

Recent Progress in Synthetic and Natural Catechol-*O*-methyltransferase Inhibitors for Neurological Disorders

Sandeep Bindra, Ananya Datta, Haya Khader Ahmad Yasin,* Riya Rachel Thomas, Shailesh Verma, Ankita Patel, Della Grace Thomas Parambi, Suraj N. Mali, T. M. Rangarajan,* and Bijo Mathew*



Cite This: *ACS Omega* 2024, 9, 44005–44018



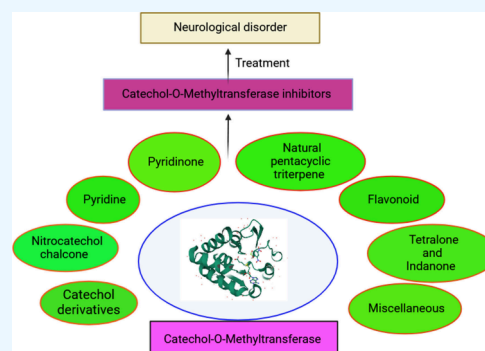
Read Online

ACCESS |

Metrics & More

Article Recommendations

ABSTRACT: Catechol-*O*-methyltransferase (COMT) inhibitors have played a crucial role in the development of potent and selective drugs for the treatment of Parkinson's disease, depression, and anxiety disorders. This review provides a comprehensive analysis of the structure–activity relationship (SAR) of COMT inhibitors, highlighting key structural features and pharmacophoric elements that govern their potency, selectivity, and pharmacokinetic properties. This review also discusses the application of SAR principles in the design and optimization of COMT inhibitors. Our analysis reveals the emergence of novel chemical scaffolds and the potential for COMT inhibitors to address unmet medical needs in neurology and psychiatry. This Perspective serves as a valuable resource for clinicians and researchers, providing insights into the rational design of COMT inhibitors and the development of next-generation therapeutics.



1. INTRODUCTION

Catechol-*O*-methyltransferase (COMT) is a divalent magnesium (Mg^{2+}) dependent enzyme that catalyzes the transfer of a methyl group to catechol substrates by using *S*-adenosyl-*L*-methionine (SAM) as a methyl donor, producing *O*-methylated catechol and *S*-adenosyl-*L*-homocysteine as reaction products.¹ Both endogenous and exogenous neurotransmitters, such as epinephrine, noradrenaline, and dopamine, as well as levodopa, the metabolic precursor of dopamine, are examples of COMT substrates.² The physiological actions of neurotransmitters are terminated by COMT metabolism, which makes this enzyme relevant for therapeutic purposes. Many central and peripheral nervous system disorders, such as Parkinson's disease (PD), depression, schizophrenia, and other diseases linked to dopamine deficiency, can be treated with drugs that target COMT.³

COMT exists in two isoforms that are expressed from two promoters: the membrane-bound form, MB-COMT, which is more frequently found in the brain, and the soluble S-COMT isoform, which is expressed in most tissues, including the liver, blood, and kidneys.⁴ MB-COMT is particularly noteworthy to investigate as a target due to its function in controlling extracellular dopamine levels inside the prefrontal cortex.⁵ The COMT gene is located on chromosome 22q11.2 and is 27.22 kb long.⁶ The 158th amino acid residue of the membrane-bound isoform (or the 108th amino acid of the soluble form) is changed from valine (Val) to methionine (Met) by a frequent nonsynonymous single-nucleotide polymorphism (rs4680). Because the Met variant is more thermolabile at physiological

temperature, its presence causes a 4-fold decrease in COMT enzyme activity levels in the prefrontal cortex.⁷ There are several crystal structures available for COMTs in rats and humans. Both the human and rat COMTs (Figure 1) are members of the highly structurally conserved SAM-dependent methyltransferase fold family, and they have 81% sequence

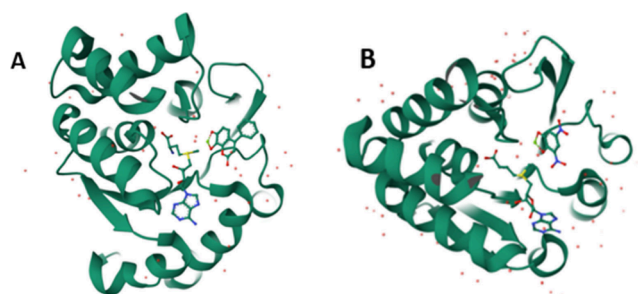


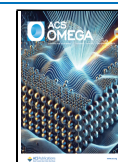
Figure 1. (A) Crystal structure of rat COMT in complex with *S*-adenosyl-methionine (SAM), dinitrocatechol (DNC), and Mg^{2+} . (B) Crystal structure of human COMT in complex with *S*-adenosyl-methionine (SAM), 7,8-dihydroxy-4-phenyl-2*H*-chromen-2-one, and Mg^{2+} .

