

Potential immunotherapy for Alzheimer disease and age-related dementia

Michal Schwartz, PhD; Michal Arad, PhD; Hila Ben-Yehuda, PhD

Emerging results support the concept that Alzheimer disease (AD) and age-related dementia are affected by the ability of the immune system to contain the brain's pathology. Accordingly, well-controlled boosting, rather than suppression of systemic immunity, has been suggested as a new approach to modify disease pathology without directly targeting any of the brain's disease hallmarks. Here, we provide a short review of the mechanisms orchestrating the cross-talk between the brain and the immune system. We then discuss how immune checkpoint blockade directed against the PD-1/PD-L1 pathways could be developed as an immunotherapeutic approach to combat this disease using a regimen that will address the needs to combat AD.

© 2019, AICH – Servier Group

Dialogues Clin Neurosci. 2019;21:21-25

Keywords: Alzheimer disease; immune checkpoint; immunotherapy; macrophage; microglia

Introduction

Alzheimer disease (AD) is a devastating age-related neurodegenerative disorder, and the most frequent cause of senile dementia.¹ The appearance of cognitive decline is associated with accumulation of misfolded proteins, as well as the presence of several additional toxic agents.² Among the common neuropathological features found in AD are synaptic and neuronal loss, intracellular neurofibrillary tangles, elevated levels of the toxic form of amyloid beta (A β),¹⁻⁴² and the accumulation of extracellular senile plaques containing misfolded A β peptide.²⁻⁴ Local inflammatory responses as well as uncontrolled astrocyte reactivity are often observed in the brains of AD patients and in animal models; these processes are not necessarily the primary causes of the disease, but are considered to be key factors in disease progression and escalation.⁵⁻⁷ The accumulated misfolded proteins and the neuroinflammatory response have led to numerous attempts over the years to arrest disease progression, either using treatments that are directed against the misfolded proteins to arrest plaque burden,^{8,9} or using systemic anti-

inflammatory drugs to arrest the brain inflammation. Inconsistent and even conflicting results were obtained, and none of the drugs tested thus far have proven effective in reversing or arresting cognitive loss in patients.¹⁰⁻¹⁶

The failure of treatments directed at A β to arrest or reverse cognitive loss could reflect the fact that by the time A β plaque burden is high, removal of plaques, while still important, may be insufficient to modify disease because numerous collateral disease-escalating factors enter into a vicious cycle and continue even after the plaques are removed. Such factors might include immune-related molecules and cells. In apparent support of such a view are, recent results demonstrating that resolution of inflammation is an active mechanism mediated by recruitment of circulating immune cells to sites of brain pathology.¹⁷⁻¹⁹

Here, we will discuss the role of brain immune communication in brain homeostasis and repair. In addition, we will discuss if and how activating the immune system by immune checkpoint blockade can contribute to disease modification.

Author affiliations: Department of Neurobiology, Weizmann Institute of Science, 7610001 Rehovot, Israel. **Address for correspondence:** Michal Schwartz, Department of Neurobiology, Weizmann Institute of Science, 7610001 Rehovot, Israel. (email: michal.schwartz@weizmann.ac.il)

Systemic leukocytes are essential players in central nervous system repair

For decades, it was commonly assumed that the brain is unable to tolerate immune cell entry, mainly due to the belief that it is a tissue behind barriers, and considered an immune privileged site.²⁰ In animal models of acute central nervous system (CNS) injuries, both monocyte-derived macrophages and CD4⁺ T cells recognizing brain antigens, are needed for coping with and helping heal parenchymal damage.²¹⁻²⁸ Moreover, T cells present in the periphery facilitate recruitment of monocyte-derived macrophages to the CNS. Such macrophages play a role in supporting neuronal survival and axonal regrowth, by resolving the local inflammatory response and facilitating local scar removal.^{25-27,29-32} Additional studies revealed that systemic T cells not only participate in CNS repair, but are also needed for life-long brain plasticity.³³⁻³⁵

Independent attempts were made to understand how T cells support healthy brain plasticity while they are excluded from the brain parenchyma, how they facilitate recruitment of monocyte-derived macrophages, and how such monocytes can gain access to the CNS without breaching the blood-brain-barrier (BBB). Such attempts have suggested that the brain's barriers, including the meningeal barrier^{36,37} and the blood-cerebrospinal fluid barrier (BCSFB) can serve as a key compartment for immune-brain crosstalk in health and disease.^{19,38,39} The BCSFB, which is comprised of the tightly connected choroid plexus (CP) epithelial cells,⁴⁰⁻⁴³ along with the accumulated evidence that immune cells are needed for brain maintenance and repair, led us to suggest that the CP is a physiological gateway that enables selective immune cell access, depending on the needs of the CNS.^{19,38}

