BRIEF REPORT



Antifungal Susceptibility Does Not Correlate With Fungal Clearance or Survival in AIDS-Associated Cryptococcal Meningitis

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We investigated the value of susceptibility testing in predicting response in AIDS-associated cryptococcal meningitis using clinical isolates from a randomized controlled trial of antifungal treatment (amphotericin monotherapy, amphotericin with flucytosine, or amphotericin with fluconazole). We found no correlation between antifungal susceptibility and either early or late survival, or fungal clearance.

Keywords. cryptococcal meningitis; susceptibility testing; HIV; outcome; mortality.

Cryptococcus neoformans causes devastating meningitis and 15% of AIDS-related deaths globally. It is an environmental saprophyte acquired through inhalation; azole-resistance may reflect exposure to agricultural pesticides. WHO guide-lines, available at https://www.who.int/hiv/pub/guidelines/ cryptococcal-disease/en/, recommend induction therapy with amphotericin and flucytosine, which delivers improved cerebrospinal fluid (CSF) sterilization and survival [1, 2]. Flucytosine is unaffordable for countries with the greatest disease burden. Consequently, many patients receive inferior treatment with amphotericin, alone or with fluconazole [1].

Broth microdilution is the current standard method for antifungal susceptibility testing (AST) of yeasts, as outlined by the Clinical Laboratory Standards Institute (CLSI) and European Committee on Antimicrobial Susceptibility Testing (EUCAST) [3]. There is substantial deviation from standardized methods in published literature, with varying media,

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incubation times, and methods of end-point determination. The Sensititre YeastOne system (Thermo Fisher Scientific, UK) is a commercially available broth microdilution method for AST with good essential agreement with CLSI and EUCAST methods [3]. It has the advantages of ease of use and interpretation via a colorimetric binary output, and consistency through central manufacturing.

There is little consensus on the value of AST in cryptococcal meningitis; few studies demonstrate any association between susceptibility and outcome [4–6], although attempts using modified methods have been more successful [6]. The variety of methods, lack of susceptibility breakpoints for *C. neoformans*, heterogeneity of induction regimens, and inconsistent assessment of baseline disease severity make these studies difficult to compare.

Previously, we reported the results of a randomized controlled trial (RCT) of induction therapy for AIDS-associated cryptococcal meningitis [1]. Here, we use isolates obtained at diagnosis in this study to determine the ability of AST to predict therapeutic response.

METHODS

Patient Population

We enrolled 299 patients into an open-label RCT of antifungal therapy for AIDS-associated cryptococcal meningitis at a single center in Vietnam between 2005 and 2010. Detailed trial methodology has been described previously [1]. Patients received induction treatment with either amphotericin monotherapy (1 mg/ kg/day for 4 weeks), amphotericin combined with flucytosine (100 mg/kg/day for 2 weeks), or amphotericin combined with fluconazole (400 mg twice daily for 2 weeks), followed by consolidation with fluconazole monotherapy (400 mg daily) until 10 weeks postrandomisation (see Supplementary Fig. 1).

Fungal Isolates and Susceptibility Testing

CSF quantitative fungal counts were determined as previously described [1]. *Cryptococcus* isolates were cultured from CSF at randomization and archived via a full plate sweep with storage on beads (Pro-Lab Diagnostics, UK) at -80° C. For AST, isolates were revived on Sabouraud plates, a single colony selected and purified by culture, and the susceptibility of this single isolate to amphotericin, fluconazole, and flucytosine determined using Sensititre YeastOne as per the manufacturer's instructions.

