

RESEARCH ARTICLE

Arabidopsis HFR1 Is a Potential Nuclear Substrate Regulated by the *Xanthomonas* Type III Effector XopD_{Xcc8004}

Choon Meng Tan^{1,2}, Meng-Ying Li¹, Pei-Yun Yang¹, Shu Heng Chang¹, Yi-Ping Ho¹, Hong Lin¹, Wen-Ling Deng³, Jun-Yi Yang^{1,2,4,5,6*}

1 Institute of Biochemistry, National ChungHsing University, Taichung, Taiwan, **2** Ph.D. Program in Microbial Genomics, National ChungHsing University and Academia Sinica, Taipei, Taiwan, **3** Department of Plant Pathology, National ChungHsing University, Taichung, Taiwan, **4** Institute of Biotechnology, National ChungHsing University, Taichung, Taiwan, **5** NCHU-UCD Plant and Food Biotechnology Center, National ChungHsing University, Taichung, Taiwan, **6** Agricultural Biotechnology Center, National ChungHsing University, Taichung, Taiwan

* jyang@nchu.edu.tw



OPEN ACCESS

Citation: Tan CM, Li M-Y, Yang P-Y, Chang SH, Ho Y-P, Lin H, et al. (2015) *Arabidopsis* HFR1 Is a Potential Nuclear Substrate Regulated by the *Xanthomonas* Type III Effector XopD_{Xcc8004}. PLoS ONE 10(2): e0117067. doi:10.1371/journal.pone.0117067

Academic Editor: Ya-Wen He, Shanghai Jiao Tong University, CHINA

Received: September 11, 2014

Accepted: December 18, 2014

Published: February 3, 2015

Copyright: © 2015 Tan et al. This is an open access article distributed under the terms of the [Creative Commons Attribution License](https://creativecommons.org/licenses/by/4.0/), which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Data Availability Statement: All relevant data are within the paper and its Supporting Information files.

Funding: This work was supported by the Ministry of Science and Technology (NSC 100-2321-B-005-007-MY3, NSC 103-2911-I-005-301, MOST 103-2321-B-005-020) and Ministry of Education (ATU plan), Taiwan. The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

Competing Interests: The authors have declared that no competing interests exist.

Abstract

XopD_{Xcc8004}, a type III effector of *Xanthomonas campestris* pv. *campestris* (Xcc) 8004, is considered a shorter version of the XopD, which lacks the N-terminal domain. To understand the functions of XopD_{Xcc8004} in *planta*, a transgenic approach combined with inducible promoter to analyze the effects of XopD_{Xcc8004} in *Arabidopsis* was done. Here, the expression of XopD_{Xcc8004} in *Arabidopsis* elicited the accumulation of host defense-response genes. These molecular changes were dependent on salicylic acid and correlated with lesion-mimic phenotypes observed in *XVE::XopD_{Xcc8004}* transgenic plants. Moreover, XopD_{Xcc8004} was able to desumoylate HFR1, a basic helix-loop-helix transcription factor involved in photomorphogenesis, through SUMO protease activity. Interestingly, the *hfr1-201* mutant increased the expression of host defense-response genes and displayed a resistance phenotype to Xcc8004. These data suggest that HFR1 is involved in plant innate immunity and is potentially regulated by XopD_{Xcc8004}.

Introduction

Post-translational modifications enable plants to rapidly modify the stability and activity of key factors that play fundamental roles in defense signaling during bacterial infections [1], [2]. Studies have shown that sumoylation, a reversible and dynamic process, appears to be essential for suppressing defense responses in non-infected plants [3]. The sumoylation machinery is composed of small ubiquitin-like modifier (SUMO)-specific E1 (SAE1/SAE2 heterodimer), E2 (SCE1), and E3 (SIZ1 and MMS21/HPY2) to make SUMO conjugates [4], [5]. Elevated accumulation of salicylic acid (SA) accompanied by localized programmed cell death in *sum1sum2* and *siz1* mutants exhibits increased resistance to *Pseudomonas syringae* and suggests that

sumoylation machinery is likely to contribute to systemic-acquired resistance (SAR), resulting in enhanced resistance against further pathogen attacks [6–8].

The plant immune system is a multilayered type of immune response, which contains pathogen-associated molecular pattern-triggered immunity and effector-triggered immunity [9], [10]. To overcome the complex immune system, pathogens secrete or inject a range of effectors into host cells to manipulate host cellular functions and alter host defense responses [11], [12]. Although the functions of these virulence factors remain largely unknown, an increasing body of evidence demonstrates that pathogens employ a strategy to structurally or functionally mimic host cellular activities [13], [14]. In the past years, several bacterial effectors have been found to share structural similarity with SUMO proteases. Because bacteria do not have a SUMO system, it would be interesting to understand the role of pathogen effectors employing SUMO protease activity.

Previous studies have shown that the *Xanthomonas* type III effector XopD possesses desumoylation activity and localizes to nuclear foci in plant cells [15–17]. The subnuclear localization of XopD suggests that XopD may target SUMO-conjugated proteins in the plant nucleus. Indeed, XopD_{XccB100} from the strain B100 of *Xcc* specifically interacts with MYB30 to suppress its activity in activating plant defense responses required for anti-*Xcc* immunity [16]; XopD_{Xcv85–10} from the strain 85–10 of *Xanthomonas campestris* pv. *vesicatoria* (*Xcv*) specifically interacts with SIERF4 to suppress its activity in activating ethylene-induced responses, which is required for anti-*Xcv* immunity [18]. XopD is composed of an N-terminal domain, ERF-associated amphiphilic repression motifs, and a C-terminal SUMO protease domain [17], [19]. Although the C-terminal domain of XopD has SUMO peptidase and isopeptidase activities, lacking the functional N-terminal domain fails to suppress MYB30-mediated defense responses or desumoylation of SIERF4 [16], [18]. Thus, the N-terminus of XopD is essential for the virulence of *Xanthomonas in planta*. However, in the *Xcc8004* strain, XopD_{Xcc8004} was considered as a XopD without N-terminal domain [17], [19]. Although XopD_{Xcc8004} has been shown to be able to be secreted via a *hrp*-dependent pathway and contains functional SUMO protease activity, the action of XopD_{Xcc8004} *in planta* is still largely unknown [19].

Recently, light has been considered as an important regulator in modulating plant immunity [20], [21]. The availability and quality of light affects the plant development, as well as influences the plant defense responses. For example, a high ratio of red to far-red light enhances plant resistance to herbivorous insects [22]; a low ratio of red to far-red light reduces plant resistance to bacterial pathogens [23], [24]. Thus, mutations in the photoreceptors greatly influence plant defense responses. In this study, an inducible expression system was used to study the functions of XopD_{Xcc8004} in transgenic *Arabidopsis* plants. Finally, we showed that HFR1, a basic helix-loop-helix transcription factor involved in light-signaling pathway, is a potential nuclear substrate regulated by XopD_{Xcc8004}.

Materials and Methods

Plant materials and growth conditions

Arabidopsis thaliana was grown at 21°C under a 16-h light/8-h dark photoperiod for *Agrobacterium* transformations, and a 12-h light/12-h dark photoperiod for *Xanthomonas* spp. inoculations. *N. benthamiana* was grown at 26°C under a 16-h light/8-h dark photoperiod for *Agrobacterium* transient expression assay. The *Arabidopsis* WT, *hfr1–201* mutant, and *nahG* transgenic plants are in the Columbia ecotype background [6], [25].

Plasmid constructions

Xcc8004 genomic DNA was used for amplification of the XC_1213 DNA fragment encoding XopD_{Xcc8004}. An *A. thaliana* cDNA library was used for the amplification of the At1g02340

DNA fragment encoding HFR1. DNA fragments amplified by PCR using AccuPrime pfx DNA polymerase (Invitrogen) were subcloned into appropriate vectors by restriction site reconstructions. For the generation of *Arabidopsis* transgenic plants, PCR products were subcloned into the pER8 vector under the control of the XVE promoter [26]. For subcellular localization assays, PCR products were subcloned into pBA-YFP or pBA-CFP vectors under the control of the *Cauliflower mosaic virus* 35S promoter [27]. For yeast two-hybrid assays, PCR products were subcloned into pGADT7 and pGBKT7 vectors (Clontech) to generate AD-HFR1 and BK-XopD_{Xcc8004} constructs. For the expression of recombinant proteins, PCR products were subcloned into pET-SUMO (Invitrogen), pMAL-c2 (New England Biolabs), or pGEX4T-1 (GE Healthcare) vectors to produce N-terminal His-SUMO-, MBP-, or GST-tagged XopD_{Xcc8004}, HFR1, or AS2 proteins. The K72A mutant of HFR1 and the C355A mutant of XopD_{Xcc8004} were generated by QuikChange site-directed mutagenesis (Stratagene) according to the manufacturer's instructions. For homologous recombination, the 985-bp upstream and 976-bp downstream regions of XopD_{Xcc8004} were amplified from Xcc8004 genomic DNA and subcloned into a pK18mobsacB vector. To establish an *in vitro* sumoylation system, DNA fragments encoding *Arabidopsis* SAE1 (SAE1b), SAE2, and SCE1 were excised from the pACYCDuet-AtSAE1b-AtSAE2 and pCDFDuet-AtSUMO1(GG)-AtSCE1 plasmids [28], and subcloned into pET28a or pET29a vectors (Invitrogen) by restriction site reconstructions to produce His-tagged SAE1, SAE2, and SCE1 proteins. All plasmids were verified by DNA sequencing.

