



Plasma membrane glycosphingolipid signaling: a turning point

Elena Chiricozzi¹

Received: 16 June 2021 / Revised: 16 June 2021 / Accepted: 25 June 2021 / Published online: 16 August 2021
© The Author(s) 2021

Abstract

Plasma membrane interaction is highly recognized as an essential step to start the intracellular events in response to extracellular stimuli. The ways in which these interactions take place are less clear and detailed. Over the last decade my research has focused on developing the understanding of the glycosphingolipids-protein interaction that occurs at cell surface. By using chemical synthesis and biochemical approaches we have characterized some fundamental interactions that are key events both in the immune response and in the maintenance of neuronal homeostasis. In particular, for the first time it has been demonstrated that a glycolipid, present on the outer side of the membrane, the long-chain lactosylceramide, is able to directly modulate a cytosolic protein. But the real conceptual change was the demonstration that the GM1 oligosaccharide chain is able, alone, to replicate numerous functions of GM1 ganglioside and to directly interact with plasma membrane receptors by activating specific cellular signaling. In this conceptual shift, the development and application of multidisciplinary techniques in the field of biochemistry, from chemical synthesis to bioinformatic analysis, as well as discussions with several national and international colleagues have played a key role.

Keywords Glycosphingolipid · Ceramide · GM1 oligosaccharide · Plasma membrane interaction · IGO award · Early career award

Abbreviations

Ganglioside nomenclature is in accordance with IUPAC-IUBB recommendations [1]

BBB	blood brain barrier
GM1	II ³ Neu5Ac-Gg ₄ Cer, β-Gal-(1-3)-β-GalNAc-(1-4)-[α-Neu5Ac-(2-3)]-β-Gal-(1-4)-β-Glc-Cer
LacCer	Lactosylceramide, β-Gal(1-4)-β-Glc-Cer
MPTP	1-Methyl-4-phenyl-1,2,3,6-tetrahydropyridine hydrochloride
NGF	nerve growth factor
OligoGM1	GM1 oligosaccharide, II ³ Neu5Ac-Gg ₄
PD	Parkinson's disease
Trk	neurotrophin tyrosin kinase receptor

Introduction

It is a great honour and privilege to be this year's recipient of the International Glycoconjugate Organization (IGO) Young Glycoscientist Award.

✉ Elena Chiricozzi
elena.chiricozzi@unimi.it

¹ Department of Medical Biotechnology and Translational Medicine, University of Milano, Milano, Italy

As a biochemist, my research focuses on lipid biochemistry and in particular characterising the sphingolipids involvement in the regulation of the cell homeostasis [2–4]. Although my work in this area only began during my graduate studies in 2010, I have been amazed to be, together with my research team, the author of important discoveries in the field of membrane interactions between proteins and lipids in the last decade. These advancements have been the result of multidisciplinary collaboration in both domestic and international research institutes: the broad collaborative network I sowed along my career represents for my research a lifeline.

My key contributions in this research field have been the application of approaches based both on the molecular study thanks to proteomics and bioinformatics techniques, and the application of chemical synthesis products, such as photoactivable compounds, in which, historically, my research group is on top of the world. Using these approaches, we have greatly improved our understanding of the mechanisms by which membrane lipids interact with specific proteins to ensure adequate cell signalling.

Therefore, in this summary of my contributions I will try to highlight these important and new discoveries so that they may be the key to opening a new perspective on lipid-mediated signalling across the plasma membrane.

Glycosphingolipids chemistry and their biochemical role

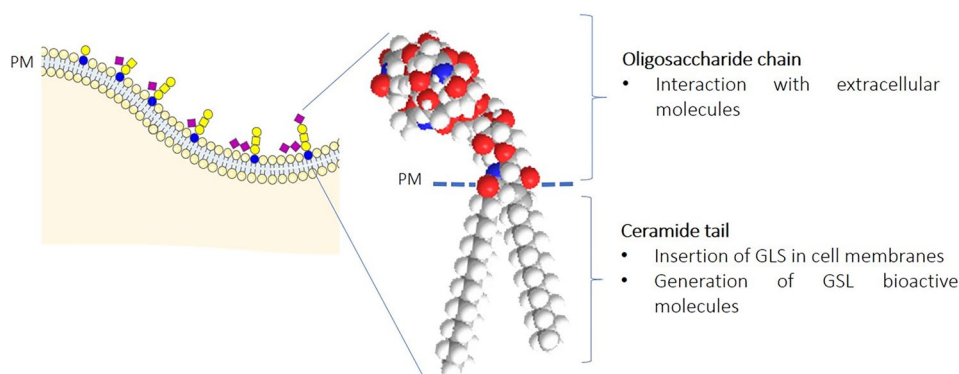
Glycosphingolipids are a family of membrane lipids with important roles in the regulation of the fluidity and subdomain structure of the lipid bilayer characterized by particular physical-chemical properties that allow to act as organizers of cell membrane [5–8]. These molecules are strongly amphiphilic: they are characterized by a hydrophobic portion, embedded in the lipidic core of biological membranes, and by a hydrophilic portion protruding in the extracellular milieu (Fig. 1).