Received: July 4, 2024

Revised: October 3, 2024

Accepted: October 11, 2024

Published: October 23, 2024



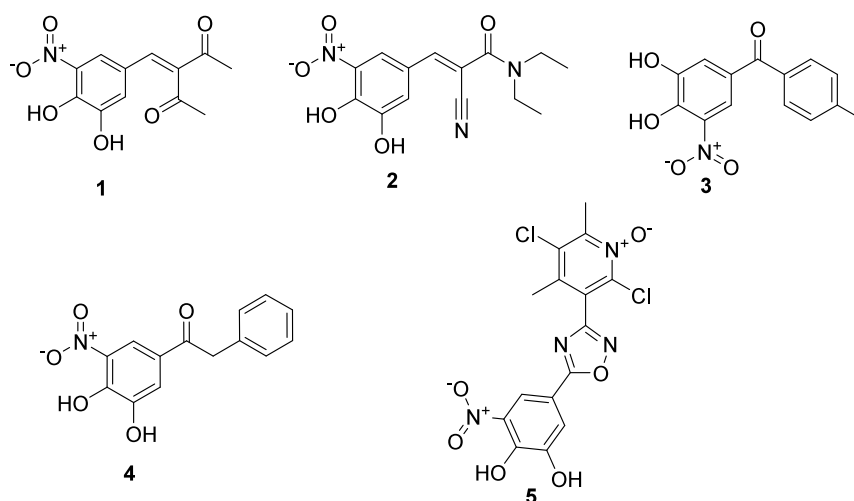


Figure 2. Structures of clinically used COMT inhibitors: nitecapone (1), entacapone (2), tolcapone (3), nebicapone (4), and opicapone (5).

similarity.⁸ Specifically, the COMT enzyme is made up of two sets of α -helices (helices $\alpha 1$ – $\alpha 5$ on one side and helices $\alpha 6$ – $\alpha 8$ on the other) sandwiched around a seven-stranded β -sheet core (ordered in an order of 3, 2, 1, 4, 5, 7, 6). Strand 7 is antiparallel to the others in the β -sheet. Along the first half of the core β -sheet ($\beta 1$ – $\beta 4$), the cofactor SAM interacts with conserved residues.⁹ E90 ($\beta 2$) makes hydrogen bonds with SAM's ribose hydroxyl groups. SAM's adenine ring interacts via van der Waals interactions with residues I91 ($\beta 2$), A118 ($\beta 3$ – $\alpha 6$ loop), and W143 ($\beta 4$ – $\alpha 7$ loop) and forms hydrogen bonds with S119 and Q120 ($\alpha 6$). The coordination of methionine in SAM is achieved through hydrophobic contacts with M40, V42, and Y68, and hydrogen bonds with residues V42 ($\alpha 2$ – $\alpha 3$ loop), S72 ($\alpha 4$), and D141 ($\beta 4$). The methyl group (CH_3) of the SAM methionine sulfur atom is orientated toward the substrate binding site, more precisely toward the catechol oxygen atom that has to be methylated. The “gatekeeper” residues W38 and P174, together with M40, L198, and W143, define the shallow pocket that serves as the substrate-binding site. All of these residues being hydrophobic implies that van der Waals interactions are the primary mechanisms underlying ligand binding. Every single crystal structure of COMT complexed with a ligand contains the magnesium ion (Mg^{2+}), a cofactor essential for methylation.¹⁰ The side chains of D141, D169, and N170, the two hydroxyl groups of the catechol substrate, and a water molecule all work together to coordinate the Mg^{2+} ion octahedrally in the active site. This is unambiguous evidence that Mg^{2+} aids in the enzymatic activity by making the substrate more soluble. This explains why the ion is necessary for catalysis mediated by COMT. It is noteworthy that the $\text{p}K_a$ of the catechol hydroxyl is lowered by the Mg^{2+} ion, causing it to be more ionized.¹¹ In general, human COMT has lower K_m (Michaelis constant) values for catechol substrate methylation than rat COMT.¹² An analysis comparing the human and rat active sites explains why the K_m values of these two proteins and other kinetic characteristics differ. Human COMT has shorter Mg^{2+} ligand distances in its crystal structure than rat COMT, suggesting that substrate binding is more robust (lower K_m value) in the human protein.⁹ The majority of residues in the sites are identical, though three residues are found to differ in the SAM binding sites of these two proteins: M89, M91, and Y95 in the rat protein and I89, I91, and C95 in the human protein. In

