Rab25 GTPase: Functional roles in cancer

Sisi Wang¹, Chunhong Hu¹, Fang Wu¹ and Shasha He¹

¹Department of Oncology, The Second Xiangya Hospital, Central South University, Changsha, Hunan, China

Correspondence to: Shasha He, email: laishiweisha611@126.com

Keywords: Rab25, small GTPase, vesicle trafficking, ovarian cancer, renal cancer

Received: March 21, 2017 **Accepted:** July 19, 2017 **Published:** July 26, 2017

Copyright: Wang et al. This is an open-access article distributed under the terms of the Creative Commons Attribution License 3.0 (CC BY 3.0), which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

ABSTRACT

Rab25, a small GTPase belongs to the Rab protein family, has a pivotal role in cancer pathophysiology. Rab25 governs cell-surface receptors recycling and cellular signaling pathways activation, allowing it to control a diverse range of cellular functions, including cell proliferation, cell motility and cell death. Aberrant expression of Rab25 was linked to cancer development. Majority of research findings revealed that Rab25 is an oncogene. Elevated expression of Rab25 was correlated with poor prognosis and aggressiveness of renal, lung, breast, ovarian and other cancers. However, tumor suppressor function of Rab25 was reported in several cancers, such as colorectal cancer, indicating the tumor type-specific function of Rab25. In this review, we recapitulate the current knowledge of Rab25 in cancer development and therapy.

INTRODUCTION

Rab25 (Ras-related protein Rab-25) belongs to the Rab protein family. Until now, 70 Homo sapiens Rab proteins have been identified [1, 2] and they form the largest branch of the Ras superfamily [3]. Rabs are evolutionary conserved and their homologs are found in organisms ranging from yeast to human. Rabs are small GTPases involved in the control of vesicle trafficking. They govern correct transport of substances between different cellular compartments and have been implicated in various cellular functions, including cell proliferation, cell mobility, signal transduction and protein transport [4, 5]. Since they are key regulators of major cellular functions, some of them have been reported to be involved in tumorigenesis and cancer progression. For example, Rab5a and Rab7 were both up-regulated and have a higher degree of membrane association in autonomous thyroid adenomas [6].

Each Rab protein has different subcellular localization and controls specific membrane transport pathways [7–9]. Rab25 belongs to the Rab11 subfamily (Rab11a, Rab11b, and Rab25) that regulates apical transport and/or recycle of vesicles to the plasma membrane [10, 11]. Some Rab proteins are ubiquitously expressed in human tissues, whereas Rab25 is specifically expressed in epithelial cells. Rab25 functions either as

an oncogene or a tumor suppressor with a cancer typedependent manner. It was predominantly reported to have oncogenic function and elevated expression in cancer, including ovarian cancer [12, 13], breast cancer [12–14], renal cancer [15, 16], gastric cancer [17], liver cancer [18], non-small cell lung cancer [19], bladder cancer [20], glioblastoma multiforme [21] and prostate cancer [22]. Tumor suppressor function of Rab25 has also been found in several types of cancer, including colon cancer [23], head and neck cancer [24], esophageal squamous cell carcinoma [25] and oral and oropharyngeal squamous cell carcinoma [26]. The function of Rab25 in cancer is, therefore, multifactorial and tumor type-specific.

This review recapitulates our current knowledge of the role of Rab25 in cancer, including the involvement of Rab25 in cancer prognosis, cancer progression, functional mechanisms and future research directions.

Structure of Rab25

Structure and biological function of Rab25

Rab25 is a 23kDa small GTPase with general GTPase tertiary structure, a central barrel composed of a six-stranded b-sheet surrounded by a-helixs [27]. The control of vesicle trafficking is the major function of Rab proteins and was first reported by Salminen *et al.* [28]. The function of Rab25 is determined by two structural features, the guanine nucleotide binding motif and the carboxyl-

terminus region. Guanine nucleotide binding motif allows Rab25 to bind with GTP/GDP, which controls the activity of Rab25. The carboxyl-terminus of Rab25 contains CCXXX motif. Post-translational modification of the C-terminus determines the binding of Rab25 on specific vesicles and regulates the entire membrane trafficking process [4, 29].

Each Rab protein recognizes and controls the transportation of different membrane vesicles. The C-terminus of Rab proteins contain XXXCC, XXCCX, XCCXX, CCXXX or XXCXC motif. Prenylation of the two cysteine residues at the motif, together with the addition of geranylgeranyl groups to one or two cysteine residues near the C-terminus, is required for tight binding of Rab proteins to their specific vesicles [30–32]. Post-translational modifications of the CCXXX motif of Rab25 allow the protein to bind to apical recycling endosomes and regulate recycle of the vesicles to the apical plasma membrane [11].

The Rab proteins cycle between two states, the GTP-bound active state and the GDP-bound inactive state (Figure 1). The inactive Rab proteins located in the cytosol and are recognized by guanine nucleotide exchange factor (GEF), which catalyzes the exchange of GDP for GTP at the guanine nucleotide binding motif to form the active Rab proteins [33]. After coordinating the transport of vesicles to their destination, the active Rab proteins are converted back to the GDP-bound inactive form through hydrolysis of GTP, which is stimulated by a GTPase-activating protein (GAP) [34].