The *ceramide* backbone confers some physical-chemical properties to the sphingolipids that differ from those of other membrane lipids. In fact, the amidic linkage, with the contemporary presence in the same molecule of a hydrogen bond donor and an acceptor group (the carbonyl oxygen and the amidic hydrogen), allows the formation of a thick network of hydrogen bonds among different sphingolipid molecules in the lipid bilayer. The presence of hydrogen linkages considerably stabilizes the sphingolipid segregation in specific membrane areas, which appear enriched with this lipid family and for this reason they have been defined “glycosphingolipid-rich membrane domains” [9–12]. In addition, the numerous hydrogen bonds in these domains confer rigidity and resistance to the membrane allowing their differentiation by physical-chemical properties from the remaining membrane.

On the other side of the molecule there is the hydrophilic head that faces the extracellular environment: the *saccharide core*. This is small and composed by one or two saccharides in neutral sphingolipids, while it is composite and more complex in gangliosides, glycosylated sphingolipids characterized by the presence of sialic acid. For its part, the oligosaccharide head can interact through mild hydrophilic bonds, *i.e.*, hydrogen bonds, with neighbouring membrane components or extracellular ligands.

Recent results suggest new aspects of lipid function: not only structural components of cell membrane but also fundamental actors of signalling and regulatory pathways.

Fig. 1 Plasma membrane glycosphingolipid structure: the oligosaccharide head and the ceramide tail. PM: plasma membrane. Glycosphingolipid sugar code is according to Varki *et al.* [49]



Unexpectedly, advances in biochemical and molecular studies of glycosphingolipid functions during the past 3 decades, revealed that biological active glycosphingolipids (ceramide, ceramide 1 phosphate, glucosyl-ceramide, lactosylceramide, galactosylceramide, sphingosine, sphingosine 1 phosphate, ganglioside) have key roles in the regulation of several fundamental biological processes: they act also as effector molecules and not only as inert precursor and products of glycosphingolipids metabolism [13–21].

In fact, once understood the conformation of these molecules, it's easy to image how the monomeric glycosphingolipids can interact, through the lipid component and by the oligosaccharide one, both with the membrane components (*i.e.*, proteins and lipids), segregating and clustering into the membrane and with extracellular actors. Therefore, these molecules forming cell type specific profiles have essential roles in several aspects of cell homeostasis: cell growth, differentiation, morphogenesis, cell to matrix interaction, cell to cell communication and so on.

Below I report two of my researches that highlight the importance of both the *ceramide tail* and the *saccharide head* of these molecules as a starting point for the modulation of the plasma membrane signalling.

Ceramide tail role in the formation of Lactosylceramide lipid rafts in neutrophil cells

The biochemical study on glycosphingolipid-mediated signalling started in collaboration with the laboratory of Professor Kazuisa Iwabuchi, at the Juntendo University of Tokyo, where I spent 9 months carrying out a project aimed at understanding the role of a specific glycosphingolipid called Lactosylceramide (LacCer) in modulating the cell signalling involved in neutrophils response upon bacteria's infection [22].

The innate immune system is the first line of defence against pathogenic microorganisms, such as bacteria, fungi, and viruses. Phagocytes, such as neutrophils and

macrophages, play an important role in the innate immune system by recognizing, engulfing, and eliminating pathogens. It has been suggested that lipid membrane microdomains of phagocytes are involved in these innate immune responses, including superoxide generation, cell migration, and phagocytosis. Glycosphingolipids are highly enriched in specialized membrane microdomains where they participate to the process of transduction of information across the membrane. Recent proteomic analyses of microdomains from phagocytes have provided insight into membrane microdomain-mediated functions in the processes of phagocytosis [23–26]. LacCer, a neutral glycosphingolipid, is abundantly expressed on human neutrophils and, specifically, recognizes several pathogenic microorganisms [27, 28]. LacCer forms membrane microdomains coupled with the Src family kinase Lyn on the plasma membrane, and ligand binding to LacCer activates Lyn, resulting in neutrophil functions, such as superoxide generation and migration [22, 29, 30]. In contrast, neutrophilic-differentiated HL-60 cells, highly used in basic research instead of human neutrophils, do not have Lyn-associated LacCer-enriched microdomains and lack LacCer-mediated functions. In neutrophil plasma membranes, the very long fatty acid C24:0 and C24:1 chains are the main components of LacCer, whereas plasma membrane of D-HL-60 cells mainly includes C16-LacCer species. LacCer species containing very long fatty acid chains are thus indispensable for the association of Lyn with LacCer-enriched microdomains and LacCer-mediated functions [22, 29, 30].