comparison to the human protein, the rat protein residues are larger and interact with SAM more intimately. Additionally, two changes in charge (R201 and E202 in humans compared to M201 and K202 in rats) are present at the substrate-binding site. It seems unlikely that the two residues will interact directly with the substrate. However, because of their position at the pocket entry, they might have an impact on V_{max} in addition to substrate binding and release.

Several COMT inhibitors have been developed to treat neurodegenerative disorders to date. The early developed COMT inhibitors, often referred to as “first-generation” inhibitors, inhibited COMT from extending the physiological action of endogenous neurotransmitters, albeit their effectiveness on noradrenergic, adrenergic, and dopaminergic pathways was limited.¹³ The derivatives of pyrogallol and catechols, such as gallic acid, caffeic acid, U-0521, 2-hydroxy estrogens, or flavonoids like quercetin or rutin are examples of first-generation COMT inhibitors. Moreover, it was questioned how “first-generation” COMT inhibitors affected the inactivation of neurotransmitters at receptor sites. During the 1970s, there was escalating interest in COMT following the discovery that the enzyme was involved in the metabolic breakdown of the popular antiparkinsonian medication L-DOPA (levodopa). PD is a chronic degenerative neurological disorder caused by the reduction of dopamine levels due to the death of dopaminergic cells in the substantia nigra pars compacta. It is possible to control the levels of this neurotransmitter in the brain by using L-DOPA, a biological precursor of dopamine.¹⁴ L-DOPA's short *in vivo* half-life is one of its main disadvantages, despite its advantageous capacity to regulate dopamine levels in the brain. COMT inhibitors prolong the pharmacological impact of L-DOPA and reduce the therapeutic dose by inhibiting COMT from converting L-DOPA to 3-*O*-methyl-L-DOPA (3-OMD) in peripheral tissues. In order to target COMT and provide a more continuous and prolonged distribution of dopamine to the striatum, various research groups from academia and the pharmaceutical industry have been working in the last 30 years to optimize L-DOPA therapy. In the process of finding novel COMT inhibitors, “second-generation” inhibitors were discovered, which had better pharmacokinetic characteristics than their “first-generation” predecessors. Second generation inhibitor examples of clinically relevant COMT inhibitors for

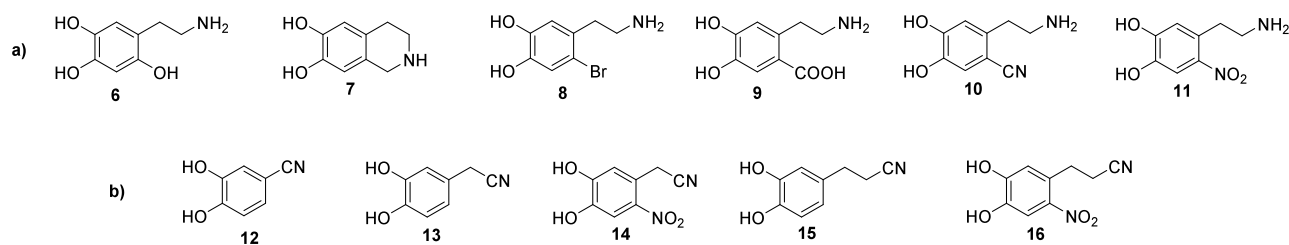


Figure 3. Designed and synthesized (a) dopamine and (b) catechol derivatives.