The active GTP-bound Rab protein coordinates the entire 5-step vesicle trafficking process, including vesicle sorting, uncoating, motility, tethering and fusion, by recruiting effector proteins for distinct membrane trafficking steps [35]. GTP-bound Rab proteins activate sorting adaptor to sort a receptor into a budding vesicle [36]. The active Rab proteins promote the vesicle uncoating through the recruitment of phosphoinostide (PI) kinases or phosphatases [37]. After that, the Rab proteins recruit motor adaptors to enhance vesicle transport along the cytoskeletal tracts (actin filaments or microtubules) [38]. When the vesicle cargos are transported to the destination membrane, Rab proteins mediate vesicle tethering by recruiting tethering factors that interact with molecules on the acceptor membrane. Those factors promote membrane fusion through their interaction with soluble N-ethylmaleimide-sensitive factor attachment protein receptors (SNAREs) [39].

Precise control of endocytic and exocytic trafficking of receptors and their ligands is essential in the regulation of cellular signaling pathways, which impact gene expression and biological functions. Since Rab25 is involved in the control of vesicle trafficking and signaling pathways, it is frequently reported to have altered expression in cancer [40, 41]. Rab25 associated protein, FIP1C, was also reported to alter cancer progression [42]. Rab25 coordinated the recycling of α 5 β 1 integrin vesicles and the activation of various signaling pathways.

Down-stream effectors of Rab25

Current evidence suggested that Rab25 is predominantly an oncogene. It promotes oncogenic functions through the regulation of integrin recycling and intracellular signaling pathways (Figure 2). Direct interaction between Rab25 and $\alpha 5\beta 1$ integrin was linked to increase tumor cell metastasis and aggressiveness [43]. Modified Rab25, which does not interact with β 1 integrin, did not promote cell invasion [43]. Active recycling of α 5 β 1 integrin containing vesicle to the plasma membrane in the presence of Rab25 strongly promotes cell migration/ invasion through 3D matrices [43]. Rab25 contributes to tumor progression by directing the localization of integrin-recycling vesicles and thereby enhancing the ability of tumor cells to invade the extracellular matrix. Nevertheless, it was found that Rab25 regulates integrin recycling no matter when it acts as an oncogene or a tumor suppressor. In the study of colorectal cancer, Rab25 acts as a tumor suppressor gene. Rab25-deficient mice have more tumor formation and less β 1 integrin recycled to the lateral membrane [44]. However, whether suppression of β 1 integrin recycling contributed to reduction of tumor formation in Rab25-deficient mice and the mechanisms involved have to be further investigated.

Rab25 activates intracellular signaling pathways, including Akt, Wnt and Src pathways and suppresses apoptotic pathway. Ectopic overexpression of Rab25 increased phospho-Akt level, which may mediate the cell proliferation-stimulating effect of Rab25 on ovarian cancer cells [12]. Rab25 knockdown reduced phospho-Akt level, leading to the reduction of cell migration/invasion in bladder cancer cell [20], hepatocellular carcinoma cells [45] and glioblastoma multiforme cells [21]. Rab25 knockdown or Akt inhibitor was found to reduce cisplatin resistance in ovarian cancer cells [46]. Rab25 was reported to activate Wnt pathway. Depletion of Rab25 inhibited the expressions of Wnt pathway target genes, including cyclin D1, c-Myc and MMP7 in hepatocellular carcinoma [45]. Rab25 promoted the expression of CLIC3, which in turn activated Src signaling pathway [47]. Ectopic overexpression of Rab25 induced phospho-ERK1/2, which is one of the downstream signaling molecules of Src, and may lead to cisplatin resistance of ovarian cancer [48]. Knockdown of Rab25 reduced phospho-ERK1/2 level and promoted cell proliferation in breast cancer [49]. Moreover, Rab25 suppressed apoptotic cell death by reducing the expression of pro-apoptotic molecules, BAX and BAK, in ovarian cancer [12]. Knockdown of Rab25 was found to reduce anti-apoptotic protein, Bcl-2, in tobacco carcinogeninduced lung cancer [50]. Taken together, activation of growth-stimulating signaling pathways and suppression of apoptotic cell death pathways mediate the effect of Rab25 on enhancing cancer progression.

Up-stream regulators of Rab25

Altered expression of Rab25 is commonly observed in various types of cancer. However, the mechanisms leading to Rab25 expression alteration are not fully understood. Recently, altered Rab25 expression was reported to be associated with changes in microRNA expression, Rab25 gene copy number and epigenetic regulation.

Elevated expression of several Rabs is due to downregulation of miRNAs. For example, up-regulation of Rab34 in gastric cancer is related to decreased expression of miR-9 [51]. Increase of a series of Rab proteins, including Rab21, Rab23, Rab18 and Rab3B, is due to reduction of miR-200b in breast cancer [52]. Li *et al.* has showed that Rab25 is a direct target of let-7d, which is a down-regulated miRNA in renal cell carcinoma tissues as compared with the corresponding non-tumor tissues [16].

Copy number amplification is another cause of Rab25 up-regulation. Rab25 is encoded by *RAB25* gene located at chromosome 1q22. The minimum region of recurrent amplification at the 1q22 amplicon containing *RAB25* was found in 54% of advanced serous epithelial

ovarian cancers by high-resolution array comparative genomic hybridization profiling [12]. Analysis of 21 epithelial ovarian cancers showed a direct relationship between copy number and expression of Rab25, indicating mRNA overexpression of Rab25 is likely due to amplification of *RAB25* gene [12].

The expression of Rab25 has been reported to be controlled by epigenetic regulation. Rab25, an oncogene of ovarian cancer, shows a negative correlation between promoter methylation and mRNA expression [53]. In the study of ovarian cancer TCGA data set, decrease in Rab25 promoter methylation was associated with increase in Rab25 mRNA level, suggesting the elevation of Rab25 level by epigenetic regulation [53]. DNA methylation inhibitor was reported to induce Rab25 expression. The induction effect was further increased by co-treatment of DNA methylation inhibitor and histone deacetylases inhibitor in breast cancer cell [54] and lung squamous adenocarcinoma cells [55], proofing the elevation of Rab25 level by promoter demethylation and acetylation. Epigenetic regulation also led to down-regulation of Rab25 in oral and oropharyngeal squamous cell carcinoma (OOSCC) and esophageal squamous cell carcinoma (ESCC), in which Rab25 acts as a tumor suppressor.