With this project we want to study the role of very long fatty acid-LacCer species in the physical and functional coupling with Lyn and in microdomain-mediated functions in the processes of phagocytosis to determine the molecular mechanisms underlying these functions. For this purpose, our chemical group developed the tritium-labelled and photoactivable LacCer derivatives with the aim to give a name to these proteins. To understand what happens at the level of plasma membrane microdomain, we prepared the plasma membrane microdomains to analyse and to recognize the protein associated to the LacCer environment. By means of cross linking and immunoprecipitation experiments with photoactivable and radioactive [^3H]-C24/C16-(N₃)-LacCer we showed that Lyn and LacCer with long acyl chain have a direct interaction in LacCer-enriched membrane domain from neutrophilic cells (Fig. 2). First biological results with analogues are encouraging. In fact, they seem to confirm the initial hypothesis related to the requirement of long chains LacCer for the innate responses in human neutrophils.

Our results prove that glycosphingolipids with a long fatty acid chain of Ceramide moiety, in our specific case LacCer, could be the key-player on the signal transduction across the cellular plasma membrane, modulating membrane interdigitation, reducing the membrane thickness and forming a

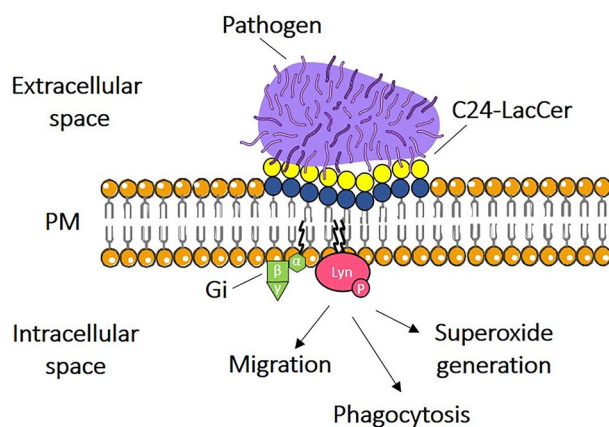


Fig. 2 Proposed mechanism diagram for LacCer-lipid raft mediated function in inflammatory response. C24-LacCer directly associates with the cytoplasmic protein Lyn via myristic/palmitic chains promoting its phosphorylation and activation of other proteins (Gi family protein) within plasma membranes lipid rafts of neutrophil cells. This specific C:24-LacCer/Lyn interaction finally leads to phagocytosis, superoxide generation and cell migration following CD11b/CD18 integrin translocation in C:24-LacCer microdomain in response to bacterial infection. Modified from [22]. Glycosphingolipid sugar code is according to Varki *et al.* [49]

specific plasma membrane microdomains. We know that, for LacCer mediated, Lyn activation is necessary: LacCer molecular species with ceramide contain a very long fatty acid chain, but also β -Gal(1-4)- β -Glc disaccharide structure of LacCer. This means that, in turn, the oligosaccharide chain is able to recognize some specific proteins belonging to the extracellular space. The synthesis of new LacCer photoactivable analogues will certainly help in the identification of these new players opening a new research field.

Oligosaccharide role in the GM1 ganglioside neuronal functions

Since 2015, my research activity has mainly been focused on GM1 ganglioside in relation to both neuronal physiological and pathological implication [31–41]. Together with my research group, I give a very important contribution to the understanding of the role of GM1 oligosaccharide as the bioactive portion of the GM1 molecule, opening a new perspective on the ganglioside-mediated signalling across the plasma membrane. Furthermore, the identification of the GM1 oligosaccharide as a membrane-permeable GM1 analogue with superior ability to access intraneuronal GM1 functional sites may hold considerable therapeutic promises for sporadic Parkinson's disease (PD). Like all the important discoveries, it derived from the randomness of the moment while we were carrying out a new research on the synthesis of GM1.

