

Variations in Seasonal Influenza Vaccine Effectiveness due to Study Characteristics: A Systematic Review and Metaanalysis of Test-Negative Design Studies

George N. Okoli,^{1,2,3} Florentin Racovitan,³ Christiaan H. Righolt,³ and Salaheddin M. Mahmud³

¹George and Fay Yee Centre for Healthcare Innovation, Max Rady College of Medicine, Rady Faculty of Health Sciences, University of Manitoba, Winnipeg, MB, Canada, ²College of Pharmacy, Rady Faculty of Health Sciences, University of Manitoba, Winnipeg, MB, Canada, and ³Vaccine and Drug Evaluation Centre, Rady Faculty of Health Sciences, University of Manitoba, Winnipeg, MB, Canada, Canada

Background. Study characteristics influence vaccine effectiveness (VE) estimation. We examined the influence of some of these on seasonal influenza VE estimates from test-negative design (TND) studies.

Methods. We systematically searched bibliographic databases and websites for full-text publications of TND studies on VE against laboratory-confirmed seasonal influenza in outpatients after the 2009 pandemic influenza. We followed the Cochrane Handbook for Systematic Reviews of Interventions guidelines. We examined influence of source of vaccination information, respiratory specimen swab time, and covariate adjustment on VE. We calculated pooled adjusted VE against H1N1 and H3N2 influenza subtypes, influenza B, and all influenza using an inverse-variance random-effects model.

Results. We included 70 full-text articles. Pooled VE against H1N1 and H3N2 influenza subtypes, influenza B, and all influenza was higher for studies that used self-reported vaccination than for those that used medical records. Pooled VE was higher with respiratory specimen collection within \leq 7 days vs \leq 4 days of symptom onset, but the opposite was observed for H1N1. Pooled VE was higher for studies that adjusted for age but not for medical conditions compared with those that adjusted for both. There was, however, a lack of statistical significance in almost all differences in pooled VE between compared groups.

Conclusions. The available evidence is not strong enough to conclude that influenza VE from TND studies varies by source of vaccination information, respiratory specimen swab time, or adjustment for age/medical conditions. The evidence is, however, indicative that these factors ought to be considered while designing or evaluating TND studies of influenza VE.

Keywords. seasonal influenza; vaccine effectiveness; test-negative design; outpatients; systematic review; meta-analysis.

Vaccination is the most effective prevention for seasonal influenza. Observational studies, rather than randomized controlled trials, are used to examine seasonal influenza vaccine effectiveness (VE) due to feasibility and ethical considerations. Continuous changes that occur in influenza viruses (antigenic drift) [1] mean that influenza vaccines have to be re-formulated every influenza season and that vaccine virus strains may be mismatched with circulating virus strains. Influenza VE studies are conducted each season in many jurisdictions worldwide to assess vaccine performance and to inform subsequent influenza season vaccine development.

Open Forum Infectious Diseases[®]

Studies on influenza VE often have differences in their design. Studies approach participant recruitment differently, and influenza vaccination status may be determined by either self-report or medical record ascertained. Clinic presentation and timing of respiratory specimen swab collection differ across study participants. The characteristics of study participants, such as age and health status, also vary and may impact VE [2]. Adjustment in analysis of VE varies across studies, and adjustment for specific potential confounders such as age and medical conditions may lead to differences in VE estimations. Due to these variations and other factors, influenza VE estimates vary between jurisdictions.

The test-negative design (TND), an observational study design type, is an increasingly popular design for estimating influenza VE [3, 4]. In a TND study, patients presenting with influenza-like symptoms are tested for influenza. Those with a positive test result become the cases, and those with a negative test result become the controls. Influenza VE (represented as a percentage) is calculated as 1 minus the adjusted ratio of the odds of vaccination in those with positive test results to the odds of vaccination in those with negative test results, multiplied by 100. The TND has been credited with reducing biases due to

Received 26 February 2020; editorial decision 12 May 2020; accepted 19 May 2020.

Correspondence: Salaheddin M. Mahmud, MD, PhD, Vaccine and Drug Evaluation Centre, University of Manitoba, 339-750 McDermot Avenue, Winnipeg, R3E 0T5, MB, Canada (salah. mahmud@gmail.com).

[©] The Author(s) 2020. Published by Oxford University Press on behalf of Infectious Diseases Society of America. This is an Open Access article distributed under the terms of the Creative Commons Attribution-NonCommercial-NoDerivs licence (http://creativecommons.org/licenses/ by-nc-nd/4.0/), which permits non-commercial reproduction and distribution of the work, in any medium, provided the original work is not altered or transformed in any way, and that the work is properly cited. For commercial re-use, please contact journals.permissions@oup.com DOI: 10.1093/ofid/ofaa177

differential health care-seeking behavior between vaccinated and unvaccinated individuals and differential misclassification of influenza infection status [3]. However, if stringent methods for study participants' enrollment and influenza testing are not applied, the TND may fail to correct for differential health careseeking behavior among vaccinated and unvaccinated individuals [5].

We systematically identified, critically appraised, and summarized the findings of published TND studies that examined seasonal influenza VE in primary care settings since the 2009 pandemic influenza. We conducted a systematic review and meta-analysis following the Cochrane Handbook for Systematic Reviews of Interventions guidelines [6], and we reported our findings following the Preferred Reporting Items for Systematic Reviews and Meta-Analysis (PRISMA) guidelines [7].

METHODS

Search Strategy and Selection Criteria

We developed and registered a review protocol in the international prospective register of systematic reviews (PROSPERO) before commencement of this review (registration number CRD42017064595). We searched the MEDLINE (Ovid), Embase (Ovid), PubMed, Scopus (Elsevier), Web of Science, and Google Scholar bibliographic databases. Our literature search strategy (Supplementary Table 1) was reviewed by a knowledge synthesis librarian using the PRESS checklist [8]. The literature search was first conducted in April 2017 and updated in July 2018. Corresponding authors of regional influenza surveillance studies were contacted to check if our searches missed any relevant studies. Identified literature citations were imported and screened in a specially designed Microsoft Access 2016 database (Microsoft Corporation, Redmond, WA, USA).

We were interested in community-based TND studies conducted in primary care settings (outpatients) after the 2009 pandemic influenza (from influenza season 2010/2011 onwards). Only studies that reported multivariable-adjusted influenza VE estimates against laboratory-confirmed influenza of any type or subtype were considered for inclusion in the review. We included only studies with influenza confirmation based on reverse transcriptase polymerase chain reaction (RT-PCR) assay or viral culture of a respiratory specimen and only full-text study publications, irrespective of language of publication. We also included only studies in which patients deemed to have received influenza vaccination did so at least 14 days before their symptom onset, and their symptom onset must not have been >7 days before medical consultation, specimen collection, and study enrollment. Studies involving only hospitalized patients and studies that reported results from mixed hospitalized patients and outpatients without reporting separate results for the 2 patient groups were excluded. We also excluded studies based on retrospective analysis of respiratory samples obtained for clinical diagnostic testing. Furthermore, we excluded studies conducted in military barracks, prisons, care homes, schools, and in subgroups such as individuals with chronic diseases. The outcomes of our interest were adjusted influenza VE against the H1N1 and H3N2 influenza subtypes, influenza B, and all influenza. Two reviewers independently screened the identified citations against the eligibility criteria using a 2-stage sifting approach to review titles/abstracts and full-text articles. Disagreements during this process were resolved through discussion between the 2 reviewers or by involvement of a third reviewer. The number of ineligible citations at the title/abstract screening stage and both the number and reasons for ineligibility at the full-text article screening stage were documented.

Data Extraction

We extracted data in MS Excel 2016 (Microsoft Corporation, Redmond, WA, USA). One reviewer independently extracted data from the included articles, and a second reviewer independently checked the extracted data for errors. Disagreements during this process were resolved through discussion between the 2 reviewers or by involvement of a third reviewer. We extracted study details such as name of the first author, publication year, country, and funding source; study characteristics such as influenza season, participant recruitment strategy, number of participants, source of vaccination information, respiratory specimen swab time, influenza vaccine type, influenza diagnostic test, and the adjusted covariates in analysis; study outcome: influenza VE against the H1N1 and H3N2 influenza subtypes, influenza B, and all influenza; and study results: multivariable-adjusted influenza VE and associated 95% confidence interval (CI). Vaccine antigenic similarity with circulating virus strains was determined from articles, where reported. Where incidence of confirmed influenza was reported, we considered the season's vaccine to be antigenically similar if the strain that caused a majority of the cases (at least 75%) was similar to that contained in the vaccine, antigenically partially similar if there was modest similarity with strains covered in the vaccine, and antigenically dissimilar if circulating strains were not similar to the strains covered in the vaccine.

Data Synthesis and Analysis

The main study characteristics were synthesized in tabular form. We pooled reported multivariable-adjusted influenza VE estimates and their associated 95% CIs using inverse-variance random-effects models implemented in STATA (version 13; StataCorp LP, Texas, USA). Heterogeneity between the pooled adjusted VE estimates was assessed and quantified statistically using the I^2 statistic [9]. The chi-square statistic (χ^2) was used to assess the statistical significance (*P* value) of the difference between 2 groups of pooled adjusted results. We assessed publication bias (where appropriate) visually using funnel plots and, statistically, using the Egger's regression test [10]. Subgroup analysis was conducted according to the source of participants' influenza vaccination status, respiratory specimen swab time, and whether studies included age or age and medical conditions in their multivariable adjustment models. Subgroup analyses were conducted for all patients, and for each of the following age groups: <5 years, 5 to 17 years, 18 to 49 years, 50 to 64 years, and \geq 65 years. We included only results for age groups that clearly fell within these predefined age groups without overlapping with another age group.