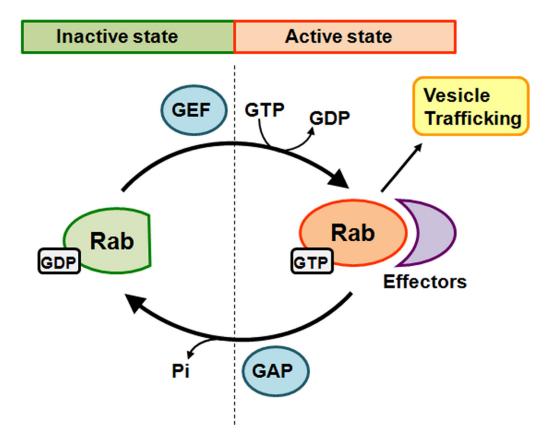


Figure 1: Schematic diagram of Rab GTPase cycle. Rabs switch between two conformations, an inactive GDP-bound form and an active GTP-bound form. A guanine nucleotide exchange factor (GEF) catalyzes the conversion from GDP-bound to GTP-bound and leads to activation of Rab. Activated Rab binds with specific effectors to promote vesicle trafficking. The inherent GTP hydrolysis of Rabs together with the enhancing effect of GTPase-activating protein (GAP) leads to Rab inactivation. Conversion of Rabs to the inactive state leads to effector dissociation from the Rab protein.

Methylation status of Rab25 promoter was significantly higher and correlated with lower Rab25 mRNA levels in OOSCC by analyzing 147 OOSCC samples in TCGA database [26]. In ESCC, demethylation treatment and bisulfite genomic sequencing analyses revealed that down-regulation of Rab25 expression in both ESCC cell lines and clinical samples was associated with promoter hypermethylation [25]. Up till now, the cause of hypermethylation of Rab25 promoter when Rab25 acts as a tumor suppressor, and vice versa, remains elusive. Further investigation is needed to understand the epigenetic regulation mechanisms of Rab25.

Function of Rab25

Oncogenic function of Rab25

Rab25 is recognized to play a crucial role in tumorigenesis and cancer progression [56]. Rab25 was reported to initiate cancer formation, promote cell proliferation, suppress apoptotic cell death, enhance cell migration/invasion and increase drug resistance upon chemotherapy of various types of cancer [40]. The oncogenic function of Rab25 is likely due to its function on regulating vesicle trafficking. Rab25 increases integrin recycling to the plasma membrane and stimulating intracellular signaling pathways, which in turn regulates the oncogenic functions [41].

Tumorigenesis

Recent reports indicated that Rab25 is related to transformed phenotype of neoplastic cells. Overexpression of Rab25 in non-transformed rat intestinal epithelial cells led to morphological and physiological transformation according to Lapierre *et al.*'s report [57]. The transformed cells changed from circular to more spindle-shaped. They were found to grow in soft agar and formed tumors in immunocompromised mice. The transformed cells also demonstrated disruption of integrin-based focal adhesions and alteration in modified microtubule subsets, indicating the transformed phenotype is microtubule-dependent. Treatment with nocodazole, a microtubule polymerization inhibitor, reversed the transformed morphology and internalization of $\alpha 5\beta$ 1-integrin [57].

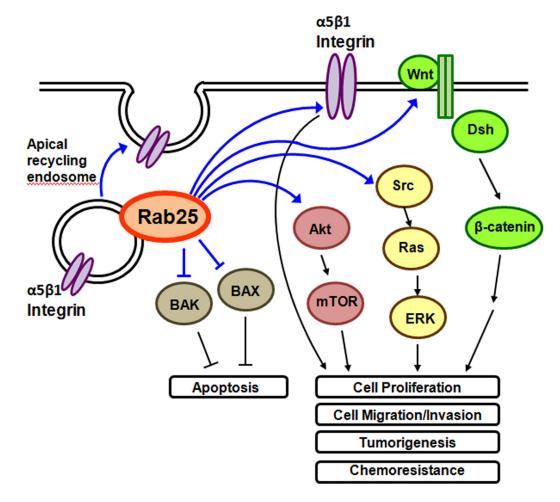


Figure 2: Overview of Rab25 functions in the cells. Rab25 regulates apical recycling of α 5 β 1 integrin containing endosome. Suppressive effect of Rab25 on BAX and BAK leads to reduction of apoptotic cell death. Direct Rab25 / α 5 β 1 integrin interaction and Rab25 induced activation of Akt, Src and Wnt pathways are associated with cell growth, metastasis and chemoresistance of cancer cells.

Cell growth/death and tumor development

It has been demonstrated that Rab25 enhances cell proliferation and tumor development. Knockdown of Rab25 expression by siRNA / shRNA transfection inhibited in vitro cell growth of renal cell carcinoma cells (786-O and A-498) [16], prostate cancer cells (LNCaP) [22] and hepatocellular carcinoma cells (Bel7402 and SK-Hep-1) [45]. Suppression of both in vitro cell growth and in vivo xenograft development were observed after knockdown of Rab25 in glioblastoma multiforme cells (U87MG) [21] and breast cancer cells (MCF7) [58]. Knockdown of Rab25 was reported to inhibit tumor growth in tobacco carcinogen-induced lung cancer model [50]. Conversely, ectopic overexpression of Rab25 was found to induce in vitro cell growth of breast cancer cells (MCF7) [12, 58] and ovarian cancer cells (A2780, DOV13, HEY, OCC1) [12]. Rab25 ectopic overexpression was found to increase development of xenograft derived from breast cancer cells (MCF7) [58] and ovarian cancer cells (A2780, HEY) [12, 59].