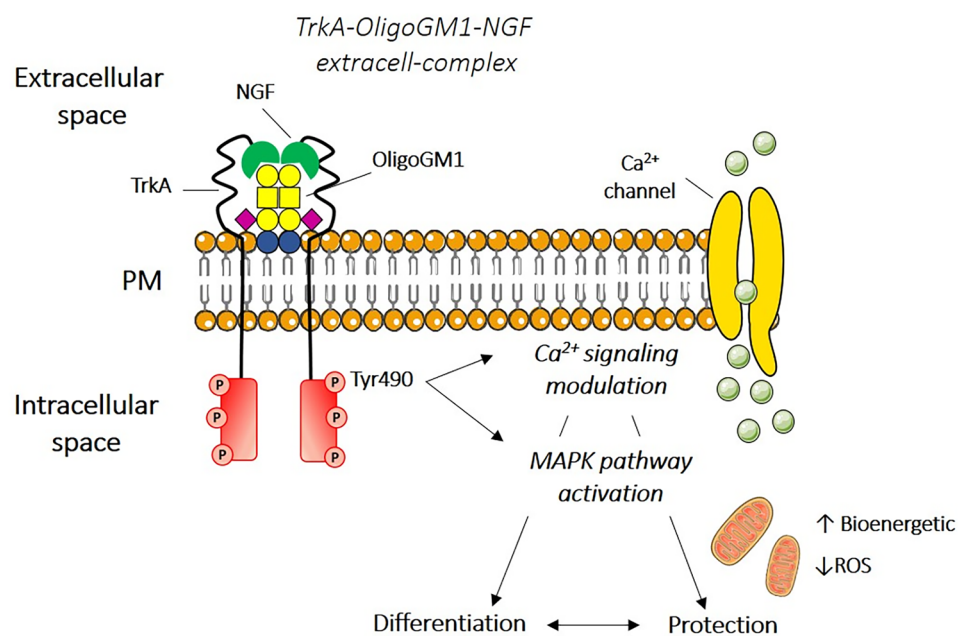
In vitro and *in vivo* studies have indirectly suggested us that the oligosaccharide portion of the GM1 (OligoGM1) could provide an important contribution. In 1988, a pioneering paper by Shengrund and Prouty observed that the OligoGM1 promoted neurogenesis [42]. In 2012, Ledeen and coworkers identified LIGA20, a membrane brain permeable analogue of GM1, as an effective (albeit toxic in the long-term) alternative to GM1 [43]. Treatment of GM1-deficient parkinsonian mice with LIGA20 induced beneficial results, including the reduction of *substantia nigra* α -synuclein aggregates. The most important finding of this study was the identification of a hydrophilic GM1 derivative, modified on the ceramide moiety with a dichloroacetyl group instead of the acyl chain, but keeping the entire oligosaccharide intact, which still maintained trophic potential. This suggested that the ceramide structure is not critically related to GM1 modulatory effects. In 2015, Schneider noted that the plasma membrane GM1 increased by intraventricular injection of *Vibrio cholerae* sialidase exerted a neuroprotective effect on the damaged nigrostriatal dopaminergic system of MPTP mice [44]. This enzyme removes sialic acid residues from brain polysialogangliosides (*i.e.*, GD1a), thus increasing plasma membrane GM1, further evidence that the oligosaccharide may act as the mediator of GM1 function.

Accordingly, my collaborators and I recently reported that the soluble OligoGM1 (II³Neu5Ac-Gg₄) replicates the neurotrophic and neuroprotective properties of the entire GM1 molecule: GM1 exerts its bioactivity through its hydrophilic head, which protrudes into the extracellular environment and therefore acts at the cell surface by interacting with plasma membrane proteins [31–41].

In fact, I reported in neuronal cells that, within the entire GM1 molecule, OligoGM1 is the actual moiety responsible for GM1 neurotrophic properties [31–41]. OligoGM1 directly interacts with the NGF receptor TrkA, leading to MAPK (*i.e.* ERK1/2) downstream pathway activation [31, 33, 38], to cell differentiation [31, 38] and to sustain MPTP neuroprotection [32] through mitochondria bioenergetic [32, 37] and calcium modulation [32, 40] (Fig. 3). Following its injection in wild-type mice, OligoGM1 was found to reach brain areas [34], and by using an *in vitro* human blood brain barrier (BBB) model, OligoGM1 showed a 20-fold higher crossing rate than that of entire GM1 and crossed the BBB directly by a passive-paracellular mechanism [36]. Importantly, OligoGM1 systemically administered to a PD mouse model (the GM1 deficient *B4galnt1*^{+/-} one) was found to completely rescue the physical symptoms, to reduce α S aggregates, to restore tyrosine hydroxylase neurons and to recover the neurotransmitters' levels [34]. Altogether, this evidence confirms that the specific role of ganglioside GM1 in neuronal homeostasis is mediated by its oligosaccharide: this bioactive portion, protruding in the extracellular environment, acts at the cell surface by a direct interaction with specific proteins.

From this premise, it emerged our idea that the plasma membrane OligoGM1 decline could be the trigger for sustained pathway impairment as the etiopathogenetic target event of sporadic PD, which is characterized by a systemic reduction of GM1. Supported by extensive research into the physiological function and pathological implications of ganglioside GM1 [39, 45–48] and by solid data on OligoGM1 functions [31–40], I propose a new key to untying the knot of complex neurodegenerative

Fig. 3 Proposed mechanism diagram of the GM1 oligosaccharide mediated functions. GM1 ganglioside through its oligosaccharide chain stabilizes the TrkA-NGF complex on the cell surface triggering the phosphorylation of TrkA on tyrosine 490 (Tyr490) promoting MAPK signaling. This induces the activation of multiple intracellular pathways that finally lead to neuronal differentiation, protection and restoration. Modified from [31, 33, 37, 40]. Glycosphingolipid sugar code is according to Varki *et al.* [49]