RESULTS

From a total of 10 041 identified citations, 70 full-text articles met our eligibility criteria (Figure 1) [11–80]. The main characteristics of these articles are summarized in Table 1. There were 11 articles each from the United States and Spain, 8 articles from Australia, 7 articles from the I-MOVE group (involving

multiple European countries), and 6 articles each from the United Kingdom and Canada. There were 3 articles from China and 2 articles each from Germany, Israel, Netherlands, Romania, and South Africa. One article each was from Austria, Croatia, Italy, Japan, New Zealand, Portugal, Taiwan, and Turkey. The sample size from the studies in these articles ranged from 197 to 11 430 participants. All the studies were funded by nonindustry sources, and 1 study received funding from both industry and nonindustry sources.

Pooled Adjusted VE by Method of Confirmation of Vaccination Status

Although not statistically significant, we observed a 10% higher pooled VE against H1N1 (P = .191), 7% against H3N2 (P = .626), and 5% against both influenza B (P = .529) and

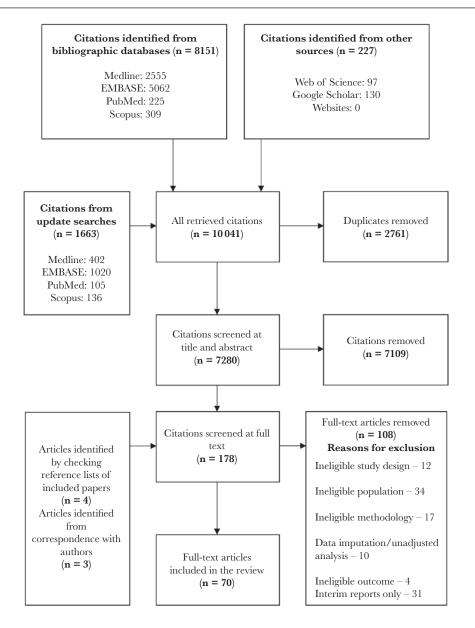


Figure 1. Modified Preferred Reporting Items for Systematic Reviews and Meta-Analysis flowchart.

Study	Country	Influenza Season (Study Period)	Respiratory Specimen (Diagnostic Test)	No. of Participants	Circulating Influ- enza Type(s)	Dominant Influenza Type	VE Outcomes Assessed
kissling et al. (2011) [11]	Europe	2010/2011	Nasal or throat swab (PCR & culture)	3254	H1N1, H3N2, influenza B	H1N1	All influenza H1N1 Influenza B
Jimenez-Jorge et al. (2012) [12]	Spain	2010/2011	Not reported (PCR)	1369	H1N1, H3N2, influenza B	H1N1	All influenza H1N1 Influenza B
Fielding et al. (2012) [13]	Australia	2011	Nose and/or throat swab (PCR)	529	H1N1, H3N2, influenza B	H1N1 first half, H3N2 mid to later season, influenza B throughout	All influenza H1N1 H3N2 Influenza B
Treanor et al. (2012) [14]	NSA	2010/2011	Nasal and throat swabs (children aged <2 years provided nasal swabs only; PCR)	4757	H1N1, H3N2, influenza B	H1N1	All influenza H1N1 H3N2 Influenza A Influenza B
Skowronski et al. (2012) [15]	Canada	2010/2011	Nasal/nasopharyngeal specimen (PCR)	1718	H1N1, H3N2, influenza B	H3N2	All influenza H1N1 H3N2 Influenza A Influenza B
Pitigoi et al. (2012) [16]	Romania	2010/2011	Not reported (PCR)	255	H1N1, H3N2, influenza B	H1N1 and influenza B	All influenza H1N1 Influenza B
Castilla et al. (2013) [17]	Spain	2011/2012	Nasopharyngeal and pharyngeal swabs (PCR)	588	H3N2, influenza B	H3N2	All influenza
Kelly et al. (2013) [18]	Australia	2010 & 2011	Combined nose and throat swab specimens (nose swab spe- cimens were only obtained from children aged <2 years; PCR)	309 (2010) 398 (2011)	H1N1, H3N2, influenza B	2010 H1N1, 2011 H3N2	All influenza H1N1 H3N2 Influenza B
Sullivan et al. (2013) [1 <mark>9]</mark>	Australia	2010, 2011, & 2012	Not reported (PCR)	420 (2010) 630 (2011) 678 (2012)	H1N1, H3N2, influenza B	2010 H1N1, 2011 influenza B, 2012 H3N2	All influenza
Martínez-Baz et al. (2013) [20]	Spain	2010/2011	Nasopharyngeal swabbing (PCR)	530	H1N1, H3N2, influenza B	H1N1	All influenza
Kissling et al. (2013) [21]	Europe	2011/2012	Nasopharyngeal swab (PCR & culture)	4362	H1N1, H3N2, influenza B	H3N2	H3N2
Jimenez-Jorge et al. (2013) [<mark>22</mark>]	Spain	2011/2012	Not reported (PCR & culture)	378	H1N1, H3N2, influenza B	H3N2	All influenza H3N2
^D ebody et al. (2013) [<mark>23</mark>]	UK	2011/2012	Respiratory samples (PCR)	3560	H1N1, H3N2, influenza B	H3N2	H3N2
Bateman et al. (2013) [24]	NSA	2010/2011	Nasal and oropharyngeal swab (PCR)	1549	H1N1, H3N2, influenza B	H3N2	H1N1 H3N2

Table 1. Summary of Characteristics of Included Studies

Study	Country	Influenza Season (Study Period)	Respiratory Specimen (Diagnostic Test)	No. of Participants	Circulating Influ- enza Type(s)	Dominant Influenza Type	VE Outcomes Assessed
Englund et al. (2013) [<mark>25</mark>]	Germany	2010/2011	Nasal or pharyngeal swabs or nasopharyngeal aspirates (PCR)	1866	H1N1, H3N2, influenza B	H1N1	All influenza H1N1 Influenza B
Lo et al. (2013) [26]	Taiwan	2011/2012	Throat or nasal swabs (PCR & culture)	918	H1N1, H3N2, influenza B	Influenza B	All influenza Influenza A Influenza B
Pebody et al. (2013) [27]	СК	2010/2011	Mouth swab (PCR)	7121	H1N1, influenza B	HINI	H1N1 Influenza B
Sullivan et al. (2014) [28]	Australia	2012	Nasal and throat samples (PCR)	600	H1N1, H3N2, influenza B	H3N2	All influenza H3N2
Levy et al. (2014) [29]	Australia	2010 to 2012	Two nose swabs and 1 throat swab (PCR)	448 (2010) 351 (2011) 1361 (2012)	H1N1, H3N2, influenza B	H1N1 in 2010 and 2011, H3N2 in 2012	All influenza H1N1 H3N2 Influenza B
Ohmit et al. (2014) [30]	USA	2011/2012	Throat swab and nasal swab (or nasal swab only in patients aged <2 years; PCR)	4771	H1N1, H3N2, influenza B	H3N2	All influenza H1N1 H3N2 Influenza A Influenza B
Kissling et al. (2014) [31]	Europe	2012/2013	Nasopharyngeal swab (PCR & culture)	6009	H1N1, H3N2, influenza B	Influenza B	H1N1 H3N2 Influenza B
Suzuki et al. (2014) [32]	Japan	2011/2012	Nasopharyngeal swab (PCR)	309	H1N1, H3N2, influenza B	H3N2	All influenza Influenza A
Skowronski et al. (2014) [33]	Canada	2011/2012	Nasal/nasopharyngeal swabs (PCR)	1507	H1N1, H3N2, influenza B	Influenza B	All influenza H1N1 H3N2 Influenza A Influenza B
Savulescu et al. (2014) [34]	Spain	2010/2011	Not reported (PCR & culture)	5057	H1N1, H3N2, influenza B	H1N1 and influenza B	H1N1 Influenza B
Nunes et al. (2014) [35]	Portugal	2012/2013	Nasopharyngeal swab or a com- bined nasopharyngeal and oropharyngeal swab (PCR & culture)	335	H1N1, H3N2, influenza B	LNL	All influenza
Skowronski et al. (2014) [36]	Canada	2012/2013	Nasal or nasopharyngeal swabs (PCR)	1501	H1N1, H3N2, influenza B	H3N2	All influenza H1N1 H3N2 Influenza A Influenza B
Yang et al. (2014) [<mark>37</mark>]	China	2012/2013	Pharyngeal swabs (culture)	1998	H1N1, H3N2, influenza B	H1N1	All influenza H1N1 H3N2
Andrews et al. (2014) [38]	ЧK	2012/2013	Not reported (PCR)	3286	H1N1, H3N2, influenza B	Influenza B	H1N1 H3N2 Influenza A Influenza B

