



Platelet-Derived Growth Factor Receptor- β Antagonism Restores Morphine Analgesic Potency against Neuropathic Pain

Courtney L. Donica¹, Yan Cui¹, Shanping Shi¹, Howard B. Gutstein^{1,2*}

¹ Department of Anesthesiology, The University of Texas – MD Anderson Cancer Center, Houston, Texas, United States of America, ² Department of Biochemistry and Molecular Biology, Genes and Development Graduate Program, The University of Texas – MD Anderson Cancer Center, Houston, Texas, United States of America

Abstract

Background: Chronic, intractable pain is a problem of pandemic proportions. Pain caused by nerve injuries (neuropathic pain) is extremely difficult to treat. For centuries, opiates such as morphine have been the first-line treatment for severe chronic pain. However, opiates are often ineffective against neuropathic pain, leaving few options for suffering patients. We previously demonstrated that platelet-derived growth factor- β (PDGFR- β) inhibition completely eliminated morphine tolerance. In these studies, we determined whether PDGFR- β inhibition could improve the effectiveness of morphine for neuropathic pain treatment.

Results and Findings: Spinal nerve ligation was performed in male Sprague-Dawley rats. The clinically used PDGFR antagonist imatinib did not relieve mechanical pain in a nerve injury model as determined by Von Frey assay. Surprisingly, combining imatinib with a previously ineffective dose of morphine led to complete pain relief. Scavenging released PDGF-B also markedly augmented the analgesic effect of morphine.

Conclusions: These findings suggest the novel hypothesis that PDGF-B released by injured nerves renders animals resistant to morphine, implying that PDGFR- β inhibition could potentially eliminate the tremendous suffering caused by neuropathic pain.

Citation: Donica CL, Cui Y, Shi S, Gutstein HB (2014) Platelet-Derived Growth Factor Receptor- β Antagonism Restores Morphine Analgesic Potency against Neuropathic Pain. PLoS ONE 9(5): e97105. doi:10.1371/journal.pone.0097105

Editor: Shilpa J Buch, University of Nebraska Medical Center, United States of America

Received: January 19, 2014; **Accepted:** April 14, 2014; **Published:** May 12, 2014

Copyright: © 2014 Donica et al. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Funding: This work was funded by grant number RP 120408 from the Cancer Prevention and Research Institute of Texas to H.B.G. The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

Competing Interests: The authors have declared that no competing interests exist.

* E-mail: hgutstein@mdanderson.org

Introduction

In the United States alone, chronic pain afflicts over 100 million people at an estimated yearly cost to society of over 500 billion dollars [1]. Throughout recorded history, opioid drugs such as morphine have been a mainstay of treatment for severe, chronic pain [2]. However, over time tolerance to opioid analgesia develops. Because there are few alternatives to opioids for the treatment of intractable pain, marked increases in opioid dose may be required to compensate for inadequate analgesia as tolerance develops. However, tolerance to the unpleasant or potentially life-threatening side effects of opioids such as respiratory depression, constipation, urinary retention and delirium, does not occur as rapidly as analgesic tolerance [2,3]. Therefore, patients face increased risk as well as suffering when opioids lose effectiveness.

It has been proposed that pain and tolerance utilize common signaling mechanisms [4]. We recently discovered that the platelet derived growth factor beta (PDGFR- β) is a highly selective and specific mediator of morphine tolerance [5]. We established that PDGFR- β signaling is both necessary and sufficient to cause morphine tolerance, and that morphine induced the release of PDGF-B, which caused tolerance to occur. Pain due to nerve injury (neuropathic pain) is particularly resistant to opioids,

although high doses of morphine can temporarily relieve neuropathic pain in rodents [6–8]. It is estimated that 40–60% of people suffering from neuropathic pain have inadequate pain relief [9]. It is thought that one of the most difficult features of neuropathic pain to treat is mechanical hypersensitivity caused by nerve injury [10]. We hypothesized that mechanical hypersensitivity could be resistant to the analgesic effects of morphine because the nerve injury itself induced analgesic tolerance. In an animal model of neuropathic pain [11], we found that not only did PDGFR- β inhibition block analgesic tolerance, but also markedly improved the analgesic effectiveness of morphine against mechanical hypersensitivity. Additional experiments suggested the hypothesis that PDGF-B release by injured nerves could render neuropathic pain resistant to the analgesic effects of morphine.