diseases: the OligoGM1 as an entirely new neurotrophic player, able to counteract all the multifactorial aspects of PD. Indeed, the soluble OligoGM1 loses the amphiphilicity of the entire ganglioside and gains the possibility to efficiently cross the BBB, reaching the target neurons. Here, without entering the cells but interacting with cell surface proteins, OligoGM1 should be enough to restore and maintain the correct neuronal homeostasis, through the activation of trophic plasma membrane signal and alpha-synuclein clearance, with significant therapeutic implications. The prompt restoration of all the mechanisms (*i.e.*, oxidative stress and mitochondrial dysfunction, inflammation, excitotoxicity, loss of trophic support and protein accumulation/aggregation) that are likely to contribute and to exacerbate the pathogenesis of Parkinson and in general to multifactorial neurodegenerative disorders (*i.e.*, Alzheimer's disease) is fundamental to the recovery of injured cells and tissues, particularly those with delicate equilibria (*i.e.*, dopaminergic neurons). Otherwise, OligoGM1 should drive plasma membrane trophic signalling inducing a reduction in glia/microglia activation, oxidative stress, mitochondria dysfunction, excitotoxicity and α -synuclein aggregation resulting in the recovery of damaged neurons and the restoration of normal healthy phenotype [39]. These concepts bring out the OligoGM1 as a cutting-edge molecule able to recover and cope with all aspects of neurodegenerative diseases, for which, to date, there are no effective therapies.

Overall, my aim is to provide a significant advance in the understanding of the molecular mechanism by which GM1, and in particular its oligosaccharide portion, is responsible for the onset occurring in sporadic PD. I expect to provide novel insights into the GM1-Parkinson pathogenesis, as well as into the mechanism occurring at the plasma membrane level that might account for the reduced neuroactivity in Parkinson neurons, but also in the extra-central tissues.

These findings will open a new perspective on the ganglioside mediated signalling across the plasma membrane. Furthermore, the identification of a novel membrane-permeable GM1 analogue with superior ability to access intraneuronal GM1 functional sites holds considerable therapeutic promises.

What's next

The field of glycobiology and, in particular, the studies concerning the molecular mechanisms underlying the interaction between membrane lipids and proteins are fields of research that represent the starting engine of everything that happens inside our cells in response to external stimuli. Even if it seems that everything has already been discovered, characterizing in a

fine and more detailed way, with the new technical approaches available today, will allow unexpected discoveries which may have even more unexpected applications.

A key challenge within this field will continue to be the identification of previously unknown interactions such as the one of GM1 oligosaccharide. The goal of my research group is to see OligoGM1 applied to the clinic since its ability to recover the neuro-degenerated Parkinson's phenotype is truly sensational and at the same time to develop a tool for the large-scale synthesis of OligoGM1. The hope is to verify how this molecule can be useful for all other diseases where there is a deficiency of GM1 or where GM1 has been used in the past but with poor results due to the impossibility of GM1 to overcome an intact BBB.

Acknowledgments My thanks go to everyone I have met in my short but busy career. Starting with the Professors and Researchers of the University of Perugia (Italy) and the University of Salamanca (Spain), where I took my first steps as an undergraduate student. The Juntendo University of Tokyo (Japan), Nara Institute of Science and Technology (Japan), Artois University (France) and the Rutgers University (Newark, NJ, US) where, as a guest researcher, I was able to deal with international realities that allowed me to obtain a good degree of independence and a large capability of discussion and contacts. Finally, my University of Milan, which gave me the opportunity and the tools to become a researcher and to compare myself with very valid colleagues. My heartfelt thanks go to my *OligoTeam* with whom I discuss every day and thanks to whom I find new incentives to continue this fascinating work.

Funding Open access funding provided by Università degli Studi di Milano within the CRUI-CARE Agreement.

Declarations

Conflict of interest The author declares that he has no conflicts of interest.

Ethical approval This article does not contain any studies with human participants or animals performed by the author.

Open Access This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit <http://creativecommons.org/licenses/by/4.0/>.

References

1. Chester, M.A.: IUPAC-IUB Joint Commission on Biochemical Nomenclature (JCBN). Nomenclature of glycolipids--recommendations 1997. *Eur. J. Biochem.* **257**, 293–8 (1998)

