)):1120–30.
- Cho H-W, Ouh Y-T, Hong JH, Min KJ, So KA, Kim TJ, et al. Comparison of urine, selfcollected vaginal swab, and cervical swab samples for detecting human papillomavirus (HPV) with Roche Cobas HPV, Anyplex II HPV, and RealTime HR-S HPV assay. J Virol Methods 2019;269:77–82.
- Clad A, Reuschenbach M, Weinschenk J, Grote R, Rahmsdorf J, Freudenberg N. Performance of the Aptima high-risk human papillomavirus mRNA assay in a referral population in comparison with Hybrid Capture 2 and cytology. J Clin Microbiol 2011;49(Mar (3)):1071–6.
- Cuzick J, Clavel C, Petry K-U, Meijer CJLM, Hoyer H, Ratnam S, et al. Overview of the European and North American studies on HPV testing in primary cervical cancer screening. Int J Cancer 2006;119(Sep (5)):1095–101.
- De Alba I, Anton-Culver H, Hubbell FA, Ziogas A, Hess JR, Bracho A, et al. Selfsampling for human papillomavirus in a community setting: feasibility in Hispanic women. Cancer Epidemiol Biomarkers Prev 2008;17(Aug (8)):2163–8.
- Denny LA, Franceschi S, de Sanjosé S, Heard I, Moscicki AB, Palefsky J. Human papillomavirus, human immunodeficiency virus and immunosuppression. Vaccine 2012;30(Nov (Suppl 5)):F168–74.
- Denslow SA, Rositch AF, Firnhaber C, Ting J, Smith JS. Incidence and progression of cervical lesions in women with HIV: a systematic global review. Int J STD AIDS 2014;25(Mar (3)):163–77.
- Dezzutti CS, Hendrix CW, Marrazzo JM, Pan Z, Wang L, Louissaint N, et al. Performance of swabs, lavage, and diluents to quantify biomarkers of female genital tract soluble mucosal mediators. PLoS One 2011;6(8):e23136.
- Engels EA, Pfeiffer RM, Goedert JJ, Virgo P, McNeel TS, Scoppa SM, et al. Trends in cancer risk among people with AIDS in the United States 1980–2002. AIDS 2006;20(Aug (12)):1645–54.
- Engesæter B, van Diermen Hidle B, Hansen M, Moltu P, Staby KM, Borchgrevink-Persen S, et al. Quality assurance of human papillomavirus (HPV) testing in the implementation of HPV primary screening in Norway: an inter-laboratory reproducibility study. BMC Infect Dis 2016;16(Dec (1)):698.
- Gyllensten U, Gustavsson I, Lindell M, Wilander E. Primary high-risk HPV screening for cervical cancer in post-menopausal women. Gynecol Oncol 2012;125(May (2)):343–5.
- Huh WK, Ault KA, Chelmow D, Davey DD, Goulart RA, Garcia FAR, et al. Use of primary high-risk human papillomavirus testing for cervical cancer screening: interim clinical guidance. Obstet Gynecol 2015;125(Feb (2)):330–7.
- IARC Working Group on the Evaluation of Carcinogenic Risks to Humans. Biological agents. Volume 100 B. A review of human carcinogens. IARC Monogr Eval Carcinog Risks Hum 2012;100(Pt B):1–441.

- Knoepp SM, Kuebler DL, Wilbur DC. Resolution of equivocal results with the Hybrid Capture II high-risk HPV DNA test: a cytologic/histologic review of 191 cases. Diagn Mol Pathol 2007;16(Sep (3)):125–9.
- Kojic EM, Cu-Uvin S, Conley L, Bush T, Onyekwuluje J, Swan DC, et al. Human papillomavirus infection and cytologic abnormalities of the anus and cervix among HIV-infected women in the study to understand the natural history of HIV/AIDS in the era of effective therapy (the SUN study). Sex Transm Dis 2011;38(Apr (4)):253–9.
- Leinonen MK, Nieminen P, Lönnberg S, Malila N, Hakama M, Pokhrel A, et al. Detection rates of precancerous and cancerous cervical lesions within one screening round of primary human papillomavirus DNA testing: prospective randomised trial in Finland. BMJ 2012;345(Nov):e7789.
- Levi JE, Kleter B, Quint WGV, Fink MCS, Canto CLM, Matsubara R, et al. High prevalence of human papillomavirus (HPV) infections and high frequency of multiple HPV genotypes in human immunodeficiency virus-infected women in Brazil. J Clin Microbiol 2002;40(Sep (9)):3341–5.
- Malila N, Leinonen M, Kotaniemi-Talonen L, Laurila P, Tarkkanen J, Hakama M. The HPV test has similar sensitivity but more overdiagnosis than the Pap test – a randomised health services study on cervical cancer screening in Finland. Int J Cancer 2013;132(May (9)):2141–7.
- Menon S, Wusiman A, Boily MC, Kariisa M, Mabeya H, Luchters S, et al. Epidemiology of HPV genotypes among HIV positive women in Kenya: a systematic review and meta-analysis. Chung MH, editor. PLoS One 2016;11(Oct (10)):e0163965.
- Moscicki A, Ellenberg JH, Farhat S, Xu J. Persistence of human papillomavirus infection in HIV-infected and -uninfected adolescent girls: risk factors and differences, by phylogenetic type. J Infect Dis 2004;190(Jul (1)):37–45.
- Moss CF, Chou B, Coleman JS. Home screening for human papillomavirus falls short in initial application, remains promising. JAMA Netw Open 2019;2(Nov (11)): e1914704.