Methods

Animals

Male Sprague Dawley rats (175–200 g, Harlan) were housed in groups of three and were maintained on a 12 hr light/dark cycle with *ad libitum* access to food and water. Rats habituated to the colony room for one week prior to experimental manipulations.

Left L5 spinal nerve ligations were performed as described [11]. All protocols were approved by the MD Anderson Cancer Center Institutional Animal Care and Use Committee.

Drug Administration

Drugs were dissolved in a solution of 10% β -cyclodextrin sulfobutyl ether (Captisol[®], CyDex, Lenexa, KS) solution and 0.45% saline. Morphine sulfate was obtained from the MD Anderson Cancer Center Pharmacy, imatinib from LC Laboratories (Woburn, MA) and recombinant human PDGFR- β -Fc from R&D Systems (Minneapolis, MN). PDGFR- β -Fc was re-constituted in phosphate buffered saline (PBS) with 0.1% bovine serum albumin (BSA) at 100 μ g/mL and stored at -80° C until use. Drugs were administered daily via subcutaneous injection or lumbar puncture as previously described [12].

Nociceptive Testing

Animals were placed in Plexiglas cages on a mesh surface and habituated for 30 min per day for 3 days prior to testing. Mechanical sensitivity was assessed by Von Frey filaments using the up-down method of Dixon and median 50% threshold determined as described [13,14].

Statistical Analyses

Data were analyzed using GraphPad v 5.0 and was considered statistically significant if $P < 0.05$ by two-way analysis of variance (ANOVA).

Results

We initially administered morphine in the presence or absence of the PDGFR inhibitor imatinib [15] daily for 4 days in rats that underwent spinal nerve ligation (SNL). A morphine dose that is analgesic in thermal assays of nociception in non-ligated animals [5] (2 nmol, injected intrathecally (i.t.)) had no effect on mechanical hypersensitivity (Figure 1a). Imatinib alone also had no effect. Surprisingly, administering morphine and imatinib together completely eliminated the mechanical allodynia induced by SNL (Figure 1a; Treatment $F_{(3,31)} = 1009$, Day $F_{(5,155)} = 339.2$, Interaction $F_{(15,155)} = 92.55$; all $p < 0.0001$ by two-way ANOVA).

We then determined whether imatinib would block mechanical allodynia from the initiation of nerve injury when given systemically. Animals were treated with a morphine dose that is analgesic in tests of thermal nociception (2.5 mg/kg, injected subcutaneously (s.c.)), imatinib (5 mg/kg, s.c.), the combination, or vehicle starting the day after SNL. Neither morphine nor imatinib alone altered mechanical hypersensitivity. Similar to the results obtained above after a two week surgical recovery (Figure 1a), combining morphine and imatinib produced immediate and complete reversal of mechanical allodynia from the day after nerve injury until the study was concluded 9 days later (Figure 1b; Drug $F_{(4,190)} = 443.9$, Day $F_{(4,190)} = 11.13$, Interaction $F_{(16,190)} = 16.61$; all $p < 0.0001$ by two-way ANOVA).

These results suggested the hypothesis that release of PDGF by ligated nerves could render the resulting mechanical hypersensitivity resistant to the analgesic effects of morphine. To test this hypothesis, animals that had undergone SNL were treated with either 2 nmol morphine i.t., 500 ng of a fusion protein constructed of the extracellular domains of the PDGFR- β fused to antibody Fc fragments (PDGFR- β -Fc), which scavenges released PDGF-B (5), the combination, or vehicle. Administering vehicle, morphine, or PDGFR- β -Fc alone had no effect on mechanical hypersensitivity. Remarkably, combining morphine with PDGFR- β -Fc completely eliminated mechanical allodynia (Figure 1c; Treatment

$F_{(4,29)} = 186.9$, Day $F_{(9,261)} = 43.29$, Interaction $F_{(36, 261)} = 12.04$; all $p < 0.0001$ by two-way ANOVA). Similar to the combination of morphine and imatinib, repeated doses of PDGFR- β -Fc and morphine continued to block mechanical allodynia. These results support the hypotheses that 1) PDGF-B released by injured nerves reduces the effectiveness of morphine against mechanical allodynia and 2) that imatinib augments the effectiveness of morphine against mechanical allodynia by blocking PDGFR- β -mediated signaling.

