



# Article From Automated Synthesis to In Vivo Application in Multiple Types of Cancer—Clinical Results with [<sup>68</sup>Ga]Ga-DATA<sup>5m</sup>.SA.FAPi

Lukas Greifenstein <sup>1,\*,†</sup>, Carsten S. Kramer <sup>1,†</sup>, Euy Sung Moon <sup>2</sup>, Frank Rösch <sup>2</sup>, Andre Klega <sup>1</sup>, Christian Landvogt <sup>1</sup>, Corinna Müller <sup>1</sup> and Richard P. Baum <sup>1,\*</sup>

- <sup>1</sup> CURANOSTICUM Wiesbaden-Frankfurt, Center for Advanced Radiomolecular Precision Oncology, 65191 Wiesbaden, Germany
- <sup>2</sup> Department of Chemistry–TRIGA, Institute of Nuclear Chemistry, Johannes Gutenberg University, 55128 Mainz, Germany
- \* Correspondence: greifenstein@curanosticum.de (L.G.); baum@curanosticum.de (R.P.B.)
- † These authors contributed equally to this work.

**Abstract:** Radiolabeled FAPI (fibroblast activation protein inhibitors) recently gained attention as widely applicable imaging and potential therapeutic compounds targeting CAF (cancer-associated fibroblasts) or DAF (disease-associated fibroblasts in benign disorders). Moreover, the use of FAPI has distinct advantages compared to FDG (e.g., increased sensitivity in regions with high glucose metabolism, no need for fasting, and rapid imaging). In this study, we wanted to evaluate the radiochemical synthesis and the clinical properties of the new CAF-targeting tracer [<sup>68</sup>Ga]Ga-DATA<sup>5m</sup>.SA.FAPi. The compound consists of a (radio)chemically easy to use hybrid chelate DATA.SA, which can be labeled at low temperatures, making it an interesting molecule for 'instant kit-type' labeling, and a squaric acid moiety that provides distinct advantages for synthesis and radiolabeling. Our work demonstrates that automatic synthesis of the FAP inhibitor [<sup>68</sup>Ga]Ga-DATA<sup>5m</sup>.SA.FAPi for the unambiguous detection of cancer-associated fibroblasts of various carcinomas and their metastases (NSCLC, liposarcoma, parotid tumors, prostate cancer, and pancreas adenocarcinoma), while physiological uptake in brain, liver, intestine, bone, and lungs was very low.

**Keywords:** nuclear medicine; PET; Ga-68; FAP; FAPI; DATA; squaric acid; molecular imaging; chelator; first-in-human study

# 1. Introduction

Since targeted radiopharmaceuticals against certain cancer-related antigens (such as PSMA and SSTR) have demonstrated the clinical impact of radiomolecular precision oncology, the demand became apparent to find new targets with a suitable profile for targeted imaging and therapy. An ideal neoantigen should be highly expressed on malignant tissue and not only be restricted to a small subset of cancer types (pan-cancer target). The fibroblast activation protein (FAP) can be seen as a very interesting candidate as it is not only expressed on several tumor types, but also on cancer-associated fibroblasts (CAFs) that are part of the tumor microenvironment (TME) [1–7]. FAP is a transmembrane glycoprotein (type II) and belongs to the serine protease family [8,9]. The unique feature of FAP is that it is nearly absent in healthy tissues as well as in benign tumors. In contrast, FAP is overexpressed in over 90% of the most common human epithelial tumors, such as breast, pancreatic, colon, and lung cancers; it is also associated with conditions or diseases such as wound healing, chronic inflammation, liver cirrhosis, rheumatoid arthritis, pulmonary



**Citation:** Greifenstein, L.; Kramer, C.S.; Moon, E.S.; Rösch, F.; Klega, A.; Landvogt, C.; Müller, C.; Baum, R.P. From Automated Synthesis to In Vivo Application in Multiple Types of Cancer—Clinical Results with [<sup>68</sup>Ga]Ga-DATA<sup>5m</sup>.SA.FAPi. *Pharmaceuticals* **2022**, *15*, 1000. https://doi.org/10.3390/ ph15081000

Academic Editor: Irina Velikyan

Received: 30 April 2022 Accepted: 9 August 2022 Published: 14 August 2022

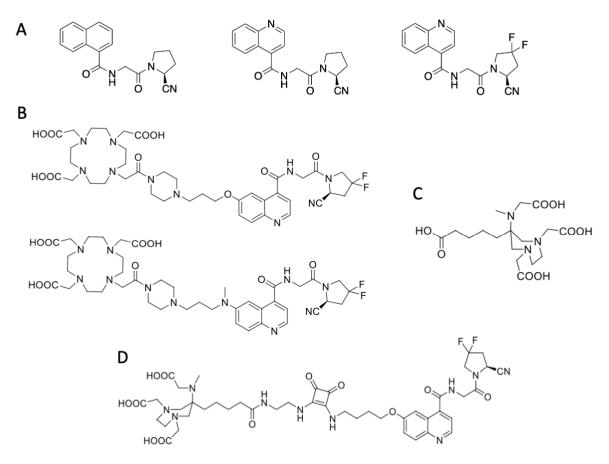
**Publisher's Note:** MDPI stays neutral with regard to jurisdictional claims in published maps and institutional affiliations.



**Copyright:** © 2022 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (https:// creativecommons.org/licenses/by/ 4.0/). fibrosis, bone and soft tissue sarcomas, osteoarthritis, and Crohn's disease [8,10–30]. For a recent review, see Imlimthan et al. [31].

Currently, most FAP-targeted radiopharmaceuticals are highly affine inhibitors of the FAP enzyme (FAPI, FAP inhibitors) that possess a low off-target selectivity towards other peptidases.

So far, the best known FAP inhibitors are boronic acid-based and glycine-2cyanopyrrolidine-based FAPIs; for those, the boronic acid acts as the 'warhead', while the nitrile group in the (cyano)proline-derived FAPIs takes over this function (Figure 1A). In particular, the gly-(2-cyano)Pro inhibitors show very high selectivity for FAP and additionally sufficient non-specificity for competing peptidases [32–34]. The most potent inhibitor is (*S*)-N-(2-(2-cyano-4,4-difluoropyrrolidin-1-yl)-2-oxoethyl)quinoline-4-carboxamide (UAMC1110), developed by Pieter van der Veken et al. [35–37].



**Figure 1.** Schematic representation of the relevant chemical structures. (**A**): Gly-2-cyanopyrrolidine FAP inhibitors: (N-(1-naphthyl)-gly-2-cyanopyrrolidine; (4-quinolinyl)glycyl-2-cyanopyrrolidine and the quinoline-gly-2-cyano-4,4-difluoroPro-based FAPi, UAMC1110; (**B**) DOTA-conjugated FAPI conjugates FAPI-04 (top) and FAPI-46 (bottom); (**C**) DATA<sup>5m</sup> (**D**) DATA<sup>5m</sup>. SA.FAPi.

Accordingly, the first FAP-targeting inhibitor compounds that found application for radiomolecular precision imaging were DOTA-conjugated variants derived from UAMC1110 [38–40]. In preclinical studies, especially FAPI-04 and FAPI-46 conjugates gained attention as the most promising FAP-addressing radiopharmaceuticals to date (Figure 1B) [41–43]. Since then, FAPI derivatives found their way into several clinical applications for a wide variety of tumor types, primarily with a focus on diagnosis by <sup>68</sup>Ga-PET/CT. However, increasingly, other PET nuclides, such as <sup>18</sup>F and <sup>64</sup>Cu [44–54], as well as beta and alpha emitters, are tested in preclinical studies, e.g., <sup>177</sup>Lu [55–60], while therapeutical nuclides such as <sup>153</sup>Sm [61], <sup>90</sup>Y, <sup>177</sup>Lu and <sup>225</sup>Ac [62–67] and combinations thereof ("TANDEM" approach) have entered clinical phases [68].

In recent years, several groups have tried to design molecules that combine the advantages of both cyclic and acyclic chelators as so-called hybrid chelators. Hybrid chelators are generally considered to be particularly fast in their radiolabeling kinetics, and temperature independent when compared to acyclic chelators. Furthermore, kinetic stability should be comparable to cyclic complexes [69–71]. A very promising approach is the usage of a diazepine scaffold which contributes with two endocyclic amines to complex formation. Introduction of an additional exocyclic amine function provides a third coordination unit. Further functionalization of these amines with carboxylic acids allows the introduction of more donor units and final formation of the DATA (6-amino-1,4-diazepine-triacetic acid) chelator (Figure 1C) [72,73]. For the development of these structures to bifunctional systems, many derivatives were investigated. In this study, DATA<sup>5m</sup> is used: herein, a C5-linker with a terminal carboxylic acid is introduced to provide bifunctionality (Figure 1C). In previous studies, it could be shown that DATA possess excellent properties for complexing gallium at room temperature resulting in highly stable complexes (note: even though reaction is possible at room temperature, we applied slightly elevated temperatures in our clinical routine work due to validated production methods).