The paradoxical fate of the "leukocyte gate" to the brain in Alzheimer disease models

Several independent studies in animal models have shown that recruitment of circulating monocyte-derived macrophages,⁴⁴⁻⁵² possibly together with additional immunoregulatory leukocytes, can modify AD pathology.^{31,53,54} Such cells can help remove misfolded protein including A β -

plaques,^{48,55,56} balance the local inflammatory milieu,^{46,47,57} reduce gliosis,⁵⁸ and protect synaptic structures.^{46,57,59}

Analyzing the fate of the CP with respect to its ability to support leukocyte trafficking revealed that its activity is impaired in animal models of brain aging and AD.^{60,61} It was further discovered that reducing systemic immune suppression in AD animal models, by transiently depleting peripheral Foxp3⁺ regulatory T cells has a beneficial effect in mitigating disease pathology.⁶² These results are consistent with an independent observation, showing that the adaptive immune system plays an important role in the progression of AD in animal models. For example, it was demonstrated that genetic ablation of B, T, and natural killer cells in the 5xFAD mouse model by crossing these mice with Rag2/Il2rc double knockout animals (Rag-5xFAD), resulted in increased plaque load and increased soluble A β levels.⁶³

Importantly, immunoregulatory T cells and anti-inflammatory cells are needed in the brain as a source of anti-inflammatory cytokines for reducing the inflammatory response. Homing of such immunomodulating cells requires well-controlled boosting, rather than suppression of systemic

immunity. Accordingly, special care must be taken when viewing immunosuppressive cells (such as FoxP3) as uniformly beneficial or harmful in neurodegenerative diseases, without considering their localization and kinetics.

Taken together, the results summarized above created the basis for our approach of empowering the systemic immune system, by transiently blocking inhibitory immune checkpoints, to drive a cascade of immune events that starts outside the brain, induces activation of the CP, and culminates in immune-dependent brain repair processes.^{61,64}

Immune checkpoint blockade for mitigating Alzheimer disease pathology

Inhibitory immune checkpoints restrain the activity of memory T cells, mainly those directed against self-compounds, to avoid autoimmune diseases. Among such checkpoints are the programmed cell death protein 1

AD is a devastating age-related neurodegenerative disorder, and the most frequent cause of senile dementia

Original article

Alzheimer disease immunotherapy - *Schwartz et al*

(PD-1), a member of the B7-CD28 family, expressed by a variety of activated effector memory immune cells, including CD4+ T cells.⁶⁵ The PD-1 ligand is expressed by dendritic cells and regulatory T cells,⁶⁶ as well as by non-immune cells such as endothelial and epithelial cells,^{67,68} and astrocytes.⁶⁶ The interaction between PD-1 and PD-L1 suppresses memory T-cell responses, including proliferation, and cytokine production.^{65,69} Blocking the PD-L1/PD-1 pathway potentially results in an increase in T cell activation.⁷⁰⁻⁷² Based on our new understanding, we envisioned that targeting systemic PD-1/PD-L1 might be a way to activate such a protective/repairative immune response.

Our studies using anti-PD-1 or anti-PD-L1 antibody in the 5xFAD mouse model of AD, as well as in a dementia model of tau pathology, revealed that such treatments are effective in helping and even reversing cognitive impairments and reducing disease pathology. This process was associated with monocyte-derived macrophages homing to the brain.^{61,62} These macrophages locally express numerous molecules including scavenger receptors for removal of dead cells as well as misfolded or aggregated proteins, anti-inflammatory cytokines, and growth factors.^{61,64}

Importantly, a single injection of antibody directed against either PD-1 or PD-L1 initiated a chain of events that started outside the brain, and led to alterations in several processes within the brain that together resulted in disease modification.⁶⁴

Notably, in most mouse models of AD, disease symptoms begin earlier in females than in males. In humans there is no clear scientific consensus regarding gender differences in AD, though most studies have shown that men and women exhibit differences in the development and progression of the disease.⁷³ Generally, women are considered at greater risk and show more rapid progression.⁷⁴ Notably, both female and male mice of tau-driven models

of dementia and amyloid β -driven pathology similarly responded to treatment with immune checkpoint blockade directed to PD-1 or PD-L1.⁶⁴