2. Chiricozzi, E., Niemir, N., Aureli, M., Magini, A., Loberto, N., Prinetti, A., Bassi, R., Polchi, A., Emiliani, C., Caillaud, C., Sonnino, S.: Chaperone therapy for GM2 gangliosidosis: effects of pyrimethamine on β -hexosaminidase activity in Sandhoff fibroblasts. *Mol. Neurobiol.* **50**, 159–67 (2013)
3. Prinetti, A., Prioni, S., Chiricozzi, E., Schuchman, E.H., Chigorno, V., Sonnino, S.: Secondary alterations of sphingolipid metabolism in lysosomal storage diseases. *Neurochem. Res.* **36**, 1654–68 (2011)
4. Aureli, M., Bassi, R., Prinetti, A., Chiricozzi, E., Pappalardi, B., Chigorno, V., Di Muzio, N., Loberto, N., Sonnino, S.: Ionizing radiations increase the activity of the cell surface glycohydrolases and the plasma membrane ceramide content. *Glycoconj. J.* **29**, 585–97 (2012)
5. Thudichum, J.L.W.: A Treatise on the chemical constitution of the brain: based throughout upon original researches. London: Bailliere, Tindall, and Cox. Biomed. J. Digit. Proj. (1884)
6. Sonnino, S., Prinetti, A., Mauri, L., Chigorno, V., Tettamanti, G.: Dynamic and structural properties of sphingolipids as driving forces for the formation of membrane domains. *Chem. Rev.* **106**, 2111–25 (2006)
7. Andreev, K.: The Structural Role of Gangliosides: Insights from x-ray scattering on model membranes. *Curr. Med. Chem.* **27**, 6548–70 (2020)
8. Schnaar, R.L., Kinoshita, T.: Glycosphingolipids. In: Varki A, Cummings RD, Esko JD, Stanley P, Hart GW, Aebi M, Darvill AG, Kinoshita T, Packer NH, Prestegard JH, Schnaar RL, Seeberger PH, editors. *Essentials of Glycobiology* [Internet]. 3rd ed. Cold Spring Harbor (NY): Cold Spring Harbor Laboratory Press; 2015–2017. **11**, (2017)
9. Sonnino, S., Chiricozzi, E., Grassi, S., Mauri, L., Prioni, S., Prinetti, A.: Gangliosides in membrane organization. *Prog. Mol. Biol. Transl. Sci.* **156**, 83–120 (2018)
10. Aureli, M., Grassi, S., Prioni, S., Sonnino, S., Prinetti, A.: Lipid membrane domains in the brain. *Biochim. Biophys. Acta.* **1851**, 1006–16 (2015)
11. Sonnino, S., Aureli, M., Mauri, L., Ciampa, M.G., Prinetti, A.: Membrane lipid domains in the nervous system. *Front. Biosci. (Landmark Ed)* **20**, 280–302 (2015)
12. Wenekes, T., van den Berg, R.J., Boot, R.G., van der Marel, G.A., Overkleeft, H.S., Aerts, J.M.: Glycosphingolipids—nature, function, and pharmacological modulation. *Angew Chem. Int. Ed. Engl.* **48**, 8848–69 (2009)
13. Lunghi, G., Fazzari, M., Di Biase, E., Mauri, L., Chiricozzi, E., Sonnino, S.: The structure of gangliosides hides a code for determining neuronal functions. *FEBS Open Bio.* (2021)
14. Chiricozzi, E., Loberto, N., Schiumarini, D., Samarani, M., Mancini, G., Tamanini, A., Lippi, G., Dechecchi, M.C., Bassi, R., Giussani, P., Aureli, M.: Sphingolipids role in the regulation of inflammatory response: From leukocyte biology to bacterial infection. *J. Leukoc. Biol.* **103**, 445–456 (2018)
15. Coskun, Ü., Simons, K.: Cell membranes: The lipid perspective. *Structure* **19**, 1543–1548 (2011)
16. Russo, D., Parashuraman, S., D'Angelo, G.: Glycosphingolipid–protein interaction in signal transduction. *Int. J. Mol. Sci.* **17**, (2016)
17. D'Angelo, G., Capasso, S., Sticco, L., Russo, D.: Glycosphingolipids: synthesis and functions. *FEBS J.* **280**, 6338–53 (2013)
18. Sasaki, N., Toyoda, M., Ishiwata, T.: Gangliosides as Signaling Regulators in Cancer. *Int. J. Mol. Sci.* **22**, 5076 (2021)
19. Ledeen, R.W., Kopitz, J., Abad-Rodríguez, J., Gabius, H.J.: Glycan chains of gangliosides: functional ligands for tissue lectins (Siglecs/Galectins). *Prog. Mol. Biol. Transl. Sci.* (2018)
20. Gomez-Larrauri, A., Presa, N., Dominguez-Herrera, A., Ouro, A., Trueba, M., Gomez-Muñoz, A.: Role of bioactive sphingolipids in physiology and pathology. *Essays Biochem.* **64**, 579–89 (2020)
21. Wu, A.M.: Glycan structures and their recognition roles in the human blood group ABH/i, Lea, b, x, y and Sialyl Lea, x active cyst glycoproteins. *Glycoconj. J.* **36**, 495–507 (2019)
22. Chiricozzi, E., Ciampa, M.G., Brasile, G., Compostella, F., Prinetti, A., Nakayama, H., Ekyalongo, R.C., Iwabuchi, K., Sonnino, S., Mauri, L.: Direct interaction, instrumental for signaling processes, between LacCer and Lyn in the lipid rafts of neutrophil-like cells. *J. Lipid. Res.* **56**, 129–41 (2015)
23. Nakayama, H., Ogawa, H., Takamori, K., Iwabuchi, K.: GSL-enriched membrane microdomains in innate immune responses. *Arch. Immunol. Ther. Exp. (Warsz)*. **61**, 217–28 (2013)
24. Iwabuchi, K., Nakayama, H., Masuda, H., Kina, K., Ogawa, H., Takamori, K.: Membrane microdomains in immunity: glycosphingolipid-enriched domain-mediated innate immune responses. *Biofactors* **38**, 275–83 (2012)
25. Iwabuchi, K., Nakayama, H., Iwahara, C., Takamori, K.: Significance of glycosphingolipid fatty acid chain length on membrane microdomain-mediated signal transduction. *FEBS Lett.* **584**, 1642–52 (2009)
26. Sonnino, S., Prinetti, A., Nakayama, H., Yangida, M., Ogawa, H., Iwabuchi, K.: Role of very long fatty acid-containing glycosphingolipids in membrane organization and cell signaling: the model of lactosylceramide in neutrophils. *Glycoconj. J.* **26**, 615–21 (2009)
27. Brackman, D., Lund-Johansen, F., Aarskog, D.: Expression of leukocyte differentiation antigens during the differentiation of HL-60 cells induced by 1,25-dihydroxyvitamin D3: Comparison with the maturation of normal monocytic and granulocytic bone marrow cells. *J. Leukoc. Biol.* **58**, 547–55 (1995)
28. Kniep, B., Skubitz, K.M.: Subcellular localization of glycosphingolipids in human neutrophils. *J. Leukoc. Biol.* **63**, 83–8 (1998)
29. Iwabuchi, K., Nagaoka, I.: Lactosylceramide-enriched glycosphingolipid signaling domain mediates superoxide generation from human neutrophils. *Blood* **100**, (2002)
30. Iwabuchi, K., Prinetti, A., Sonnino, S., Mauri, L., Kobayashi, T., Ishii, K., Kaga, N., Murayama, K., Kurihara, H., Nakayama, H., Yoshizaki, F., Takamori, K., Ogawa, H., Nagaoka, I.: Involvement of very long fatty acid-containing lactosylceramide in lactosylceramide-mediated superoxide generation and migration in neutrophils. *Glycoconj. J.* **25**, 357–74 (2008)
31. Chiricozzi, E., Pome, D.Y., Maggioni, M., Di Biase, E., Parravicini, C., Palazzolo, L., Loberto, N., Eberini, I., Sonnino, S.: Role of the GM1 ganglioside oligosaccharide portion in the TrkA-dependent neurite sprouting in neuroblastoma cells. *J. Neurochem.* **143**, 645–59 (2017)
32. Chiricozzi, E., Maggioni, M., di Biase, E., Lunghi, G., Fazzari, M., Loberto, N., Elisa, M., Scalvini, F.G., Tedeschi, G., Sonnino, S.: The Neuroprotective role of the GM1 oligosaccharide, I13Neu5Ac-Gg4, Neuroblastoma cells. *Mol. Neurobiol.* **56**, 6673–702 (2019)
33. Chiricozzi, E., Biase, E.D., Maggioni, M., Lunghi, G., Fazzari, M., Pome, D.Y., Casellato, R., Loberto, N., Mauri, L., Sonnino, S.: GM1 promotes TrkA-mediated neuroblastoma cell differentiation by occupying a plasma membrane domain different from TrkA. *J. Neurochem.* **149**, 231–41 (2019)
34. Chiricozzi, E., Mauri, L., Lunghi, G., Di Biase, E., Fazzari, M., Maggioni, M., Valsecchi, M., Prioni, S., Loberto, N., Pome, D.Y., Ciampa, M.G., Fato, P., Verlengia, G., Cattaneo, S., Assini, R., Wu, G., Alselehdar, S., Ledeen, R.W., Sonnino, S.: Parkinson's disease recovery by GM1 oligosaccharide treatment in the B4galnt1^{+/−} mouse model. *Sci. Rep.* **9**, 19330 (2019)
35. Chiricozzi, E., Lunghi, G., Di Biase, E., Fazzari, M., Sonnino, S., Mauri, L.: GM1 ganglioside is a key factor in maintaining the mammalian neuronal functions avoiding neurodegeneration. *Int. J. Mol. Sci.* **21**, 868 (2020)