In the present study, we used the DATA<sup>5m</sup>-conjugated precursor DATA<sup>5m</sup>.SA.FAPi, which was first introduced by Moon et al. [74] (Figure 1D). The synthesis of the precursor takes advantage of a squaric acid (SA) moiety that conveniently allows the coupling of two amine bearing units [75,76], such as the chelator and FAP-targeting motif. SA is not only acting as a connector, the influence of the SA unit on the pharmacophore and the biological activity of the final compound must be considered as well. Favorable kinetics by SA have been shown for other molecules before [77,78]. For the coupling of chelators, it is also noteworthy that SA has complexing properties itself [79] and may therefore influence the complexation properties of the bound chelator.

While the FAP-targeted and radiolabeled SA derivatives DOTA.SA.FAPi and AAZTA<sup>5</sup>. SA.FAPi have been investigated preclinically and clinically in several studies [74,80–83], there are only two clinical case reports by Kreppel et al. that investigated [<sup>68</sup>Ga]Ga-DATA<sup>5m</sup>.SA.FAPi in fourteen patients with focal nodular hyperplasia (FNH) (one patient) and neuroendocrine tumors (13 patients) [84,85]. In this study, we present our results on automated radiolabeling of DATA<sup>5m</sup>.SA.FAPi with Ga-68 and furthermore our first experience in patients with different cancers such as small-cell cancer of the prostate, parotid gland tumor, liposarcoma, pancreatic adenocarcinoma and non-small-cell lung cancer (NSCLC).

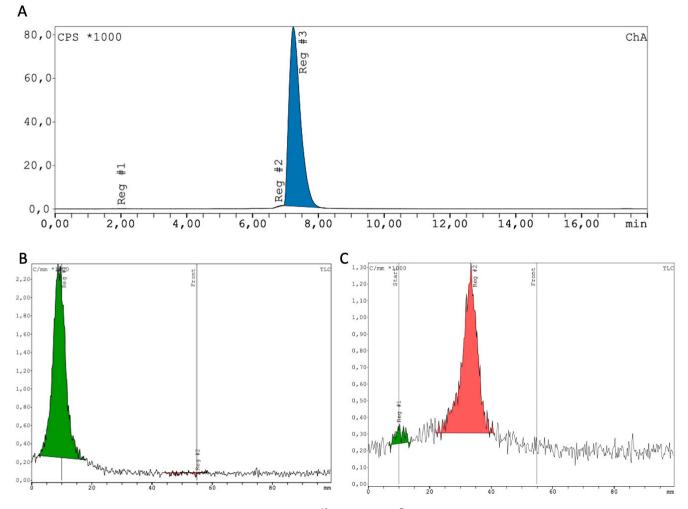
## 2. Results

## 2.1. Synthesis and Radiochemistry

Synthesis of the molecule was performed in accordance with the literature published by Moon et al. [74].

Radiolabeling of the new FAP inhibitor DATA<sup>5m</sup>.SA.FAPi with gallium-68 was performed automatically using a mini-All-in-One cassette-based module from Trasis. Reaction at 50 °C in sodium acetate buffer (0.7 M, pH 5.5) provided RCY  $\geq$  95% within 10 min. An example of the radio-HPLC and radio-TLC is shown in Figure 2, demonstrating a radiochemical purity (RCP)  $\geq$  98% (avg. RCP 99.28  $\pm$  0.99%). The automated synthesis yielded activities between 616 and 654 MBq (avg. 634  $\pm$  16 MBq; *n* = 4; radiochemical yield (RCY = 84.4  $\pm$  5.0%) after 10 min of labeling with 50 µg of DATA<sup>5m</sup>.SA.FAPi and 20 min of quality control.

No indices for tracer instability were detected in the period between synthesis and image acquisition.



**Figure 2.** Quality control of  $[^{68}Ga]Ga$ -DATA<sup>5m</sup>.SA.FAPi. (**A**) radio-HPLC; (**B**) radio-TLC in citric acid buffer; (**C**) radio-TLC in ammoniumacetate buffer and methanol (1:1 v/v).

#### 2.2. Clinical Safety

All patients tolerated the tracer application (median: 168 MBq, range: 126–316 MBq) extremely well without any acute adverse effects. No symptoms were noticed by any patient during the follow-up period of approx. 4 weeks (by consultation and clinical examination), and blood tests (CBC, liver, and kidney parameters).

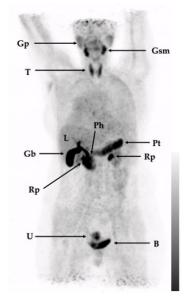
## 2.3. In-Human Tracer Uptake and Biodistribution

Mild physiological uptake was predominantly found in salivary glands and the thyroid, gall bladder (and consequently in intestines), frequently in the uterus and—due to excretion—in the kidneys and the urinary bladder. Pancreatic uptake was inhomogeneous and varied considerably between patients, with accentuation in the head and/or tail. Table 1 summarizes the physiological uptake (SUV<sub>max</sub> and SUV<sub>mean</sub>) in selected tissues and organs (collected data from all examined patients). Figure 3 illustrates a patient with sarcoidosis that did not show any disease-related tracer uptake. As patients were not imaged with other FAPI tracers, data for direct comparison are not available. To benchmark [<sup>68</sup>Ga]Ga-DATA<sup>5m</sup>.SA.FAPi with other FAP-directed tracers, we compared the tracer distribution pattern of [<sup>68</sup>Ga]Ga-DATA<sup>5m</sup>.SA.FAPi with published data from FAPI-02, FAPI-46, and FAPI-74 at comparable time points [86,87]. At best, the physiological distribution of the squaric acid-based analog is similar to the pattern of FAPI-02 but with less kidney uptake and less blood pool activity. Blood pool and muscle uptake are significantly better compared to FAPI-46 and FAPI-74, while uptakes in the thyroid gland, glandular \_

submandibularis, and pancreas are higher for [<sup>68</sup>Ga]Ga-DATA<sup>5m</sup>.SA.FAPi as well as for FAPI-02 (compared to FAPI-46 and FAPI-74).

**Table 1.** Physiological uptake (SUV<sub>max</sub> and SUV<sub>mean</sub>) of the tracer [average  $\pm$  SD] in selected organs and tissues.

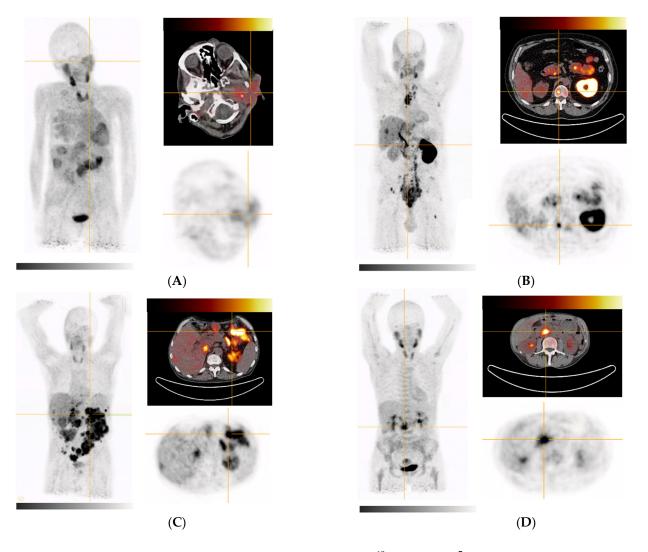
Tissue/Organ	SUVmax [Average $\pm$ SD]	SUVmean [Average $\pm$ SD]
Spleen	$2.4\pm0.5$	$1.6\pm0.5$
Liver	$2.5\pm0.4$	$1.9\pm0.3$
Red marrow (vertebrae)	$2.3\pm0.4$	$1.5\pm0.5$
Kidney	$3.0 \pm 1.1$	$2.4\pm0.9$
Brain (frontal cortex)	$0.4\pm0.2$	$0.1\pm0.1$
Pituitary gland	$1.7\pm0.6$	$1.0\pm0.3$
Submandibular gland	$6.3 \pm 1.2$	$4.3\pm2.1$
Thyroid	$6.1 \pm 1.8$	$3.9\pm1.2$
Pancreas	$7.8\pm2.5$	$4.5\pm1.5$
Lung	$0.8\pm0.3$	$0.6\pm0.2$
Muscle (quadriceps)	$0.9\pm0.2$	$0.6\pm0.2$
Blood pool (aorta)	$1.8\pm0.9$	$1.5\pm0.7$



**Figure 3.** Maximum-intensity projection (MIP) of [<sup>68</sup>Ga]Ga-DATA<sup>5m</sup>.SA.FAPi in a patient without any disease-related tumor uptake. Gp: glandula parotis; Gsm: glandula submandibularis; T: thyroid; L: liver; Ph: pancreas head; Pt: pancreas tail; Gb: gall bladder; Rp: renal pelvis; U: uterus; B: urinary bladder.