Variations in Seasonal Influenza Vaccine Effectiveness • OFID • 5

Study	Country	Influenza Season (Study Period)	Respiratory Specimen (Diagnostic Test)	No. of Participants	Circulating Influ- enza Type(s)	Dominant Influenza Type	VE Outcomes Assessed
McAnerney et al. (2015) [39]	South Africa	2010 to 2013	Nasopharyngeal swab (PCR)	5344	H1N1, H3N2, influenza B	2010 influenza B, 2011 H1N1, 2012 H3N2, 2013 H1N1	All influenza
Pitigoi et al. (2015) [40]	Romania	2012/2013	Not reported (PCR)	197	H1N1, H3N2, influenza B	Influenza B	All influenza H1N1
Valenciano et al. (2015) [41]	Europe	2013/2014	Nasopharyngeal swab (PCR)	3020	H1N1, H3N2, influenza B	H3N2	H1N1
Helmeke et al. (2015) [42]	Germany	2012/2013	Throat or nasopharyngeal swab (PCR)	834	H1N1, H3N2, influenza B	Influenza B	All influenza H1N1 H3N2 Influenza B
Carville et al. (2015) [43]	Australia	2013	Nose or throat swab (PCR)	262	H1N1, H3N2, influenza B	Influenza A and B	All influenza H1N1 Influenza B
Chen et al. (2015) [44]	USA	2010/2011 & 2011/2012	One nasal and 1 throat swab (PCR)	927	H1 N1, H3N2, influenza B	H1N1	All influenza
VicLean et al. (2015) [45]	USA	2012/2013	Nasal and throat specimens (for children aged <2 years, only nasal specimens were obtained; PCR)	6452	H1N1, H3N2, influenza B	H3N2	All influenza H3N2
Jimenez-Jorge et al. (2015) [46]	Spain	2010/2011, 2011/2012, & 2012/2013	Nasal or nasopharyngeal (PCR & culture)	3180:SISS, 1369:cycEVA (2010/2011) 3484:SISS, 1446:cycEVA (2011/2012) 3357:SISS, 1432:cycEVA (2012/2013)	H1 N1, H3N2, influenza B	2010/2011 H1N1, 2011/2012 H3N2, 2012/2013 influenza B	H1N1 H3N2 Influenza B
Jimenez-Jorge et al. (2015) [47]	Spain	2010/2011, 2011/2012, 2012/2013, & 2013/2014	Nasal or nasopharyngeal (PCR & culture)	(cycEVA)	H1N1, H3N2, influenza B	2010/2011 H1N1, 2011/2012 H3N2, 2012/2013 influenza B, 2013/2014 H3N2 and H1N1	All influenza H1N1 H3N2 Influenza B
Kurecic- Filipovic et al. (2015) [48]	Croatia	2010/2011	Not reported (PCR)	495	H1N1, influenza B	H1N1	All influenza H1N1
Martinez- Baz et al. (2015) [49]	Spain	2012/2013	Nasopharyngeal and pharyngeal swabs (PCR)	522	H1N1, H3N2, influenza B	Influenza B	All influenza Influenza B
Skowronski et al. (2015) [<mark>50</mark>]	Canada	2013/2014	Nasal/nasopharyngeal speci- mens (PCR)	1700	H1 N1, H3N2, influenza B	H1N1	All influenza H1N1
Pebody et al. (2015) [51]	лк	2014/2015	Not reported (PCR)	2931	H1N1, H3N2, influenza B	H3N2	All influenza H3N2 Influenza A Influenza B
Gherasim et al. (2016) [<mark>52</mark>]	Spain	2014/2015	Not reported (PCR)	5044	H3N2, influenza B	H3N2	H3N2 Influenza B

Study	Country	Influenza Season (Study Period)	Respiratory Specimen (Diagnostic Test)	No. of Participants	Circulating Influ- enza Type(s)	Dominant Influenza Type	VE Outcomes Assessed
Fielding et al. (2016) [53]	Australia	2015	Nose/throat swabs (PCR)	2443	H1 N1, H3N2, influenza B	Influenza B	All influenza H1N1 H3N2 Influenza B
Pebody et al. (2016) [54]	N	2015/2016	Respiratory samples (PCR)	3841	H1N1, H3N2, influenza B	H1N1	All influenza H1N1 Influenza B
Rizzo et al. (2016) [55]	Italy	2014/2015	Nasal or throat swab (PCR)	1193	H1N1, H3N2, influenza B	H1N1 and H3N2	All influenza H1N1 H3N2 Influenza B
Castilla et al. (2016) [56]	Spain	2014/2015	Double swabs, nasopharyngeal and pharyngeal (PCR)	660	H1N1, H3N2, influenza B	H3N2 and influenza B	All influenza H3N2 Influenza B
Redlberger- Fritz et al. (2016) [57]	Austria	2014/2015	Nasopharyngeal swabs (PCR)	8 15	H1N1, H3N2, influenza B	H3N2	All influenza H1N1 H3N2 Influenza B
Thompson et al. (2016) [58]	USA	2011/2012 & 2012/2013	Nasal and throat specimens (or nasal specimens only for children aged <2 years; PCR)	1441 (2011/2012) 1327 (2012/2013)	H1N1, H3N2, influenza B	H3N2 in both seasons	All influenza H3N2 Influenza B
Pierse et al. (2016) <mark>[59]</mark>	New Zea- land	2014	Nasopharyngeal or throat swab (PCR)	1154	H1N1, H3N2, influenza B	1 Z	All influenza H1N1 H3N2 Influenza A Influenza B
Van Doorn et al. (2017) <mark>[60]</mark>	The Nether- lands	2010/2011, 2011/2012, 2012/2013, & 2013/2014	Nose and throat swabs (PCR & culture)	Unclear	H1N1, H3N2, influenza B	2010/2011 H1N1; 2011/2012, 2012/2013, and 2013/2014 H3N2	All influenza
Kelly et al. (2016) <mark>[61]</mark>	Australia	2011, 2012, & 2013	Not reported (PCR)	642 (2012/2013) 684 (2012) 354 (2013)	H1N1, H3N2, influenza B	Not reported	All influenza
Wang et al. (2016) [62]	China	2011/2012	Nasopharyngeal specimen (PCR)	668	Not reported	Not reported	All influenza
Cowling et al. (2016) [63]	USA	2010/2011, 2011/2012, & 2012/2013	Nasopharyngeal, oropharyngeal or nasal swab (PCR)	4208 (2010/2011) 2164 (2011/2012) 4278 (2012/2013)	H1N1, H3N2, influenza B	H1N1, H3N2, and influenza B in 2010/2011; H3N2 in 2011/2012; H3N2 and influenza B in 2012/2013	All influenza H1N1 H3N2 Influenza B
Skowronski et al. (2016) [64]	Canada	2014/2015	Nasal/nasopharyngeal speci- mens	1930	H1N1, H3N2, influenza B	H3N2	All influenza H3N2 Influenza B
Zimmerman et al. (2016) [65]	NSA	2014/2015	Nasal and throat swabs (children aged <2 years provided nasal swabs only; PCR)	9311	H3N2, influenza B	H3N2	All influenza H3N2