Discussion

Our findings demonstrate that PDGFR- β inhibition, either spinally or systemically, enhances morphine effectiveness against neuropathic pain. Combining a previously ineffective morphine dose with PDGFR- β antagonists can reverse established allodynia as well as prevent allodynia from developing after nerve injury. In addition, similar to our previously reported results [5], PDGFR- β inhibition prevents tolerance to this effect from developing. Interestingly, imatinib alone does not affect morphine analgesia [5]. Further, repeated administration of morphine with imatinib remains effective through at least 9 days (figure 1B). This suggests that there is no “tolerance” to the PDGFR- β antagonist rescue of morphine effectiveness. These results suggest that lower doses of opioids could be used to treat neuropathic pain if PDGFR- β inhibitors were given concurrently. In addition, the dose escalation that commonly occurs [9] could be avoided. Since tolerance to unpleasant or potentially life-threatening side effects of opioids such as respiratory depression, constipation, urinary retention and delirium, does not occur as rapidly as analgesic tolerance [3], PDGFR- β antagonism could potentially reduce the risks associated with chronic opioid treatment [16] and the suffering that results when opioids lose effectiveness.

We previously demonstrated that administration of PDGF-B to naïve animals rendered them tolerant to subsequent morphine doses [5]. PDGF-B is localized in the dorsal root ganglion and dorsal horn of the spinal cord and PDGFR- β is located in myelinated and unmyelinated nerves, dorsal root ganglion neurons and the spinal dorsal horn [17–20]. Given the distribution of PDGF-B and PDGFR- β , it is possible that PDGFR- β inhibition blocks opioid tolerance by modulating opioid effects on peptidergic primary afferent fibers. It has been suggested that pain and opioid tolerance utilize common signaling pathways [4,21]. Our current observation that PDGF-B scavenging augmented the analgesic effect of morphine against neuropathic pain indicates that nerve ligation also induces PDGF-B release. PDGF-B alone does not appear to induce pain: PDGF-BB administration does not induce thermal sensitivity [5] and scavenging released PDGF-B alone does not alleviate neuropathic pain (Fig 1C). Rather, our data suggests that released PDGF-B affects the ability of morphine to induce analgesia. Taken together, these findings suggest that PDGF-B released by injured nerves causes resistance to the analgesic effect of morphine, possibly through similar translational and post-translational modifications of analgesic effectors induced by prior opioid exposure. In effect, PDGF-B renders animals “pre-tolerant” to morphine analgesia. Previous work has shown that neuropathic pain reduces the efficacy of morphine, supporting this hypothesis [6–8]. Our results also suggest that blockade of PDGFR- β signaling could become an important therapeutic approach for relieving the suffering of untold numbers of patients living with intractable neuropathic pain.