Conclusion

In conclusion, results from animal studies suggest that treatment with PD-1/PD-L1 blockade evokes a series of immunological events that start outside the brain, and, in synergy with inflammatory signals emerging from the diseased brain, restore the immunological communication between the brain and the immune system.⁷⁵ The resulting modification of the immunological milieu of the brain culminates in reduction of cognitive deficits and disease pathological manifestations. The treatment protocol going forward to clinical trials will require intermittent administration of the antibody. Such a protocol is likely to reduce adverse immunological effects. Moreover, since the treatment is not directed against a single factor within the brain that contributes to disease escalation, but rather affects common immunological pathways, it is expected to have a higher efficacy than past attempts, and to overcome disease heterogeneity and some translational obstacles. ■

Disclosure/Acknowledgements: We thank Dr Shelley Schwarzbaum for editing the manuscript. Research in the M.S. lab is supported by Advanced European Research Council grants (232835 and 741744), and by the EU Seventh Framework Program HEALTH-2011 (279017); Israel Science Foundation (ISF) research grant no. 991/16; and ISF-Legacy Heritage Biomedical Science Partnership-research grant no. 1354/15. M.A. fellowship is supported by Ministry of Science and Technology. We wish to thank the Adelis Foundation and Fisher Center for Alzheimer Research Foundation, for their generous support of our AD research. M.S. holds the Maurice and Ilse Katz Professorial Chair in Neuroimmunology. M.S. serves as a consultant of ImmunoBrain Checkpoint, Ltd.

References

1. Ballard C, Gauthier S, Corbett A, Brayne C, Aarsland D, Jones E. Alzheimer's disease. *Lancet*. 2011;377:1019-1031.
2. Hardy J, Selkoe DJ. The amyloid hypothesis of Alzheimer's disease: progress and problems on the road to therapeutics. *Science*. 2002;297:353-356.
3. Glenner GG, Wong CW, Quaranta V, Eanes ED. The amyloid deposits in Alzheimer's disease: their nature and pathogenesis. 1984; *Appl Pathol* 2:357-369.
4. Price DL, Whitehouse PJ, Struble RG. Alzheimer's disease. *Annu Rev Med*. 1985;36: 349-356.
5. Akiyama H, Barger S, Barnum S, et al. Inflammation and Alzheimer's disease. *Neurobiol Aging*. 2000;21:383-421.
6. Wyss-Coray T. Inflammation in Alzheimer disease: driving force, bystander or beneficial response? *Nat Med*. 2006;12:1005-1015.