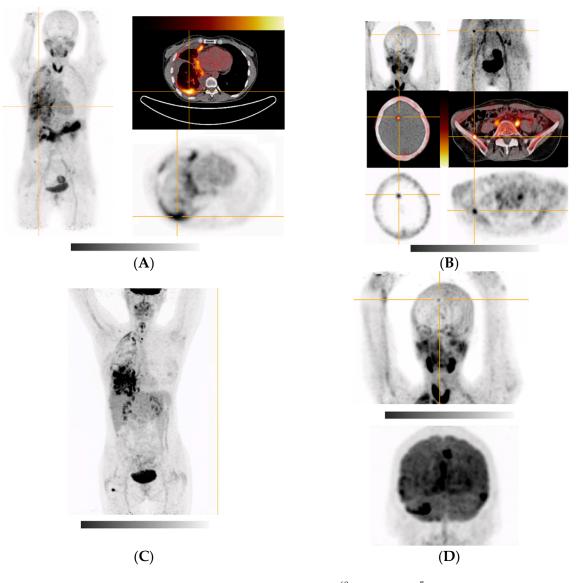
In contrast, malignant lesions showed a higher uptake of [ $^{68}$ Ga]Ga-DATA<sup>5m</sup>.SA.FAPi with an average SUV<sub>max</sub> of 9.1  $\pm$  3.3, resulting in a mean tumor-to-background ratio (TBR) of 4.7. The highest SUV<sub>max</sub> of 12.7 was found in a bone metastasis of a patient with prostate cancer, while the highest tumor uptake could be detected in a liver metastasis with a SUV<sub>max</sub> of 5.0 in a parotid cancer patient. Notably, these observations are not statistically significant as only one patient for each tumor entity was examined.

Figure 4A demonstrates tracer uptake in a patient with heavily metastatic parotid gland tumor: while uptake of the primary did not exceed physiological accumulation in the salivary glands, liver metastasis could be easily demarcated. In small-cell prostate cancer patients (Figure 4B), soft tissue and bone metastases could be visualized with high contrast and a remarkably high tumor SUV<sub>max</sub> of 13.7. Additionally, liposarcoma-derived peritoneal metastases as well as a adenocarcinoma of the pancreas showed excellent tracer uptake with SUV<sub>max</sub> values of approx. 10 (Figure 4C,D).



**Figure 4.** MIPs and transversal images of [ $^{68}$ Ga]Ga-DATA $^{5m}$ .SA.FAPi in cancer patients. Crosshair indicates target lesion (highest tumor SUV<sub>max</sub>); (**A**) hepatic metastases of parotid gland tumor; target lesion (primary tumor/liver metastasis): 5.0; (**B**) metastasized prostate cancer; target lesion (bone metastasis in L1 vertebra): 12.7; (**C**) metastasized liposarcoma; target lesion (peritoneal metastases): 10.6; (**D**) primary pancreatic head adenocarcinoma tumor: 10.1.

In a case with metastasized NSCLC, the primary could be easily visualized (Figure 5) as well as the known cerebral metastases (Figure 5B left) (previously known from FDG-PET/CT that was performed 2 weeks before, Figure 5C,D). Notably, in [<sup>68</sup>Ga]Ga-DATA<sup>5m</sup>. SA.FAPi PET/CT, new bone lesions were detected that were previously not known (Figure 5B right).



**Figure 5.** (**A**) MIP and transversal image of [<sup>68</sup>Ga]Ga-DATA<sup>5m</sup>.SA.FAPi in a NSCLC patient. Crosshair indicates target lesion with a SUV<sub>max</sub> of 7.3. (**B**) Cerebral (**left**) and bone (**right**) metastases demonstrated by FAPI-PET/CT. (**C**,**D**) MIPs, FDG scan performed two weeks before imaging with [<sup>68</sup>Ga]Ga-DATA<sup>5m</sup>.SA.FAPi; the upper picture in (**D**) is the enlarged image of the head shown in (**B**).

Table 2 summarizes additional findings, clinical details as well as information regarding relevant pretreatments.

Patient	Disease	Pretreatments(Relevant)	Tracer Accumulation
1 (Figure 3)	Sarcoidosis (left cervical lymph node manifestation), in the past inflammatory pulmonary activity, the activity of the sarcoidosis during image acquisition was not clear. History of DCIS in the left breast	Lymph node extirpation, modified radical mastectomy on left side, prophylactic radical modified mastectomy on right side	Only physiological tracer distribution with no further suspect tracer accumulations
2 (Figure 4A)	Metastasized parotid gland tumor (adenoid-cystic subtype)	Subtotal parotidectomy (left), partial liver resection of segments II/III	Parotid gland, multiple masses in the right lung, accumulation in several liver segments

**Table 2.** Findings in all six patients during imaging with [ $^{68}$ Ga]Ga-DATA $^{5m}$ .SA.FAPi.

Detter	<b>D:</b>	Duratura dan an ta (D. 1	Transa Assess 1. Com
Patient 3 (Figure 4B)	Disease Metastasized prostate cancer (Gleason 4 + 3 = 7)	Pretreatments(Relevant) IMRT to prostate and seminal vesicles	Tracer Accumulation Extensive bone and bone marrow involvement (including the extremities), large lymph node metastases in the retroperitoneum, mediastinum (bulky), and cervical region as well as in soft tissue. Additional findings: Severely impaired function of left kidney (slow wash out of activity with high parenchymal contrast and increased activity in the ureter and pelvis)
4 (Figure 4C)	Metastasized liposarcoma	Tumor enucleation on the duodenum, pancreatic head resection, omentectomy, pancreaticogastrostomy	Extensive peritoneal tumor foci in all abdominal quadrants, caudal liver margin segment VI (or adjacent peritoneal foci)
5 (Figure 4D)	Primary pancreatic head adenocarcinoma, moderately differentiated, ductal	ERCP with stenting	Inhomogeneous in the pancreas with emphasis in pancreatic head, peritoneal (extensively in the left mid to lower abdomen), anterior margin of liver (peritoneal or lymph nodes), mammaria interna lymph node, segment VI of the liver (most likely biliary excreted tracer), uterus (nonspecific/physiological), muscle attachments at the hip joints (nonspecific/inflammatory/bursitis)
6 (Figure 5A,B)	Poorly differentiated NSCLC of right lower lobe (pleural, cerebral, hepatic, and osseous metastases)	VATS with partial pleurectomy and talc pleurodesis	Right hemithorax and right lung, diffuse cerebral (mainly focal in the right frontal cortex in the region of the great longitudinal fissure and periventricular) right and right ilium, left sacrum, right thigh (after surgical removal of a hibernoma), uterus (possible myoma) FDG-PET (Figure 5C,D): whole right hemithorax (especially basolateral, in the myelon, intracranially in the area of the meninges, (around the temporal poles and on the tentorium), in the right frontal cortex in the area of the great longitudinal fissure, right thigh (after surgical removement of a hibernoma)

#### Table 2. Cont.

## 3. Discussion

The automatic synthesis allows a fast, easy and reproducible preparation of [<sup>68</sup>Ga]Ga-DATA<sup>5m</sup>.SA.FAPi at low temperatures and in high RCPs and RCYs. These favorable conditions allow the 'kit-type synthesis' of this tracer, making it a very convenient imaging agent for FAP—even for PET centers that possess only basic technical equipment and infrastructure.

Currently, only two clinical case reports with [<sup>68</sup>Ga]Ga-DATA<sup>5m</sup>.SA.FAPi, focusing on FNH and on metastasized neuroendocrine tumors, were published [84]. Our study confirms the clinical safety of this compound as all patients tolerated the application of the tracer very well. Moreover, we demonstrate its first application in patients with a diversity of solid tumors.