In addition to promote cell growth, Rab25 inhibited apoptotic and autophagic cell death. Elevated expression of Rab25 decreased UV-induced apoptosis in ovarian cancer [12]. Knockdown of Rab25 was reported to increase apoptosis in ovarian cancer cells [60] and tobacco carcinogen-induced lung cancer model [50]. Moreover, Rab25 down-regulation was reported to induce autophagic cell death in ovarian cancer from two independent research studies [60, 61]. The ability of Rab25 in cell growth stimulation and cell death suppression implicates the important role of Rab25 in supporting tumor growth.

Cell migration/invasion and tumor metastasis

Current reports indicated the involvement of Rab25 in promoting cell migration and invasion. Knockdown of Rab25 suppressed in vitro cell migration and cell invasion in renal cell carcinoma cells [16], advanced non-small cell lung cancer cells [19], prostate cancer cells [22] and glioblastoma multiforme cells [21]. Suppression of cell invasion after Rab25 knockdown was observed in gastric cancer cells [17]. The finding was further supported by Geng et al.'s study, which showed that Rab25 depletion negatively regulated the invasion ability of hepatocellular carcinoma cells [45]. In the study of bladder cancer, Rab25 knockdown not only suppress in vitro cell migration but also reduce in vivo tumor metastasis [20]. Tail vein injection of Rab25 knockdown bladder cancer cells (EJ, T24) into immune-compromised mice has resulted in less tumor nodule formation in lung compared with the mice injected with parental bladder cancer cells [20]. Rab25 is, therefore, important in determining the metastatic ability of cancer cells.

Chemoresistance

Rab25 induces chemoresistance towards cisplatin, a first-line chemotherapeutic agent, commonly used in the treatment of patients with various types of cancer, such as advanced non-small cell lung cancer patients [62, 63], metastatic breast cancer [64] and ovarian cancer [65]. However, cisplatin resistance limits the efficacy of chemotherapy in cancer patients [66, 67]. Rab25 expression level was significantly higher in cisplatin-resistance A549 NSCLC cells when compared with the cisplatin-sensitive A549 cells [19]. Knockdown of Rab25 by siRNA transfection decreased cisplatin resistant of NSCLC cells [19] and ovarian cancer cells [46]. On the contrary, ectopic overexpression of Rab25 in A2780 ovarian cancer cells increased cisplatin resistance in *in vitro* cell culture [48] and *in vivo* i.p. tumor xenograft [59], indicating the functional role of Rab25 in inducing cisplatin resistance in conventional chemotherapy.

Tumor suppressor function of Rab25

Rab25 has tumor suppressor function in several types of cancer, including colorectal cancer, esophageal squamous cell carcinoma and head and neck squamous cell carcinoma [23-25]. Loss of Rab25 in human colon cancers was associated with poorer patient prognosis [44]. Rab25 deficiency promotes intestinal/colon adenoma formation in ApcMin/+ mice [44]. The result was further supported by another Rab25 deficiency study in colon cancer [68]. Rab25 knockdown Caco2-BBE colorectal cancer cells demonstrated twofold increase in the number of soft agar colonies formation and a significant increase in colony size than the control cell line [68]. Ectopic overexpression of Rab25 in esophageal squamous cell carcinoma inhibited in vivo xenograft tumor development and angiogenesis [25]. In the study of head and neck squamous cell carcinoma, Rab25 ectopic overexpression reduced in vitro cell invasion and in vivo tumor metastasis to cervical lymph node [24].

Dual character of Rab25 in cancer

Rab25 has been reported to both enhance and suppress cancer progression. The role of Rab25 is cancer type-dependent. However, the mechanisms that lead to the discrepancies of Rab25 function remain elusive. One of the hypotheses suggested that the role of Rab25 depends on its ability to induce chloride intracellular channel protein 3 (CLIC3), which is involved in α 5 β 1 integrin trafficking necessary for cancer cell invasion [47]. Pancreatic cancer patients with high Rab25 and high CLIC3 levels were associated with significantly shorter survival time. Conversely, patients with high Rab25 level and low CLIC3 levels predicted better clinical outcomes [47]. It is suggested that in the presence of CLIC3, Rab25 acts as an oncogene, whereas, in the absence of CLIC3, Rab25 acts as a tumor suppressor. When Rab25 acts as an oncogene, Rab25 enhances $\alpha 5\beta 1$ integrin recycling to the plasma membrane, leading to increase in cancer progression of ovarian cancer [43]. When Rab25 acts as a tumor suppressor, Rab25-deficiency reduced \u00df1 integrin recycling to the plasma membrane and decreased tumor formation in colorectal adenocarcinomas [44].

Future research direction

Clinical implication of Rab25 in cancer

A large majority of clinicopathological findings indicate the oncogenic role of Rab25 in cancer. Elevated expression of Rab25 was significantly associated with shorter survival time of patients with bladder cancer [20], advanced non-small cell lung cancer [19], ovarian cancer [12], breast cancer [12, 49], clear cell renal cell carcinoma [15] and prostate cancer [22]. It is indicated that Rab25 is a potential prognostic marker for patients with various types of cancer.