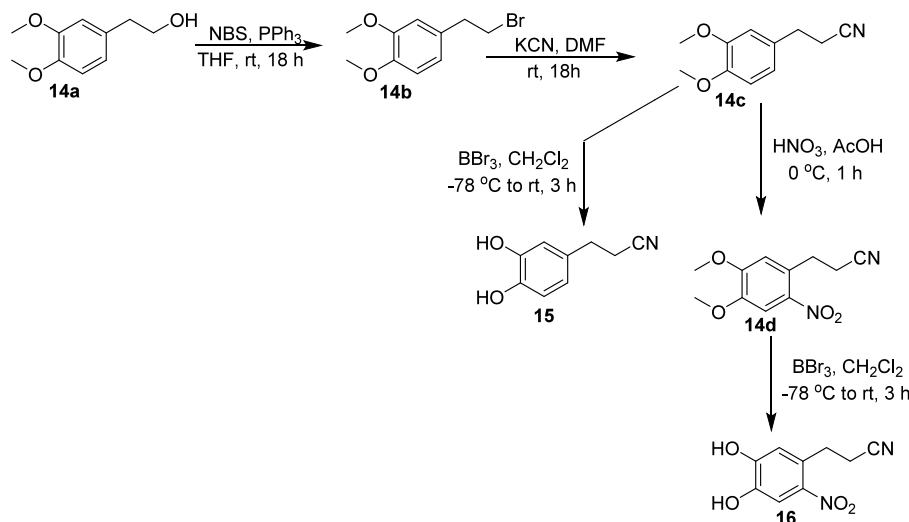


Figure 4. Synthesis of the catechol derivatives.

the adjunctive therapy of Parkinson's disease (PD) are nitecapone **1** (OR-462), entacapone **2** (OR-611), tolcapone **3** (Ro40-7592), and nebicapone **4** (BIA 3-202) (Figure 2), while opicapone **5** (BIA 9-1067) (Figure 2) is an example of a third-generation COMT inhibitor having lower toxicity and higher inhibitory activity. Recently, Akhtar et al. summarized the last 50 years of research on COMT inhibitors from 2019 as a review article.¹⁵ A timely update is necessary for the further design and development of the novel COMT inhibitors. Therefore, in this review, we cover the COMT inhibitors published from 2019 to 2024.

2. COMT INHIBITOR STRUCTURE BASED DESIGN

2.1. Catechol Derivatives. Several FDA-approved drugs, for example, entacapone, as COMT inhibitors for the treatment of Parkinson's disease showed poor performance due to various pharmacokinetic and pharmacodynamics reasons such as their low bioavailability and low metabolic stability that have resulted in increased and repeated doses throughout the day. Also, some side effects, such as hepatitis, make therapy less efficient. Therefore, continuous efforts have been made to deliver a good therapeutic drug molecule. Recently, Peterson et al. designed and synthesized six substituted dopamine analogues (**6–11**) and five catechol derivatives (**12–16**) (Figure 3) for their COMT inhibition studies.¹⁶ The authors undertook the work based on an *ab initio* study of the designed compounds' binding energies in the enzyme's active site.¹⁷

Dopamine derivatives were synthesized by methods previously reported by the authors,¹⁶ and the catechol derivatives were achieved from the corresponding commercially available starting materials by nitration and deprotection

of hydroxyl groups by BBr_3 . The side chain $-\text{CN}$ group was introduced from corresponding 2-phenyl ethanol to 2-phenyl ethyl bromide by the Appel reaction and subsequent conversion to nitrile by KCN. Finally, the deprotection of methoxy groups afforded the catechol derivatives (Figure 4).