Arabidopsis transformations

To obtain *Arabidopsis* transgenic plants, plasmids were introduced into the *Agrobacterium tumefaciens* strain ABI by the freeze-thaw method [29] and then transformed into *A. thaliana* Col-0 using the floral-dip method [30]. *Arabidopsis* seeds were grown on half-strength Murashige and Skoog (1/2× MS) medium containing hygromycin (12.5 μg mL⁻¹) and carbenicillin (100 μg mL⁻¹) to obtain transgenic lines. Homozygous seeds were further selected and amplified for analyses.

Trypan blue staining

To characterize the lesion-mimic phenotype, *Arabidopsis* transgenic plants expressing XopD_{Xcc8004} were examined by trypan blue staining [31]. Briefly, transgenic seeds were germinated on 1/2× MS medium containing DMSO or 20 μM β-estradiol. Three-week-old seedlings were stained by boiling in a solution containing 10 mL of lactic acid, 10 mL of glycerol, 10 g of phenol, and 10 mg of trypan blue (dissolved in 10 mL of distilled water), and further destained in 2.5 g mL⁻¹ chloral hydrate solution. Images were collected with a Leica ZM75 microscope.

qRT-PCR

To measure *Arabidopsis* gene expression levels, total RNA was extracted by using the Trizol reagent and reverse transcribed into cDNAs using a Superscript III first-strand synthesis supermix according to the manufacturer's instructions (Invitrogen). The qRT-PCR reactions were performed on an Eco real-time PCR system (Illumina) with the KAPA SYBR fast qPCR kit (Kapa Biosystems). Relative amounts of transcripts were normalized to the transcript level of a house keeping gene, *EF1α*. Experiments were repeated at least 3 times.

Recombinant protein purifications and antibody production

To produce recombinant proteins, all constructs were transformed into *Escherichia coli* BL21 (DE3) cells and cultured at 24°C until the optical density at 600 nm reached 0.4. Then,

isopropyl β -D-1-thiogalactopyranoside was added to a final concentration of 0.2 mM and cells were further incubated overnight. After cell lysis, bacterial cell extracts were purified using appropriate resins according to the manufacturer's instruction. For *in vitro* sumoylation assays, *Arabidopsis* SAE1, SAE2, SCE1, and AtSUMO1 (with Gly-Gly at the C-terminus) proteins were purified using Ni²⁺-NTA resin (Qiagen). For *in vitro* pull-down assays, MBP- and GST-tagged proteins were purified using an amylose resin (New England Biolabs) and glutathione-Sepharose 4B (GE Healthcare), respectively. To generate a specific antibody against XopD_{Xcc8004}, the His-SUMO-XopD_{Xcc8004} protein was purified using a Ni²⁺-NTA resin and cleaved with Ubl-specific protease 1 (Ulp1) to remove the His-SUMO tag. After cleavage, proteins were purified by using a Sephacryl S-200 HR gel filtration column (GE Healthcare) to obtain the XopD_{Xcc8004} protein alone. Finally, a rabbit polyclonal antibody raised against XopD_{Xcc8004} was obtained by affinity purification using a polyvinylidene difluoride membrane as a coupling matrix [32].

Bacterial strains and inoculations

The Xcc8004 Δ XopD mutant strain was obtained using the sacB system [33]. Plasmid for homologous recombination was introduced into Xcc8004, and deletion mutant was verified by PCR. For bacterial inoculations, Xcc8004 spp. were cultured in nutrient broth supplemented with yeast extract (3 g of beef extract, 5 g of peptone, and 3 g of yeast extract in 1 liter of water) at 28°C. Four- to five-week-old *Arabidopsis* plants were used for bacterial growth assays. For XVE::XopD_{Xcc8004} transgenic plants, leaves were infiltrated with a bacterial suspension (2×10^6 CFU mL⁻¹) by using a syringe at 24 h after plants had been sprayed with 20 μ M β -estradiol. After inoculation, plants were kept at 21°C in a growth chamber, and bacterial populations in leaves were determined at indicated time intervals using agar plates of nutrient broth supplemented with yeast extract containing rifampin (50 μ g mL⁻¹). Experiments were repeated at least 3 times.

Yeast two-hybrid assays

AD-HFR1 and BK-XopD_{Xcc8004} constructs were transformed into the yeast strain AH109 by using the lithium acetate/single-stranded carrier DNA/polyethylene glycol method [34]. First, yeast cells were grown on synthetic-defined minimal yeast medium lacking leucine and tryptophan (Clontech) to maintain plasmids. Transformed colonies were further plated on synthetic-defined minimal yeast medium lacking leucine, tryptophan, and histidine (Clontech) to test the interaction between XopD_{Xcc8004} and HFR1.

Subcellular localization assays

To examine the subcellular localization of HFR1-CFP and XopD_{Xcc8004}-YFP, agroinfiltration was performed [35]. Briefly, *A. tumefaciens* strains carrying the 35S::HFR1-CFP or 35S::XopD_{Xcc8004}-YFP plasmid were inoculated into *N. benthamiana* leaves. Fluorescence signals were observed by confocal laser scanning microscopy at 36 h after agroinfiltration, and images were collected with the Olympus Fluoview FV1000 system.

In vitro pull-down assays

GST pull-down experiments were performed by inoculating 2 μ g of GST alone or GST-tagged proteins with 2 μ g of MBP alone or MBP-tagged proteins in binding buffer (50 mM Tris-HCl at pH 7.5, 100 mM NaCl, 0.25% Triton X-100, 35 mM β -mercaptoethanol) for 2 h at room temperature. Next, 25 μ L of glutathione-Sepharose 4B (GE Healthcare) were added, and

samples were further incubated for 1 h at room temperature. After extensive wash, pulled down proteins were eluted with 2.5× sample buffer and separated on a 10% sodium dodecyl sulfate-polyacrylamide gel. Western blotting was performed using an anti-MBP antibody to detect MBP-tagged proteins (Amersham). The chemiluminescence signals generated by the ECL reagent were further examined with the ImageQuant LAS4000 mini (GE Healthcare).

In vitro sumoylation assays

In vitro sumoylation was performed using purified recombinant proteins in a reaction buffer (50 mM Tris-HCl, pH7.4, 100 mM NaCl, 4 mM ATP, 10 mM MgCl₂, 4 mM DTT) in a total volume of 30 μL. After incubation for 2 h at 30°C, the reaction mixtures were separated on 10% sodium dodecyl sulfate-polyacrylamide gels. MBP-HFR1 and sumoylated MBP-HFR1 were detected by western blotting using an anti-MBP antibody. The chemiluminescence signals generated by the ECL reagent were further examined with the ImageQuant LAS4000 mini (GE Healthcare).

RNA-Seq analysis

To identify differentially expressed genes in *XVE::XopD_{Xcc8004}* transgenic plants upon β-estradiol treatment, next-generation sequencing was done on the HiSeq 2000 (Illumina) using total RNA samples extracted with the RNeasy plant mini kit (Qiagen). For transcriptome analysis, sequence reads were aligned using CLC bio and gene expression levels were normalized as reads per kilobase of exon model per million mapped reads. Finally, the differentially expressed genes were identified by DEseq [36].

Primers

Primer sequences for plasmid constructions and qRT-PCR analyses are listed in [S1 Table](#).

Results

Expression of XopD_{Xcc8004} elicits a SA-mediated defense response in *Arabidopsis*

XopD_{Xcc8004} from the *Xcc* strain 8004 is a shorter version of XopD, which lacks the N-terminal domain (Fig. 1A). To characterize the functions of XopD_{Xcc8004} in plant cells, *Arabidopsis* transgenic plants carrying an inducible *XVE::XopD_{Xcc8004}* transgene were generated. Here, a lesion-mimic phenotype associated with localized, necrotic spots was observed in *XVE::XopD_{Xcc8004}* transgenic plants after β-estradiol (inducer) treatment (Fig. 1B, C). Further examination using trypan blue staining confirmed that lesion-mimics were formed because of cell death (Figs. 1B, S1), whereas transgenic plants harboring empty vector (*XVE*) did not show cell death phenotype upon β-estradiol treatment (S2 Fig.).