- Naucler P, Ryd W, Törnberg S, Strand A, Wadell G, Elfgren K, et al. Human papillomavirus and papanicolaou tests to screen for cervical cancer. N Engl J Med 2007;357(Oct (16)):1589–97.
- Nelson EJ, Hughes J, Oakes JM, Thyagarajan B, Pankow JS, Kulasingam SL. Human papillomavirus infection in women who submit self-collected vaginal swabs after internet recruitment. J Commun Health 2015;40(Jun (3)):379–86.
- Ogilvie GS, Krajden M, van Niekerk DJ, Martin RE, Ehlen TG, Ceballos K, et al. Primary cervical cancer screening with HPV testing compared with liquid-based cytology: results of round 1 of a randomised controlled trial the HPV FOCAL Study. Br J Cancer 2012;107(Dec (12)):1917–24.
- Oster AM, Sullivan PS, Blair JM. Prevalence of cervical cancer screening of HIVinfected women in the United States. J Acquir Immune Defic Syndr 2009;51(Aug (4)):430–6.
- Panel on Opportunistic Infections in Adults and Adolescents with HIV. Guidelines for the prevention and treatment of opportunistic infections in adults and adolescents with HIV: recommendations from the Centers for Disease Control and Prevention, the National Institutes of Health, and the HIV Medicine Association of the Infectious Diseases Society of America. MMWR Recomm Rep 2009; Available at http://aidsinfo.nih.gov/contentfiles/lvguidelines/adult_oi. pdf.
- Peprah S, Coleman JS, Rositch AF, Vanden Bussche CJ, Moore R, D'Souza G. Utilization of Pap testing among women living with HIV enrolled in primary care in Baltimore, Maryland: a 10-year longitudinal study, 2005–2014. Papillomavirus Res 2018;6(Dec):52–7.
- Racey CS, Withrow DR, Gesink D. Self-collected HPV testing improves participation in cervical cancer screening: a systematic review and meta-analysis. Can J Public Health 2013;104(Feb (2)):e159–66.
- Ronco G, Dillner J, Elfström KM, Tunesi S, Snijders PJF, Arbyn M, et al. Efficacy of HPV-based screening for prevention of invasive cervical cancer: follow-up of four European randomised controlled trials. Lancet 2014;383(Feb (9916)):524– 32.
- Ronco G, Giorgi-Rossi P, Carozzi F, Confortini M, Palma PD, Del Mistro A, et al. Efficacy of human papillomavirus testing for the detection of invasive cervical cancers and cervical intraepithelial neoplasia: a randomised controlled trial. Lancet Oncol 2010;11(Mar (3)):249–57.Sargent A, Bailey A, Almonte M, Turner A, Thomson C, Peto J, et al. Prevalence of
- Sargent A, Bailey A, Almonte M, Turner A, Thomson C, Peto J, et al. Prevalence of type-specific HPV infection by age and grade of cervical cytology: data from the ARTISTIC trial. Br J Cancer 2008;98(May (10)):1704–9.
- Saville M, Hawkes D, Mclachlan E, Anderson S, Arabena K. Self-collection for underscreened women in a National Cervical Screening Program: pilot study. Curr Oncol 2018;25(Feb (1)):27.
- Siegel RL, Miller KD, Jemal A. Cancer statistics, 2019. CA Cancer J Clin 2019;69(Jan (1)):7-34.
- Smith JS, Des Marais AC, Deal AM, Richman AR, Perez-Heydrich C, Yen-Lieberman B, et al. Mailed human papillomavirus self-collection with papanicolaou test referral for infrequently screened women in the United States. Sex Transm Dis 2018;45(Jan (1)):42–8.
- Tello MA, Jenckes M, Gaver J, Anderson JR, Moore RD, Chander G. Barriers to recommended gynecologic care in an urban United States HIV clinic. J Womens Health (Larchmt) 2010;19(Aug (8)):1511–8.
- US Preventive Services Task Force, Bibbins-Domingo K, Grossman DC, Curry SJ, Barry MJ, Davidson KW, et al. Screening for gynecologic conditions with pelvic examination: US Preventive Services Task Force Recommendation Statement. JAMA 2017;317(Mar (9)):947.
- Walboomers JM, Jacobs MV, Manos MM, Bosch FX, Kummer JA, Shah KV, et al. Human papillomavirus is a necessary cause of invasive cervical cancer worldwide. J Pathol 1999;189(Sep (1)):12–9.

- Winer RL, Gonzales AA, Noonan CJ, Cherne SL, Buchwald DS. Collaborative to Improve Native Cancer Outcomes (CINCO). Assessing acceptability of selfsampling kits. prevalence, and risk factors for human papillomavirus infection in American Indian women. J Commun Health 2016;41(5):1049–61.
- Winer RL, Lin J, Tiro JA, Miglioretti DL, Beatty T, Gao H, et al. Effect of mailed human papillomavirus test kits vs usual care reminders on cervical cancer screening uptake. Precancer detection, and treatment: a randomized clinical trial. JAMA Netw Open 2019;2(Nov (11)):e1914704.
- Wright TC, Stoler MH, Behrens CM, Sharma A, Zhang G, Wright TL. Primary cervical cancer screening with human papillomavirus: end of study results from the ATHENA study using HPV as the first-line screening test. Gynecol Oncol 2015;136(Feb (2)):189–97.
- Zhao F-H, Lewkowitz AK, Chen F, Lin MJ, Hu S-Y, Zhang X, et al. Pooled analysis of a self-sampling HPV DNA test as a cervical cancer primary screening method. J Natl Cancer Inst 2012;104(Feb (3)):178–88.