Original article

Alzheimer disease immunotherapy - Schwartz et al

7. Wyss-Coray T, Mucke L. Inflammation in neurodegenerative disease—a double-edged sword. *Neuron*. 2002;35:419-432.
8. Schenk D, Barbour R, Dunn W, et al. Immunization with amyloid-beta attenuates Alzheimer-disease-like pathology in the PDAPP mouse. *Nature*. 1999;400:173-177.
9. Weiner HL, Lemere CA, Maron R, et al. Nasal administration of amyloid-beta peptide decreases cerebral amyloid burden in a mouse model of Alzheimer's disease. *Ann Neurol*. 2000;48:567-579.
10. Group AR, Martin BK, Szekely C, et al. Cognitive function over time in the Alzheimer's Disease Anti-inflammatory Prevention Trial (ADAPT): results of a randomized, controlled trial of naproxen and celecoxib. *Arch Neurol*. 2008;65:896-905.
11. Arvanitakis Z, Grodstein F, Bienias JL, Schneider JA, Wilson RS, Kelly JF, Evans DA, Bennett DA. Relation of NSAIDs to incident AD, change in cognitive function, and AD pathology. *Neurology*. 2008;70:2219-2225.
12. Breitner JC, Hanuse SJ, Walker R, Dublin S, Crane PK, Gray SL, Larson EB. Risk of dementia and AD with prior exposure to NSAIDs in an elderly community-based cohort. *Neurology*. 2009;72:1899-1905.
13. Orgogozo JM, Gilman S, Dartigues JF, et al. Subacute meningoencephalitis in a subset of patients with AD after Abeta42 immunization. *Neurology*. 2003;61:46-54.
14. Senior K. Dosing in phase II trial of Alzheimer's vaccine suspended. *Lancet Neurol*. 2002;1:3.
15. Doody RS, Raman R, Farlow M, et al. A phase 3 trial of semagacestat for treatment of Alzheimer's disease. *N Engl J Med*. 2013;369:341-350.
16. Gilman S, Koller M, Black RS, et al. Clinical effects of Abeta immunization (AN1792) in patients with AD in an interrupted trial. *Neurology*. 2005;64:1553-1562.
17. Schwartz M, Baruch K. The resolution of neuroinflammation in neurodegeneration: leukocyte recruitment via the choroid plexus. *EMBO J*. 2014;33:7-22.
18. Theriault P, ElAli A, Rivest S. The dynamics of monocytes and microglia in Alzheimer's disease. *Alzheimers Res Ther*. 2015;7:41.
19. Kunis G, Baruch K, Rosenzweig N, Kertser A, Miller O, Berkutzki T, Schwartz M. IFN-gamma-dependent activation of the brain's choroid plexus for CNS immune surveillance and repair. *Brain*. 2013;136: 3427-3440.
20. Medawar PB. Immunity to homologous grafted skin; the fate of skin homografts transplanted to the brain, to subcutaneous tissue, and to the anterior chamber of the eye. *Br J Exp Pathol*. 1948;29:58-69.
21. Rapalino O, Lazarov-Spiegler O, Agranov E, et al. Implantation of stimulated homologous macrophages results in partial recovery of paraplegic rats. *Nat Med*. 1998;4: 814-821.
22. Hauben E, Agranov E, Gothilf A, et al. Post-traumatic therapeutic vaccination with modified myelin self-antigen prevents complete paralysis while avoiding autoimmune disease. *J Clin Invest*. 2001;108:591-599.
23. Moalem G, Leibowitz-Amit R, Yoles E, Mor F, Cohen IR, Schwartz M. Autoimmune T cells protect neurons from secondary degeneration after central nervous system axotomy. *Nat Med*. 1999;5:49-55.
24. Zhao W, Xie W, Xiao Q, Beers DR, Appel SH. Protective effects of an anti-inflammatory cytokine, interleukin-4, on motoneuron toxicity induced by activated microglia. *J Neurochem*. 2006;99:1176-1187.
25. Shechter R, London A, Varol C, et al. Infiltrating blood-derived macrophages are vital cells playing an anti-inflammatory role in recovery from spinal cord injury in mice. *PLoS Med*. 2009;6:e1000113.
26. London A, Itskovich E, Benhar I, et al. Neuroprotection and progenitor cell renewal in the injured adult murine retina requires healing monocyte-derived macrophages. *J Exp Med*. 2011;208:23-39.
27. Benowitz LI, Popovich PG. Inflammation and axon regeneration. *Curr Opin Neurol*. 2011;24:577-583.
28. Louveau A, Harris TH, Kipnis J. Revisiting the mechanisms of CNS immune privilege. *Trends Immunol*. 2015;36:569-577.