Style Control Reparticity Sectors Reparticity Sectors Control Reparticity Sectors Repartitsectors Repartitsectors <threpart< th=""><th></th><th></th><th></th><th></th><th></th><th></th><th></th><th></th></threpart<>								
USA D013/C014 Control control control transmic formers from some some some some some some some so	Study	Country	Influenza Season (Study Period)	Respiratory Specimen (Diagnostic Test)	No. of Participants	Circulating Influ- enza Type(s)	Dominant Influenza Type	VE Outcomes Assessed
Europe 2014/2015 Nappliantyngel spectmens 653 HINI, H3N2, Influenza HAN2, Influenza HAN3, Influenza HAN3, Influenza	Gaglani et al. (2016) [66]	USA	2013/2014	Combined nose and throat swab specimens (nose swab spe- cimens were only obtained from children aged <2 years; PCR)	5637	H1N1, H3N2, influenza B	H INI	H1N1
South Africa 2015 Thread and on near layed and nose seed 840 HIN1, HON2, Influences HIN1, HON2, In	Valenciano et al. (2016) [<mark>67</mark>]	Europe	2014/2015	Nasopharyngeal specimens (PCR)	6524	H1N1, H3N2, influenza B	H3N2	H1N1 H3N2 Influenza B
The Merline 2017/2013. The Merline 2017/2013. Mill Hall Na Addition 2012/2013. Mill China 2017/2013. (FG)	McAnerney et al. (2017) [68]	South Africa	2015	Throat and/or nasal swabs (PCR)	668	H1N1, H3N2, influenza B	L Z I	All influenza H1N1 H3N2 Influenza B
China 2014/2015 Ontophonential 9297 HNL, Influenza B HNL HNL HNL UK 2016/2016 (PCR) Not reported 2881 HNL, HSN2, HSN2, HSN2, HSN2, UK 2015/2016 Nasi/naopharyngeal swab 208 HNL, HSN2, HSN2, HSN2, HSN1, UK 2015/2016 Nasi/naopharyngeal swab 208 HNL, HSN2, HNL, HSN1, USA 2015/2016 Nasi/naopharyngeal swab 6879 HINL, HSN2, HINL, All USA 2015/2016 Nasi/naopharyngeal swab 6879 HINL, Influenza B HINL, HINL USA 2015/2016 Not reported 661 HINL, Influenza B HINL, Influenza B HINL I. Isael 2016/2016 Nos and throat swabs 1068 1068 HINL, Influenza B HINL I. Isael 2016/2016 HINL, HSN2, HINL, Influenza B HINL HINL I. Isael 2016/2016 HINL, HSN2, HINL, Influenza B HINL I. Isael 2016/2016 HINL, HSN2, HINL, INTERPARA HINL I. Isael 2016/2016 HINL HINL HINL H	Darvishian et al. (2017) [69]	The Nether- lands	2010/2011, 2011/2012, & 2012/2013	Throat swab and nose swab (PCR)	Not reported	H1N1, H3N2, influenza B	H3N in 2011/2012, influenza B in 2012/2013, H3N2 in 2013/2014, influenza B in 2010/2011	All influenza H1N1 H3N2 Influenza B
UK 2016/2017 Not reported 2881 HINI, H3N2, influenza B HINI, H3N2, influenza B H3N1, H3N2, influenza B Granda 2015/2016 Nasal/nasopharyngeal swab 2008 H1NI, H3N2, influenza B H1NI, H3N2, influenza B H1NI, H3N2, influenza B USA 2015/2016 Nasal/nasopharyngeal swab 6879 H1NI, H3N2, influenza B H1NI, H3N2, influenza B H1NI, H3N2, influenza B Isable 2015/2016 Not reported 661 H1NI, H3N2, influenza B H1NI, H3N2, influenza B H1NI, H3N2, influenza B H1NI, H3N2, influenza B I Isable 2015/2016 Not reported 661 H1NI, H3N2, influenza B H1NI, H3N2, influenza B H1NI, H3N2, influenza B I Isable 2015/2016 Not reported 1068 H1NI, H3N2, influenza B H1NI, H3N2, influenza B H1NI, H3N2, influenza B H1NI, H3N2, influenza B I Isable 2015/2016 Nose and throat swabs 1006 (2015/2016) H1NI, H3N2, influenza B A1 I Isable 2015/2016 Nasal swab 1005 (2015/2016) H1NI, H3N2, influenza B A1 I USA 2015/2016 Nasal swab 1005 (2015/2016) H1NI, H3N2, influenza B A1	Ma et al. (2017) [<mark>70</mark>]	China	2014/2015	Oral pharyngeal swab (PCR)	9297	H3N2, influenza B	H3N2	All influenza H3N2 Influenza B
Canada 2015/2016 Nasal/nacopharyngeal swab 2008 HINI, HSN2, Intiuenza B AII USA 2014/2015, ICCR Intise 1005 (2014/2015), Intiuenza B HINI, HSN2, Intiuenza B HINI, HSN2, Intiuenza B AII USA 2014/2015, ICCR Intiuenza B Intiuenza B HINI, HSN2, Intiuenza B AII	Pebody et al. (2017) [71]	х С	2016/2017	Not reported (PCR)	2881	H1N1, H3N2, influenza B	H3N2	All influenza Influenza A H3N2 Influenza B
USA 2015/2016 Nasal/oropharyngeal swab 6879 H1N1, H3N2, Influenza B H1N1, H3N2, Influenza B H1N1, H3N2, H1 H1N1, H3N2, H1 H1N1, H3N2, H1 H1N1, H3N2, H1 . Spain 2015/2016 Not reported 661 H1N1, Influenza B H1 . Spain 2016/2016 Not reported 661 H1N1, H3N2, Influenza B H3N2 H3 . Israel 2016/2016 Nose and throat swabs 1006 (2014/2015) H1N1, H3N2, Influenza B H3N2, Influenza B H3N2, Influenza B H3 tal. Israel 2014/2015 Nose and throat swabs 1005 (2014/2016) H1N1, H3N2, Influenza B H3N2, Influenza B AII USA 2015/2016 Nasal swab 1012 H1N1, influenza B AII	Skowronski et al. (2017) [72]	Canada	2015/2016	Nasal/nasopharyngeal swab (PCR)	2008	H1N1, H3N2, influenza B	Z	All influenza Influenza A H1N1 H3N2 Influenza B
Figure Spain 2015/2016 Not reported 661 H1N1, influenza B Influenza B Influenza B H1N1, influenza B H1N1, H3N2, H3N2 H3N2 H3 Israel 2016/2017 Nasal and throat swabs 1088 H1N1, H3N2, H3N2, H3N2 H3 H1 H3 H1	Jackson et al. (2017) [73]	NSA	2015/2016	Nasal/oropharyngeal swab (PCR)	6879	H1N1, H3N2, influenza B	E N	All influenza H1N1 H3N2 Influenza B
Israel 2016/2017 Nasal and throat swabs 108 H1N1, H3N2, h3N2 H3N2, h3N2 H3N2, h3N2 H3 et al. Israel 2014/2015 Nose and throat swabs 1005 (2014/2015) H1N1, H3N2, H3N2 in 2014/2015, and	Gherasim et al. (2017) [74]	Spain	2015/2016	Not reported (PCR & culture)	661	H1N1, influenza B	Influenza B	IZa
et al. Israel 2014/2015 Nose and throat swabs 1005 (2014/2015) H1N1, H3N2, H3N2 in 2014/2015, All (PCR) 1658 (2015/2016) influenza B H1N1 & influenza B in 2015/2016 All . USA 2015/2016 Nasal swab 1012 H1N1, influenza B H1N1 influenza B H1N1 M1 (PCR)	Stein et al. (2018) [75]	Israel	2016/2017	Nasal and throat swabs (PCR)	1088	H1N1, H3N2, influenza B	H3N2	H3N2
USA 2015/2016 Nasal swab 1012 H1N1, influenza B H1N1 (PCR)	Yaron-Yakoby et al. (2018) [<mark>76</mark>]	Israel	2014/2015	Nose and throat swabs (PCR)	1005 (2014/2015) 1658 (2015/2016)	H1N1, H3N2, influenza B	H3N2 in 2014/2015, H1N1 & influenza B in 2015/2016	All influenza & H3N2 in 2014/2015 All influenza, H1N1, & in- fluenza B in 2015/2016
	Poehling et al. (2018) [77]	USA	2015/2016	Nasal swab (PCR)	1012	H1N1, influenza B	H1N1	All influenza H1N1 Influenza B

Study	Country	Influenza Season (Study Period)	Respiratory Specimen (Diagnostic Test)	No. of Participants	Circulating Influ- enza Type(s)	Dominant Influenza Type	VE Outcomes Assessed
Valenciano et al. (2018) [78]	Europe	2011/2012 to 2016/2017	Nasopharyngeal swab (PCR)	Not clear	H1N1, influenza B (2015/16) H3N2 (2016/17)	H3N2	H1N1 H3N2 Influenza B
Hekimoglu et al. (2018) [79]	Turkey	2014/2015	Nasal, nasopharyngeal, throat, nasal plus throat, nasopha- ryngeal plus throat, nasal plus nasopharyngeal (PCR)	2561	H1N1, H3N2, influenza B	Influenza B	All influenza H1N1 H3N2 Influenza B
Kissling et al. (2018) [80]	Europe	2015/2016	Nasopharyngeal or combined naso- and oropharyngeal specimens (PCR)	11 430	H1N1, H3N2, influenza B	H1N1 Influenza B	H1N1 Influenza B

all influenza (P = .554) (Figure 2) for self-reported vaccination compared with medical record vaccination confirmation (Table 2). Almost all of the studies with self-reported vaccination were, however, from 1 research group in Canada. More of the studies with self-reported vaccination compared with those with medical record vaccination confirmation adjusted for both age and medical conditions. Zero percent (for H1N1), 20% (for H3N2, and influenza B), and 14% (for all influenza) of the studies with self-reported vaccination were from seasons in which vaccine virus strains were antigenically dissimilar to the circulating strains. In contrast, 8.3% (for H1N1), 30.8% (for H3N2), 23.1% (for influenza B), and 16% (for all influenza) of the studies with medical record vaccination confirmation were from seasons in which vaccine virus strains were antigenically dissimilar. Similar observations were made against H1N1 in 18- to 49-year-olds and against all influenza in ≥65-year-olds (Supplementary Table 2).

Pooled Adjusted VE by Timing of Respiratory Specimen Swab Collection

Despite a lack of statistical significance, we observed a 10% higher pooled adjusted VE against H3N2 (P = .596) and influenza B (P = .491), and 8% against all influenza (P = .447) (Figure 3), for swab collection within ≤ 7 days compared with ≤4 days of symptom onset (Table 2). In contrast, a 5% higher pooled adjusted VE was observed against H1N1 (P = .410) for swab collection within ≤4 days compared with swab collection within ≤ 7 days of symptom onset. There was no meaningful difference between studies with swab collection within ≤ 7 days and ≤ 4 days with regards to adjustment for both age and medical conditions in their analyses. Fifteen percent (for influenza B) and 18.5% (for all influenza) of the studies with swab collection \leq 7 days were, however, from seasons in which vaccine virus strains were antigenically dissimilar to the circulating strains. In contrast, 22.2% (for influenza B) and 27.3% (for all influenza) of the studies with swab collection within ≤ 4 days were from seasons in which vaccine virus strains were antigenically dissimilar. Similarly, 5% (for H1N1) of the studies with swab collection within \leq 7 days were from seasons in which vaccine strains were antigenically dissimilar, whereas 0% of the studies with swab collection within ≤ 4 days were from seasons in which vaccine strains were antigenically dissimilar. Evidence was conflicting across age groups (Supplementary Table 2).

Pooled Adjusted VE by Covariate Adjustment

Notwithstanding a lack of statistical significance apart from for H3N2, we observed a 4% higher pooled adjusted VE against H1N1 (P = .375), 13% against H3N2 (P = .029), 10% against influenza B (P = .144), and 4% against all influenza (P = .427) (Figure 4) for studies that included age among the adjusted covariates compared with those that included both age and medical conditions (Table 2). Three point eight percent (for H1N1), 13% (for H3N2), 13.6% (for influenza B), and 6.7%

