



Genome Sequence of *Oenococcus oeni* OE37, an Autochthonous Strain Isolated from an Italian White Wine

 Antonella Costantini,^a Ana B. Blazquez,^b  Francesco Cerutti,^c Enrico Vaudano,^a Simone Peletto,^c Juan-Carlos Saiz,^b Emilia Garcia-Moruno^a

^aConsiglio per la Ricerca in Agricoltura e l'Analisi dell'Economia Agraria, Research Centre for Viticulture and Enology, Asti, Italy

^bDepartamento de Biotecnología, Instituto Nacional de Investigación y Tecnología Agraria y Alimentaria, Madrid, Spain

^cIstituto Zooprofilattico Sperimentale del Piemonte, Liguria e Valle d'Aosta, Torino, Italy

ABSTRACT *Oenococcus oeni* OE37 is an autochthonous strain that was isolated from a Chardonnay wine from Piedmont (Italy) during spontaneous malolactic fermentation. Here, the OE37 genome sequence is presented, and a brief description of the main genes is reported.

Malolactic fermentation (MLF) is a biological process in which malic acid is converted to lactic acid and carbon dioxide by decarboxylation through the NADH-malic enzyme (1). *Oenococcus oeni* is the main bacterium responsible for conducting this process because of its ability to survive the harsh wine conditions and its production of desirable wine sensory attributes. Currently, the most significant studies have been focused on describing the occurrence of MLF, lactic acid bacteria, and *O. oeni* starter selection (2, 3). The number of reported *O. oeni* genomes is increasing, but only 4 of 242 records currently available in GenBank are complete genome sequences.

OE37 is an autochthonous strain that was isolated from a Chardonnay wine during spontaneous MLF by plating diluted wine onto MRS agar plates, as described by Doria et al. (4). In this study, the OE37 strain was multiplied in MRS broth, and DNA was purified using the ArchivePure yeast/Gram-positive DNA kit (Eppendorf, Milan, Italy). Whole-genome sequencing was performed by Macrogen (South Korea). A library was prepared using the TruSeq Nano DNA kit, and sequencing was performed using the Illumina HiSeq 2500 platform, with a paired-end read length of 101 bp. Trimmomatic v0.36 (5) was used to remove adapter sequences; after trimming, a total of 23,084,810 reads, with 96.72% of bases having a quality score above Q30, were found. The GC content was 37.53%, and the mean coverage depth was 1,132.21×. Reads were mapped, using *Oenococcus* PSU-1 as the reference genome (GenBank accession number [NC_008528.1](https://doi.org/10.1128/MRA.00582-20)), with BWA v0.7.17 (6) and SAMtools v1.9-1 (7). For all software, default parameters were used.

The sequence was annotated by the National Center for Biotechnology Information (NCBI) Prokaryotic Genome Annotation Pipeline (PGAP). The resulting draft complete sequence is 1,718,702 bp, with a GC content of 37.99%, and contains 1,720 coding sequences, 46 tRNAs, and 6 rRNA-like motifs. Assembly completeness was checked with Benchmarking Universal Single-Copy Orthologs (BUSCO) v.1 (8) on the gVolante server (9), using bacteria as the selected reference gene set, which resulted in 97.5% completeness.

In winemaking, different aspects are considered for starter selection, such as the ability to tolerate the harsh wine conditions and physiological, biochemical, and technological properties (10). The observation of the genes contained in the OE37 genome confirmed the ability of this bacterium to conduct MLF, since malic enzyme is present; genes related to citrate metabolism are also present, as is a gene coding for

Citation Costantini A, Blazquez AB, Cerutti F, Vaudano E, Peletto S, Saiz J-C, Garcia-Moruno E. 2020. Genome sequence of *Oenococcus oeni* OE37, an autochthonous strain isolated from an Italian white wine. *Microbiol Resour Announc* 9:e00582-20. <https://doi.org/10.1128/MRA.00582-20>.

Editor Kenneth M. Stedman, Portland State University

Copyright © 2020 Costantini et al. This is an open-access article distributed under the terms of the [Creative Commons Attribution 4.0 International license](https://creativecommons.org/licenses/by/4.0/).

Address correspondence to Antonella Costantini, antonella.costantini@crea.gov.it.

Received 28 May 2020

Accepted 3 September 2020

Published 24 September 2020

diacetyl reductase. Diacetyl is the main aromatic compound associated with MLF and is derived from citrate consumption (11). Concerning the organoleptic quality, other genes are involved and are present in this strain, such as various glucosidase enzyme genes, including that for a β -glucosidase (12, 13).

Regarding the mechanism of the stress response of *O. oeni*, several genes previously reported as being implicated in the response are present in the OE37 genome, such as those involved in cell wall biosynthesis (e.g., *N*-acetylmuramoyl-L-alanine amidase and D-alanyl-D-alanine carboxypeptidase), whose expression is strongly influenced by the presence of ethanol (14, 15). Genes coding for exopolysaccharides, which are important for the adaptation of *O. oeni* to its ecological niche (16), are also present.

The genome sequence of this *O. oeni* strain confirms the current knowledge regarding malolactic bacteria and their impact on wine.

Data availability. The genome sequence was deposited in GenBank under accession number [CP053280](https://doi.org/10.1093/bioinformatics/btp698). The raw sequencing data were deposited in the SRA database under accession number [PRJNA607182](https://doi.org/10.1093/bioinformatics/btp698).