29. Shechter R, Raposo C, London A, Sagi I, Schwartz M. The glial scar-monocyte interplay: a pivotal resolution phase in spinal cord repair. *PLoS One*. 2011;6:e27969.
30. Cohen M, Matcovitch O, David E, et al. Chronic exposure to TGFbeta1 regulates myeloid cell inflammatory response in an IRF7-dependent manner. *EMBO J*. 2014;33: 2906-2921.
31. Raposo C, Graubardt N, Cohen M, et al. CNS repair requires both effector and regulatory T cells with distinct temporal and spatial profiles. *J Neurosci*. 2014;34: 10141-10155.
32. Kigerl KA, Gensel JC, Ankeny DP, Alexander JK, Donnelly DJ, Popovich PG. Identification of two distinct macrophage subsets with divergent effects causing either neurotoxicity or regeneration in the injured mouse spinal cord. *J Neurosci*. 2009;29: 13435-13444.
33. Ziv Y, Ron N, Butovsky O, et al. Immune cells contribute to the maintenance of neurogenesis and spatial learning abilities in adulthood. *Nat Neurosci*. 2006;9:268-275.
34. Miller AH. 2010. Depression and immunity: a role for T cells? *Brain Behav Immun*. 2010; 24:1-8.
35. Wolf SA, Steiner B, Akpınarlı A, et al. CD4-positive T lymphocytes provide a neuroimmunological link in the control of adult hippocampal neurogenesis. *J Immunol*. 2009; 182:3979-3984.
36. Herz J, Filiano AJ, Smith A, Yogev N, Kipnis J. Myeloid cells in the central nervous system. *Immunity*. 2017;46:943-956.
37. Louveau A, Plog BA, Antila S, Alitalo K, Nedergaard M, Kipnis J. Understanding the functions and relationships of the glymphatic system and meningeal lymphatics. *J Clin Invest*. 2017;127:3210-3219.
38. Shechter R, Miller O, Yovel G, et al. Recruitment of beneficial M2 macrophages to injured spinal cord is orchestrated by remote brain choroid plexus. *Immunity*. 2013;38:555-569.
39. Baruch K, Ron-Harel N, Gal H, et al. CNS-specific immunity at the choroid plexus shifts toward destructive Th2 inflammation in brain aging. *Proc Natl Acad Sci U S A*. 2013;110: 2264-2269.
40. Emerich DF, Skinner SJ, Borlongan CV, Vascunellos AV, Thanos CG. The choroid plexus in the rise, fall and repair of the brain. *Bioessays*. 2005;27:262-274.
41. Engelhardt B, Wolburg-Buchholz K, Wolburg H. Involvement of the choroid plexus in central nervous system inflammation. *Microsc Res Tech*. 2001;52:112-129.
42. Marques F, Sousa JC, Correia-Neves M, Oliveira P, Sousa N, Palha JA. The choroid plexus response to peripheral inflammatory stimulus. *Neuroscience*. 2007;144:424-430.
43. Redzic ZB, Segal MB. The structure of the choroid plexus and the physiology of the choroid plexus epithelium. *Adv Drug Deliv Rev*. 2004;56:1695-1716.
44. Lampron A, Elali A, Rivest S. Innate immunity in the CNS: redefining the relationship between the CNS and Its environment. *Neuron*. 2013;78:214-232.
45. Butovsky O, Bukshpan S, Kunis G, Jung S, Schwartz M. Microglia can be induced by IFN-gamma or IL-4 to express neural or dendritic-like markers. *Mol Cell Neurosci*. 2007;35:490-500.
46. Butovsky O, Koronyo-Hamaoui M, Kunis G, et al. Glatiramer acetate fights against Alzheimer's disease by inducing dendritic-like microglia expressing insulin-like growth factor 1. *Proc Natl Acad Sci U S A*. 2006;103:11784-11789.
47. Koronyo-Hamaoui M, Ko MK, Koronyo Y, et al. Attenuation of AD-like neuropathology by harnessing peripheral immune cells: local elevation of IL-10 and MMP-9. *J Neurochem*. 2009;111:1409-1424.
48. Simard AR, Soulet D, Gowing G, Julien JP, Rivest S. Bone marrow-derived microglia play a critical role in restricting senile plaque formation in Alzheimer's disease. *Neuron*. 2006;49: 489-502.
49. Derecki NC, Cronk JC, Lu Z, et al. Wild-type microglia arrest pathology in a mouse model of Rett syndrome. *Nature*. 2012;484:105-109.
50. Town T, Laour Y, Pittenger C, et al. Blocking TGF-beta-Smad2/3 innate immune signaling mitigates Alzheimer-like pathology. *Nat Med*. 2008;14:681-687.
51. Varvel NH, Grathwohl SA, Baumann F, et al. Microglial repopulation model reveals a robust