To examine whether XopD_{Xcc8004} can trigger a SA-mediated defense response which may contribute to the hypersensitive cell death, genes involved in the SA-mediated defense-signaling network [37] were selected for quantitative reverse transcription-polymerase chain reaction (qRT-PCR) validation. Compared with dimethyl sulfoxide (DMSO) control treatment, genes involved in defense response (*AIG1* and *DOX1*), SA biosynthesis (*EDS5* and *SID2*), SA accumulation and downstream signaling (*WIN3*, *PAD4* and *WRKY70*), as well as SA-standard marker genes (*PR1*, *PR2* and *PR5*) were highly elevated in *XVE::XopD_{Xcc8004}* transgenic plants after β-estradiol treatment (Figs. 1D, S1). By contrast, no significant difference was observed in *XVE* (vector only) transgenic plants (S2 Fig.). Next, we transfected *XVE::XopD_{Xcc8004}* into an *Arabidopsis* transgenic line overexpressing bacterial salicylate hydroxylase (*nahG*). With the

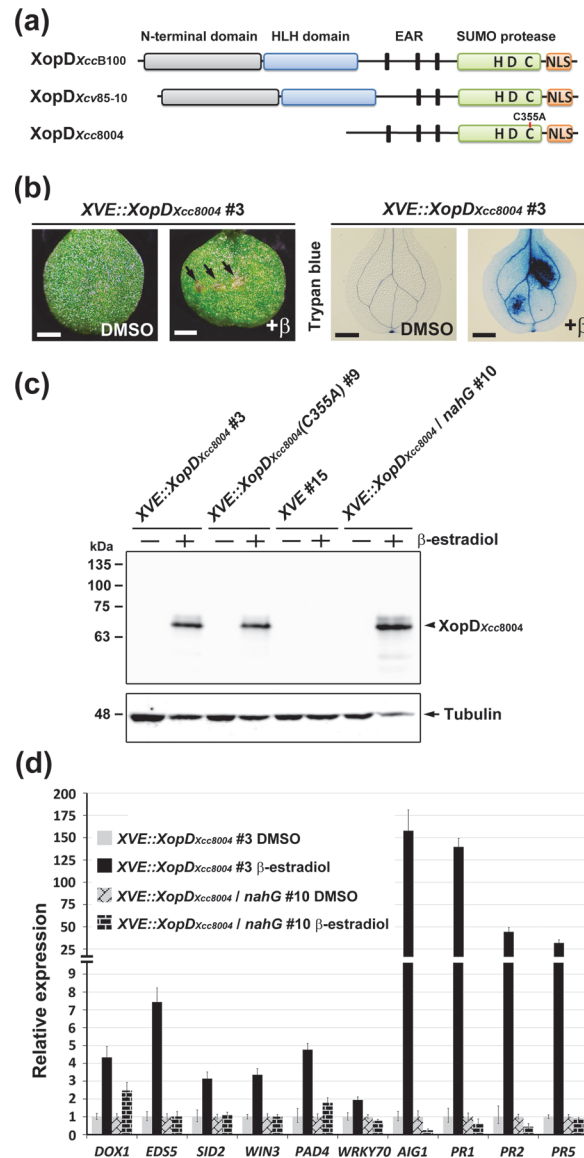


Figure 1. Salicylic acid-dependent defense responses were elicited by the expression of XopD_{Xcc8004} in *Arabidopsis*. (a) Schematic representation of XopD proteins. (b) Morphological examination and trypan blue staining of two-week-old leaves of *Arabidopsis* XVE::XopD_{Xcc8004} transgenic plants. Lesion-mimic phenotypes were indicated by arrows. Scale bar: 1 mm. (c) Translated products of XopD_{Xcc8004} and XopD_{Xcc8004}(C355A) were examined by western blotting using a specific antibody against XopD_{Xcc8004}. Anti-tubulin was used for loading control. (d) The expression levels of genes involved in the SA-mediated defense signaling network were examined by qRT-PCR and normalized to *EF1α*. The relative expression levels of each gene in the DMSO control were set at 1.

doi:10.1371/journal.pone.0117067.g001

expression of *nahG*, the cell death phenotype and SA response-related genes induced by XopD_{Xcc8004} were all inhibited by *nahG* (Figs. 1D, S3). These results suggest that XopD_{Xcc8004} elicits a SA-dependent defense response which may contribute to the lesion-mimic phenotype observed in XVE::XopD_{Xcc8004} transgenic plants.

To investigate the genome-wide expression profile of mRNAs in response to the expression of XopD_{Xcc8004} in *Arabidopsis*, the total RNA from XVE::XopD_{Xcc8004} transgenic seedlings upon DMSO and β-estradiol treatments were extracted for a comparative transcriptome

Table 1. Annotation of the differentially expressed genes ($p < 0.001$) involved in plant defense responses.

Name	AGI number	Base mean		log ₂ Fold Change	Functional annotations
		β-estradiol	DMSO		
SBT3.3	At1g32960	2.19	434.53	7.63	Subtilase family protein
DIR5	At1g64160	2.19	354.19	7.34	Disease resistance-responsive family protein
AIG1	At1g33960	20.81	850.80	5.35	AvrRPT2-induced gene 1
PCC1	At3g22231	14.24	544.07	5.26	Pathogen and circadian controlled 1
TPS4	At1g61120	5.48	262.91	5.58	Terpene synthase 4
	At5g10760	25.20	670.96	4.74	Eukaryotic aspartyl protease family protein
GSTF3	At2g02930	16.43	406.23	4.63	Glutathione S-transferase F3
	At5g03350	39.44	746.73	4.24	Legume lectin family protein
GSTU4	At2g29460	23.00	461.91	4.33	Glutathione S-transferase tau 4
NIT2	At3g44300	261.81	3868.75	3.89	Nitrilase 2
RLP23	At2g32680	12.05	272.95	4.50	Receptor-like protein 23
ELI3	At4g37990	5.48	175.27	5.00	Elicitor-activated gene 3–2
	At1g13609	4.38	157.93	5.17	Defensin-like (DEFL) family protein
ANK	At5g54610	6.57	188.05	4.84	Ankyrin
CRK7	At4g23150	1.10	102.24	6.54	Cysteine-rich receptor-like protein kinase 7
PR2	At3g57260	47.10	709.30	3.91	Beta-1,3-glucanase 2
FMO1	At1g19250	18.62	340.50	4.19	Flavin-dependent monooxygenase 1
CRK4	At3g45860	8.76	189.88	4.44	Cysteine-rich receptor-like protein kinase 4
CYP71A13	At2g30770	15.34	262.91	4.10	Cytochrome P450, family 71, subfamily A, polypeptide 13
WAKL10	At1g79680	8.76	179.84	4.36	Wall-associated kinase-like 10
PDF1.4	At1g19610	27.39	355.11	3.70	Arabidopsis defensin-like protein
PNP-A	At2g18660	12.05	191.70	3.99	Plant natriuretic peptide A
RLP38	At3g23120	2.19	85.81	5.29	Receptor-like protein 38
	At5g24200	4.38	109.54	4.64	Alpha/beta-hydrolases superfamily protein
	At3g04220	9.86	163.40	4.05	Disease resistance protein (TIR-NBS-LRR) family

AGI, *Arabidopsis* Genome Initiative; Base mean, the number of reads divided by the size factor (normalization constant) of sample; Fold change, β-estradiol base mean/DMSO base mean.

doi:10.1371/journal.pone.0117067.t001

analysis. In summary, a total of 23.1 million reads mapped to the *Arabidopsis* genome were generated after quality trim and a total of 103 differentially expressed genes with $p < 0.001$ were identified using the DESeq method (S2 and S3 Tables). Among them, 85 genes were up-regulated and 18 genes were downregulated after β-estradiol treatment. Functional annotations on the 85 up-regulated genes revealed that a total of 25 genes associated with defense responses including the SA-mediated response were highly induced by XopD_{Xcc8004} (Table 1).

Suppression of Xcc8004 growth by XopD_{Xcc8004}

Because the lesion-mimic phenotype accompanied by the up-regulation of SA response-related genes in XopD_{Xcc8004}-transgenic plants resembles the hypersensitive response (HR) in pathogen infection, we hypothesize that the expression of XopD_{Xcc8004} in *Arabidopsis* may result in enhanced resistance against bacterial pathogens. In order to examine the effect of XopD_{Xcc8004} on the resistance of *Arabidopsis*, multiplication of the Xcc8004 strain on XVE::XopD_{Xcc8004} transgenic plants was tested after β-estradiol treatment. Compared with XVE transgenic plants, multiplication of the Xcc8004 strain was suppressed by the expression of XopD_{Xcc8004} in planta at 5 days post inoculation (dpi) (Fig. 2A).