Statistical Analysis

We analyzed the joint effect of the relevant drug Minimum Inhibitory Concentration (MIC) for patients on combination therapy and the

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effect of amphotericin MIC for patients on monotherapy. Primary outcome was survival until 70 days analyzed using Cox regression. Secondary outcomes were survival until 14 days and 6 months, and CSF fungal decline over the first 14 days (estimated from longitudinal measurements during that period and a linear mixed-effects model). The Cox model was also analyzed with adjustment for baseline fungal burden and Glasgow Coma Score (GCS), these factors being associated with worse outcome [1]. Data were included for all strains with a valid MIC result, and are presented for participants by trial arm in terms of the effect that a 2-fold increase in MIC has on the outcome measure of interest. We also defined isolates as fully sensitive or not, using breakpoints from published literature and as suggested in CLSI guidelines; MIC $\leq 0.512 \,\mu$ g/mL for amphotericin B, $\leq 4 \mu g/mL$ for flucytosine, $\leq 8 \mu g/mL$ for fluconazole at 72 hours [4]. All analyses were performed using R software version 2.13.1 (https://www.r-project.org/.).

RESULTS

Fluconazole

Of 299 study participants, 23 were excluded (no viable baseline isolate n = 9; inadequate growth by 72 hours n = 12; missing purity plate data, n = 2). Baseline characteristics of the primary analysis population are in Supplementary Table 1. Drug susceptibilities were similar between treatment arms (Supplementary Table 2); the range of susceptibilities is illustrated in Supplementary Figure 2.

Primary Outcomes (Patient Survival)

Table 1 shows the estimated effect on survival of decreasing antifungal susceptibility by 70 days postrandomization, without adjustment for disease severity. There was no consistent trend in hazard ratios (HR) produced by the model. Due to the multiplicity of analyses, individual HR estimates and *P*-values should be interpreted with caution. The adjusted model produced similar results.

The Kaplan-Meier curves in Supplementary Figure 3 illustrate the estimated effect of antifungal susceptibility on time to death up to 6 months when patients' isolates are categorized as either "fully sensitive" or "nonsusceptible" (Supplementary Table 3). We found no evidence that this categorization affected risk of death, including in an exploratory analysis of patients with high fungal loads (defined as $>6 \times 10^6$ colony forming units/mL CSF).

Secondary Outcomes

We found no evidence that antifungal susceptibility affected either the early (day 14) or late (6 month) hazard of death

Table 1. The estimated effect, defined by hazard ratio (HR) and 95% confidence intervals, of a 2-fold increase in the MICs estimated at 72 hours for amphotericin, fluconazole, and flucytosine on survival outcomes at 14, 70, and 182 days postrandomisation for the primary analysis population and the effect on mycological outcome (the rate of decline of CSF fungal count, log10 Colony Forming Units/mL CSF/day)

	HR		Group 2 Amphotericin and Flucytosine (n = 96)		Group 3 Amphotericin and Fluconazole (n = 88)	
		P value	HR	P value	HR	<i>P</i> value
Death by day 14						
Effect on the rate of dea	th per 2-fold increase in t	he MIC (95% CI) of				
Amphotericin	0.64 (.28, 1.44)	.28	0.86 (.35, 2.13)	.75	0.69 (.27, 1.75)	.43
Flucytosine	-	-	0.70 (.33, 1.47)	.34	-	-
Fluconazole	-	-	-	-	1.23 (.63, 2.43)	.54
Death by day 70						
Effect on the rate of dea	ath per 2-fold increase in t	he MIC (95% CI) of				
Amphotericin	0.94 (.65, 1.86)	.83	0.58 (.31, 1.09)	.09 ^a	0.97 (.47, -2.01)	.94
Flucytosine	-	-	0.88 (.55, 1.39)	.58	-	-
Fluconazole	-	-	-	-	0.87 (.51, 1.49)	.61
Death by 6 months (da	ay 182)					
Effect on the rate of dea	ath per2-fold increase in th	ne MIC (95% CI) of:				
Amphotericin	1.10 (.65, 1.86)	.72	0.62 (.34, 1.11)	.11 ^b	1.28 (.71, 2.30)	.42
Flucytosine	-	-	0.89 (.58, 1.39)	.62	-	-
Fluconazole	-	-	-	-	0.86 (.54, 1.36)	.51
Change in CSF fungal	decline in first 14 days (log10 CFU/mL of C	SF per day) per 2-fold incr	ease in MIC		
Numbers in brackets are	95% Confidence Interva	als				
	Effect estimate	P value	Effect estimate	P value	Effect estimate	P value
Amphotericin	-0.01 (07, .04)	.59	0.02 (03, .07)	.40	0.00 (04, .04)	.95
Flucytosine	-	-	1.10 (.80, 1.49)	.63	-	-

^aWhen adjusted for baseline CSF log-quantitative fungal count, GCS below 15 and Cryptococcus genotype at recruitment the results for a 2-fold increase in 72 hour MIC for amphotericin in group 2 were 0.55 (95% CI, .30–1.01), P value .053.