The synthesized compounds were checked for their stability under the assay conditions in a phosphate buffer at pH 7.4 for 1 h and then analyzed by HPLC before the inhibition study, revealing that all of the compounds experienced little to no degradation over the time except 5-(2-aminoethyl)benzene-1,2,4-triol (**6**), 1,2,3,4-tetrahydroisoquinoline-6,7-diol (**7**), 2-(4,5-dihydroxy-2-nitrophenyl)acetonitrile (**14**), and 3-(3,4-dihydroxyphenyl)propanenitrile (PCN) (**15**). The *in vitro* S-COMT inhibition by the synthesized dopamine derivatives (**6–11**) was tested at two concentrations, 10 and 100 μM , with reference to 3,5-dinitrocatechol (DNC) as a positive control. The authors did not report the IC_{50} values for the compounds. Dopamine derivatives (**6–11**) did not show any inhibition effect at 10 μM . At 100 μM , the catechols 2-(2-aminoethyl)-4,5-dihydroxybenzonitrile (**10**) and 4-(2-aminoethyl)-5-nitrobenzene-1,2-diol (**11**) exhibited about 30% to 50% COMT inhibitory activity. At 10 μM , only 2-(4,5-dihydroxy-2-nitrophenyl)acetonitrile (**14**) showed a significant decrease in the COMT activity. However, at 100 μM , all the nitrile derivatives inhibited COMT activity to some degree with 2-(4,5-dihydroxy-2-nitrophenyl)acetonitrile (**14**) and 3-(4,5-dihydroxy-2-nitrophenyl)propanenitrile (**16**) demonstrating complete inhibition of COMT. The pK_a values and docking studies supported trends of the COMT inhibition effect of these compounds. The compounds with an electron-withdrawing nitro group help to lower the pK_a , but the neutral

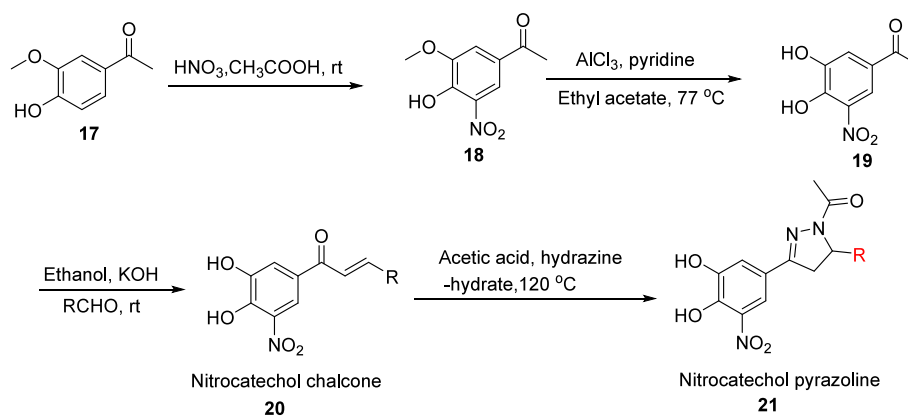


Figure 5. Synthesis of nitrocatechol chalcone derivatives.

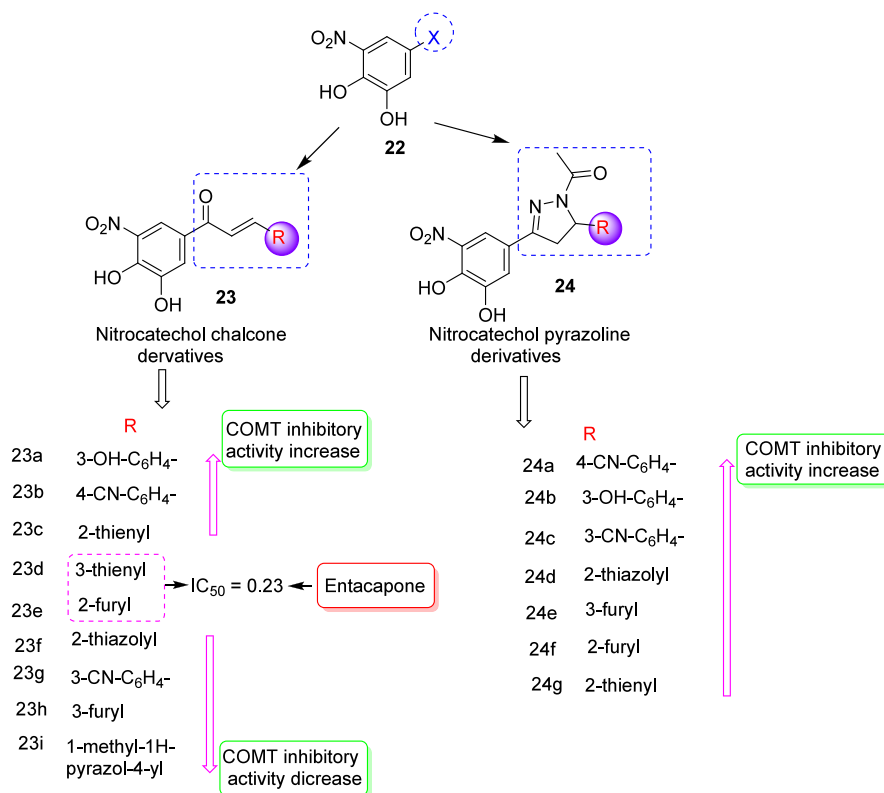


Figure 6. SAR of nitrocatechol chalcone and pyrazoline derivatives.

nitrile tail interacts more favorably with the residues at the back of the active site.