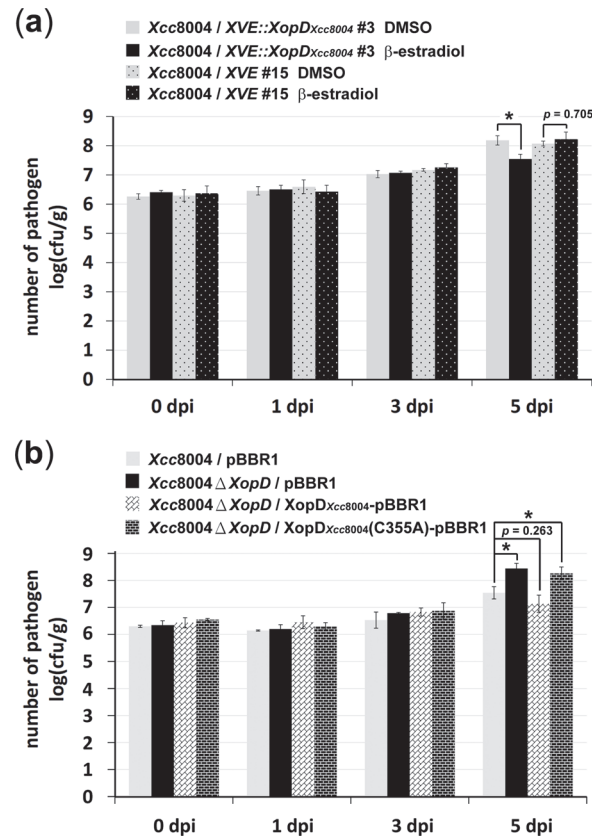


Figure 2. XopD_{Xcc8004} suppresses the virulence of the Xcc8004. (a) Bacterial growth of the Xcc8004 strain in *Arabidopsis* transgenic plants. DMSO or β-estradiol was applied 24 h before bacterial inoculation. Hand-infiltrated leaves were collected at the indicated times for measuring the *in planta* growth of bacterial populations. (b) Bacterial growth of Xcc8004 spp. in *Arabidopsis* plants were measured to examine the effects of XopD_{Xcc8004} on the virulence of Xcc8004. Statistically significant differences were determined using one-way ANOVA (* indicates $p < 0.05$).

doi:10.1371/journal.pone.0117067.g002

We further generated a *xopD* null mutant by homologous recombination to validate the function of XopD_{Xcc8004} in suppressing the virulence of Xcc8004. Here, *Arabidopsis* WT leaves inoculated with Xcc8004 Δ*XopD* strain exhibited a higher titer of bacteria at 5 dpi than those inoculated with the Xcc8004 strain (Fig. 2B). This phenotype was able to be complemented when Xcc8004 Δ*XopD* strain was transformed with a broad host plasmid (pBBR1) expressing XopD_{Xcc8004} (Fig. 2B). Taken together, these results suggest that XopD_{Xcc8004} acts as a negative factor in suppressing the growth of Xcc8004.

XopD_{Xcc8004}-eliciting defense responses are mainly dependent on the SUMO protease activity

To examine whether the SUMO protease activity of XopD_{Xcc8004} is required for eliciting the plant immunity, XVE::XopD_{Xcc8004}(C355A) transgenic plants were generated, in which mutation of the conserved cysteine (C355A) in XopD_{Xcc8004} was not able to hydrolyse the SUMO substrates. Compared with XVE::XopD_{Xcc8004} transgenic plants, no hypersensitive cell death was observed in XVE::XopD_{Xcc8004}(C355A) transgenic plants after β-estradiol treatment (Figs. 1B, 3A, S4). Further investigation of the gene expression involved in defense responses revealed that the C355A mutation largely suppressed the ability of XopD_{Xcc8004} in eliciting the

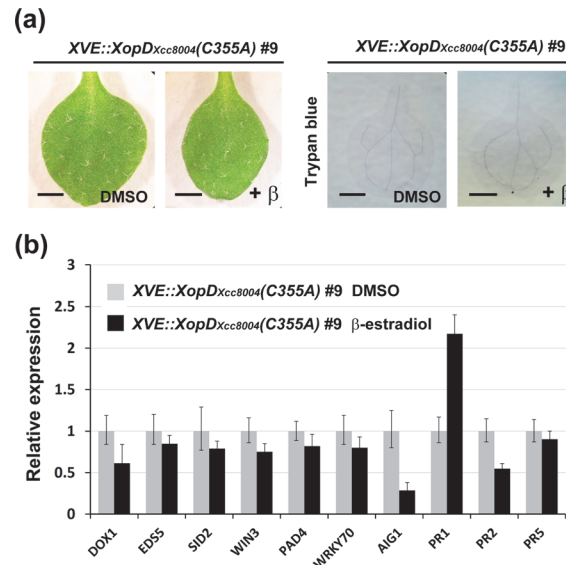


Figure 3. XopD_{Xcc8004}(C355A) loses the activity for activating plant immunity. (a) Morphological examination and trypan blue staining of two-week-old leaves of *Arabidopsis XVE::XopD_{Xcc8004}(C355A)* transgenic plants. Scale bar: 1 mm. (b) The expression levels of genes involved in the SA-mediated defense signaling network were examined by qRT-PCR and normalized to *EF1α*. The relative expression levels of each gene in the DMSO control were set at 1.

doi:10.1371/journal.pone.0117067.g003

expression of SA response-related genes in *Arabidopsis* (Figs. 3B, S4). Next, we transformed *Xcc8004 ΔXopD* strain with a broad host plasmid to express XopD_{Xcc8004}(C355A). However, *Arabidopsis* WT leaves inoculated with *Xcc8004 ΔXopD/XopD_{Xcc8004}(C355A)* strain still exhibited a higher titer of bacteria at 5 dpi as well as those inoculated with *Xcc8004 ΔXopD* strain (Fig. 2B). These results suggest that the SUMO protease activity of XopD_{Xcc8004} is required for eliciting a SA-mediated defense response in *Arabidopsis* and suppressing the virulence of *Xcc8004*.

XopD_{Xcc8004} interacts with *Arabidopsis* HFR1

Unexpectedly, a long hypocotyl phenotype associated with the lesion-mimic phenotype was observed with the expression of XopD_{Xcc8004} in *Arabidopsis* (Fig. 4A). Compared with *XVE* transgenic plants, the hypocotyl length of *XVE::XopD_{Xcc8004}* seedlings reached to 4.5 mm at 14 days after germination in medium containing β-estradiol (Figs. 4B, S5). We noted that the long hypocotyl phenotype elicited by the expression of XopD_{Xcc8004} similar to those observed in *siz1-2* carrying a partially functional *SIZ1^{phd}* [38]. *SIZ1^{phd}* contains a C134Y mutation in the zinc finger motif required for the SUMO E3 ligase activity of SIZ1. Cheong et al. found that expression of *SIZ1^{phd}* in the *siz1-2* mutant created a light-dependent long hypocotyl phenotype. Therefore, we investigated whether light signaling components were potential substrates of XopD_{Xcc8004}. Here, several key components involved in the light signaling pathway including HY5, LAF1, Fin219, PAT3, SPA1, and HFR1 were analyzed for their interaction with XopD_{Xcc8004} using the yeast two-hybrid assay. Among them, HFR1, a basic helix-loop-helix transcription factor, showed a positive interaction with XopD_{Xcc8004} in selection medium lacking tryptophan/leucine/histidine (Fig. 5A).

Next, we investigated whether XopD_{Xcc8004} can colocalize with HFR1 in plant cells. HFR1 has been shown to localize to nucleus in subnuclear foci [27], whereas XopD_{Xcc8004} was localized to the nucleus in a homogeneous pattern [17]. If XopD_{Xcc8004} can interact with HFR1, the

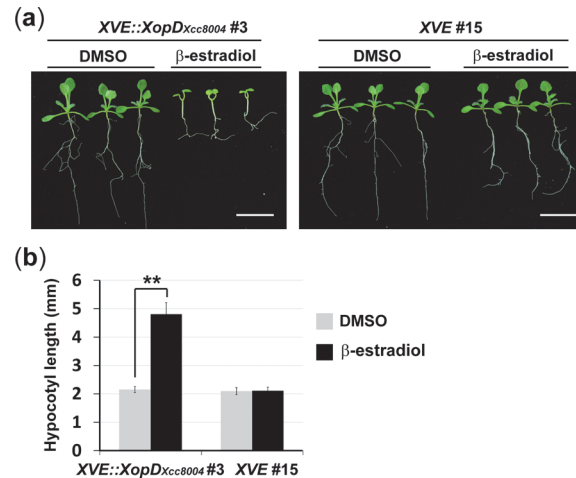


Figure 4. Expression of XopD_{Xcc8004} induces a long hypocotyl phenotype in *Arabidopsis*. (a) Phenotypes of 12-day-old *Arabidopsis* transgenic plants carrying a XopD_{Xcc8004} gene driven by the inducible XVE promoter. Scale bars: 8 mm. (b) Average hypocotyl lengths of seedlings grown on medium containing DMSO or 20 μM β-estradiol. Statistically significant differences were determined using one-way ANOVA (** indicates $p < 0.005$).

doi:10.1371/journal.pone.0117067.g004

coexpression of HFR1 may cause the relocalization of XopD_{Xcc8004} to HFR1-containing nuclear foci. Here, we coexpressed XopD_{Xcc8004}-yellow fluorescence protein (YFP) and HFR1-cyan fluorescent protein (CFP) in *Nicotiana benthamiana* cells using agroinfiltration and found that XopD_{Xcc8004}-YFP was colocalized with HFR1-CFP in nuclear foci (Fig. 5B). To further examine whether XopD_{Xcc8004} can directly interact with HFR1 *in vitro*, a pull-down assay was performed with purified recombinant proteins. Fig. 5C shows that maltose-binding protein (MBP)-HFR1 was specifically pulled down by glutathione S-transferase (GST)-XopD_{Xcc8004} but not by GST alone. By contrast, no signal was observed when the negative control proteins MBP and MBP-AS2 were used in the assay (Fig. 5C).