^bWhen adjusted as before, results for a 2-fold increase in 72 hour MIC for amphotericin in group 2 were 0.58 (95% Cl, .33–1.03), P value .06.

0.01(-.02..04)

.53

(Table 1). The adjusted model produced similar results for all outcome measures. We found no consistent effect of drug susceptibility on the rate of fungal clearance from CSF for any of the 3 drugs tested.

DISCUSSION

The most effective induction regimen for cryptococcal meningitis, amphotericin combined with flucytosine, results in mortality rates of 15% to 40% [1, 2]. Trial data suggest that amphotericin accelerates CSF fungal clearance, but amphotericin toxicity contributes to mortality when therapy continues for more than 1 week [2]. Given the high mortality rate, toxicity, and cost of combination induction therapy, the ability to predict a patient's response to antifungal therapy at diagnosis would enable optimization of the limited therapeutic options available [6]. However, we found no evidence that AST, measured using Sensititre YeastOne, can help guide treatment choices in cryptococcal meningitis.

Some small studies report an association between *Cryptococcus* susceptibility and survival. Witt and colleagues [7] found that fluconazole susceptibility was an independent predictor of treatment outcome (survival at 10 weeks with sterile CSF) in HIV-associated disease; however, they failed to demonstrate this association using the CLSI macrotiter method, used an amphotericin-free treatment regimen, and did not adjust for baseline disease severity. A small, retrospective study by Lee (n = 46) found an association between fluconazole susceptibility and survival, but none for amphotericin B or flucytosine. This study included a heterogeneous patient population managed with multiple treatment regimens, which were not adjusted for in the statistical analysis, potentially confounding results [6].

In contrast, larger studies, including ours, with more robust sampling and less selection bias by analysis of subsets of RCTs, fail to show an association between susceptibility and survival [4, 5, 8-10]. The strengths of our study are its size, the randomized allocation of induction therapy (removing bias in treatment selection), standardization of drug formulations, care delivery within a single institution, and AST within a single laboratory. We found no evidence that AST of isolates at the point of diagnosis predicts mycological response or survival, even in patients with high fungal burdens. This was true for all 3 key antifungal drugs (amphotericin, flucytosine, and fluconazole), for survival at both early (14 day) and late (70, 182 day) time-points, and following adjustment for baseline factors associated with severity, including fungal burden. We must conclude that AST has no utility in optimizing therapy for patients with a first presentation of cryptococcal meningitis.

There are several possible explanations for the poor correlation between antifungal susceptibility of *C. neoformans* and therapeutic outcomes. These include significant differences between in vivo infection and in vitro AST environments. *Cryptococcus neoformans* variably expresses its phenotype in different models and culture systems, including virulence factors (melanization, polysaccharide capsule size, titan cell formation) which may influence susceptibility. Second, host–drug interactions may play a role in clinical response; amphotericin may have immunomodulatory effects promoting yeast clearance that cannot be reflected in vitro, and may be variably expressed in AIDS patients [6].

A potential weakness of our study is that we tested only single purified isolates from our patients. While the majority of immunosuppressed patients have infections from a single strain of *C. neoformans*, multiple strain infections may occur in up to 18%, challenging the concept of correlating outcome with the susceptibility of a single isolate [11]. A further potential weakness is that we tested only baseline isolates; *C. neoformans* displays the phenomenon of heteroresistance to azoles whereby a resistant subpopulation can emerge from within a predominantly susceptible single strain following azole exposure; this may contribute to disease relapse [12, 13].