The radiopharmaceutical demonstrated excellent diagnostic usability for the unambiguous detection of CAFs of various carcinomas (such as NSCLC, prostate, and pancreas) as well as in liposarcoma and in soft tissue and bone metastases. High tumor uptake was observed, and very low accumulation in brain, liver, bone, and lungs. In the patient with a pancreas head carcinoma (Figure 4D), the primary tumor could be undoubtfully visualized despite some physiological uptake in the pancreas. In line with the findings of Kreppel et al. [84], liver metastases could also readily be detected with this tracer. A clear advantage of FAP-targeting tracers in comparison to FDG is that metastases are not masked by high physiological FDG uptake. Therefore, imaging with [<sup>68</sup>Ga]Ga-DATA<sup>5m</sup>.SA.FAPi allowed also visualization of cerebral metastasis (Figure 5B left). Furthermore, bone lesions that were visible on [<sup>68</sup>Ga]Ga-DATA<sup>5m</sup>.SA.FAPi-PET/CT had not been detected by FDG-PET/CT. With regard to physiological biodistribution, the here presented tracer has a similar distribution pattern to FAPI-02 but with less kidney uptake and less blood pool activity, while radiolabeling can be achieved under more suitable conditions. As this study shows only one patient per cancer entity, tumor uptakes cannot be compared in a statistically significant manner.

A limitation of our study is the heterogenous patient group and small sample size, making a general statement about sensitivity and specificity in specific malignancies difficult. Furthermore, a bigger sample size is necessary to allow comparisons between DATA<sup>5m</sup>.SA.FAPi, other FAP-targeting tracers, and FDG-PET. Additionally, the potential of the tracer to image disease-associated fibroblasts (DAFs) needs to be further explored: although Kreppel et al. [84,85] revealed that [<sup>68</sup>Ga]Ga-DATA<sup>5m</sup>.SA.FAPi-PET/CT visualizes FNH (a benign disease), no active lesion was seen in a patient with a past history of sarcoidosis (Figure 3). During the time of the PET scan, the activity status was unclear while the patient suffered from several seizures; therefore, the missing tracer uptake cannot be correlated with disease (in) activity.

In conclusion, our results demonstrate that the FAP-targeting tracer [<sup>68</sup>Ga]Ga-DATA<sup>5m</sup>. SA.FAPi can conveniently be synthesized in an automatized process and that malignant lesions of solid tumors could be detected in a non-preselected heterogenous patient population. At this point, further studies need to be performed to compare the sensitivity of this molecule with established imaging agents. As FAP-imaging has clear advantages over FDG imaging, the synthetic accessibility to [<sup>68</sup>Ga]Ga-DATA<sup>5m</sup>.SA.FAPi, along with the excellent first clinical experience, can provide a good opportunity even for PET centers with basic equipment to implement this tracer in daily routine.

#### 4. Materials and Methods

#### 4.1. Radiochemistry

Gallium-68 was eluted from a  ${}^{68}$ Ge/ ${}^{68}$ Ga generator (Eckard und Ziegler AG, Braunschweig, Germany) and used without further purification. Radiolabeling was performed in 2.0 mL of 0.7 M ammonium acetate buffer at pH 5.5 with 50 µg precursor at 50 °C in an All-In-One mini synthesis module (Trasis, Ans, Belgium) with a disposable cassette. Subsequently, an Oasis HLB Plus cartridge was used to agitate the final compound. Elution of the product was performed with 0.5 mL ethanol. The product was then formulated via a sterile filter with 10 mL of saline.

The pH was controlled at the start and after the labeling. For reaction control, radio-TLC (TLC Silica gel 60 F254 Merck) with (1) citrate buffer pH 4 and (2) a 1:1 mixture of ammonium acetate buffer and MeOH (*v:v*) as the mobile phase was used. Additionally, radio-HPLC using an analytical HPLC from Agilent (Infinity 1200) with a Ramona \* radiodetector (Elysisa-Raytest, Angleur, Belgium) (Column: VDSpher PUR 150 C18-E 5  $\mu$ m, 100 × 40 m), linear gradient of 5–45% MeCN (+0.1% TFA)/95–55% Water (+0.1% TFA) in 10 min). TLCs were measured with a TLC miniGita (Elysia-Raytest, Angleur, Belgium) with the analysis software GINA (Elysia-Raytest, Angleur, Belgium).

# 4.2. Clinical

Five patients had histologically confirmed malignant disease, while one patient was diagnosed with sarcoidosis. All patients had an ECOG performance status <3, good renal function, and no allergies against any of the ingredients of the radiopharmaceutical. All patients signed written informed consent before this study.

Tracer uptake and biodistribution were analyzed in 6 patients (mean age: 56 y; range: 49–63 y) with various malignancies: small-cell cancer of the prostate (male, 63 y), malignant parotid gland tumor (adenoid-cystic subtype) (female, 54 y), liposarcoma (male, 62 y), pancreatic adenocarcinoma (female, 54 y), and NSCLC (female, 51 y). A sixth patient had a history of sarcoidosis (with no information about active lesions during the time of the scan) (female, 49 y).

After injection of [<sup>68</sup>Ga]Ga-DATA<sup>5m</sup>.SA.FAPi (median: 168 MBq, range: 126–316 MBq), PET/CT images (Siemens Biograph Vision 600 Edge) were acquired (malignant patients: from the vertex to mid-thigh; sarcoidosis patient: from skull to mid-thigh) after an average uptake time of 70 min (range: 48–83 min). A low-dose CT scan (120 keV, 30 mAs, CareDose; reconstructed with a soft-tissue kernel to a slice thickness of 2 mm) was used for attenuation correction and anatomical mapping of the tracer. An infusion of 500 mL saline with 20 mg of furosemide was infused from 15 min before to 150 min after tracer application.

Tumor uptake was quantified by  $SUV_{mean}$  and  $SUV_{max}$ .  $SUV_{max}$  data were calculated on the reconstructed images by drawing circular regions of interests around various organs and tumor lesions using Affinity Viewer (Hermes Medical Solutions). The tumor-tobackground ratio (TBR) was defined as  $SUV_{max}$  of the target lesions, and the  $SUV_{mean}$  of healthy liver parenchyma.

Blood samples were drawn before tracer application (CBC, liver, and kidney parameters) if no recent blood test results were available. During application, patients were monitored by medical personnel and proactively asked about side effects. Patients that were not seen in a follow-up period of 4 weeks were requested to contact us if side effects appeared. Blood results after the scan were acquired in our center or inquired from the patient.

#### 5. Patents

L.G. and F.R. have filed a patent (WO2020083853A1) on [68Ga]Ga-DATA<sup>5m</sup>.SA.FAPi.

Author Contributions: Conceptualization R.P.B., L.G. and C.S.K.; methodology L.G., C.S.K. and E.S.M.; software L.G., C.S.K., A.K., C.M., C.L. and R.P.B.; validation L.G., C.S.K., E.S.M., F.R., A.K., C.L., C.M. and R.P.B.; formal analysis L.G., C.S.K. and R.P.B.; investigation L.G., C.S.K., E.S.M., A.K., C.M., C.L. and R.P.B.; resources F.R., C.M. and C.L.; data curation F.R., C.L., C.M. and R.P.B.; writing—original draft preparation L.G. and C.S.K.; writing—review and editing E.S.M., F.R., A.K., C.L., C.M. and R.P.B.; visualization L.G. and C.S.K.; supervision R.P.B. and F.R.; project administration L.G. All authors have read and agreed to the published version of the manuscript.

Funding: This research received no external funding.

**Institutional Review Board Statement:** This study did not require ethical approval due to the retrospective nature of this study. Additionally, all imaging procedures were part of routine follow-up examinations. This study was conducted in accordance with the Declaration of Helsinki. [<sup>68</sup>Ga]Ga-DATA<sup>5m</sup>.SA.FAPi were administered in compliance with The German Medicinal Products Act (AMG § 13 2b) and in accordance with the responsible regulatory body.

Informed Consent Statement: Informed consent was obtained from all subjects involved in this study.

Data Availability Statement: The data presented in this study are available in the article.

**Acknowledgments:** We thank the medical personnel involved in scanning and data acquisition as well as the patients that underwent the imaging procedure.