Rab25 is up-regulated among the 52 Rab GTPases analyzed in renal cell carcinoma (RCC) according to Li *et al.*'s study [16]. High Rab25 expression level was significantly correlated with high invasion classification, lymph-node metastasis and pathological stage in RCC [16]. Moreover, high level of Rab25 expression was associated with lymph node/distant metastasis in bladder cancer [20], prostate cancer [22], gastric cancer [17] and hepatocellular carcinoma [45], suggesting the involvement of Rab25 in promoting tumor metastasis. The role or Rab25 in promoting cancer metastasis is confirmed by *in vitro* experiments in which knockdown of Rab25 decreased cell migration and invasion of 786-O and A-498 RCC cells [16].

Rab25 expression was associated with response rate of cisplatin-based chemotherapy of advanced non-small cell lung cancer (NSCLC) patients. The cisplatin-based chemotherapy response rate of NSCLC patients with Rab25-positive expression was 30%, as opposed to 52% for patients with Rab25-negative expression [19].

Elevated expression of Rab25 may be due to amplification of *RAB25* gene located in chromosome 1q22. Amplification of *RAB25* was reported to associate with markedly decreased disease-free survival or overall survival in ovarian cancer in Cheng *et al.*'s study [12]. Their report indicated an increase (at least 1.3-fold) in DNA copy number in a 1.1-Mb region located on chromosome 1q22 (which contain *RAB25* gene) in 28 of 52 (54%) of advanced serous epithelial ovarian cancers. Ovarian cancer patients with elevated *RAB25* amplification either did not enter a disease free state following surgery and chemotherapy or showed very short disease-free survival, implicating *RAB25* as potential driver gene correlated with poor prognosis and aggressive behavior of ovarian cancer [12].

Rab25 is predominantly an oncogene, but it also acts as tumor suppressor in certain types of cancer. Lower expression of Rab25 correlated with significantly shorter survival time of patients with colorectal adenocarcinomas [44] and esophageal squamous cell carcinoma (ESCC) [25]. In the study of oral and oropharyngeal squamous cell carcinoma (OOSCC), lower Rab25 expression was associated with lymph node metastasis status [26].

Conclusion and future research direction

Rab25 GTPase regulates crucial biological functions and is important in maintaining proper function of cells. Biological and clinicopathological findings revealed that alteration of Rab25 level has high impact on cancer progression and patient survival. Rab25 may act as an oncogene or a tumor suppressor.

Rab25 overexpression and gene copy number amplification was reported in various types of cancer. Elevated expression of Rab25 correlated with poor prognosis and tumor metastatic potential of cancer patients, suggesting Rab25 is a potential marker for cancer progression. Rab25 enhances cell growth, cell migration/invasion, chemoresistance and in vivo tumor growth. The mechanisms for Rab25 to regulate oncogenic functions involved the control of extracellular receptors recycling and intracellular signaling pathways, including Akt, Wnt, Src and apoptotic pathways. Further investigation on the mechanisms for Rab25 mediated oncogenic functions is important for the development of novel therapeutic strategies or inhibitors targeting Rab25. Moreover, it is important to identify the mechanisms that lead to copy number amplification of RAB25 in order to find out the potential methods for cancer prevention.

The function of Rab25 differs greatly in different types of cancer. The mechanisms that lead to the tumor suppressing function of Rab25 have not been fully understood. CLIC3 expression level was reported to be an important factor in determining the function of Rab25. In pancreatic cancer, Rab25 acts as an oncogene in the presence of CLIC3, whereas, Rab25 acts as a tumor suppressor in the absence of CLIC3 [47]. The correlation of Rab25/CLIC3 expression and clinical outcome in different types of cancer has to be investigated. In addition, the downstream mechanisms activated by Rab25 when it acts as an oncogene or a tumor suppressor are not yet clear. When Rab25 acts as an oncogene, Rab25 enhances $\alpha 5\beta 1$ integrin recycling to the plasma membrane, leading to increase in cancer progression of ovarian cancer [43]. When Rab25 acts as a tumor suppressor, Rab25-deficiency reduced B1 integrin recycling to the plasma membrane and decreased tumor formation in colorectal adenocarcinomas [44]. In-depth study is needed to find out the downstream effectors that mediate the distinct function of Rab25 in cancer.

Taken together, better understanding of Rab25 function under different circumstances, together with development of Rab25 specific targeting agents, will be required for development of novel therapeutic strategy targeting Rab25 in cancer treatment.

CONFLICTS OF INTEREST

None.