In conclusion, we present robust data that AST of baseline isolates of *C. neoformans* in AIDS-associated cryptococcal meningitis does not correlate with survival or mycological clearance; it has no place in routine clinical use in first cases of cryptococcal meningitis. However, several aspects warrant further investigation. Our study enrolled only severely immunosuppressed patients, which may confound any effect of susceptibility on outcome; similar data should be generated for immunocompetent patients. Furthermore, because of the phenomenon of heteroresistance, AST may be more informative if measured following a few days of treatment [12].

Supplementary Data

Supplementary materials are available at *Clinical Infectious Diseases* online. Consisting of data provided by the authors to benefit the reader, the posted materials are not copyedited and are the sole responsibility of the authors, so questions or comments should be addressed to the corresponding author.

Notes

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References

 Day JN, Chau TTH, Wolbers M, et al. Combination antifungal therapy for cryptococcal meningitis. N Engl J Med 2013; 368:1291–302.

- Molloy SF, Kanyama C, Heyderman RS, et al; ACTA Trial Study Team. Antifungal combinations for treatment of cryptococcal meningitis in Africa. N Engl J Med 2018; 378:1004–17.
- Delma FZ, Al-Hatmi AMS, Buil JB, et al. Comparison of MIC test strip and sensititre YeastOne with the broth microdilution reference methods CLSI and EUCAST for in vitro antifungal susceptibility testing of *Cryptococcus neoformans*. Antimicrob Agents Chemother 2020; 64:e02261–19.
- Arechavala AJ, Ochiuzzi ME, Borgnia MD, Santiso GM. Fluconazole and amphotericin B susceptibility testing of *Cryptococcus neoformans*: results of minimal inhibitory concentrations against 265 isolates from HIV-positive patients before and after two or more months of antifungal therapy. Rev Iberoam Micol 2009; 26:194–7.
- Manosuthi W, Sungkanuparph S, Thongyen S, et al. Antifungal susceptibilities of *Cryptococcus neoformans* cerebrospinal fluid isolates and clinical outcomes of cryptococcal meningitis in HIV-infected patients with/without fluconazole prophylaxis. J Med Assoc Thai 2006; 89:795–802.
- Grossman NT, Casadevall A. Physiological differences in *Cryptococcus neoformans* strains in vitro versus in vivo and their effects on antifungal susceptibility. Antimicrob Agents Chemother 2017; 61:e00091–10.
- 7. Witt MD, Lewis RJ, Larsen RA, et al. Identification of patients with acute AIDS-associated cryptococcal meningitis who can be effectively treated with

fluconazole: the role of antifungal susceptibility testing. Clin Infect Dis 1996; 22:322-8.

- Nasri H, Kabbani S, Alwan MB, et al. Retrospective study of Cryptococcal meningitis with elevated minimum inhibitory concentration to fluconazole in immunocompromised patients. Open Forum Infect Dis 2016; 3:ofw076.
- Dannaoui E, Abdul M, Arpin M, et al; French Cryptococcosis Study Group. Results obtained with various antifungal susceptibility testing methods do not predict early clinical outcome in patients with cryptococcosis. Antimicrob Agents Chemother 2006; 50:2464–70.
- Nascimento E, Vitali LH, von Zeska Kress MR, Martinez R. Cryptococcus neoformans and C. Gattii isolates from both HIV-infected and uninfected patients: Antifungal susceptibility and outcome of cryptococcal disease. Rev Inst Med Trop Sao Paulo 2017; 59:e49.
- Desnos-Ollivier M, Patel S, Spaulding AR, et al. Mixed infections and in vivo evolution in the human fungal pathogen *Cryptococcus neoformans*. MBio 2010; 1:e00091–10.
- 12. Hope W, Stone NRH, Johnson A, et al. Fluconazole monotherapy is a suboptimal option for initial treatment of cryptococcal meningitis because of emergence of resistance. MBio **2019**; 10:e02575–19.
- Sionov E, Chang YC, Garraffo HM, Kwon-Chung KJ. Heteroresistance to fluconazole in *Cryptococcus neoformans* is intrinsic and associated with virulence. Antimicrob Agents Chemother 2009; 53:2804–15.