To examine whether the long hypocotyl phenotype observed in XVE::XopD_{Xcc8004} seedlings is dependent on the SUMO protease activity of XopD_{Xcc8004}, we investigated the phenotype of XVE::XopD_{Xcc8004}(C355A) transgenic plants. However, a long hypocotyl phenotype was also observed in XVE::XopD_{Xcc8004}(C355A) seedlings (S5 Fig.). This result suggests that the long hypocotyl phenotype caused by XopD_{Xcc8004} is not simply due to the SUMO protease activity.

K72 in HFR1 is desumoylated by XopD_{Xcc8004} *in vitro*

Because XopD_{Xcc8004} has been shown to possess SUMO protease activity [16], [17], the interaction between XopD_{Xcc8004} and HFR1 prompted us to examine whether HFR1 can be modified with SUMO and further desumoylated by XopD_{Xcc8004}. Here, the examination of the deduced amino acid sequence of HFR1 revealed a probable sumoylation site at lysine 72 (K72) in the consensus motif ΨKxE/D (where Ψ is a large and hydrophobic amino acid and x is any amino acid) (Fig. 6A). To address whether HFR1 can be modified with SUMO, an *in vitro* sumoylation assay was performed with *Arabidopsis* SAE1/SAE2 (SUMO-activating E1), SCE1 (SUMO-conjugating E2), AtSUMO1, and MBP-HFR1. Fig. 6B shows that a clear mobility shift of MBP-HFR1 was detected after incubation with the components of the *Arabidopsis* sumoylation cascade, and the shift was in a molecular mass consistent with mono-SUMO modification. Compared with MBP-HFR1, no mobility shift was detected for MBP-HFR1(K72A) in the *in vitro* sumoylation system (Fig. 6B). These results indicate that the K72 of HFR1 is the principal site

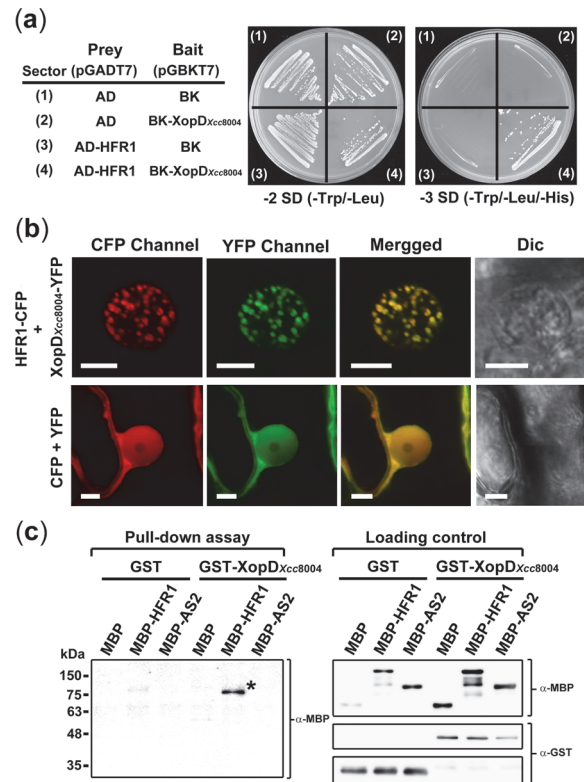


Figure 5. XopD_{Xcc8004} interacts with HFR1. (a) Investigation of the interaction between HFR1 and XopD_{Xcc8004} by yeast two-hybrid analysis. Yeast strains transformed with two plasmids (prey and bait) were plated onto synthetic-defined (SD) minimal medium lacking tryptophan/leucine or tryptophan/leucine/histidine. (b) *Nicotiana benthamiana* leaves were co-infiltrated with agro bacterium carrying 35S::XopD_{Xcc8004}-YFP and 35S::HFR1-CFP. Fluorescence and differential interference contrast (DIC) images were obtained by confocal laser scanning microscopy. 35S::YFP and 35S::CFP were used for control. Scale bars: 15 μm. (c) Investigation of the interaction between HFR1 and XopD_{Xcc8004} by using pull-down assays. Briefly, 2 μg of GST or GST fusion proteins were used to pull down 2 μg of MBP or MBP fusion proteins, and an anti-MBP antibody was used to detect the associated proteins by western blotting (the left panel). The right panels present the input proteins examined by anti-MBP or anti-GST antibodies. The asterisk indicates the protein signal of MBP-HFR1.

doi:10.1371/journal.pone.0117067.g005

for SUMO conjugation. Next, we examined whether the sumoylated HFR1 can be desumoylated by XopD_{Xcc8004}. Fig. 6C shows that no sumoylated MBP-HFR1 was detected when XopD_{Xcc8004}, but not the catalytic mutant XopD_{Xcc8004}(C355A), was present in the *in vitro* sumoylation reaction. These results indicate that XopD_{Xcc8004} catalyzes the SUMO hydrolysis from the K72 of HFR1.

hfr1-201 increases resistance to Xcc8004

Based on the interaction between HFR1 and XopD_{Xcc8004}, we propose that HFR1 may play a role in the plant immune response. To this end, we monitored the expression levels of genes involved in the SA-mediated defense-signaling network in *hfr1-201* mutants and WT plants. Irrespective of treatment with or without 2 mM SA, we found that the levels of *PR2*, *WRKY70*, *WIN3*, *EDS5*, *AIG1*, *PUB54*, *WRKY18*, *PR1*, and *WRKY51* transcripts are higher in the *hfr1-201* mutant than in WT plants (Fig. 7A). We further investigated the growth of Xcc8004 WT and ΔXopD mutant strains in the *hfr1-201* mutant. Compared with WT plants, multiplication of Xcc8004 and Xcc8004 ΔXopD strains was suppressed in the *hfr1-201* mutant (Fig. 7B).

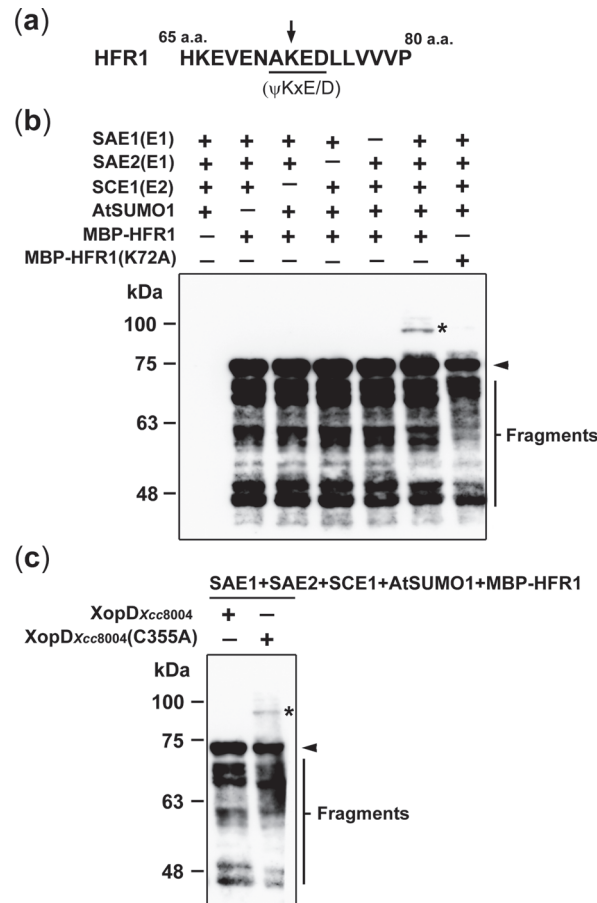


Figure 6. K72 in HFR1 is desumoylated by XopD_{Xcc8004} *in vitro*. (a) The deduced amino acid sequence of HFR1 from 65 to 80 a.a. The arrow indicates a putative sumoylation site at K72 of the consensus motif ΨKxE/D of HFR1. (b) An *in vitro* sumoylation system was established by using purified recombinant proteins, including *Arabidopsis* SAE1, SAE2, SCE1, and AtSUMO1. MBP-HFR1 or MBP-HFR1(K72A) were used as potential substrates for sumoylation and detected with an anti-MBP antibody. (c) Together with SAE1, SAE2, SCE1, AtSUMO1, and MBP-HFR1, XopD_{Xcc8004} was added to the reaction mixture to investigate the sumoylation of HFR1. XopD_{Xcc8004}(C355A), a SUMO protease mutant, was used as control. Asterisks indicate sumoylated MBP-HFR1 proteins. Arrowheads indicate unmodified MBP-HFR1 proteins. Signals below the unmodified MBP-HFR1 proteins were degraded products of purified MBP-HFR1 proteins.

doi:10.1371/journal.pone.0117067.g006

These results suggest that HFR1 is required for modulating the defense response in *Arabidopsis*, and the loss-of-function mutant in the *HFR1* increases resistance to *Xcc8004* spp.