Original article

Alzheimer disease immunotherapy - Schwartz et al

- homeostatic process for replacing CNS myeloid cells. *Proc Natl Acad Sci U S A*. 2012;109:18150-18155.
52. El Khoury J, Toft M, Hickman SE, et al. Ccr2 deficiency impairs microglial accumulation and accelerates progression of Alzheimer-like disease. *Nat Med*. 2007;13:432-438.
53. Beers DR, Henkel JS, Zhao W, Wang J, Appel SH. CD4+ T cells support glial neuroprotection, slow disease progression, and modify glial morphology in an animal model of inherited ALS. *Proc Natl Acad Sci U S A*. 2008;105:15558-15563.
54. Kunis G, Baruch K, Miller O, Schwartz M. Immunization with a myelin-derived antigen activates the brain's choroid plexus for recruitment of immunoregulatory cells to the CNS and attenuates disease progression in a mouse model of ALS. *J Neurosci*. 2015;35:6381-6393.
55. Wisniewski HM, Barcikowska M, Kida E. Phagocytosis of beta/A4 amyloid fibrils of the neuritic neocortical plaques. *Acta Neuropathol*. 1991;81:588-590.
56. Akiyama H, Kondo H, Mori H, et al. The amino-terminally truncated forms of amyloid beta-protein in brain macrophages in the ischemic lesions of Alzheimer's disease patients. *Neurosci Lett*. 1996;219:115-118.
57. Koronyo Y, Salumbides BC, Sheyn J, et al. Therapeutic effects of glatiramer acetate and grafted CD115+ monocytes in a mouse model of Alzheimer's disease. *Brain*. 2015;138(Pt 8):2399-422.
58. Rolls A, Shechter R, Schwartz M. The bright side of the glial scar in CNS repair. *Nat Rev Neurosci*. 2009;10:235-241.
59. Simard AR, Rivest S. Neuroprotective properties of the innate immune system and bone marrow stem cells in Alzheimer's disease. *Mol Psychiatry*. 2006;11:327-335.
60. Baruch K, Deczkowska A, David E, et al. Aging. Aging-induced type I interferon response at the choroid plexus negatively affects brain function. *Science*. 2014;346:89-93.
61. Baruch K, Deczkowska A, Rosenzweig N, et al. PD-1 immune checkpoint blockade reduces pathology and improves memory in mouse models of Alzheimer's disease. *Nat Med*. 2016;22:135-137.
62. Baruch K, Rosenzweig N, Kertser A, et al. Breaking immune tolerance by targeting Foxp3+ regulatory T cells mitigates Alzheimer's disease pathology. *Nat Commun*. 2015;6:7967.
63. Marsh SE, Abud EM, Lakatos A, et al. The adaptive immune system restrains Alzheimer's disease pathogenesis by modulating microglial function. *Proc Natl Acad Sci U S A*. 2016;113:E1316-1325.
64. Rosenzweig N, Dvir-Sternfeld R, Tsitsou-Kampeli A, et al. PD-1/PD-L1 checkpoint blockade harnesses monocyte-derived macrophages to combat cognitive impairment in a mouse model of tau-associated dementia. *Nat Commun*. 2019;10:465.
65. Gotsman I, Grabie N, Dacosta R, Sukhova G, Sharpe A, Lichtman AH. Proatherogenic immune responses are regulated by the PD-1/PD-L pathway in mice. *J Clin Invest*. 2007;117:2974-2982.
66. Pittet CL, Newcombe J, Antel JP, Arbour N. The majority of infiltrating CD8 T lymphocytes in multiple sclerosis lesions is insensitive to enhanced PD-L1 levels on CNS cells. *Glia*. 2011;59:841-856.
67. Ansari MJ, Salama AD, Chitnis T, et al. The programmed death-1 (PD-1) pathway regulates autoimmune diabetes in nonobese diabetic (NOD) mice. *J Exp Med*. 2013;198:63-69.
68. Yang W, Li H, Chen PW, et al. PD-L1 expression on human ocular cells and its possible role in regulating immune-mediated ocular inflammation. *Invest Ophthalmol Vis Sci*. 2009;50:273-280.
69. Carter L, Fouser LA, Jussif J, et al. PD-1:PD-L inhibitory pathway affects both CD4(+) and CD8(+) T cells and is overcome by IL-2. *Eur J Immunol*. 2002;32:634-643.
70. Freeman GJ, Long AJ, Iwai Y, et al. Engagement of the PD-1 immunoinhibitory receptor by a novel B7 family member leads to negative regulation of lymphocyte activation. *J Exp Med*. 2002;192:1027-1034.
71. Fife BT, Pauken KE, Eagar TN, et al. Interactions between PD-1 and PD-L1 promote tolerance by blocking the TCR-induced stop signal. *Nat Immunol*. 2009;10:1185-1192.
72. Karwacz K, Bricogne C, MacDonald D, Arce F, Bennett CL, Collins M, Escors D. PD-L1 co-stimulation contributes to ligand-induced T cell receptor down-modulation on CD8+ T cells. *EMBO Mol Med*. 2011;3:581-592.
73. Mielke MM, Vemuri P, Rocca WA. Clinical epidemiology of Alzheimer's disease: assessing sex and gender differences. *Clin Epidemiol*. 2014;6:37-48.
74. Alzheimer's A. 2013 Alzheimer's disease facts and figures. *Alzheimers Dement*. 2013;9:208-245.
75. Deczkowska A, Schwartz M. Targeting neuro-immune communication in neurodegeneration: Challenges and opportunities. *J Exp Med*. 2018;215:2702-2704.