Study	Country	Season		VE (95% CI)	% Weigh
Medical records					
Pitigoi 2012	Romania	2010/11		83 (25, 96)	0.62
Jimenez-Jorge 2012	Spain	2010/11	+	39(-18, 68)	2.04
Fielding 2012	Australia	2011/12		59(-2, 81)	1.50
Treanor 2012	USA	2010/11		59 (51, 66)	4.20
Castilla 2013	Spain	2011/12		29(-37, 63)	2.03
Kelly 2013	Australia	2010/11		87 (59, 96)	0.96
Kelly 2013	Australia	2011/12	- -	59 (5, 82)	1.51
Sullivan 2013	Australia	2010/11		73 (47, 86)	2.00
Sullivan 2013	Australia	2011/12	⊢	48 (-0, 73)	2.04
Sullivan 2013	Australia	2012/13		44 (14, 64)	3.00
Martinez-Baz 2013	Spain	2010/11		68 (19, 87)	1.31
Jimenez-Jorge 2013	Spain	2011/12		47 (7, 70)	2.38
Lo 2013	Taiwan	2011/12	_ _	-31(-79, 4)	3.59
Sullivan 2014	Australia	2012/13		45 (10, 67)	2.67
Ohmit 2014	USA	2012/13		47 (36, 56)	4.20
Suzuki 2014	Japan	2011/12	*	5(-66, 44)	2.54
Nunes 2014	Portugal	2011/12	ľ	68(21, 87)	1.32
Pitigoi 2015	Romania			95 (59, 99)	0.36
0		2012/13		(- ,	2.82
Helmeke 2015	Germany	2012/13		38 (1, 61)	
Carville 2015	Australia	2013/14		55 (-12, 82)	1.34
McLean 2015	USA	2012/13	•	49 (431, 55)	4.45
Kurecic-Filipovic 2015	Croatia	2010/11		21(-71, 63)	1.68
Martinez-Baz 2015	Spain	2012/13		55 (-0, 80)	1.60
Pebody 2015	UK	2014/15		34 (18, 47)	4.04
Fielding 2016	Australia	2015/16		54 (42, 63)	4.03
Pebody 2016	UK	2015/16		52(41, 62)	4.08
Rizzo 2016	Italy	2014/15		6(-36, 35)	3.29
Castilla 2016	Spain	2014/15	_	0(-73, 42)	2.46
Redlberger-Fritz 2016	Australia	2014/15	→	70 (35, 86)	1.67
Pierse 2016	New Zealand	2014/15	→	56 (35, 70)	3.21
Kelly 2016	Australia	2011/12		37 (-33, 70)	1.74
Kelly 2016	Australia	2012/13	→	52 (20, 71)	2.60
Kelly 2016	Australia	2013/14		61 (-0, 85)	1.27
Ma 2017	China	2014/15	 +	-18(-49, 6)	4.01
Pebody 2017	UK	2016/17	→	48 (23, 53)	3.94
Jackson 2017	USA	2015/16	+	48 (40, 55)	4.40
Yaron-Yakoby 2018	Israel	2014/15		-5(-55, 29)	3.20
Yaron-Yakoby 2018	Israel	2015/16	_ _	9(-25, 34)	3.58
Hekimoglu 2018	Turkey	2013/10		51 (12, 72)	2.31
Subtotal (I-squared = 75.3%	,	2011/13	♦	43 (35, 49)	100.00
Self reported	Courd	9010711		27/17 50)	15.04
Skowroński 2012 Skowroński 2014	Canada	2010/11	→	37 (17, 52)	15.04
	Canada	2011/12		59 (43, 70)	14.27
Skowronski 2014 Vang 2014	Canada	2012/13	→	50(33, 63)	14.66
Yang 2014	China	2012/13		52 (12, 74)	9.56
Skowronski 2015	Canada	2013/14	· · · +	68 (58, 76)	14.94
Skowronski 2016	Canada	2014/15	- + +	9 (-14, 27)	15.81
Skowronski 2017	Canada	2015/16		46 (32, 57)	15.72
	p = 0.000			48(31, 61)	100.00

Figure 2. Forest plot of vaccine effectiveness against all influenza by confirmation of vaccination status. Abbreviations: CI, confidence interval; VE, vaccine effectiveness.

(for all influenza) of the studies that included age but not medical conditions were, however, from seasons in which vaccine virus strains were antigenically dissimilar to the circulating strains. In contrast, 5.3% (for H1N1), 36.8% (for H3N2), 20% (for influenza B), and 30.6% (for all influenza) of the studies that included age and medical conditions among the adjusted

Table 2. Pooled Adjusted VE for All Patients (Irrespective of Age)

Influenza Types and Subtypes Analyzed Subgroups	No. of Studies	Pooled VE Across All Seasons (95% CI)	<i>l</i> ² Statistic, %	Publication Bias, Egger's Test <i>P</i> Valu
H1N1				
Vaccination status: medical records	24	52 (45–58)	32.7	.031
Vaccination status: self-reported	6	62 (46–73)	55.0	N/A
Respiratory specimen swab: ≤7 d	39	54 (49–58)	39.5	.022
Respiratory specimen swab: ≤4 d	7	59 (47–69)	0.0	N/A
Adjusted age	26	57 (51–63)	32.1	.034
Adjusted age & medical conditions	20	53 (46–59)	43.6	.148
H3N2				
Vaccination status: medical records	26	25 (15–34)	55.0	.988
Vaccination status: self-reported	5	32 (-0–53)	76.9	N/A
Respiratory specimen swab: ≤7 d	35	28 (22–34)	57.5	.301
Respiratory specimen swab: ≤4 d	8	18 (-26–47)	63.3	N/A
Adjusted age	23	34 (28–40)	11.5	.794
Adjusted age & medical conditions	20	21 (10–30)	70.5	.997
Influenza B				
Vaccination status: medical records	26	43 (31–52)	70.3	.701
Vaccination status: self-reported	5	48 (36–59)	28.2	N/A
Respiratory specimen swab: ≤7 d	33	48 (43–53)	28.2	.974
Respiratory specimen swab: ≤4 d	10	38 (4–60)	77.5	.070
Adjusted age	22	50 (44–56)	26.5	.893
Adjusted age & medical conditions	21	40 (27–51)	70.7	.252
All influenza				
Vaccination status: medical records	39	43 (35–49)	75.3	.807
Vaccination status: self-reported	7	48 (31–61)	84.5	N/A
Respiratory specimen swab: ≤7 d	56	46 (41–51)	70.6	.152
Respiratory specimen swab: ≤4 d	12	38 (15–55)	77.3	.009
Adjusted age	32	47 (42–52)	56.5	.477
Adjusted age & medical conditions	37	43 (34–51)	79.8	.184

Abbreviations: CI, confidence interval; N/A, not applicable; VE, vaccine effectiveness.

covariates were from seasons in which vaccine virus strains were antigenically dissimilar. Evidence was conflicting across age groups (Supplementary Table 2).

DISCUSSION

Despite a lack of statistical significance, we observed differences in pooled adjusted influenza VE between sources of influenza vaccination confirmation, respiratory specimen swab timing, and adjustments for 2 key confounders in study analysis. In our analysis of all study participants (irrespective of age), small differences were found between self-reported and medical record-confirmed influenza vaccinations, with higher pooled VE observed for self-reported vaccination, contrary to our expectations. However, almost all of the studies for self-reported vaccination were conducted in Canada and by the same group of researchers. We found substantial differences between respiratory specimen swab within \leq 7 days and \leq 4 days, with higher pooled VE observed for swab within ≤ 7 days. We also found substantial differences between studies that adjusted for age and those that adjusted for both age and medical conditions, with higher pooled VE observed for studies that adjusted for age. The above findings differed across age groups.

Studies have found that exposure misclassification can lead to significant bias in VE estimation [81, 82]. Self-reported vaccination is susceptible to recall and social desirability (individuals wanting to present a vaccine-compliant image) biases, with the potential for vaccination status misclassification. Smedt and colleagues showed in their simulation study that decreased exposure sensitivity and specificity underestimate true VE when misclassification of exposure (vaccination status) is nondifferential, but that when misclassification is differential, the bias could go in either direction, with the estimated VE deviating largely from the true VE. Compared with vaccination confirmation from medical records, self-reported vaccination usually has a higher sensitivity across various populations [83, 84] but a lower specificity in some population subgroups [85, 86]. Compared with whites, Hispanics were 2.7 times more likely to claim receipt of vaccination (self-report), and compared with younger individuals, self-reported influenza vaccination in the elderly had low specificity [84]. The observed higher pooled adjusted VE for self-reported compared with medical record-confirmed influenza vaccination status in this review, although not expected, may be due to differential misclassification of vaccination status, which Smedt and colleagues

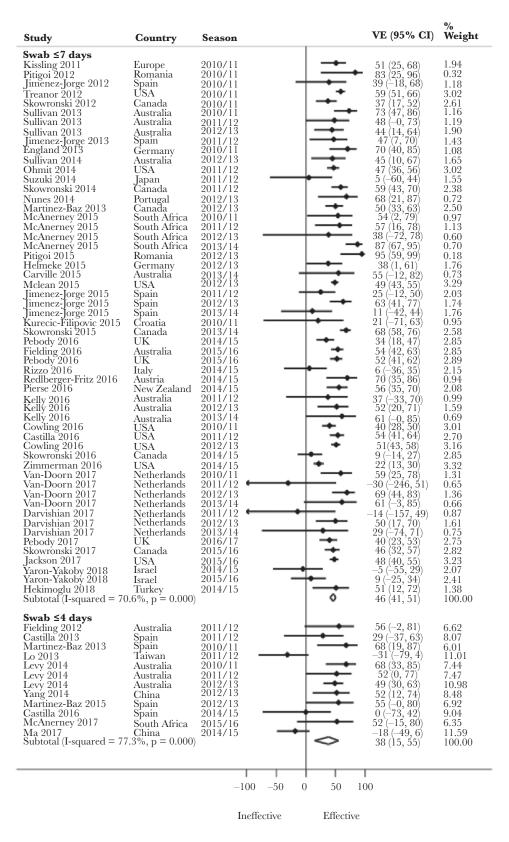


Figure 3. Forest plot of vaccine effectiveness against all influenza by timing of respiratory specimen swab collection. Abbreviations: CI, confidence interval; VE, vaccine effectiveness.

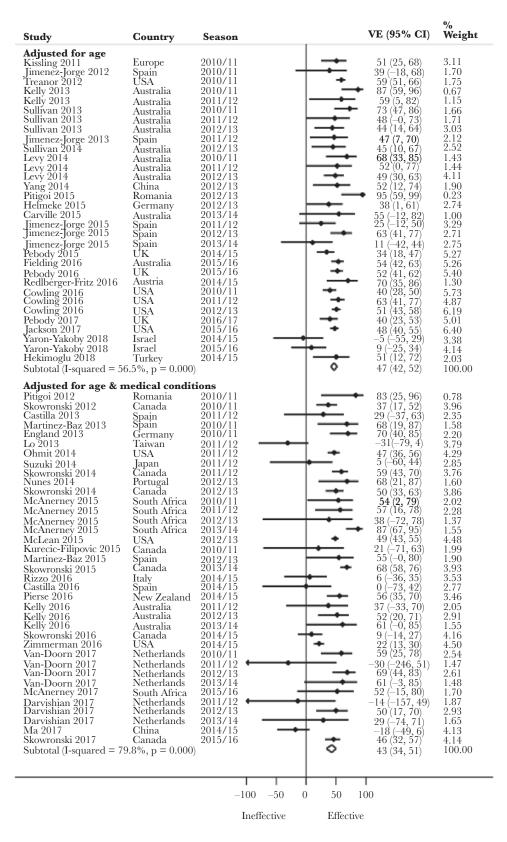


Figure 4. Forest plot of vaccine effectiveness against all influenza by covariate adjustment. Abbreviations: CI, confidence interval; VE, vaccine effectiveness.

showed could either inflate or underestimate the true VE. This becomes more plausible considering that the studies with selfreported vaccination were almost all from Canada and from the same research group. Study center influence such as characteristics of the study participants, participant recruitment strategy, and influenza testing may also explain our findings.