2.1.1. Nitrocatechol Chalcone Derivatives. Hitge et al. (2020) designed and synthesized a series of novel nitrocatechol chalcone and nitrocatechol pyrazoline derivatives as COMT inhibitors.¹⁸ The designed compounds were synthesized by nitration of 4-hydroxy-3-methoxyacetophenone (apocynin, **17**) with nitric acid in the presence of acetic acid to form the nitro derivative (5-nitro-apocynin; **18**). Demethylation of nitro derivative **18** in the presence of AlCl₃ in pyridine and ethyl acetate gave 5-acetyl-3-nitrocatechol (**19**), which on Claisen–Schmidt condensation reaction with an appropriate aldehyde in the presence of KOH in ethanol formed nitrocatechol chalcones (**20**). The nitrocatechol pyrazoline derivatives (**21**) were synthesized by reacting compounds **20** with hydrazine hydrate in the presence of acetic acid at 120 °C (Figure 5).¹⁹

Rat liver tissue was used as an enzyme source to determine the COMT inhibitory activity of the synthesized compounds.²⁰ Among the synthesized derivatives, nitrocatechol pyrazoline derivatives were found to be more potent than nitrocatechol chalcone derivatives. In a series of nitrocatechol pyrazoline derivatives, all the designed compounds demonstrated effective COMT inhibitory activity (IC₅₀ = 0.048 to 0.21 μM) compared to the standard molecule entacapone (IC₅₀ = 0.23 μM). In this series, compound **24a** was found to be the most active COMT inhibitor with an IC₅₀ value of 0.048 μM. The presence of the –CN group at the fourth position of the spacer phenyl, which attached to the R position, was responsible for the COMT inhibitory activity of compound **24a**. Substitution of the –CN group at the third position of spacer phenyl resulted in a slight decrease in COMT inhibitory activity. The replacement of the –CN group with the –OH group at the spacer phenyl also causes a decline in COMT inhibitory

activity. In this series, among the heterocyclic derivatives (24d–24g), compound 24d (2-thiazolyl substituted) was found to be the most active compound with an IC_{50} value of $0.16 \mu\text{M}$. Substitution of 2-furyl, 3-furyl, and 2-thienyl at the R position resulted in a slight decrease in COMT inhibitory activity.

In a series of nitrocatechol derivatives, compound 23a showed the highest COMT inhibitory activity with an IC_{50} value of $0.14 \mu\text{M}$. Substitution of the $-\text{OH}$ group at the third position of spacer phenyl, which attached to the R position, was responsible for the highest COMT inhibitory activity. Substitution of the $-\text{CN}$ group at the fourth position of spacer phenyl retains the COMT inhibitory activity, while substitution of the $-\text{CN}$ group at the third position of spacer phenyl results in a decrease in inhibitory activity. The substitution of heterocyclic moieties such as 3-thienyl and 2-furyl at the R position showed similar inhibitory activity compared to that of standard molecule entacapone. Substitution of 2-thiazolyl at the R position resulted in a slight decrease in COMT inhibitory activity; further substitution with 1-methyl-1H-pyrazol-4-yl at the R position resulted in the least potent COMT inhibitor 23i with an IC_{50} value of $0.29 \mu\text{M}$ (Figure 6).

2.2. Pyridine Derivatives. de Beer et al. (2021) introduced the design and synthesis of various non-nitrocatechol 3-hydroxypyridin-4-ones as COMT inhibitors.²¹ The designed compounds were synthesized by refluxing maltol with suitable primary amines in an acidified water/ethanol (0.9:0.1) mixture as a solvent to afford the 3-hydroxypyridine-4-ones (Figure 7).