Discussion

XopD is composed of an N-terminal domain, ERF-associated amphiphilic repression motifs, and a C-terminal SUMO protease domain. In XopD_{Xcv85-10} and XopD_{XccB100}, the N-terminal domain has been suggested to play a role in specifying substrate recognition and modulating SUMO protease activity [16], [18]. However, XopD_{Xcc8004} was considered as a shorter version of XopD that does not contain an N-terminal domain [17], [19]. Using a transgenic approach, we found that possessing a C-terminal SUMO protease domain, the XopD_{Xcc8004}-overexpressing transgenic plants showed morphological and molecular phenotypes similar to those of the mutants defective in the sumoylation machinery. In *Arabidopsis*, mutants defective in the sumoylation machinery, e.g., *siz1* and *sum1sum2* mutants, display localized programmed cell

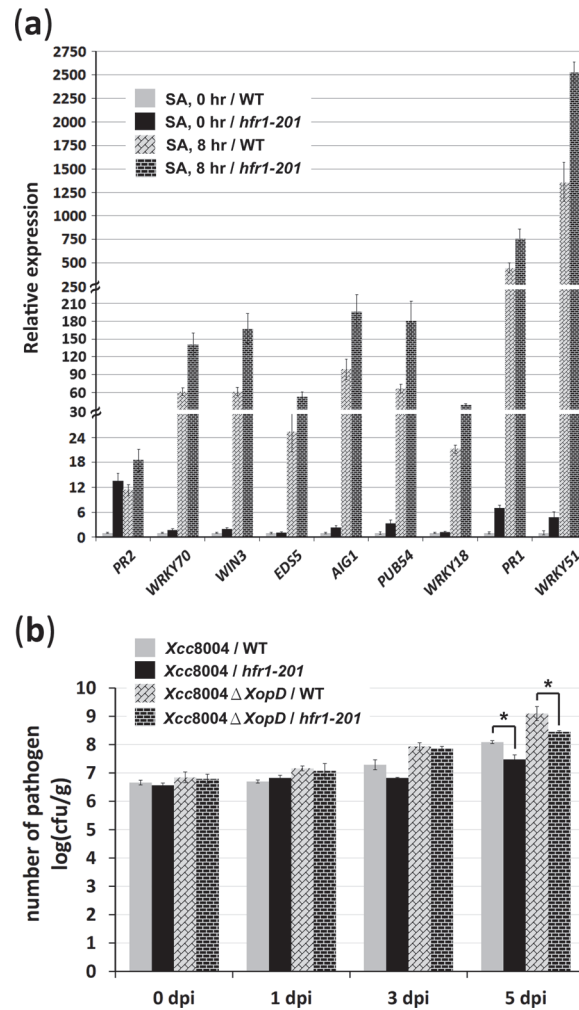


Figure 7. *hfr1-201* increases plant immunity against *Xcc8004* spp. (a) *Arabidopsis* WT and *hfr1-201* mutant plants treated with (8 hr) or without (0 hr) 2 mM SA were collected for total RNA extraction. The expression levels of genes involved in the SA-mediated defense signaling network were examined by qRT-PCR and normalized to *EF1α*. The relative expression levels of each gene in the WT plants without SA treatment were set at 1. (b) Bacterial growth in *Arabidopsis* WT and *hfr1-201* mutant plants were measured to examine the effects of HFR1 on the resistance of *Arabidopsis* against *Xcc8004* spp. Hand-infiltrated leaves were collected at the indicated times for measuring the *in planta* growth of bacterial populations. Statistically significant differences were determined using one-way ANOVA (* indicates $p < 0.05$).

doi:10.1371/journal.pone.0117067.g007

death and elevated accumulation of SA, which contribute to the enhanced resistance against bacterial attacks [6], [7]. In this study, we showed that the expression of XopD_{Xcc8004} in *Arabidopsis* elicited a lesion-mimic phenotype associated with increased expression of disease-response genes (Fig. 1B). Moreover, the multiplication of the *Xcc8004* strain was suppressed in XopD_{Xcc8004}-overexpressing transgenic plants (Fig. 2A), and *Xcc8004* Δ*XopD* displayed a higher bacterial growth rate than *Xcc8004* (Fig. 2B). Different from XopD_{Xcv} and XopD_{XccB100}, which contain a complete N-terminal domain and are successful in suppressing anti-*Xcv* and anti-*XccB100* immunities [16], [18], XopD_{Xcc8004} induced host defense responses to inhibit *Xcc8004* growth. Although it is still unknown how the N-terminal domain modulates the SUMO protease activity of XopD, it is clear that impairment in the N-terminal domain of XopD not only loses the ability to suppress host immunity, but also elicits host defense responses.

Based on the results that the catalytic mutant XopD_{Xcc8004}(C355A) was not able to elicit HR and plant immunity (Fig. 3A, B), we suggest that the activity of XopD_{Xcc8004} in eliciting host defense responses is mainly dependent on the SUMO protease activity. Yet, the substrates of XopD_{Xcc8004} functioning in plant defense responses remain largely unknown. Nevertheless, we identified that HFR1 was a potential nuclear substrate of XopD_{Xcc8004} and could be modified by sumoylation (Figs. 5, 6). With the *in vitro* assay, we found that the sumoylated-residue K72 within the consensus sumoylation motif of HFR1 was desumoylated by XopD_{Xcc8004} through the SUMO protease activity (Fig. 6B, C). Although the direct impact of SUMO protease activity of XopD_{Xcc8004} on the function of HFR1 remains unknown, we observed that the loss-of-function mutation in *HFR1* gene accelerated SA-mediated responses and increased the resistance against *Xcc8004* (Fig. 7A, B). These results clearly indicate that HFR1 plays a role in repression of defense responses in plants.

HFR1 is an important regulator involved in photomorphogenesis and regulated by ubiquitin-mediated degradation through the coat protein complex I [27], [39], [40]. Because the highly unstable character of HFR1 [41], it is a technical challenge to provide a tight connection between XopD_{Xcc8004} and HFR1 during infection. Thus, further experiments are required to demonstrate the function of sumoylation on the manipulation of the activity of HFR1. Sumoylation has been shown to play a role in mediating the transcriptional repression activity of many transcription factors [42–46]. Therefore, we do not exclude the possibility that the repression activity of HFR1 on defense responses requires a modification of SUMO. In fact, our data only explain a possible mechanism for XopD_{Xcc8004}-triggered host defense responses, and we do not exclude the possibility that XopD_{Xcc8004} may have several targets in plants. Thus, we still observed a higher growth rate of *Xcc8004* even in the *hfr1-201* mutant (Fig. 7B).

Recently, Tan et al. (2014) reported that XopD_{Xcc8004} triggers plant disease tolerance by targeting DELLA proteins [47]. In their study, they did not observe the significant difference in bacterial titers between *Xcc8004* and *Xcc8004* Δ XopD. This difference may be depending on the different experimental conditions used in our study. Here, a syringe infiltration method and a higher titer of bacterial suspension (2×10^6 CFU mL⁻¹) were used. Nevertheless, our findings are not in conflict with the fact that XopD_{Xcc8004} may act as a virulence-controlling factor by interfering with plant defense responses.

Recent studies on plant-pathogen interaction have provided new insights into fundamental cellular processes in plants [48], [49]. In this study, we identified a dual role of HFR1 in development and immunity. This finding indicates that HFR1 is required for the fine-tuning of the immune response, as well as contributes to our knowledge of the crosstalk between the light-signaling pathway and immune response.

Supporting Information

S1 Fig. Salicylic acid-dependent defense responses were elicited by the expression of XopD_{Xcc8004} in *Arabidopsis*. (a) Trypan blue staining of two-week-old leaves of *Arabidopsis* XVE::XopD_{Xcc8004} transgenic plants. Scale bar: 1 mm. (b) Translated products of XopD_{Xcc8004} were examined by western blotting using a specific antibody against XopD_{Xcc8004} and indicated by an arrow. Rubisco large subunit (RBCL) stained with coomassie brilliant blue served as a loading control. (c) The expression levels of genes involved in the SA-mediated defense signaling network were examined by qRT-PCR and normalized to *EF1 α* . The relative expression levels of each gene in the DMSO control were set at 1. (TIF)

S2 Fig. Transgenic plants harboring empty vector (XVE) did not show cell death phenotype upon β -estradiol treatment and elicited defense responses. (a) Trypan blue staining of two-

week-old leaves of *Arabidopsis XVE* transgenic plants. Scale bar: 1 mm. (b) Expression levels of genes involved in the SA-mediated defense signaling network were examined by qRT-PCR and normalized to *EF1α*. The relative expression levels of each gene in the DMSO control were set at 1. (TIF)

S3 Fig. Salicylic acid-dependent defense responses elicited by the expression of XopD_{Xcc8004} were inhibited by *nahG*. (a) Trypan blue staining of two-week-old leaves of *Arabidopsis XVE::XopD_{Xcc8004} / nahG* transgenic plants. Scale bar: 1 mm. (b) Translated products of *XopD_{Xcc8004}* were examined by western blotting using a specific antibody against XopD_{Xcc8004} and indicated by an arrow. Rubisco large subunit (RBCL) stained with coomassie brilliant blue served as a loading control. (c) The expression levels of genes involved in the SA-mediated defense signaling network were examined by qRT-PCR and normalized to *EF1α*. The relative expression levels of each gene in the DMSO control were set at 1. (TIF)