Influenza incubation averages 2 days (range, 1–4 days) [87]. To maximize influenza virus detection from respiratory specimens, it is advocated that, ideally, swabs be collected within <4 days of influenza-like symptom onset. The longer swab collection is from symptom onset, the lower the likelihood of detecting influenza and the greater the potential for falsenegative testing. Accurate reporting of symptom onset is therefore important, as a good TND study is predicated on patient symptom onset of \leq 7 days. It will also help minimize outcome misclassification bias. False-negative testing among the vaccinated leads to VE overestimation, while false-negative testing among the unvaccinated leads to VE being underestimated. The observed higher pooled adjusted VE for swab collection of ≤ 7 days compared with ≤ 4 days in this review may therefore be due to a higher proportion of false negatives among the \leq 7 days swab collection group, although this is not confirmable. Additionally, studies that included swab collection within \leq 4 days possibly used more stringent swab collection criteria, resulting in reduced precision of VE estimation.

Seasonal influenza VE can vary from person to person. Various individual factors impact the VE [88], and 2 main factors (age and medical conditions) are known to play an important role in determining the likelihood that a vaccine will protect a person against influenza and to what extent. Agedependent patterns in influenza vaccine protection have been reported from season to season, implicating the potential effect of age-related immune response in seasonal influenza VE [89]. For example, VE in the elderly population is reduced because of lower seroconversion rates that arise due to poorer immunological response to vaccination [90]. How well an individual responds to a vaccine may also be determined by underlying health conditions [91]. The observed higher pooled adjusted VE for studies that included age but not medical conditions compared with those that included both age and medical conditions among adjusted covariates in studies is in line with expectations, as adjusting for both age and medical conditions is likely to diminish VE compared with adjusting for age.

It is widely known that antigenic drift can markedly reduce seasonal influenza VE. For example, Flannery (2016) found that VE against H3N2 was almost 0 for an antigenically drifted genetic group of H3N2 viruses and 44% against a genetic group of H3N2 viruses that were antigenically similar to the seasonal vaccine strains [92]. This may explain the observed higher pooled adjusted VE in the subgroups with lower proportions of studies in which the seasonal influenza vaccine was antigenically dissimilar to the circulating virus strains. Variations in study design, sample size, vaccine type, and the demographic and temporal patterns underlying VE estimates from the included studies may also explain the variations observed in the pooled adjusted VE between compared groups. This, together with vaccine antigenic similarity with the circulating virus strains, may explain the high heterogeneity in many of the pooled adjusted

14 • OFID • Okoli et al

VE. Where there were adequate numbers of studies for exploration of heterogeneity using metaregression, the available covariates tended to be highly collinear, thus limiting the usefulness of metaregression. Second, it was impossible to disentangle the effects of vaccine type and the underlying patient-level variations, as the analysis was conducted at the study level and these were not clearly reported in studies.

To our knowledge, our review is the first to evaluate differences in VE due to source of influenza vaccination status, respiratory specimen swab time, and confounder adjustments in statistical models for analysis. Irving et al. (2009) evaluated influenza vaccination status determined by self-report and by a real-time vaccination registry and found that the sensitivity and specificity of self-reported influenza vaccination compared with vaccination registry records were 95% and 90%, respectively, and that self-reported vaccination status was a sensitive and somewhat specific indicator of actual vaccine status, with misclassification being more common among young people [83]. However, the study did not compare influenza VE from these 2 sources of vaccination. No reviews seem to have compared seasonal influenza VE by respiratory specimen swab time and inclusion of main confounders in statistical models for analysis as we have done.

Our decision to include only influenza seasons after the 2009 pandemic influenza may have limited the number of potentially relevant TND studies for this review. However, it allowed us to focus on studies conducted from when public funding of influenza vaccination increased in most Western jurisdictions. It should be noted that some eligible studies conducted during this stated period may not have been published by the time we conducted our literature search, and therefore would not have been included in this review. Despite growing evidence to suggest that VE may be influenced by prior vaccinations [93, 94], the included studies did not report whether the study participants received the previous season's influenza vaccination; hence, we could not assess the impact on VE estimates in our analyses. Furthermore, due to insufficient data, we could not examine VE against all outcomes for our subgroup analyses and for all age groups. We could also not separate individual study participants' effects from study center effects (eg, effectiveness of vaccine policies and programs, participant recruitment strategy, and slight differences in symptom definitions), as the studies were conducted in different jurisdictions with potentially unique jurisdictional characteristics. Finally, we could not assess the reliability of reported estimates from the included studies because we could not ascertain if the studies met all of the assumptions that well-conducted TND studies are expected to meet to ensure that effect size estimates from the studies are not biased [5]. Although many of the studies adjusted for age or age and medical conditions, there were differences in the other covariates adjusted for in the studies. This may have contributed to the high heterogeneity observed in some of our pooled VE estimates.

Our review has many merits. We developed and registered a detailed protocol in PROSPERO before the execution of our search strategy, and we fully complied with the Cochrane Handbook for Systematic Reviews of Interventions guidelines throughout the review. We utilized the expertise of a methodologist trained in evidence synthesis literature searching to develop a comprehensive search strategy for the review, and this was subsequently reviewed by a professional knowledge synthesis librarian using the PRESS checklist. We searched appropriate bibliographic databases for literature and properly screened retrieved citations (against the eligibility) following the standards specified in the Cochrane Handbook for Systematic Reviews of Interventions. Where necessary, we requested additional data from the corresponding authors of the included studies to ensure completeness of the analyzed data. We included only studies in which influenza testing was conducted using the gold standard tests (PCR or viral culture). Furthermore, we examined variations in seasonal influenza VE across all clinically relevant age groups (<5 years, 5 to <18 years, 18 to 49 years, 50 to 64 years, and \geq 65 years). We conducted the review to the highest expected standards and have reported in accordance with the PRISMA guidelines.

CONCLUSIONS

The available evidence from TND studies conducted after the 2009 pandemic influenza is not strong enough to conclude that influenza VE varies by source of vaccination status, respiratory specimen swab time, or adjustment for age/medical conditions. However, the evidence is indicative that these factors should be considered while designing or evaluating influenza VE from this study type. There is a need for researchers to ensure that age and medical conditions are both adjusted for in influenza VE estimations from TND studies, while uniformity in covariate adjustments across studies would help reduce heterogeneity and increase precision of pooled VE.

Supplementary Data

Supplementary materials are available at *Open Forum Infectious Diseases* online. Consisting of data provided by the authors to benefit the reader, the posted materials are not copyedited and are the sole responsibility of the authors, so questions or comments should be addressed to the corresponding author.

Acknowledgments

Financial support. No external funding was obtained for this study. S.M.M. is supported, in part, by funding from the Canada Research Chairs Program. G.N.O. is a current recipient of the Manitoba Training Program Fellowship Award and the Evelyn Shapiro Award, both for health services research.

Potential conflicts of interest. S.M.M. has received unrestricted research grants from GlaxoSmithKline, Merck, Sanofi Pasteur, Pfizer, and Roche-Assurex for unrelated studies. The other authors declare that they have no conflicts of interest. All authors have submitted the ICMJE Form for Disclosure of Potential Conflicts of Interest. Conflicts that the editors consider relevant to the content of the manuscript have been disclosed.