REFERENCES

- 1. Pereira-Leal JB, Seabra MC. The mammalian Rab family of small GTPases: definition of family and subfamily sequence motifs suggests a mechanism for functional specificity in the Ras superfamily. J Mol Biol. 2000; 301:1077–87.
- 2. Colicelli J. Human RAS superfamily proteins and related GTPases. Sci STKE. 2004; 2004:RE13.
- Schwartz SL, Cao C, Pylypenko O, Rak A, Wandinger-Ness A. Rab GTPases at a glance. J Cell Sci. 2007; 120:3905–10.
- 4. Bhuin T, Roy JK. Rab proteins: the key regulators of intracellular vesicle transport. Exp Cell Res. 2014; 328:1–19.
- 5. Chavrier P, Goud B. The role of ARF and Rab GTPases in membrane transport. Curr Opin Cell Biol. 1999; 11:466–75.
- Croizet-Berger K, Daumerie C, Couvreur M, Courtoy PJ, van den Hove MF. The endocytic catalysts, Rab5a and Rab7, are tandem regulators of thyroid hormone production. Proc Natl Acad Sci U S A. 2002; 99:8277–82.
- 7. Ferro-Novick S, Novick P. The role of GTP-binding proteins in transport along the exocytic pathway. Annu Rev Cell Biol. 1993; 9:575–99.
- 8. Novick P, Zerial M. The diversity of Rab proteins in vesicle transport. Curr Opin Cell Biol. 1997; 9:496–504.
- 9. Zerial M, Stenmark H. Rab GTPases in vesicular transport. Curr Opin Cell Biol. 1993; 5:613–20.
- 10. Zerial M, McBride H. Rab proteins as membrane organizers. Nat Rev Mol Cell Biol. 2001; 2:107–17.
- Casanova JE, Wang X, Kumar R, Bhartur SG, Navarre J, Woodrum JE, Altschuler Y, Ray GS, Goldenring JR. Association of Rab25 and Rab11a with the apical recycling system of polarized Madin-Darby canine kidney cells. Mol Biol Cell. 1999; 10:47–61.
- Cheng KW, Lahad JP, Kuo WL, Lapuk A, Yamada K, Auersperg N, Liu J, Smith-McCune K, Lu KH, Fishman D, Gray JW, Mills GB. The RAB25 small GTPase determines aggressiveness of ovarian and breast cancers. Nat Med. 2004; 10:1251–6.
- Cheng KW, Lu Y, Mills GB. Assay of Rab25 function in ovarian and breast cancers. Methods Enzymol. 2005; 403:202–15.
- Yin YX, Shen F, Pei H, Ding Y, Zhao H, Zhao M, Chen Q. Increased expression of Rab25 in breast cancer correlates with lymphatic metastasis. Tumour Biol. 2012; 33:1581–7.
- Liu L, Ding G. Rab25 expression predicts poor prognosis in clear cell renal cell carcinoma. Exp Ther Med. 2014; 8:1055–8.
- Li Y, Jia Q, Zhang Q, Wan Y. Rab25 upregulation correlates with the proliferation, migration, and invasion of renal cell carcinoma. Biochem Biophys Res Commun. 2015; 458:745–50.
- Cao C, Lu C, Xu J, Zhang J, Li M. Expression of Rab25 correlates with the invasion and metastasis of gastric cancer. Chin J Cancer Res. 2013; 25:192–9.

- He H, Dai F, Yu L, She X, Zhao Y, Jiang J, Chen X, Zhao S. Identification and characterization of nine novel human small GTPases showing variable expressions in liver cancer tissues. Gene Expr. 2002; 10:231–42.
- Ma YF, Yang B, Li J, Zhang T, Guo JT, Chen L, Li M, Chu J, Liang CY, Liu Y. Expression of Ras-related protein 25 predicts chemotherapy resistance and prognosis in advanced non-small cell lung cancer. Genet Mol Res. 2015; 14:13998–4008.
- 20. Zhang J, Wei J, Lu J, Tong Z, Liao B, Yu B, Zheng F, Huang X, Chen Z, Fang Y, Li B, Chen W, Xie D, et al. Overexpression of Rab25 contributes to metastasis of bladder cancer through induction of epithelial-mesenchymal transition and activation of Akt/GSK-3β/Snail signaling. Carcinogenesis. 2013; 34:2401–8.
- Ding B, Cui B, Gao M, Li Z, Xu C, Fan S, He W. Knockdown of Ras-Related Protein 25 (Rab25) Inhibits the *In Vitro* Cytotoxicity and *In Vivo* Antitumor Activity of Human Glioblastoma Multiforme Cells. Oncol Res. 2017; 25:331–40.
- 22. Hu C, Chen B, Zhou Y, Shan Y. High expression of Rab25 contributes to malignant phenotypes and biochemical recurrence in patients with prostate cancer after radical prostatectomy. Cancer Cell Int. 2017; 17:45.
- 23. Goldenring JR, Nam KT. Rab25 as a tumour suppressor in colon carcinogenesis. Br J Cancer. 2011; 104:33–6.
- Amornphimoltham P, Rechache K, Thompson J, Masedunskas A, Leelahavanichkul K, Patel V, Molinolo A, Gutkind JS, Weigert R. Rab25 regulates invasion and metastasis in head and neck cancer. Clin Cancer Res. 2013; 19:1375–88.
- 25. Tong M, Chan KW, Bao JY, Wong KY, Chen JN, Kwan PS, Tang KH, Fu L, Qin YR, Lok S, Guan XY, Ma S. Rab25 is a tumor suppressor gene with antiangiogenic and antiinvasive activities in esophageal squamous cell carcinoma. Cancer Res. 2012; 72:6024–35.
- 26. Clausen MJ, Melchers LJ, Mastik MF, Slagter-Menkema L, Groen HJ, Laan BF, van Criekinge W, de Meyer T, Denil S, van der Vegt B, Wisman GB6, Roodenburg JL, Schuuring E. RAB25 expression is epigenetically downregulated in oral and oropharyngeal squamous cell carcinoma with lymph node metastasis. Epigenetics. 2016; 11:653–63.
- Scapin SM, Carneiro FR, Alves AC, Medrano FJ, Guimarães BG, Zanchin NI. The crystal structure of the small GTPase Rab11b reveals critical differences relative to the Rab11a isoform. J Struct Biol. 2006; 154:260–8.
- Salminen A, Novick PJ. A ras-like protein is required for a post-Golgi event in yeast secretion. Cell. 1987; 49:527–38.
- 29. Fukuda M. Regulation of secretory vesicle traffic by Rab small GTPases. Cell Mol Life Sci. 2008; 65:2801–13.
- Brennwald P, Novick P. Interactions of three domains distinguishing the Ras-related GTP-binding proteins Ypt1 and Sec4. Nature. 1993; 362:560–3.
- Stenmark H, Valencia A, Martinez O, Ullrich O, Goud B, Zerial M. Distinct structural elements of rab5 define its functional specificity. EMBO J. 1994; 13:575–83.