S4 Fig. XopD_{Xcc8004}(C355A) loses the activity for activating plant immunity. (a) Morphological examination and trypan blue staining of two-week-old leaves of *Arabidopsis XVE::XopD_{Xcc8004}(C355A)* transgenic plants. Scale bar: 1 mm. (b) Translated products of *XopD_{Xcc8004}(C355A)* were examined by western blotting using a specific antibody against XopD_{Xcc8004} and indicated by an arrow. Rubisco large subunit (RBCL) stained with coomassie brilliant blue served as a loading control. (c) The expression levels of genes involved in the SA-mediated defense signaling network were examined by qRT-PCR and normalized to *EF1α*. The relative expression levels of each gene in the DMSO control were set at 1. (TIF)

S5 Fig. Expression of XopD_{Xcc8004} induces a long hypocotyl phenotype in *Arabidopsis*. (a, b) Average hypocotyl lengths of seedlings grown on medium containing DMSO (grey bars) and 20 μM β-estradiol (black bars). Statistically significant differences were determined using one-way ANOVA (** indicates $p < 0.005$). (TIF)

S1 Table. Primer sequences for plasmid constructions and qRT-PCR.

(XLS)

S2 Table. Reads counting (mapped read sequence statistic) for Illumina sequencing data.

(XLS)

S3 Table. Differentially expressed genes with $p < 0.001$. Identification genes that were differentially expressed in *Arabidopsis* after XopD_{Xcc8004} were induced by β-estradiol.

(XLS)

Acknowledgments

We thank M. Dow (National University of Ireland, Cork, Ireland) for the *X. campestris* pv. *campestris* 8004 strain; K. Tanaka (Kwansei Gakuin University, Sanda, Japan) for the *in vivo* sumoylation system; K. Miura (University of Tsukuba, Tsukuba, Japan) and D.-J. Yun (Gyeong-sang National University, Jinju, Korea) for the *Arabidopsis nahG* transgenic line.

Author Contributions

Conceived and designed the experiments: CMT JYY. Performed the experiments: CMT MYL PYY SHC YPH HL JYY. Analyzed the data: CMT JYY. Contributed reagents/materials/analysis tools: WLD. Wrote the paper: JYY.

References

1. Stulemeijer IJ, Joosten MH (2008) Post-translational modification of host proteins in pathogen-triggered defence signalling in plants. *Mol Plant Pathol* 9: 545–560. doi: [10.1111/j.1364-3703.2008.00468.x](https://doi.org/10.1111/j.1364-3703.2008.00468.x) PMID: [18705867](https://pubmed.ncbi.nlm.nih.gov/18705867/)
2. Ribet D, Cossart P (2010) Post-translational modifications in host cells during bacterial infection. *FEBS Lett* 584: 2748–2758. doi: [10.1016/j.febslet.2010.05.012](https://doi.org/10.1016/j.febslet.2010.05.012) PMID: [20493189](https://pubmed.ncbi.nlm.nih.gov/20493189/)
3. van den Burg HA, Takken FL (2010) SUMO-, MAPK-, and resistance protein-signaling converge at transcription complexes that regulate plant innate immunity. *Plant Signal Behav* 5: 1597–1601. PMID: [21150289](https://pubmed.ncbi.nlm.nih.gov/21150289/)
4. Colby T, Matthäi A, Boeckelmann A, Stuible HP (2006) SUMO-conjugating and SUMO-deconjugating enzymes from *Arabidopsis*. *Plant Physiol* 142: 318–332. PMID: [16920872](https://pubmed.ncbi.nlm.nih.gov/16920872/)
5. Park H, Kim WY, Park H, Lee S, Bohnert H, et al. (2011) SUMO and SUMOylation in plants. *Mol Cells* 32: 305–316. doi: [10.1007/s10059-011-0122-7](https://doi.org/10.1007/s10059-011-0122-7) PMID: [21912873](https://pubmed.ncbi.nlm.nih.gov/21912873/)
6. Lee J, Nam J, Park HC, Na G, Miura K, et al. (2007) Salicylic acid-mediated innate immunity in *Arabidopsis* is regulated by SIZ1 SUMO E3 ligase. *Plant J* 49: 79–90. PMID: [17163880](https://pubmed.ncbi.nlm.nih.gov/17163880/)
7. van den Burg HA, Kini RK, Schuurink RC, Takken FL (2010) *Arabidopsis* small ubiquitin-like modifier paralogs have distinct functions in development and defense. *Plant Cell* 22: 1998–2016. doi: [10.1105/tpc.109.070961](https://doi.org/10.1105/tpc.109.070961) PMID: [20525853](https://pubmed.ncbi.nlm.nih.gov/20525853/)
8. Park BS, Song JT, Seo HS (2011) *Arabidopsis* nitrate reductase activity is stimulated by the E3 SUMO ligase AtSIZ1. *Nat Commun* 2: 400. doi: [10.1038/ncomms1408](https://doi.org/10.1038/ncomms1408) PMID: [21772271](https://pubmed.ncbi.nlm.nih.gov/21772271/)
9. Göhre V, Robatzek S (2008) Breaking the barriers: microbial effector molecules subvert plant immunity. *Annu Rev Phytopathol* 46: 189–215. doi: [10.1146/annurev.phyto.46.120407.110050](https://doi.org/10.1146/annurev.phyto.46.120407.110050) PMID: [18422429](https://pubmed.ncbi.nlm.nih.gov/18422429/)
10. Dodds PN, Rathjen JP (2010) Plant immunity: towards an integrated view of plant–pathogen interactions. *Nat Rev Genet* 11: 539–548. doi: [10.1038/nrg2812](https://doi.org/10.1038/nrg2812) PMID: [20585331](https://pubmed.ncbi.nlm.nih.gov/20585331/)
11. Grant SR, Fisher EJ, Chang JH, Mole BM, Dangl JL (2006) Subterfuge and manipulation: type III effector proteins of phytopathogenic bacteria. *Annu Rev Microbiol* 60: 425–449. PMID: [16753033](https://pubmed.ncbi.nlm.nih.gov/16753033/)
12. Lee AHY, Middleton MA, Guttman DS, Desveaux D (2013) Phytopathogen type III effectors as probes of biological systems. *Microb Biotechnol* 6: 230–240. doi: [10.1111/1751-7915.12042](https://doi.org/10.1111/1751-7915.12042) PMID: [23433088](https://pubmed.ncbi.nlm.nih.gov/23433088/)
13. Hotson A, Mudgett MB (2004) Cysteine proteases in phytopathogenic bacteria: identification of plant targets and activation of innate immunity. *Curr Opin Plant Biol* 7: 384–390. PMID: [15231260](https://pubmed.ncbi.nlm.nih.gov/15231260/)
14. Desveaux D, Singer AU, Dangl JL (2006) Type III effector proteins: doppelgangers of bacterial virulence. *Curr Opin Plant Biol* 9: 376–382. PMID: [16713730](https://pubmed.ncbi.nlm.nih.gov/16713730/)
15. Hotson A, Chosed R, Shu H, Orth K, Mudgett MB (2003) *Xanthomonas* type III effector XopD targets SUMO-conjugated proteins *in planta*. *Mol Microbiol* 50: 377–389. PMID: [14617166](https://pubmed.ncbi.nlm.nih.gov/14617166/)
16. Canonne J, Marino D, Jauneau A, Pouzet C, Brière C, et al. (2011) The *Xanthomonas* type III effector XopD targets the *Arabidopsis* transcription factor MYB30 to suppress plant defense. *Plant Cell* 23: 3498–3511. doi: [10.1105/tpc.111.088815](https://doi.org/10.1105/tpc.111.088815) PMID: [21917550](https://pubmed.ncbi.nlm.nih.gov/21917550/)
17. Kim JG, Taylor KW, Mudgett MB (2011) Comparative analysis of the XopD type III secretion (T3S) effector family in plant pathogenic bacteria. *Mol Plant Pathol* 12: 715–730. doi: [10.1111/j.1364-3703.2011.00706.x](https://doi.org/10.1111/j.1364-3703.2011.00706.x) PMID: [21726373](https://pubmed.ncbi.nlm.nih.gov/21726373/)
18. Kim JG, Stork W, Mudgett MB (2013) *Xanthomonas* type III effector XopD desumoylates tomato transcription factor SIERF4 to suppress ethylene responses and promote pathogen growth. *Cell Host Microbe* 13: 143–154. doi: [10.1016/j.chom.2013.01.006](https://doi.org/10.1016/j.chom.2013.01.006) PMID: [23414755](https://pubmed.ncbi.nlm.nih.gov/23414755/)
19. Canonne J, Marino D, Noël LD, Arechaga I, Pichereaux C, et al. (2010) Detection and functional characterization of a 215 amino acid N-terminal extension in the *Xanthomonas* type III effector XopD. *PLoS ONE* 5: e15773. doi: [10.1371/journal.pone.0015773](https://doi.org/10.1371/journal.pone.0015773) PMID: [21203472](https://pubmed.ncbi.nlm.nih.gov/21203472/)
20. Ballare CL, Mazza CA, Austin AT, Pierik R (2012) Canopy light and plant health. *Plant Physiol* 160: 145–155. doi: [10.1104/pp.112.200733](https://doi.org/10.1104/pp.112.200733) PMID: [22802612](https://pubmed.ncbi.nlm.nih.gov/22802612/)
21. Kangasjärvi S, Neukermans J, Li S, Aro EM, Noctor G (2012) Photosynthesis, photorespiration, and light signalling in defence responses. *J Exp Bot* 63: 1619–1636. doi: [10.1093/jxb/err402](https://doi.org/10.1093/jxb/err402) PMID: [22282535](https://pubmed.ncbi.nlm.nih.gov/22282535/)
22. Moreno JE, Tao Y, Chory J, Ballaré CL (2009) Ecological modulation of plant defense via phytochrome control of jasmonate sensitivity. *Proc Natl Acad Sci USA* 106: 4935–4940. doi: [10.1073/pnas.0900701106](https://doi.org/10.1073/pnas.0900701106) PMID: [19251652](https://pubmed.ncbi.nlm.nih.gov/19251652/)
23. Genoud T, Buchala AJ, Chua NH, Métraux JP (2002) Phytochrome signalling modulates the SA-perceptive pathway in *Arabidopsis*. *Plant J* 31: 87–95. PMID: [12100485](https://pubmed.ncbi.nlm.nih.gov/12100485/)