References

- Boni MF. Vaccination and antigenic drift in influenza. Vaccine 2008; 26(Suppl 3):C8-14.
- Gomez Lorenzo MM, Fenton MJ. Immunobiology of influenza vaccines. Chest 2013; 143:502–10.
- Jackson ML, Nelson JC. The test-negative design for estimating influenza vaccine effectiveness. Vaccine 2013; 31:2165–8.
- Sullivan SG, Tchetgen Tchetgen EJ, Cowling BJ. Theoretical basis of the testnegative study design for assessment of influenza vaccine effectiveness. Am J Epidemiol 2016; 184:345–53.
- Lewnard JA, Tedijanto C, Cowling BJ, Lipsitch M. Measurement of vaccine direct effects under the test-negative design. Am J Epidemiol 2018; 187:2686–97.
- Higgins JPT, Thomas J, Chandler J, et al. Cochrane Handbook for Systematic Reviews of Interventions. The Cochrane Collaboration; Chichester, UK: John Wiley & Sons; 2019.
- Moher D, Liberati A, Tetzlaff J, Altman DG; PRISMA Group. Preferred Reporting Items for Systematic Reviews and Meta-Analyses: the PRISMA statement. Int J Surg 2010; 8:336–41.
- McGowan J, Sampson M, Salzwedel DM, et al. PRESS peer review of electronic search strategies: 2015 guideline statement. J Clin Epidemiol 2016; 75:40–6.
- Higgins JP, Thompson SG. Quantifying heterogeneity in a meta-analysis. Stat Med 2002; 21:1539–58.
- Sterne JA, Egger M, Smith GD. Systematic reviews in health care: investigating and dealing with publication and other biases in meta-analysis. BMJ 2001; 323:101–5.
- Kissling E, Valenciano M, Cohen JM, et al. I-MOVE Multi-Centre Case Control Study 2010–11: overall and stratified estimates of influenza vaccine effectiveness in Europe. PLoS One 2011; 6:e27622.
- Jiménez-Jorge S, Savulescu C, Pozo F, et al; cycEVA Study Team; Spanish Influenza Sentinel Surveillance System. Effectiveness of the 2010-11 seasonal trivalent influenza vaccine in Spain: cycEVA study. Vaccine 2012; 30:3595–602.
- Fielding JE, Grant KA, Tran T, et al. Moderate influenza vaccine effectiveness in Victoria, Australia, 2011. Eurosurveillance 2012; 17:20115.
- Treanor JJ, Talbot HK, Ohmit SE, et al; US Flu-VE Network. Effectiveness of seasonal influenza vaccines in the United States during a season with circulation of all three vaccine strains. Clin Infect Dis 2012; 55:951–9.
- Skowronski DM, Janjua NZ, De Serres G, et al. A sentinel platform to evaluate influenza vaccine effectiveness and new variant circulation, Canada 2010-2011 season. Clin Infect Dis 2012; 55:332–42.
- Pitigoi D, Ivanciuc AE, Necula G, et al. Influenza vaccine effectiveness to prevent medically attended laboratory confirmed influenza during season 2010–2011 in Romania: a case control study. Rev Romana Med Lab 2012; 20:127–34.
- Castilla J, Martínez-Baz I, Martínez-Artola V, et al. Decline in influenza vaccine effectiveness with time after vaccination, Navarre, Spain, season 2011/12. Eurosurveillance 2013; 18:20388.
- Kelly HA, Sullivan SG, Grant KA, Fielding JE. Moderate influenza vaccine effectiveness with variable effectiveness by match between circulating and vaccine strains in Australian adults aged 20-64 years, 2007-2011. Influenza Other Respir Viruses 2013; 7:729–37.
- Sullivan SG, Kelly H. Late season interim estimates of influenza vaccine effectiveness reliably predict end of season estimates in Victoria, Australia, 2007 to 2012. Euro Surveill 2013; 18:20605.
- Martínez-Baz I, Martínez-Artola V, Reina G, et al. Effectiveness of the trivalent influenza vaccine in Navarre, Spain, 2010–2011: a population-based test-negative case-control study. BMC Public Health 2013; 13:191.
- Kissling E, Valenciano M, Larrauri A, et al. Low and decreasing vaccine effectiveness against influenza A(H3) in 2011/12 among vaccination target groups in Europe: results from the I-MOVE multicentre case-control study. Eurosurveillance 2013; 18:20390.
- 22. Jiménez-Jorge S, de Mateo S, Delgado-Sanz C, et al; Spanish Influenza Sentinel Surveillance System. Effectiveness of influenza vaccine against laboratoryconfirmed influenza, in the late 2011-2012 season in Spain, among population targeted for vaccination. BMC Infect Dis 2013; 13:441.
- 23. Pebody RG, Andrews N, McMenamin J, et al. Vaccine effectiveness of 2011/12 trivalent seasonal influenza vaccine in preventing laboratory-confirmed influenza in primary care in the United Kingdom: evidence of waning intra-seasonal protection. Eurosurveillance 2013; 18:20389.
- Bateman AC, Kieke BA, Irving SA, et al. Effectiveness of monovalent 2009 pandemic influenza A virus subtype H1N1 and 2010–2011 trivalent inactivated influenza vaccines in Wisconsin during the 2010–2011 influenza season. J Infect Dis 2013; 207:1262–9.
- 25. Englund H, Campe H, Hautmann W. Effectiveness of trivalent and monovalent influenza vaccines against laboratory-confirmed influenza infection in persons with medically attended influenza-like illness in Bavaria, Germany, 2010/2011 season. Epidemiol Infect 2013; 141:1807–15.

- Lo YC, Chuang JH, Kuo HW, et al. Surveillance and vaccine effectiveness of an influenza epidemic predominated by vaccine-mismatched influenza B/Yamagatalineage viruses in Taiwan, 2011-12 season. PLoS One 2013; 8:e58222.
- Pebody RG, Andrews N, Fleming DM, et al. Age-specific vaccine effectiveness of seasonal 2010/2011 and pandemic influenza A(H1N1) 2009 vaccines in preventing influenza in the United Kingdom. Epidemiol Infect 2013; 141:620–30.
- Sullivan SG, Komadina N, Grant K, et al. Influenza vaccine effectiveness during the 2012 influenza season in Victoria, Australia: influences of waning immunity and vaccine match. J Med Virol 2014; 86:1017–25.
- 29. Levy A, Sullivan SG, Tempone SS, et al. Influenza vaccine effectiveness estimates for Western Australia during a period of vaccine and virus strain stability, 2010 to 2012. Vaccine **2014**; 32:6312–8.
- Ohmit SE, Thompson MG, Petrie JG, et al. Influenza vaccine effectiveness in the 2011–2012 season: protection against each circulating virus and the effect of prior vaccination on estimates. Clin Infect Dis 2014; 58:319–27.
- 31. Kissling E, Valenciano M, Buchholz U, et al. Influenza vaccine effectiveness estimates in Europe in a season with three influenza type/subtypes circulating: The I-MOVE multicentre case-control study, influenza season 2012/13. Eurosurveillance 2014; 19:20701.
- Suzuki M, Minh LN, Yoshimine H, et al. Vaccine effectiveness against medically attended laboratory-confirmed influenza in Japan, 2011–2012 season. PLoS One 2014; 9:e88813.
- Skowronski DM, Janjua NZ, Sabaiduc S, et al. Influenza A/subtype and B/lineage effectiveness estimates for the 2011–2012 trivalent vaccine: cross-season and cross-lineage protection with unchanged vaccine. J Infect Dis 2014; 210:126–37.
- 34. Savulescu C, Jiménez-Jorge S, Delgado-Sanz C, et al; Spanish Influenza Surveillance System. Higher vaccine effectiveness in seasons with predominant circulation of seasonal influenza A(H1N1) than in A(H3N2) seasons: testnegative case-control studies using surveillance data, Spain, 2003-2011. Vaccine 2014; 32:4404–11.
- Nunes B, Machado A, Guiomar R, et al. Estimates of 2012/13 influenza vaccine effectiveness using the case test-negative control design with different influenza negative control groups. Vaccine 2014; 32:4443–9.
- 36. Skowronski DM, Janjua NZ, De Serres G, et al. Low 2012-13 influenza vaccine effectiveness associated with mutation in the egg-adapted H3N2 vaccine strain not antigenic drift in circulating viruses. PLoS One 2014; 9:e92153.
- Yang P, Thompson MG, Ma C, et al. Influenza vaccine effectiveness against medically-attended influenza illness during the 2012-2013 season in Beijing, China. Vaccine 2014; 32:5285–9.
- Andrews N, McMenamin J, Durnall H, et al. Effectiveness of trivalent seasonal influenza vaccine in preventing laboratory-confirmed influenza in primary care in the United Kingdom: 2012/13 end of season results. Euro Surveill 2014; 19:5–13.
- McAnerney JM, Walaza S, Cohen AL, et al. Effectiveness and knowledge, attitudes and practices of seasonal influenza vaccine in primary healthcare settings in South Africa, 2010-2013. Influenza Other Respir Viruses 2015; 9:143–50.
- Pitigoi D, Necula G, Alexandrescu V, et al. Circulating influenza viruses and the effectiveness of seasonal influenza vaccine in Romania, season 2012–2013. Rev Rom Med Lab 2015; 23:9–20.
- 41. Valenciano M, Kissling E, Reuss A, et al; I-MOVE Multicentre Case Control Study Team. The European I-MOVE Multicentre 2013-2014 Case-Control Study. Homogeneous moderate influenza vaccine effectiveness against A(H1N1)pdm09 and heterogenous results by country against A(H3N2). Vaccine 2015; 33:2813–22.
- 42. Helmeke C, Gräfe L, Irmscher HM, et al. Effectiveness of the 2012/13 trivalent live and inactivated influenza vaccines in children and adolescents in Saxony-Anhalt, Germany: a test-negative case-control study. PLoS One 2015; 10:e0122910.
- Carville KS, Grant KA, Sullivan SG, et al. Understanding influenza vaccine protection in the community: an assessment of the 2013 influenza season in Victoria, Australia. Vaccine 2015; 33:341–5.
- Chen Q, Griffin MR, Nian H, et al. Influenza vaccine prevents medically attended influenza-associated acute respiratory illness in adults aged ≥50 years. J Infect Dis 2015; 211:1045–50.
- McLean HQ, Thompson MG, Sundaram ME, et al. Influenza vaccine effectiveness in the United States during 2012–2013: variable protection by age and virus type. J Infect Dis 2015; 211:1529–40.
- Jimenez-Jorge S, de Mateo S, Delgado-Sanz C, et al. Estimating influenza vaccine effectiveness in Spain using sentinel surveillance data. Euro Surveill Bulletin 2015; 20:21187.
- Jimenez-Jorge S, Pozo F, Larrauri A. Interim influenza vaccine effectiveness: a good proxy for final estimates in Spain in the seasons 2010–2014. Vaccine 2015; 33:3276–80.
- Kurečić Filipović S, Gjenero-Margan I, Kissling E, et al. Influenza vaccine effectiveness estimates in Croatia in 2010-2011: a season with predominant circulation of A(H1N1)pdm09 influenza virus. Epidemiol Infect 2015; 143:2596–603.
- 49. Martínez-Baz I, Navascués A, Pozo F, et al; Primary Health Care Sentinel Network and Network for Influenza Surveillance in Hospitals of Navarra. Influenza

vaccine effectiveness in preventing inpatient and outpatient cases in a season dominated by vaccine-matched influenza B virus. Hum Vaccin Immunother **2015**; 11:1626–33.