**Conflicts of Interest:** L.G. and F.R. have filed a patient on [<sup>68</sup>Ga]Ga-DATA<sup>5m</sup>.SA.FAPi. The other authors declare no conflict of interest.

## References

- Sollini, M.; Kirienko, M.; Gelardi, F.; Fiz, F.; Gozzi, N.; Chiti, A. State-of-the-art of FAPI-PET imaging: A systematic review and meta-analysis. *Eur. J. Nucl. Med. Mol. Imaging* 2021, 48, 4396–4414. [CrossRef] [PubMed]
- 2. Li, M.; Younis, M.H.; Zhang, Y.; Cai, W.; Lan, X. Clinical summary of fibroblast activation protein inhibitor-based radiopharmaceuticals: Cancer and beyond. *Eur. J. Nucl Med. Mol. Imaging* **2022**, *49*, 2844–2868. [CrossRef] [PubMed]

- 3. Gilardi, L.; Airò Farulla, L.S.; Demirci, E.; Clerici, I.; Omodeo Salè, E.; Ceci, F. Imaging Cancer-Associated Fibroblasts (CAFs) with FAPi PET. *Biomedicines* **2022**, *10*, 523. [CrossRef] [PubMed]
- Fortunati, E.; Argalia, G.; Zanoni, L.; Fanti, S.; Ambrosini, V. New PET Radiotracers for the Imaging of Neuroendocrine Neoplasms. *Curr. Treat. Options Oncol.* 2022, 23, 703–720. [CrossRef] [PubMed]
- Te Beek, E.T.; Burggraaf, J.; Teunissen, J.J.M.; Vriens, D. Clinical pharmacology of radiotheranostics in oncology. *Clin. Pharmacol. Ther.* 2022. [CrossRef] [PubMed]
- Lindner, T.; Giesel, F.L.; Kratochwil, C.; Serfling, S.E. Radioligands Targeting Fibroblast Activation Protein (FAP). *Cancers* 2021, 13, 5744. [CrossRef]
- Kamali Zonouzi, S.; Pezeshki, P.S.; Razi, S.; Rezaei, N. Cancer-associated fibroblasts in colorectal cancer. *Clin. Transl. Oncol.* 2022, 24, 757–769. [CrossRef]
- Aertgeerts, K.; Levin, I.; Shi, L.; Snell, G.P.; Jennings, A.; Prasad, G.S.; Zhang, Y.; Kraus, M.L.; Salakian, S.; Sridhar, V.; et al. Structural and kinetic analysis of the substrate specificity of human fibroblast activation protein alpha. *J. Biol. Chem.* 2005, 280, 19441–19444. [CrossRef]
- Yadav, D.; Ballal, S.; Yadav, M.P.; Tripathi, M.; Roesch, F.; Bal, C. Evaluation of [<sup>68</sup>Ga]Ga-DATA-TOC for imaging of neuroendocrine tumours: Comparison with [<sup>68</sup>Ga]Ga-DOTA-NOC PET/CT. *Eur. J. Nucl. Med. Mol. Imaging* 2020, 47, 860–869. [CrossRef]
- 10. Puré, E. The road to integrative cancer therapies: Emergence of a tumor-associated fibroblast protease as a potential therapeutic target in cancer. *Expert Opin. Ther. Targets* **2009**, *13*, 967–973. [CrossRef]
- 11. Fitzgerald, A.A.; Weiner, L.M. The role of fibroblast activation protein in health and malignancy. *Cancer Metastasis Rev.* 2020, 39, 783–803. [CrossRef] [PubMed]
- 12. Zi, F.; He, J.; He, D.; Li, Y.; Yang, L.; Cai, Z. Fibroblast activation protein α in tumor microenvironment: Recent progression and implications (review). *Mol. Med. Rep.* **2015**, *11*, 3203–3211. [CrossRef] [PubMed]
- 13. Yu, D.M.T.; Yao, T.-W.; Chowdhury, S.; Nadvi, N.A.; Osborne, B.; Church, W.B.; McCaughan, G.W.; Gorrell, M.D. The dipeptidyl peptidase IV family in cancer and cell biology. *FEBS J.* **2010**, *277*, 1126–1144. [CrossRef]
- 14. Hamson, E.J.; Keane, F.M.; Tholen, S.; Schilling, O.; Gorrell, M.D. Understanding fibroblast activation protein (FAP): Substrates, activities, expression and targeting for cancer therapy. *Proteom.-Clin. Appl.* **2014**, *8*, 454–463. [CrossRef] [PubMed]
- Xu, W.; Meng, T.; Shang, Q.; Pang, Y.; Chen, H. Uncommon Metastases From Occult Breast Cancer Revealed by <sup>18</sup>F-FDG and <sup>68</sup>Ga-FAPI PET/CT. *Clin. Nucl. Med.* **2022**, *47*, 751–753. [CrossRef] [PubMed]
- Pang, Y.; Zhao, L.; Shang, Q.; Meng, T.; Zhao, L.; Feng, L.; Wang, S.; Guo, P.; Wu, X.; Lin, Q.; et al. Positron emission tomography and computed tomography with [<sup>68</sup>Ga]Ga-fibroblast activation protein inhibitors improves tumor detection and staging in patients with pancreatic cancer. *Eur. J. Nucl. Med. Mol. Imaging* 2022, 49, 1322–1337. [CrossRef]
- Shang, Q.; Zhao, L.; Pang, Y.; Meng, T.; Chen, H. Differentiation of Reactive Lymph Nodes and Tumor Metastatic Lymph Nodes With <sup>68</sup>Ga-FAPI PET/CT in a Patient With Squamous Cell Lung Cancer. *Clin. Nucl. Med.* **2022**, 47, 458–461. [CrossRef]
- Giesel, F.L.; Kratochwil, C.; Schlittenhardt, J.; Dendl, K.; Eiber, M.; Staudinger, F.; Kessler, L.; Fendler, W.P.; Lindner, T.; Koerber, S.A.; et al. Head-to-head intra-individual comparison of biodistribution and tumor uptake of (68)Ga-FAPI and (18)F-FDG PET/CT in cancer patients. *Eur. J. Nucl. Med. Mol. Imaging* 2021, 48, 4377–4385. [CrossRef]
- Can, C.; Gündoğan, C.; Güzel, Y.; Kaplan, I.; Kömek, H. <sup>68</sup>Ga-FAPI Uptake of Thyroiditis in a Patient With Breast Cancer. *Clin. Nucl. Med.* 2021, 46, 683–685. [CrossRef]
- Deng, M.; Chen, Y.; Cai, L. Comparison of <sup>68</sup>Ga-FAPI and <sup>18</sup>F-FDG PET/CT in the Imaging of Pancreatic Cancer with Liver Metastases. *Clin. Nucl. Med.* 2021, 46, 589–591. [CrossRef]
- Zhang, X.; Song, W.; Qin, C.; Liu, F.; Lan, X. Non-malignant findings of focal (68)Ga-FAPI-04 uptake in pancreas. Eur. J. Nucl. Med. Mol. Imaging. 2021, 48, 2635–2641. [CrossRef] [PubMed]
- Röhrich, M.; Naumann, P.; Giesel, F.L.; Choyke, P.L.; Staudinger, F.; Wefers, A.; Liew, D.P.; Kratochwil, C.; Rathke, H.; Liermann, J.; et al. Impact of (68)Ga-FAPI PET/CT Imaging on the Therapeutic Management of Primary and Recurrent Pancreatic Ductal Adenocarcinomas. *J. Nucl. Med.* 2021, 62, 779–786. [CrossRef] [PubMed]
- Luo, Y.; Pan, Q.; Yang, H.; Peng, L.; Zhang, W.; Li, F. Fibroblast Activation Protein-Targeted PET/CT with (68)Ga-FAPI for Imaging IgG4-Related Disease: Comparison to (18)F-FDG PET/CT. J. Nucl. Med. 2021, 62, 266–271. [CrossRef]
- Roth, K.S.; Voltin, C.A.; van-Heek, L.; Wegen, S.; Schomaecker, K.; Fischer, T.; Marnitz, S.; Drzezga, A.E.; Kobe, C. Dual-tracer PET/CT protocol with [<sup>18</sup>F]-FDG and [<sup>68</sup>Ga]Ga-FAPI-46 for cancer imaging—A proof of concept. J. Nucl. Med. 2022. [CrossRef]
- Backhaus, P.; Burg, M.C.; Roll, W.; Büther, F.; Breyholz, H.-J.; Weigel, S.; Heindel, W.; Pixberg, M.; Barth, P.; Tio, J.; et al. Simultaneous FAPI PET/MRI Targeting the Fibroblast-Activation Protein for Breast Cancer. *Radiology* 2022, 302, 39–47. [CrossRef] [PubMed]
- Röhrich, M.; Leitz, D.; Glatting, F.M.; Wefers, A.K.; Weinheimer, O.; Flechsig, P.; Kahn, N.; Mall, M.A.; Giesel, F.L.; Kratochwil, C.; et al. Fibroblast Activation Protein-Specific PET/CT Imaging in Fibrotic Interstitial Lung Diseases and Lung Cancer: A Translational Exploratory Study. J. Nucl. Med. 2022, 63, 127–133. [CrossRef]
- 27. Yang, T.; Ma, L.; Hou, H.; Gao, F.; Tao, W. FAPI PET/CT in the Diagnosis of Abdominal and Pelvic Tumors. *Front. Oncol.* 2021, 11, 797960. [CrossRef]
- Dendl, K.; Schlittenhardt, J.; Staudinger, F.; Kratochwil, C.; Altmann, A.; Haberkorn, U.; Giesel, F.L. The Role of Fibroblast Activation Protein Ligands in Oncologic PET Imaging. *PET Clin.* 2021, 16, 341–351. [CrossRef]