24. Faigón-Soverna A, Harmon FG, Storani L, Karayekov E, Staneloni RJ, et al. (2006) A constitutive shade-avoidance mutant implicates TIR-NBS-LRR proteins in *Arabidopsis* photomorphogenic development. *Plant Cell* 18: 2919–2928. PMID: [17114357](#)
25. Kim YM, Woo JC, Song PS, Soh MS (2002) HFR1, a phytochrome A-signalling component, acts in a separate pathway from HY5, downstream of COP1 in *Arabidopsis thaliana*. *Plant J* 30: 711–719. PMID: [12061902](#)
26. Zuo J, Niu QW, Chua NH (2000) An estrogen receptor-based transactivator XVE mediates highly inducible gene expression in transgenic plants. *Plant J* 24: 265–273. PMID: [11069700](#)
27. Jang IC, Yang JY, Seo HS, Chua NH (2005) HFR1 is targeted by COP1 E3 ligase for post-translational proteolysis during phytochrome A signaling. *Genes Dev* 19: 593–602. PMID: [15741320](#)
28. Okada S, Nagabuchi M, Takamura Y, Nakagawa T, Shinmyozu K, et al. (2009) Reconstitution of *Arabidopsis thaliana* SUMO pathways in *E. coli*: functional evaluation of SUMO machinery proteins and mapping of SUMOylation sites by mass spectrometry. *Plant Cell Physiol* 50: 1049–1061. doi: [10.1093/pcp/pcp056](#) PMID: [19376783](#)
29. Wise A, Liu Z, Binns A (2006) Three methods for the introduction of foreign DNA into *Agrobacterium*. *Methods Mol Biol* 343: 43–53. PMID: [16988332](#)
30. Clough SJ, Bent AF (1998) Floral dip: a simplified method for *Agrobacterium*-mediated transformation of *Arabidopsis thaliana*. *Plant J* 16: 735–743. PMID: [10069079](#)
31. Ho YP, Tan CM, Li MY, Lin H, Deng WL, et al. (2012) The AvrB_AvrC domain of AvrXccC of *Xanthomonas campestris* pv. *campestris* is required to elicit plant defense responses and manipulate ABA homeostasis. *Mol Plant-Microbe Interact* 26: 419–430.
32. Nishino H, Ishibashi T, Nakamura H (1994) Preparation method of highly-purified antibodies from antisera by using polyvinylidene difluoride membrane. Application to fatty acid binding protein in rat liver. *Biochem Mol Biol Int* 34: 409–417. PMID: [7849652](#)
33. Schäfer A, Tauch A, Jäger W, Kalinowski J, Thierbach G, et al. (1994) Small mobilizable multi-purpose cloning vectors derived from the *Escherichia coli* plasmids pK18 and pK19: selection of defined deletions in the chromosome of *Corynebacterium glutamicum*. *Gene* 145: 69–73. PMID: [8045426](#)
34. Daniel Gietz R, Woods RA (2002) Transformation of yeast by lithium acetate/single-stranded carrier DNA/polyethylene glycol method. *Methods Enzymol* 350: 87–96. PMID: [12073338](#)
35. Vaghchhipawala Z, Rojas C, Senthil-Kumar M, Mysore K (2011) Agroinoculation and agroinfiltration: simple tools for complex gene function analyses. *Methods Mol Biol* 678: 65–76. doi: [10.1007/978-1-60761-682-5_6](#) PMID: [20931373](#)
36. Anders S, Huber W (2010) Differential expression analysis for sequence count data. *Genome Bio* 11: R106.
37. Lu H (2009) Dissection of salicylic acid-mediated defense signaling networks. *Plant Signal Behav* 4: 713–717. PMID: [19820324](#)
38. Cheong MS, Park HC, Hong MJ, Lee J, Choi W, et al. (2009) Specific domain structures control abscisic acid-, salicylic acid-, and stress-mediated SIZ1 phenotypes. *Plant Physiol* 151: 1930–1942. doi: [10.1104/pp.109.143719](#) PMID: [19837819](#)
39. Duek PD, Elmer MV, van Oosten VR, Fankhauser C (2004) The degradation of HFR1, a putative bHLH class transcription factor involved in light signaling, is regulated by phosphorylation and requires COP1. *Curr Biology* 14: 2296–2301.
40. Yang J, Lin R, Sullivan J, Hoecker U, Liu B, et al. (2005) Light regulates COP1-mediated degradation of HFR1, a transcription factor essential for light signaling in *Arabidopsis*. *Plant Cell* 17: 804–821. PMID: [15705947](#)
41. Jang IC, Niu QW, Deng S, Zhao P, Chua NH (2012) Enhancing protein stability with retained biological function in transgenic plants. *Plant J* 72: 345–354. doi: [10.1111/j.1365-313X.2012.05060.x](#) PMID: [22631228](#)
42. Perdomo J, Verger A, Turner J, Crossley M (2005) Role for SUMO modification in facilitating transcriptional repression by BKLF. *Mol Cell Biol* 25: 1549–1559. PMID: [15684403](#)
43. Lin DY, Huang YS, Jeng JC, Kuo HY, Chang CC, et al. (2006) Role of SUMO-interacting motif in Daxx SUMO modification, subnuclear localization, and repression of sumoylated transcription factors. *Mol Cell* 24: 341–354. PMID: [17081986](#)
44. Wang Y, Rao VK, Kok WK, Roy DN, Sethi S, et al. (2012) SUMO modification of Stra13 is required for repression of cyclin D1 expression and cellular growth arrest. *PLoS ONE* 7: e43137. doi: [10.1371/journal.pone.0043137](#) PMID: [22905217](#)
45. Kim M, Chen Z, Shim M, Lee M, Kim J, et al. (2013) SUMO modification of NZFP mediates transcriptional repression through TBP binding. *Mol Cell* 35: 70–78.

46. Rohira AD, Chen CY, Allen JR, Johnson DL (2013) Covalent SUMO modification of Maf1 controls RNA polymerase III-dependent transcription repression. *J Biol Chem* 288: 19288–19295. doi: [10.1074/jbc.M113.473744](https://doi.org/10.1074/jbc.M113.473744) PMID: [23673667](https://pubmed.ncbi.nlm.nih.gov/23673667/)
47. Tan L, Rong W, Luo H, Chen Y, He C (2014) The *Xanthomonas campestris* effector protein XopD_{Xcc8004} triggers plant disease tolerance by targeting DELLA proteins. *New Phytol* 204: 595–608. doi: [10.1111/nph.12918](https://doi.org/10.1111/nph.12918) PMID: [25040905](https://pubmed.ncbi.nlm.nih.gov/25040905/)
48. Deslandes L, Rivas S (2012) Catch me if you can: bacterial effectors and plant targets. *Trends Plant Sci* 17: 644–655. doi: [10.1016/j.tplants.2012.06.011](https://doi.org/10.1016/j.tplants.2012.06.011) PMID: [22796464](https://pubmed.ncbi.nlm.nih.gov/22796464/)
49. Xin XF, He SY (2013) *Pseudomonas syringae* pv. *tomato* DC3000: a model pathogen for probing disease susceptibility and hormone signaling in plants. *Annu Rev Phytopathol* 51: 473–498. doi: [10.1146/annurev-phyto-082712-102321](https://doi.org/10.1146/annurev-phyto-082712-102321) PMID: [23725467](https://pubmed.ncbi.nlm.nih.gov/23725467/)