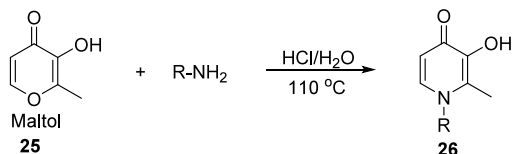


Figure 7. Synthesis of 3-hydroxypyridin-4-ones.

The COMT from porcine liver was used in the study, and (–)-norepinephrine was used as the enzyme substrate. Among the synthesized derivatives, compounds 27a and 27i were identified as active COMT inhibitors with IC_{50} values of $4.55 \mu\text{M}$ and $5.76 \mu\text{M}$, respectively, but not as active as the

reference inhibitors, entacapone and tolcapone. The spacer between the phenyl group and pyridone moiety by one to four carbon atoms significantly affected the COMT inhibitory activity. The spacers with 1- and 4-carbon atoms showed increased inhibitory effect; however, the spacer with 1-carbon, benzyl, showed the highest COMT inhibitory effect (Figure 8). The methyl substituent at the 4-position of the benzyl group exhibited slightly decreased inhibitory activity. The $-\text{Cl}$ substitution at the 4-position of the phenyl group showed a slight increase in inhibition, and the same substitution at the 3-position made no appreciable change in the activity. Docking of compound 27a indicated proper positioning for coordination with Mg^{2+} .

2.3. Pyridinone Derivatives. Ernst et al. (2019)²² designed and synthesized a series of fused bicyclic pyridinone derivatives and evaluated their inhibitory activity against COMT. The synthesis of the target molecules was started with $-\text{OH}$ protection of the hydroxyl group of kojic acid (5-hydroxy-2-(hydroxymethyl)-4H-pyran-4-one (28), to form compound 29. The N atom was installed in the kojic acid ring by ethylenediamine at $90 \text{ }^\circ\text{C}$ in ethanol through replacing the ring O atom of kojic acid (30). The cyclization to a fused heterocyclic system was then achieved by oxidation of the hydroxymethyl group and imine formation in one pot (31). The reduction of the imine with NaBH_4 yielded a pyridopyrazine fused bicyclic ring (32). Then functionalization of the N–H group (33) and successive deprotection of the benzyl group lead to the target molecules (34) as shown in Figure 9.

Initially, the COMT inhibition was assessed for the parent and seven N-functionalized compounds (Figure 10). Among them, the N-benzylated derivative exhibited COMT inhibition in the submicromolar range with an IC_{50} value of 220 nM . Others showed many fold loss in COMT inhibition. The group subsequently investigated COMT inhibition with N-benzylated compounds by modifying the electron-withdrawing groups (EWGs) and electron-releasing groups (ERGs) at the *ortho*- and *para*-positions. The result clearly showed that the EWGs and ERGs at the *ortho*-position had higher potency than the groups placed at the *para*- position. Among them, the $-\text{CH}_3$ group and $-\text{Cl}$ group at the *ortho*-position (36d,e) exhibited excellent inhibition effects with IC_{50} values of 50 and 40 nM , respectively (Figure 11).