36. Di Biase, E., Lunghi, G., Maggioni, M., Fazzari, M., Pomè, D.Y., Loberto, N., Ciampa, M.G., Fato, P., Mauri, L., Sevin, E., Gosselet, F., Sonnino, S., Chiricozzi, E.: GM1 oligosaccharide crosses the human blood-brain barrier *in vitro* by a paracellular route. *Int. J. Mol. Sci.* **21**, 2858 (2020)
37. Fazzari, M., Audano, M., Lunghi, G., Di Biase, E., Loberto, N., Mauri, L., Mitro, N., Sonnino, S., Chiricozzi, E.: The oligosaccharide portion of ganglioside GM1 regulates mitochondrial function in neuroblastoma cells. *Glycoconj. J.* **37**, 293–306 (2020)
38. Di Biase, E., Lunghi, G., Fazzari, M., Maggioni, M., Pome, D.Y., Valsecchi, M., Samarani, M., Fato, P., Ciampa, M.G., Prioni, S., Mauri, L., Sonnino, S., Chiricozzi, E.: Gangliosides in the differentiation process of primary neurons: the specific role of GM1-oligosaccharide. *Glycoconj. J.* **37**, 329–43 (2020)
39. Chiricozzi, E., Di Biase, E., Lunghi, G., Fazzari, M., Loberto, N., Aureli, M., Mauri, L., Sonnino, S.: Turning the spotlight on the oligosaccharide chain of GM1 ganglioside. *Glycoconj. J.* **38**, 101–117 (2021)
40. Lunghi, G., Fazzari, M., Di Biase, E., Mauri, L., Sonnino, S., Chiricozzi, E.: Modulation of calcium signaling depends on the oligosaccharide of GM1 in Neuro2a mouse neuroblastoma cells. *Glycoconj. J.* **37**, 713–727 (2020)
41. Mancini, G., Loberto, N., Olioso, D., Dececchi, M.C., Cabrini, G., Mauri, L., Bassi, R., Schiumarini, D., Chiricozzi, E., Lippi, G., Pesce, E., Sonnino, S., Pedemonte, N., Tamanini, A., Aureli, M.: GM1 as adjuvant of innovative therapies for cystic fibrosis disease. *Int. J. Mol. Sci.* **21**, 4486 (2020)
42. Schengrund, C.L., Prouty, C.: Oligosaccharide portion of GM1 enhances process formation by S20Y neuroblastoma cells. *J. Neurochem.* **51**, 277–82 (1988)
43. Wu, G., Lu, Z.H., Kulkarni, N., Ledeen, R.W.: Deficiency of ganglioside GM1 correlates with Parkinson's disease in mice and humans. *J. Neurosci. Res.* **90**, 1997–2008 (2012)
44. Schneider, J.S., Seyfried, T.N., Choi, H.S., Kidd, S.K.: Intraventricular sialidase administration enhances GM1 ganglioside expression and is partially neuroprotective in a mouse model of Parkinson's disease. *PLoS One* **10**, (2015)
45. Ledeen, R.W., Wu, G.: The multi-tasked life of GM1 ganglioside, a true factotum of nature. *Trends Biochem. Sci.* **40**, 407–18 (2015)
46. Aureli, M., Mauri, L., Ciampa, M.G., Prinetti, A., Toffano, G., Secchieri, C., Sonnino, S.: GM1 Ganglioside: past studies and future potential. *Mol. Neurobiol.* **53**, 1824–42 (2016)
47. Schneider, J.S.: A critical role for GM1 ganglioside in the pathophysiology and potential treatment of Parkinson's disease. *Glycoconj. J.* (2021)
48. Magistretti, P.J., Geisler, F.H., Schneider, J.S., Li, P.A., Fiumelli, H., Sipione, S.: Gangliosides: treatment avenues in neurodegenerative disease. *Front. Neurol.* **10**, 859 (2019)
49. Varki, A., Cummings, R.D., Aeby, M., Packer, N.H., Seeberger, P.H., Esko, J.D., Stanley, P., Hart, G., Darvill, A., Kinoshita, T., Prestegard, J.J., Schnaar, R.L., Freeze, H.H., Marth, J.D., Bertozzi, C.R., Etzler, M.E., Frank, M., Vliegthart, J.F., Lutteke, T., Perez, S., Bolton, E., Rudd, P., Paulson, J., Kanehisa, M., Toukach, P., Aoki-Kinoshita, K.F., Dell, A., Narimatsu, H., York, W., Taniguchi, N., Kornfeld, S.: Symbol nomenclature for graphical representations of glycans. *Glycobiology* **25**, 1323–4 (2015)