- Iakovenko A, Rostkova E, Merzlyak E, Hillebrand AM, Thomä NH, Goody RS, Alexandrov K. Semi-synthetic Rab proteins as tools for studying intermolecular interactions. FEBS Lett. 2000; 468:155–8.
- 33. Eathiraj S, Pan X, Ritacco C, Lambright DG. Structural basis of family-wide Rab GTPase recognition by rabenosyn-5. Nature. 2005; 436:415–9.
- 34. Haas AK, Yoshimura S, Stephens DJ, Preisinger C, Fuchs E, Barr FA. Analysis of GTPase-activating proteins: Rab1 and Rab43 are key Rabs required to maintain a functional Golgi complex in human cells. J Cell Sci. 2007; 120:2997–3010.
- Stenmark H. Rab GTPases as coordinators of vesicle traffic. Nat Rev Mol Cell Biol. 2009; 10:513–25.
- 36. Cremona O, Di Paolo G, Wenk MR, Lüthi A, Kim WT, Takei K, Daniell L, Nemoto Y, Shears SB, Flavell RA, McCormick DA, De Camilli P. Essential role of phosphoinositide metabolism in synaptic vesicle recycling. Cell. 1999; 99:179–88.
- 37. Semerdjieva S, Shortt B, Maxwell E, Singh S, Fonarev P, Hansen J, Schiavo G, Grant BD, Smythe E. Coordinated regulation of AP2 uncoating from clathrin-coated vesicles by rab5 and hRME-6. J Cell Biol. 2008; 183:499–511.
- 38. Seabra MC, Coudrier E. Rab GTPases and myosin motors in organelle motility. Traffic. 2004; 5:393–9.
- Ohya T, Miaczynska M, Coskun U, Lommer B, Runge A, Drechsel D, Kalaidzidis Y, Zerial M. Reconstitution of Rab- and SNARE-dependent membrane fusion by synthetic endosomes. Nature. 2009; 459:1091–7.
- 40. Mitra S, Cheng KW, Mills GB. Rab25 in cancer: a brief update. Biochem Soc Trans. 2012; 40:1404–8.
- 41. Agarwal R, Jurisica I, Mills GB, Cheng KW. The emerging role of the RAB25 small GTPase in cancer. Traffic. 2009; 10:1561–8.
- 42. Boulay PL, Mitchell L, Turpin J, Huot-Marchand JE, Lavoie C, Sanguin-Gendreau V, Jones L, Mitra S, Livingstone JM, Campbell S, Hallett M, Mills GB, Park M, et al. Rab11-FIP1C Is a Critical Negative Regulator in ErbB2-Mediated Mammary Tumor Progression. Cancer Res. 2016; 76:2662–74.
- 43. Caswell PT, Spence HJ, Parsons M, White DP, Clark K, Cheng KW, Mills GB, Humphries MJ, Messent AJ, Anderson KI, McCaffrey MW, Ozanne BW. Rab25 associates with alpha5beta1 integrin to promote invasive migration in 3D microenvironments. Dev Cell. 2007; 13:496–510.
- 44. Nam KT, Lee HJ, Smith JJ, Lapierre LA, Kamath VP, Chen X, Aronow BJ, Yeatman TJ, Bhartur SG, Calhoun BC, Condie B, Manley NR, Beauchamp RD, et al. Loss of Rab25 promotes the development of intestinal neoplasia in mice and is associated with human colorectal adenocarcinomas. J Clin Invest. 2010; 120:840–9.
- 45. Geng D, Zhao W, Feng Y, Liu J. Overexpression of Rab25 promotes hepatocellular carcinoma cell proliferation and invasion. Tumour Biol. 2016; 37:7713–8.
- 46. Fan Y, Wang L, Han X, Liu X, Ma H. Rab25 is responsible for phosphoinositide 3-kinase/AKT-mediated cisplatin

resistance in human epithelial ovarian cancer cells. Mol Med Rep. 2015; 11:2173–8.