- Chen, Y.; Zheng, S.; Zhang, J.; Yao, S.; Miao, W. Pleural Metastasis of Papillary Thyroid Cancer Depicted by <sup>68</sup>Ga-FAPI PET/CT. *Clin. Nucl. Med.* 2022, 47, 467–468. [CrossRef]
- Fu, L.; Huang, S.; Wu, H.; Dong, Y.; Xie, F.; Wu, R.; Zhou, K.; Tang, G.; Zhou, W. Superiority of [<sup>68</sup>Ga]Ga-FAPI-04/[<sup>18</sup>F]FAPI-42 PET/CT to [<sup>18</sup>F]FDG PET/CT in delineating the primary tumor and peritoneal metastasis in initial gastric cancer. *Eur. Radiol.* 2022, 1–10.
- Imlimthan, S.; Moon, E.S.; Rathke, H.; Afshar-Oromieh, A.; Rösch, F.; Rominger, A.; Gourni, E. New Frontiers in Cancer Imaging and Therapy Based on Radiolabeled Fibroblast Activation Protein Inhibitors: A Rational Review and Current Progress. *Pharmaceuticals* 2021, 14, 1023. [CrossRef] [PubMed]
- Poplawski, S.E.; Lai, J.H.; Li, Y.; Jin, Z.; Liu, Y.; Wu, W.; Wu, Y.; Zhou, Y.; Sudmeier, J.L.; Sanford, D.G.; et al. Identification of selective and potent inhibitors of fibroblast activation protein and prolyl oligopeptidase. *J. Med. Chem.* 2013, 56, 3467–3477. [CrossRef] [PubMed]
- 33. Johnson, D.S.; Weerapana, E.; Cravatt, B.F. Strategies for discovering and derisking covalent, irreversible enzyme inhibitors. *Future Med. Chem.* **2010**, *2*, 949–964. [CrossRef] [PubMed]
- Fabre, M.; Ferrer, C.; Domínguez-Hormaetxe, S.; Bockorny, B.; Murias, L.; Seifert, O.; Eisler, S.A.; Kontermann, R.E.; Pfizenmaier, K.; Lee, S.Y.; et al. OMTX705, a Novel FAP-Targeting ADC Demonstrates Activity in Chemotherapy and Pembrolizumab-Resistant Solid Tumor Models. *Clin. Cancer Res.* 2020, 26, 3420–3430. [CrossRef]
- Van Rymenant, Y.; Tanc, M.; Elzen, R.; Bracke, A.; de wever, O.; Augustyns, K.; Lambeir, A.M.; Kockx, M.; De Meester, I.; Van Der Veken, P. In Vitro and In Situ Activity-Based Labeling of Fibroblast Activation Protein with UAMC1110-Derived Probes. *Front. Chem.* 2021, 9, 640566. [CrossRef]
- De Decker, A.; Vliegen, G.; Van Rompaey, D.; Peeraer, A.; Bracke, A.; Verckist, L.; Jansen, K.; Geiss-Friedlander, R.; Augustyns, K.; De Winter, H.; et al. Novel Small Molecule-Derived, Highly Selective Substrates for Fibroblast Activation Protein (FAP). ACS Med. Chem. Lett. 2019, 10, 1173–1179. [CrossRef]
- Gunderson, A.J.; Yamazaki, T.; Mccarty, K.; Phillips, M.; Alice, A.; Bambina, S.; Zebertavage, L.; Friedman, D.; Cottam, B.; Newell, P.; et al. Blockade of fibroblast activation protein in combination with radiation treatment in murine models of pancreatic adenocarcinoma. *PLoS ONE*. 2019, 14, e0211117. [CrossRef]
- Ryabtsova, O.; Jansen, K.; Van Goethem, S.; Joossens, J.; Cheng, J.D.; Lambeir, A.-M.; De Meester, I.; Augustyns, K.; Van der Veken, P. Acylated Gly-(2-cyano)pyrrolidines as inhibitors of fibroblast activation protein (FAP) and the issue of FAP/prolyl oligopeptidase (PREP)-selectivity. *Bioorganic Med. Chem. Lett.* 2012, 22, 3412–3417. [CrossRef]
- Loktev, A.; Lindner, T.; Mier, W.; Debus, J.; Altmann, A.; Jäger, D.; Giesel, F.; Kratochwil, C.; Barthe, P.; Roumestand, C.; et al. A Tumor-Imaging Method Targeting Cancer-Associated Fibroblasts. J. Nucl. Med. 2018, 59, 1423–1429. [CrossRef]
- 40. Jansen, K.; De Winter, H.; Heirbaut, L.; Cheng, J.D.; Joossens, J.; Lambeir, A.-M.; De Meester, I.; Augustyns, K.; Van der Veken, P. Selective inhibitors of fibroblast activation protein (FAP) with a xanthine scaffold. *MedChemComm* **2014**, *5*, 1700–1707. [CrossRef]
- Kratochwil, C.; Flechsig, P.; Lindner, T.; Abderrahim, L.; Altmann, A.; Mier, W.; Adeberg, S.; Rathke, H.; Röhrich, M.; Winter, H.; et al. <sup>68</sup>Ga-FAPI PET/CT: Tracer Uptake in 28 Different Kinds of Cancer. J. Nucl. Med. 2019, 60, 801–805. [CrossRef] [PubMed]
- Da Pieve, C.; Costa Braga, M.; Turton, D.R.; Valla, F.A.; Cakmak, P.; Plate, K.H.; Kramer-Marek, G. New Fully Automated Preparation of High Apparent Molar Activity (68)Ga-FAPI-46 on a Trasis AiO Platform. *Molecules* 2022, 27, 675. [CrossRef] [PubMed]
- 43. Zhao, L.; Niu, B.; Fang, J.; Pang, Y.; Li, S.; Xie, C.; Sun, L.; Zhang, X.; Guo, Z.; Lin, Q.; et al. Synthesis, preclinical evaluation, and a pilot clinical PET imaging study of (68)Ga-labeled FAPI dimer. *J. Nucl. Med.* **2021**, *63*, 862–868. [CrossRef]
- 44. Wei, Y.; Cheng, K.; Fu, Z.; Zheng, J.; Mu, Z.; Zhao, C.; Liu, X.; Wang, S.; Yu, J.; Yuan, S. [<sup>18</sup>F]AlF-NOTA-FAPI-04 PET/CT uptake in metastatic lesions on PET/CT imaging might distinguish different pathological types of lung cancer. *Eur. J. Nucl. Med. Mol. Imaging.* **2022**, *49*, 1671–1681. [CrossRef]
- Hu, K.; Li, L.; Huang, Y.; Ye, S.; Zhong, J.; Yan, Q.; Zhong, Y.; Fu, L.; Feng, P.; Li, H. Radiosynthesis and Preclinical Evaluation of Bispecific PSMA/FAP Heterodimers for Tumor Imaging. *Pharmaceuticals* 2022, 15, 383. [CrossRef] [PubMed]
- Li, H.; Ye, S.; Li, L.; Zhong, J.; Yan, Q.; Zhong, Y.; Feng, P.; Hu, K. <sup>18</sup>F- or <sup>177</sup>Lu-labeled bivalent ligand of fibroblast activation protein with high tumor uptake and retention. *Eur. J. Nucl. Med. Mol. Imaging* 2022, 49, 2705–2715. [CrossRef]
- Wei, Y.; Zheng, J.; Ma, L.; Liu, X.; Xu, S.; Wang, S.; Jinli Pei Kai Cheng Shuanghu, Y.u.a.n.; Jinming, Y.u. [<sup>18</sup>F]AIF-NOTA-FAPI-04: FAP-targeting specificity, biodistribution, and PET/CT imaging of various cancers. *Eur. J. Nucl. Med. Mol. Imaging* 2022, 49, 2761–2773. [CrossRef]
- Naka, S.; Watabe, T.; Lindner, T.; Cardinale, J.; Kurimoto, K.; Moore, M.; Tatsumi, M.; Mori, Y.; Shimosegawa, E.; Valla, F., Jr.; et al. One-pot and one-step automated radio-synthesis of [<sup>18</sup>F]AlF-FAPI-74 using a multi purpose synthesizer: A proof-of-concept experiment. *EJNMMI Radiopharm. Chem.* 2021, *6*, 28. [CrossRef]
- Lindner, T.; Altmann, A.; Giesel, F.; Kratochwil, C.; Kleist, C.; Krämer, S.; Mier, W.; Cardinale, J.; Kauczor, H.-U.; Jäger, D.; et al. (18)F-labeled tracers targeting fibroblast activation protein. *EJNMMI Radiopharm. Chem.* 2021, 6, 26. [CrossRef]
- Hu, K.; Li, J.; Wang, L.; Huang, Y.; Li, L.; Ye, S.; Han, Y.; Huang, S.; Wu, H.; Su, J.; et al. Preclinical evaluation and pilot clinical study of [<sup>18</sup>F]AlF-labeled FAPI-tracer for PET imaging of cancer associated fibroblasts. *Acta Pharm. Sin. B.* 2022, 12, 867–875. [CrossRef]
- Kou, Y.; Yao, Z.; Cheng, Z. Hepatic Lesion of Mucosa-Associated Lymphoid Tissue Lymphoma Revealed by Al<sup>18</sup>F-NOTA-FAPI-04 PET/CT. *Clin. Nucl. Med.* 2022, 47, e49–e51. [CrossRef] [PubMed]