Publisher's Note Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.



2021 IGO Young Glycoscientist Award Elena Chiricozzi

Dr. Elena Chiricozzi's work has contributed significantly to the field of glycobiology, focusing on the role of the sphingolipids and their glycoconjugate derivatives in regulating cell homeostasis, and modulating cell surface properties through the interaction with plasma membrane proteins. She obtained her PhD at the University of Milano in 2013 where she studied the glycosphingolipids implication in several pathology ranging from neurodegenerative disorders to cancer. Then, Dr. Chiricozzi took up multiple post-doctoral experiences at Juntendo University of Tokyo (Japan), Nara Institute of Science and Technol-

ogy (Japan), Rutgers University (Newark, NJ, US), San Rafael Hospital (Milano, Italy), and University Artois (Lens, France). Thanks to these opportunities, her research activity has been developed in the context

of several national and international collaborations, which allowed her to obtain an excellent degree of independence together with a strong communicative skill and to establish new contacts. In addition, she gained extensive knowledge of multidisciplinary approaches by applying techniques of chemistry, biochemistry, cellular and molecular biology. Currently her research activity is mainly focused on GM1 ganglioside in relation to both neuronal physiological and pathological implication. She made an important contribution in understanding the role of the oligosaccharide portion of GM1 as the bioactive component of the molecule, opening a new perspective on the ganglioside-mediated signaling across the plasma membrane. Furthermore, the identification of the GM1 oligosaccharide as a blood-brain barrier permeable analogue with superior ability to reach GM1 functional neuronal sites, may hold considerable therapeutic promise in the context of neurodegenerative diseases such as Parkinson's disease. Dr. Chiricozzi now heads her own independent research group at the University of Milano, Department of Medical Biotechnology and Translational Medicine.