- Skowronski DM, Chambers C, Sabaiduc S, et al. Integrated sentinel surveillance linking genetic, antigenic, and epidemiologic monitoring of influenza vaccinevirus relatedness and effectiveness during the 2013–2014 influenza season. J Infect Dis 2015; 212:726–39.
- Pebody R, Warburton F, Andrews N, et al. Effectiveness of seasonal influenza vaccine in preventing laboratory-confirmed influenza in primary care in the United Kingdom: 2014/15 end of season results. Eurosurveillance 2015; 20.
- Gherasim A, Pozo F, de Mateo S, et al; cycEVA team and the VEVA Working Group. Waning protection of influenza vaccine against mild laboratory confirmed influenza A(H3N2) and B in Spain, season 2014-15. Vaccine 2016; 34:2371–7.
- Fielding JE, Levy A, Chilver MB, et al. Effectiveness of seasonal influenza vaccine in Australia, 2015: an epidemiological, antigenic and phylogenetic assessment. Vaccine 2016; 34:4905–12.
- Pebody R, Warburton F, Ellis J, et al. Effectiveness of seasonal influenza vaccine for adults and children in preventing laboratory-confirmed influenza in primary care in the United Kingdom: 2015/16 end-of-season results. Eurosurveillance 2016; 21:30348.
- Rizzo C, Bella A, Alfonsi V, et al. Influenza vaccine effectiveness in Italy: age, subtype-specific and vaccine type estimates 2014/15 season. Vaccine 2016; 34:3102–8.
- 56. Castilla J, Navascués A, Fernández-Alonso M, et al; Primary Health Care Sentinel Network; Network for Influenza Surveillance in Hospitals of Navarra. Effectiveness of subunit influenza vaccination in the 2014-2015 season and residual effect of split vaccination in previous seasons. Vaccine 2016; 34:1350–7.
- Redlberger-Fritz M, Kundi M, Popow-Kraupp T. Detailed report on 2014/15 influenza virus characteristics, and estimates on influenza virus vaccine effectiveness from Austria's Sentinel Physician Surveillance Network. PLoS One 2016; 11:e0149916.
- Thompson MG, Clippard J, Petrie JG, et al. Influenza vaccine effectiveness for fully and partially vaccinated children 6 months to 8 years old during 2011–2012 and 2012–2013: the importance of two priming doses. Pediatr Infect Dis J 2016; 35:299–308.
- Pierse N, Kelly H, Thompson MG, et al; SHIVERS investigation team. Influenza vaccine effectiveness for hospital and community patients using control groups with and without non-influenza respiratory viruses detected, Auckland, New Zealand 2014. Vaccine 2016; 34:503–9.
- 60. van Doorn E, Darvishian M, Dijkstra F, et al. Influenza vaccine effectiveness estimates in the Dutch population from 2003 to 2014: the test-negative design casecontrol study with different control groups. Vaccine 2017; 35:2831–9.
- Kelly HA, Lane C, Cheng AC. Influenza vaccine effectiveness in general practice and in hospital patients in Victoria, 2011e2013. Med J Australia 2016; 204:76. e71–5.
- 62. Wang Y, Zhang T, Chen L, et al. Seasonal influenza vaccine effectiveness against medically attended influenza illness among children aged 6-59 months, October 2011-September 2012: a matched test-negative case-control study in Suzhou, China. Vaccine 2016; 34:2460–5.
- Cowling BJ, Feng S, Finelli L, et al. Assessment of influenza vaccine effectiveness in a sentinel surveillance network 2010-13, United States. Vaccine 2016; 34:61–6.
- 64. Skowronski DM, Chambers C, Sabaiduc S, et al. A perfect storm: impact of genomic variation and serial vaccination on low influenza vaccine effectiveness during the 2014–2015 season. Clin Infect Dis 2016; 63:21–32.
- Zimmerman RK, Nowalk MP, Chung J, et al. 2014–2015 influenza vaccine effectiveness in the United States by vaccine type. Clin Infect Dis 2016; 63:1564–73.
- 66. Gaglani M, Pruszynski J, Murthy K, et al. Influenza vaccine effectiveness against 2009 pandemic influenza A(H1N1) virus differed by vaccine type during 2013-2014 in the United States. J Infect Dis 2016; 213:1546–56.
- 67. Valenciano M, Kissling E, Reuss A, et al. Vaccine effectiveness in preventing laboratory-confirmed influenza in primary care patients in a season of co-circulation of influenza A(H1N1)pdm09, B and drifted A(H3N2), I-MOVE Multicentre Case– Control Study, Europe 2014/15. Euro Surveill 2016; 21:pii=30139.
- McAnerney JM, Walaza S, Tempia S, et al. Estimating vaccine effectiveness in preventing laboratory-confirmed influenza in outpatient settings in South Africa, 2015. Influenza Other Respir Viruses 2017; 11:177–81.
- Darvishian M, Dijkstra F, Van Doorn E, et al. Influenza vaccine effectiveness in the Netherlands from 2003/2004 through 2013/2014: the importance of circulating influenza virus types and subtypes. PLoS One 2017; 12:e0169528.
- Ma C, Pan Y, Zhang L, et al. Influenza vaccine effectiveness against medically attended influenza illness in Beijing, China, 2014/15 season. Hum Vaccin Immunother 2017; 13:2379–84.
- Pebody R, Warburton F, Ellis J, et al. End-of-season influenza vaccine effectiveness in adults and children, United Kingdom, 2016/17. Eurosurveillance 2017; 22:10–22.

- Skowronski DM, Chambers C, Sabaiduc S, et al. Beyond antigenic match: possible agent-host and immuno-epidemiological influences on influenza vaccine effectiveness during the 2015–2016 season in Canada. J Infect Dis 2017; 216:1487–500.
- Jackson ML, Chung JR, Jackson LA, et al. Influenza vaccine effectiveness in the United States during the 2015–2016 season. New Engl J Med 2017; 377:534–43.
- Gherasim A, Martinez-Baz I, Castilla J, et al. Effect of previous and current vaccination against influenza A(H1N1)pdm09, A(H3N2), and B during the postpandemic period 2010–2016 in Spain. PLoS One 2017; 12:e0179160.
- Stein Y, Mandelboim M, Sefty H, et al. Seasonal influenza vaccine effectiveness in preventing laboratory-confirmed influenza in primary care in Israel, 2016–2017 season: insights into novel age-specific analysis. Clin Infect Dis 2018; 66:1383–91.
- Yaron-Yakoby H, Sefty H, Pando R, et al. Effectiveness of influenza vaccine in preventing medically-attended influenza virus infection in primary care, Israel, influenza seasons 2014/15 and 2015/16. Eurosurveillance 2018; 23:27–37.
- Poehling KA, Caspard H, Peters TR, et al. 2015–2016 vaccine effectiveness of live attenuated and inactivated influenza vaccines in children in the United States. Clin Infect Dis 2018; 66:665–72.
- Valenciano M, Kissling E, Larrauri A, et al. Exploring the effect of previous inactivated influenza vaccination on seasonal influenza vaccine effectiveness against medically attended influenza: results of the European I-MOVE multicentre test-negative case-control study, 2011/2012–2016/2017. Influenza Other Respir Viruses 2018; 12:567–81.
- Hekimoglu CH, Emek M, Avci E, et al. Seasonal influenza vaccine effectiveness in preventing laboratory confirmed influenza in 2014–2015 season in Turkey: a test-negative case control study. Balk Med J 2018; 35:77–83.
- Kissling E, Valenciano M, Pozo F, et al; I-MOVE/I-MOVE+ study team. 2015/16 I-MOVE/I-MOVE+ multicentre case-control study in Europe: moderate vaccine effectiveness estimates against influenza A(H1N1)pdm09 and low estimates against lineage-mismatched influenza B among children. Influenza Other Respir Viruses 2018; 12:423–37.
- De Smedt T, EM, DM, et al. Bias due to differential and non-differential diseaseand exposure misclassification in studies of vaccine effectiveness. PLoS One 2018; 13:e0199180.

- Jackson ML, Phillips CH, Benoit J, et al. The impact of selection bias on vaccine effectiveness estimates from test-negative studies. Vaccine 2018; 36:751–7.
- Irving SA, Donahue JG, Shay DK, et al. Evaluation of self-reported and registrybased influenza vaccination status in a Wisconsin cohort. Vaccine 2009; 27:6546–9.
- Rolnick SJ, Parker ED, Nordin JD, et al. Self-report compared to electronic medical record across eight adult vaccines: do results vary by demographic factors? Vaccine 2013; 31:3928–35.
- Jackson ML. Use of self-reported vaccination status can bias vaccine effectiveness estimates from test-negative studies. Vaccines 2019; 1:100003.
- Zimmerman RK, Raymund M, Janosky JE, et al. Sensitivity and specificity of patient self-report of influenza and pneumococcal polysaccharide vaccinations among elderly outpatients in diverse patient care strata. Vaccine 2003; 21:1486–91.
- Carrat F, Vergu E, Ferguson NM, et al. Time lines of infection and disease in human influenza: a review of volunteer challenge studies. Am J Epidemiol 2008; 167:775–85.
- Dhakal S, Klein SL. Host factors impact vaccine efficacy: implications for seasonal and universal influenza vaccine programs. J Virol 2019; 93:e00797–19.
- Lewnard JA, Cobey S. Immune history and influenza vaccine effectiveness. Vaccines 2018; 6:28.
- 90. Wilhelm M. Influenza in older patients: a call to action and recent updates for vaccinations. Am J Manag Care 2018; 24:15–24.
- Zhao L, Stirling R, Young K. Should individuals use influenza vaccine effectiveness studies to inform their decision to get vaccinated? Can Commun Dis Rep 2019; 45:156–8.
- Flannery B, Zimmerman RK, Gubareva LV, et al. Enhanced genetic characterization of influenza A(H3N2) viruses and vaccine effectiveness by genetic group, 2014-2015. J Infect Dis 2016; 214:1010–9.
- Sullivan SG, Kelly H. Stratified estimates of influenza vaccine effectiveness by prior vaccination: caution required. Clin Infect Dis 2013; 57:474–6.
- McLean HQ, Thompson MG, Sundaram ME, et al. Impact of repeated vaccination on vaccine effectiveness against influenza A(H3N2) and B during 8 seasons. Clin Infect Dis 2014; 59:1375–85.