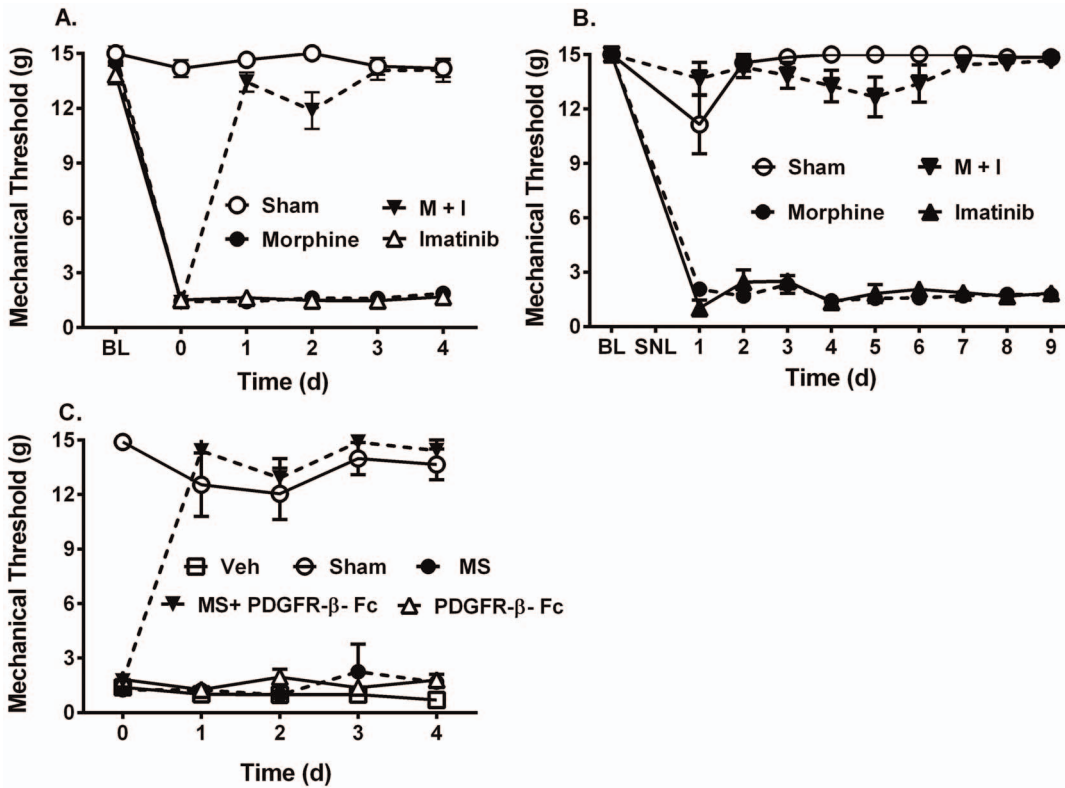


Figure 1. PDGFR inhibition restores morphine efficacy against neuropathic pain. Animals underwent left L5 SNL as described in [11]. Mechanical sensitivity was tested using von Frey filaments and 50% median response threshold determined [13,14]. Sham operated animals underwent the same surgical procedure except a suture was not tied around the L5 nerve root. **(A)** Baseline mechanical sensitivity was determined before surgery (BL). Animals then underwent SNL and were allowed to recover for two weeks. Mechanical sensitivity was tested to confirm that SNL induced mechanical allodynia (day 0). SNL animals received daily intrathecal (i.t.) injections of morphine (2 nmol), imatinib (10 μ g), or the combination and mechanical sensitivity was determined. Sham operated animals received injections of vehicle alone. Neither morphine nor imatinib alone were analgesic. Co-administration of morphine and imatinib completely reversed SNL-induced mechanical allodynia. $n=8-9$ per group. **(B)** Beginning the day after SNL, animals received daily subcutaneous (s.c.) injections of morphine (2.5 mg/kg), imatinib (5 mg/kg), morphine + imatinib or vehicle and mechanical sensitivity determined. Systemic co-administration of morphine and imatinib completely eliminated allodynia after nerve injury. $n=5-9$ per group. **(C)** Following a two week recovery after SNL, animals received daily i.t. injections of morphine (2 nmol), PDGFR- β -Fc scavenger (500 ng), the combination or vehicle. While PDGFR- β -Fc had no effect on mechanical allodynia, co-administration of morphine and PDGFR- β -Fc completely restored the effectiveness of morphine. $n=7-10$ per group. All data presented as grams \pm s.e.m; all $p<0.0001$ (2-way ANOVA). doi:10.1371/journal.pone.0097105.g001

Acknowledgments

We thank Jin Mo Chung and Shao-Rui Chen for instruction in spinal nerve ligation. All animal studies were approved by the M.D. Anderson Cancer Center Institutional Animal Care and Use Committee.

Author Contributions

Conceived and designed the experiments: HBG. Performed the experiments: CLD SS YC. Analyzed the data: CLD SS YC HBG. Wrote the paper: CLD HG.