- 47. Dozynkiewicz MA, Jamieson NB, Macpherson I, Grindlay J, van den Berghe PV, von Thun A, Morton JP, Gourley C, Timpson P, Nixon C, McKay CJ, Carter R, Strachan D, et al. Rab25 and CLIC3 collaborate to promote integrin recycling from late endosomes/lysosomes and drive cancer progression. Dev Cell. 2012; 22:131–45.
- 48. Gomez-Roman N, Sahasrabudhe NM, McGregor F, Chalmers AJ, Cassidy J, Plumb J. Hypoxia-inducible factor 1 alpha is required for the tumourigenic and aggressive phenotype associated with Rab25 expression in ovarian cancer. Oncotarget. 2016; 7:22650–64. http://doi. org/10.18632/oncotarget.7998.
- Mitra S, Federico L, Zhao W, Dennison J, Sarkar TR, Zhang F, Takiar V, Cheng KW, Mani S, Lee JS, Mills GB. Rab25 acts as an oncogene in luminal B breast cancer and is causally associated with Snail driven EMT. Oncotarget. 2016; 7:40252–65. http://doi.org/10.18632/oncotarget.9730.
- 50. Gankhuyag N, Yu KN, Davaadamdin O, Lee S, Cho WY, Park C, Jiang HL, Singh B, Chae CH, Cho MH, Cho CS. Suppression of Tobacco Carcinogen-Induced Lung Tumorigenesis by Aerosol-Delivered Glycerol Propoxylate Triacrylate-Spermine Copolymer/Short Hairpin Rab25 RNA Complexes in Female A/J Mice. J Aerosol Med Pulm Drug Deliv. 2017; 30:81–90.
- Luo H, Zhang H, Zhang Z, Zhang X, Ning B, Guo J, Nie N, Liu B, Wu X. Down-regulated miR-9 and miR-433 in human gastric carcinoma. J Exp Clin Cancer Res. 2009; 28:82.
- 52. Ye F, Tang H, Liu Q, Xie X, Wu M, Liu X, Chen B, Xie X. miR-200b as a prognostic factor in breast cancer targets multiple members of RAB family. J Transl Med. 2014; 12:17.
- 53. Wrzeszczynski KO, Varadan V, Byrnes J, Lum E, Kamalakaran S, Levine DA, Dimitrova N, Zhang MQ, Lucito R. Identification of tumor suppressors and oncogenes from genomic and epigenetic features in ovarian cancer. PLoS One. 2011; 6:e28503.
- 54. Kikuchi M, Yamashita K, Waraya M, Minatani N, Ushiku H, Kojo K, Ema A, Kosaka Y, Katoh H, Sengoku N, Enomoto T, Tanino H, Sawanobori M, et al. Epigenetic regulation of ZEB1-RAB25/ESRP1 axis plays a critical role in phenylbutyrate treatment-resistant breast cancer. Oncotarget. 2016; 7:1741–53. http://doi.org/10.18632/ oncotarget.6480.
- 55. Roche J, Nasarre P, Gemmill R, Baldys A, Pontis J, Korch C, Guilhot J, Ait-Si-Ali S, Drabkin H. Global Decrease of Histone H3K27 Acetylation in ZEB1-Induced Epithelial to Mesenchymal Transition in Lung Cancer Cells. Cancers (Basel). 2013; 5:334–56.
- 56. Goldenring JR. A central role for vesicle trafficking in epithelial neoplasia: intracellular highways to carcinogenesis. Nat Rev Cancer. 2013; 13:813–20.
- 57. Lapierre LA, Caldwell CM, Higginbotham JN, Avant KM, Hall J, Beauchamp RD, Goldenring JR. Transformation of

rat intestinal epithelial cells by overexpression of Rab25 is microtubule dependent. Cytoskeleton (Hoboken). 2011; 68:97–111.

- 58. Zhang X, Lu Y, Shen G, Li J. Effect of RAB25 gene on proliferation of human breast cancer cell line MCF-7 *in vivo* and *in vitro*. Int J Clin Exp Pathol. 2016; 9:4746–55.
- Gomez-Roman N, McGregor F, Wheate NJ, Plumb JA. Cucurbit [7] uril encapsulated cisplatin overcomes resistance to cisplatin induced by Rab25 overexpression in an intraperitoneal ovarian cancer model. J Ovarian Res. 2015; 8:62.
- Cheng KW, Agarwal R, Mitra S, Lee JS, Carey M, Gray JW, Mills GB. Rab25 increases cellular ATP and glycogen stores protecting cancer cells from bioenergetic stress. EMBO Mol Med. 2012; 4:125–41.
- Liu Y, Tao X, Jia L, Cheng KW, Lu Y, Yu Y, Feng Y. Knockdown of RAB25 promotes autophagy and inhibits cell growth in ovarian cancer cells. Mol Med Rep. 2012; 6:1006–12.
- Schiller JH, Harrington D, Belani CP, Langer C, Sandler A, Krook J, Zhu J, Johnson DH, Eastern Cooperative Oncology Group. Comparison of four chemotherapy regimens for advanced non-small-cell lung cancer. N Engl J Med. 2002; 346:92–8.
- 63. Ohe Y, Ohashi Y, Kubota K, Tamura T, Nakagawa K, Negoro S, Nishiwaki Y, Saijo N, Ariyoshi Y, Fukuoka M. Randomized phase III study of cisplatin plus irinotecan versus carboplatin plus paclitaxel, cisplatin plus

gemcitabine, and cisplatin plus vinorelbine for advanced non-small-cell lung cancer: Four-Arm Cooperative Study in Japan. Ann Oncol. 2007; 18:317–23.

- 64. Fan Y, Xu BH, Yuan P, Ma F, Wang JY, Ding XY, Zhang P, Li Q, Cai RG. Docetaxel-cisplatin might be superior to docetaxel-capecitabine in the first-line treatment of metastatic triple-negative breast cancer. Ann Oncol. 2013; 24:1219–25.
- 65. Diéras V, Guastalla JP, Ferrero JM, Curé H, Weber B, Winckel P, Lortholary A, Mayer F, Paraiso D, Magherini E, Pujade-Lauraine E. A multicenter phase II study of cisplatin and docetaxel (Taxotere) in the first-line treatment of advanced ovarian cancer: a GINECO study. Cancer Chemother Pharmacol. 2004; 53:489–95.
- 66. Rosell R, Lord RV, Taron M, Reguart N. DNA repair and cisplatin resistance in non-small-cell lung cancer. Lung Cancer. 2002; 38:217–27.
- 67. Ardizzoni A, Boni L, Tiseo M, Fossella FV, Schiller JH, Paesmans M, Radosavljevic D, Paccagnella A, Zatloukal P, Mazzanti P, Bisset D, Rosell R; CISCA (CISplatin versus CArboplatin) Meta-analysis Group. Cisplatin- versus carboplatin-based chemotherapy in first-line treatment of advanced non-small-cell lung cancer: an individual patient data meta-analysis. J Natl Cancer Inst. 2007; 99:847–57.
- Krishnan M, Lapierre LA, Knowles BC, Goldenring JR. Rab25 regulates integrin expression in polarized colonic epithelial cells. Mol Biol Cell. 2013; 24:818–31.