- Kou, Y.; Yao, Z.; Cheng, Z. Al<sup>18</sup>F-NOTA-FAPI-04 Outperforms <sup>18</sup>F-FDG PET/CT in Identifying the Primary Lesion and Rare Metastases From Gastric Cancer. *Clin. Nucl. Med.* 2021, 46, e570–e571. [CrossRef]
- Wang, S.; Zhou, X.; Xu, X.; Ding, J.; Liu, S.; Hou, X.; Li, N.; Zhu, H.; Yang, Z. Clinical translational evaluation of Al(18)F-NOTA-FAPI for fibroblast activation protein-targeted tumour imaging. *Eur. J. Nucl. Med. Mol. Imaging* 2021, 48, 4259–4271. [CrossRef] [PubMed]
- Jiang, X.; Wang, X.; Shen, T.; Yao, Y.; Chen, M.; Li, Z.; Li, X.; Shen, J.; Kou, Y.; Chen, S.; et al. FAPI-04 PET/CT Using [<sup>18</sup>F]AlF Labeling Strategy: Automatic Synthesis, Quality Control, and In Vivo Assessment in Patient. *Front. Oncol.* 2021, 11, 649148. [CrossRef]
- Loktev, A.; Lindner, T.; Burger, E.-M.; Altmann, A.; Giesel, F.; Kratochwil, C.; Debus, J.; Marme, F.; Jäger, D.; Mier, W.; et al. Development of Fibroblast Activation Protein-Targeted Radiotracers with Improved Tumor Retention. *J. Nucl. Med.* 2019, 60, 1421–1429. [CrossRef] [PubMed]
- 56. Lindner, T.; Loktev, A.; Giesel, F.; Kratochwil, C.; Altmann, A.; Haberkorn, U. Targeting of activated fibroblasts for imaging and therapy. *EJNMMI Radiopharm. Chem.* **2019**, *4*, 16. [CrossRef]
- Lindner, T.; Loktev, A.; Altmann, A.; Giesel, F.; Kratochwil, C.; Debus, J.; Jäger, D.; Mier, W.; Haberkorn, U. Development of Quinoline-Based Theranostic Ligands for the Targeting of Fibroblast Activation Protein. *J. Nucl. Med.* 2018, 59, 1415–1422. [CrossRef]
- 58. Chen, X.; Song, E. Turning foes to friends: Targeting cancer-associated fibroblasts. *Nat. Rev. Drug Discov.* **2019**, *18*, 99–115. [CrossRef] [PubMed]
- Hu, K.; Wang, L.; Wu, H.; Huang, S.; Tian, Y.; Wang, Q.; Xiao, C.; Han, Y.; Tang, G. [<sup>18</sup>F]FAPI-42 PET imaging in cancer patients: Optimal acquisition time, biodistribution, and comparison with [<sup>68</sup>Ga]Ga-FAPI-04. *Eur. J Nucl. Med. Mol. Imaging* 2022, 49, 2833–2843. [CrossRef]
- 60. Watabe, T.; Liu, Y.; Kaneda-Nakashima, K.; Shirakami, Y.; Lindner, T.; Ooe, K.; Toyoshima, A.; Nagata, K.; Shimosegawa, E.; Haberkorn, U.; et al. Theranostics Targeting Fibroblast Activation Protein in the Tumor Stroma: (64)Cu- and (225)Ac-Labeled FAPI-04 in Pancreatic Cancer Xenograft Mouse Models. *J. Nucl. Med.* **2020**, *61*, 563–569. [CrossRef]
- Kratochwil, C.; Giesel, F.L.; Rathke, H.; Fink, R.; Dendl, K.; Debus, J.; Mier, W.; Jäger, D.; Lindner, T.; Haberkorn, U. [<sup>153</sup>Sm]Samarium-labeled FAPI-46 radioligand therapy in a patient with lung metastases of a sarcoma. *Eur. J. Nucl. Med. Mol. Imaging.* 2021, 48, 3011–3013. [CrossRef] [PubMed]
- 62. Rathke, H.; Fuxius, S.; Giesel, F.L.; Lindner, T.; Debus, J.; Haberkorn, U.; Kratochwil, C. Two Tumors, One Target: Preliminary Experience With 90Y-FAPI Therapy in a Patient With Metastasized Breast and Colorectal Cancer. *Clin. Nucl. Med.* **2021**, *46*, 842–844. [CrossRef] [PubMed]
- 63. Kaghazchi, F.; Aghdam, R.A.; Haghighi, S.; Vali, R.; Adinehpour, Z. 177Lu-FAPI Therapy in a Patient With End-Stage Metastatic Pancreatic Adenocarcinoma. *Clin. Nucl. Med.* 2022, 47, e243–e245. [CrossRef] [PubMed]
- 64. Xu, M.; Zhang, P.; Ding, J.; Chen, J.; Huo, L.; Liu, Z. Albumin Binder-Conjugated Fibroblast Activation Protein Inhibitor Radiopharmaceuticals for Cancer Therapy. J. Nucl. Med. 2021, 63, 952–958. [CrossRef]
- Liu, Y.; Watabe, T.; Kaneda-Nakashima, K.; Shirakami, Y.; Naka, S.; Ooe, K.; Toyoshima, A.; Nagata, K.; Haberkorn, U.; Kratochwil, C.; et al. Fibroblast activation protein targeted therapy using [<sup>177</sup>Lu]FAPI-46 compared with [<sup>225</sup>Ac]FAPI-46 in a pancreatic cancer model. *Eur. J. Nucl. Med. Mol. Imaging* 2022, 49, 871–880. [CrossRef]
- Kuyumcu, S.; Kovan, B.; Sanli, Y.; Buyukkaya, F.; Simsek, D.H.; Özkan, Z.G.; Isik, E.G.; Ekenel, M.; Turkmen, C. Safety of Fibroblast Activation Protein-Targeted Radionuclide Therapy by a Low-Dose Dosimetric Approach Using 177Lu-FAPI04. *Clin. Nucl. Med.* 2021, 46, 641–646. [CrossRef]
- Assadi, M.; Rekabpour, S.J.; Jafari, E.; Divband, G.; Nikkholgh, B.; Amini, H.; Kamali, H.; Ebrahimi, S.; Shakibazad, N.; Jokar, N.; et al. Feasibility and Therapeutic Potential of 177Lu-Fibroblast Activation Protein Inhibitor-46 for Patients With Relapsed or Refractory Cancers: A Preliminary Study. *Clin. Nucl. Med.* 2021, 46, e523–e530. [CrossRef]
- Baum, R.P.; Schuchardt, C.; Singh, A.; Chantadisai, M.; Robiller, F.C.; Zhang, J.; Mueller, D.; Eismant, A.; Almaguel, F.; Zboralski, D.; et al. Feasibility, Biodistribution, and Preliminary Dosimetry in Peptide-Targeted Radionuclide Therapy of Diverse Adenocarcinomas Using (177)Lu-FAP-2286: First-in-Humans Results. J. Nucl. Med. 2022, 63, 415–423. [CrossRef]
- Tsionou, M.I.; Knapp, C.E.; Foley, C.A.; Munteanu, C.R.; Cakebread, A.; Imberti, C.; Eykyn, T.R.; Young, J.D.; Paterson, B.M.; Blower, P.J.; et al. Comparison of macrocyclic and acyclic chelators for gallium-68 radiolabelling. *RSC Adv.* 2017, 7, 49586–49599. [CrossRef]
- 70. Fersing, C.; Masurier, N.; Rubira, L.; Deshayes, E.; Lisowski, V. AAZTA-Derived Chelators for the Design of Innovative Radiopharmaceuticals with Theranostic Applications. *Pharmaceuticals* **2022**, *15*, 234. [CrossRef]
- Tornesello, A.L.; Buonaguro, L.; Tornesello, M.L.; Buonaguro, F.M. New Insights in the Design of Bioactive Peptides and Chelating Agents for Imaging and Therapy in Oncology. *Molecules* 2017, 22, 1282. [CrossRef] [PubMed]
- Aime, S.; Calabi, L.; Cavallotti, C.; Gianolio, E.; Giovenzana, G.B.; Losi, P.; Maiocchi, A.; Palmisano, G.; Sisti, M. [Gd-AAZTA]-: A new structural entry for an improved generation of MRI contrast agents. *Inorg. Chem.* 2004, 43, 7588–7590. [CrossRef] [PubMed]
- Waldron, B.P.; Parker, D.; Burchardt, C.; Yufit, D.S.; Zimny, M.; Roesch, F. Structure and stability of hexadentate complexes of ligands based on AAZTA for efficient PET labelling with gallium-68. *Chem. Commun.* 2013, 49, 579–581. [CrossRef] [PubMed]