References

- Board of Health Sciences Policy (2011) Relieving Pain in America: A Blueprint for Transforming Prevention, Care, Education and Research. Washington, DC: The National Academies Press. 313 p.
- Gutstein H, Akil H (2006) Opioid Analgesics. In: Brunton L, Lazo J, Parker K, editors. Goodman & Gilman's The Pharmacological Basis of Therapeutics. 11 ed. New York: McGraw-Hill. pp. 547-590.
- Collett BJ (1998) Opioid tolerance: the clinical perspective. Br J Anaesth 81: 58-68.
- Mayer D, Mao J, Holt J, Price D (1999) Cellular mechanisms of neuropathic pain, morphine tolerance, and their interactions. Proc Natl Acad Sci 96: 7731-7736.
- Wang Y, Barker KE, Shi S, Diaz MF, Mo B, et al. (2012) Blockade of PDGFR- β Activation Eliminates Morphine Analgesic Tolerance. Nature Medicine 18: 385-387.
- Bian D, Nichols ML, Ossipov MH, Lai J, Porreca F (1995) Characterization of the antiallodynic efficacy of morphine in a model of neuropathic pain in rats. Neuroreport 6: 1981-1984.
- Przewlocka B, Mika J, Labuz D, Toth G, Przewlocki R (1999) Spinal analgesic action of endomorphins in acute, inflammatory and neuropathic pain in rats. Eur J Pharmacol 367: 189-196.
- Przewlocki R, Przewlocka B (2001) Opioids in chronic pain. Eur J Pharmacol 429: 79-91.
- Dworkin RH, O'Connor AB, Backonja M, Farrar JT, Finnerup NB, et al. (2007) Pharmacologic management of neuropathic pain: evidence-based recommendations. Pain 132: 237-251.
- Woolf CJ, Mannion RJ (1999) Neuropathic pain: aetiology, symptoms, mechanisms, and management. Lancet 353: 1959-1964.
- Chung JM, Kim HK, Chung K (2004) Segmental spinal nerve ligation model of neuropathic pain. Methods Mol Med 99: 35-45.
- Xu JJ, Walla BC, Diaz MF, Fuller GN, Gutstein HB (2006) Intermittent lumbar puncture in rats: a novel method for the experimental study of opioid tolerance. Anesthesia & Analgesia 103: 714-720.
- Dixon WJ (1980) Efficient analysis of experimental observations. Annu Rev Pharmacol Toxicol 20: 441-462.
- Chaplan SR, Bach FW, Pogrel JW, Chung JM, Yaksh TL (1994) Quantitative assessment of tactile allodynia in the rat paw. J Neurosci Methods 53: 55-63.
- Buchdunger E, Gioffi CL, Law N, Stover D, Ohno-Jones S, et al. (2000) Abl protein-tyrosine kinase inhibitor STI571 inhibits in vitro signal transduction mediated by c-kit and platelet-derived growth factor receptors. J Pharmacol Exp Ther 295: 139-145.

16. Rice ASC, Hill RG (2006) New treatments for neuropathic pain. *Annual Review of Medicine* 57: 535–551.
17. Eccleston PA, Funa K, Heldin CH (1993) Expression of platelet-derived growth factor (PDGF) and PDGF alpha- and beta-receptors in the peripheral nervous system: an analysis of sciatic nerve and dorsal root ganglia. *Dev Biol* 155: 459–470.
18. Masuda J, Tsuda M, Tozaki-Saitoh H, Inoue K (2009) Intrathecal delivery of PDGF produces tactile allodynia through its receptors in spinal microglia. *Molecular Pain*. 2009/05/12 ed. pp. 23.
19. Sasahara A, Kott JN, Sasahara M, Raines EW, Ross R, et al. (1992) Platelet-derived growth factor B-chain-like immunoreactivity in the developing and adult rat brain. *Brain Res Dev Brain Res* 68: 41–53.
20. Sasahara M, Fries JW, Raines EW, Gown AM, Westrum LE, et al. (1991) PDGF B-chain in neurons of the central nervous system, posterior pituitary, and in a transgenic model. *Cell* 64: 217–227.
21. Mao J, Price DD, Mayer DJ (1995) Mechanisms of hyperalgesia and morphine tolerance: a current view of their possible interactions. *Pain* 62: 259–274.