REFERENCES

- Spano G, Massa S. 2006. Environmental stress response in wine lactic acid bacteria: beyond *Bacillus subtilis*. *Crit Rev Microbiol* 32:77–86. <https://doi.org/10.1080/10408410600709800>.
- Cañas PMI, Pérez PR, Prieto SS, Herreros MLP. 2009. Ecological study of lactic acid microbiota isolated from Tempranillo wines of Castilla-La Mancha. *J Biosci Bioeng* 108:220–224. <https://doi.org/10.1016/j.jbiosc.2009.04.001>.
- González-Arenzana L, Santamaría P, López R, López-Alfaro I. 2013. Indigenous lactic acid bacteria communities in alcoholic and malolactic fermentations of Tempranillo wines elaborated in ten wineries of La Rioja (Spain). *Food Res Int* 50:438–445. <https://doi.org/10.1016/j.foodres.2012.11.008>.
- Doria F, Napoli C, Costantini A, Berta G, Saiz J-CC, Garcia-Moruno E. 2013. Development of a new method for detection and identification of *Oenococcus oeni* bacteriophages based on endolysin gene sequence and randomly amplified polymorphic DNA. *Appl Environ Microbiol* 79:4799–4805. <https://doi.org/10.1128/AEM.01307-13>.
- Bolger AM, Lohse M, Usadel B. 2014. Trimmomatic: a flexible trimmer for Illumina sequence data. *Bioinformatics* 30:2114–2120. <https://doi.org/10.1093/bioinformatics/btu170>.
- Li H, Durbin R. 2010. Fast and accurate long-read alignment with Burrows-Wheeler transform. *Bioinformatics* 26:589–595. <https://doi.org/10.1093/bioinformatics/btp698>.
- Li H, Handsaker B, Wysoker A, Fennell T, Ruan J, Homer N, Marth G, Abecasis G, Durbin R, 1000 Genome Project Data Processing Subgroup. 2009. The Sequence Alignment/Map format and SAMtools. *Bioinformatics* 25:2078–2079. <https://doi.org/10.1093/bioinformatics/btp352>.
- Simão FA, Waterhouse RM, Ioannidis P, Kriventseva EV, Zdobnov EM. 2015. BUSCO: assessing genome assembly and annotation completeness with single-copy orthologs. *Bioinformatics* 31:3210–3212. <https://doi.org/10.1093/bioinformatics/btv351>.
- Nishimura O, Hara Y, Kuraku S. 2017. gVolante for standardizing completeness assessment of genome and transcriptome assemblies. *Bioinformatics* 33:3635–3637. <https://doi.org/10.1093/bioinformatics/btx445>.
- Capozzi V, Russo P, Beneduce L, Weidmann S, Grieco F, Guzzo J, Spano G. 2010. Technological properties of *Oenococcus oeni* strains isolated from typical southern Italian wines. *Lett Appl Microbiol* 50:327–334. <https://doi.org/10.1111/j.1472-765x.2010.02795.x>.
- Izquierdo Cañas PM, García Romero E, Gómez Alonso S, Palop Herreros MLL. 2008. Changes in the aromatic composition of Tempranillo wines during spontaneous malolactic fermentation. *J Food Compos Anal* 21:724–730. <https://doi.org/10.1016/j.jfca.2007.12.005>.
- Grimaldi A, Bartowsky E, Jiranek V. 2005. A survey of glycosidase activities of commercial wine strains of *Oenococcus oeni*. *Int J Food Microbiol* 105:233–244. <https://doi.org/10.1016/j.jfoodmicro.2005.04.011>.
- Fia G, Millarini V, Granchi L, Bucalossi G, Guerrini S, Zanoni B, Rosi I. 2018. β -Glucosidase and esterase activity from *Oenococcus oeni*: screening and evaluation during malolactic fermentation in harsh conditions. *LWT* 89:262–268. <https://doi.org/10.1016/j.lwt.2017.10.060>.
- Costantini A, Rantsiou K, Majumder A, Jacobsen S, Pessione E, Svensson B, Garcia-Moruno E, Cocolin L. 2015. Complementing DIGE proteomics and DNA subarray analyses to shed light on *Oenococcus oeni* adaptation to ethanol in wine-simulated conditions. *J Proteomics* 123:114–127. <https://doi.org/10.1016/j.jprot.2015.04.019>.
- Olguín N, Champomier-Vergès M, Anglade P, Baraige F, Cordero-Otero R, Bordons A, Zagorec M, Reguant C. 2015. Transcriptomic and proteomic analysis of *Oenococcus oeni* PSU-1 response to ethanol shock. *Food Microbiol* 51:87–95. <https://doi.org/10.1016/j.fm.2015.05.005>.
- Dimopoulou M, Vuillemin M, Campbell-Sills H, Lucas PM, Ballestra P, Miot-Sertier C, Favier M, Coulon J, Moine V, Doco T, Roques M, Williams P, Petrel M, Gontier E, Moulis C, Remaud-Simeon M, Dols-Lafargue M. 2014. Exopolysaccharide (EPS) synthesis by *Oenococcus oeni*: from genes to phenotypes. *PLoS One* 9:e98898. <https://doi.org/10.1371/journal.pone.0098898>.