- 74. Moon, E.S.; Elvas, F.; Vliegen, G.; De Lombaerde, S.; Vangestel, C.; De Bruycker, S.; Bracke, A.; Eppard, E.; Greifenstein, L.; Klasen, B.; et al. Targeting fibroblast activation protein (FAP): Next generation PET radiotracers using squaramide coupled bifunctional DOTA and DATA(5m) chelators. *EJNMMI Radiopharm. Chem.* 2020, *5*, 19. [CrossRef] [PubMed]
- 75. Chasák, J.; Šlachtová, V.; Urban, M.; Brulíková, L. Squaric acid analogues in medicinal chemistry. *Eur. J. Med. Chem.* 2021, 209, 112872. [CrossRef]
- 76. Wurm, F.R.; Klok, H.A. Be squared: Expanding the horizon of squaric acid-mediated conjugations. *Chem. Soc. Rev.* 2013, 42, 8220–8236. [CrossRef]
- 77. Greifenstein, L.; Engelbogen, N.; Lahnif, H.; Sinnes, J.P.; Bergmann, R.; Bachmann, M.; Rösch, F. Synthesis, Labeling and Preclinical Evaluation of a Squaric Acid Containing PSMA Inhibitor Labeled with (68) Ga: A Comparison with PSMA-11 and PSMA-617. *ChemMedChem* 2020, 15, 695–704. [CrossRef]
- Greifenstein, L.; Grus, T.; Nagel, J.; Sinnes, J.P.; Rösch, F. Synthesis and labeling of a squaric acid containing PSMA-inhibitor coupled to AAZTA<sup>5</sup> for versatile labeling with <sup>44</sup>Sc, <sup>64</sup>Cu, <sup>68</sup>Ga and <sup>177</sup>Lu. *Appl. Radiat. Isot.* **2020**, 156, 108867. [CrossRef]
- 79. Gauger, J.; Manecke, G. Kondensationsprodukte der Quadratsäure mit primären und sekundären Aminen, II. *Chem. Berichte* **1970**, 103, 3553–3562. [CrossRef]
- Ballal, S.; Yadav, M.P.; Kramer, V.; Moon, E.S.; Roesch, F.; Tripathi, M.; Mallick, S.; ArunRaj, S.T.; Bal, C. A theranostic approach of [<sup>68</sup>Ga]Ga-DOTA.SA.FAPi PET/CT-guided [<sup>177</sup>Lu]Lu-DOTA.SA.FAPi radionuclide therapy in an end-stage breast cancer patient: New frontier in targeted radionuclide therapy. *Eur. J. Nucl. Med. Mol. Imaging.* **2021**, *48*, 942–944. [CrossRef]
- Ballal, S.; Yadav, M.P.; Moon, E.S.; Kramer, V.S.; Roesch, F.; Kumari, S.; Bal, C. First-In-Human Results on the Biodistribution, Pharmacokinetics, and Dosimetry of [<sup>177</sup>Lu]Lu-DOTA.SA.FAPi and [<sup>177</sup>Lu]Lu-DOTAGA.(SA.FAPi)(2). *Pharmaceuticals* 2021, 14, 1212. [CrossRef] [PubMed]
- Ballal, M.S.; Yadav, M.P.; Moon, E.S.; Roesch, F.; Kumari, M.S.; Agarwal, S.; Tripathi, M.; Sahoo, R.K.; Mangu, B.S.; Tupalli, A.; et al. Novel Fibroblast Activation Protein Inhibitor-Based Targeted Theranostics for Radioiodine-Refractory Differentiated Thyroid Cancer Patients: A Pilot Study. *Thyroid* 2022, 32, 65–77. [CrossRef] [PubMed]
- Moon, E.S.; Ballal, S.; Yadav, M.P.; Bal, C.; Van Rymenant, Y.; Stephan, S.; Bracke, A.; Van der Veken, P.; De Meester, I.; Roesch, F. Fibroblast Activation Protein (FAP) targeting homodimeric FAP inhibitor radiotheranostics: A step to improve tumor uptake and retention time. *Am. J. Nucl. Med. Mol. Imaging* 2021, *11*, 476–491. [PubMed]
- 84. Kreppel, B.; Gonzalez-Carmona, M.A.; Feldmann, G.; Küppers, J.; Moon, E.S.; Marinova, M.; Bundschuh, R.A.; Kristiansen, G.; Essler, M.; Roesch, F.; et al. Fibroblast activation protein inhibitor (FAPi) positive tumour fraction on PET/CT correlates with Ki-67 in liver metastases of neuroendocrine tumours. *Nuklearmedizin* **2021**, *60*, 344–354. [CrossRef]
- Kreppel, B.; Gärtner, F.C.; Marinova, M.; Attenberger, U.; Meisenheimer, M.; Toma, M.; Kristiansen, G.; Feldmann, G.; Moon, E.S.; Roesch, F.; et al. [<sup>68</sup>Ga]Ga-DATA5m.SA.FAPi PET/CT: Specific Tracer-uptake in Focal Nodular Hyperplasia and potential Role in Liver Tumor Imaging. Nuklearmedizin 2020, 59, 387–389. [CrossRef]
- 86. Glatting, F.M.; Hoppner, J.; Liew, D.P.; van Genabith, A.; Spektor, A.M.; Steinbach, L.; Hubert, A.; Kratochwil, C.; Giesel, F.L.; Dendl, K.; et al. Repetitive early FAPI-PET acquisition comparing FAPI-02, FAPI-46 and FAPI-74: Methodological and diagnostic implications for malignant, inflammatory and degenerative lesions. J. Nucl. Med. 2022, online ahead of print.
- Giesel, F.L.; Kratochwil, C.; Lindner, T.; Marschalek, M.M.; Loktev, A.; Lehnert, W.; Debus, J.; Jäger, D.; Flechsig, P.; Altmann, A.; et al. <sup>68</sup>Ga-FAPI PET/CT: Biodistribution and Preliminary Dosimetry Estimate of 2 DOTA-Containing FAP-Targeting Agents in Patients with Various Cancers. J. Nucl. Med. 2019, 60, 386–392. [CrossRef]