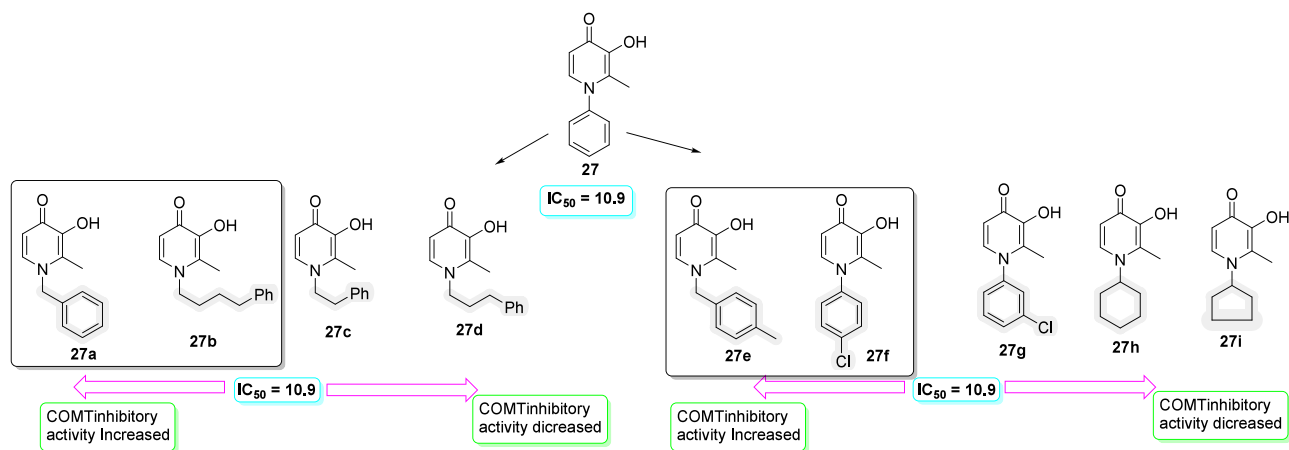


Figure 8. SAR of pyridin-4-one derivatives.

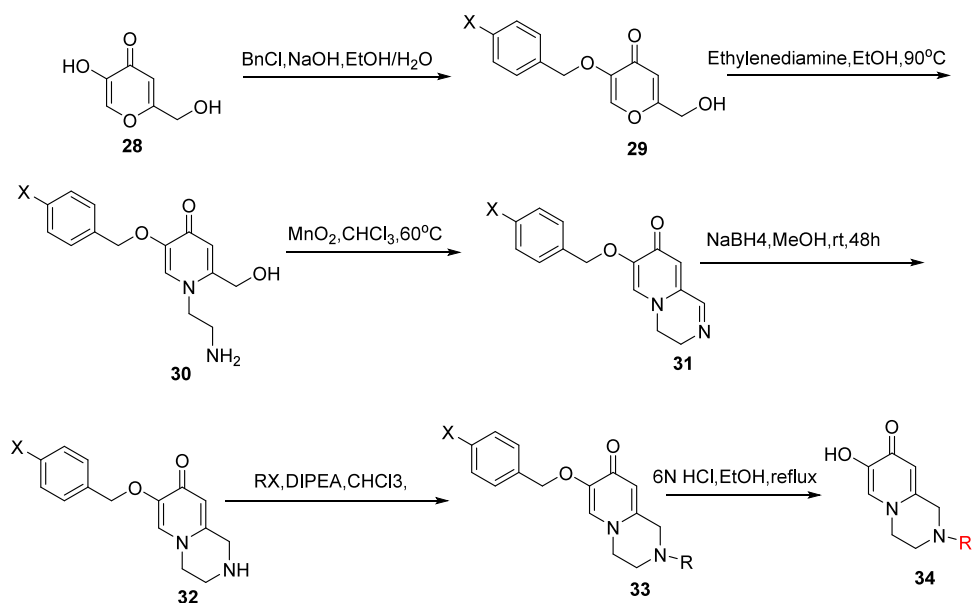


Figure 9. Synthetic route for fused bicyclic hydroxypyridones.

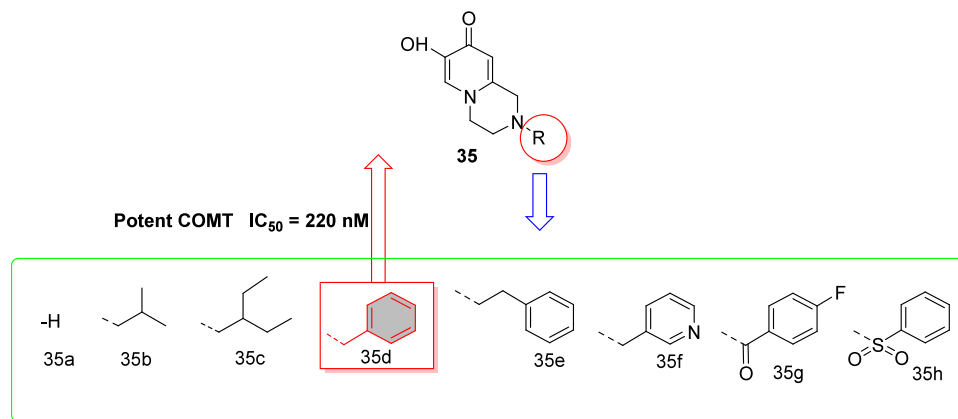


Figure 10. SAR of pyridinone derivatives.

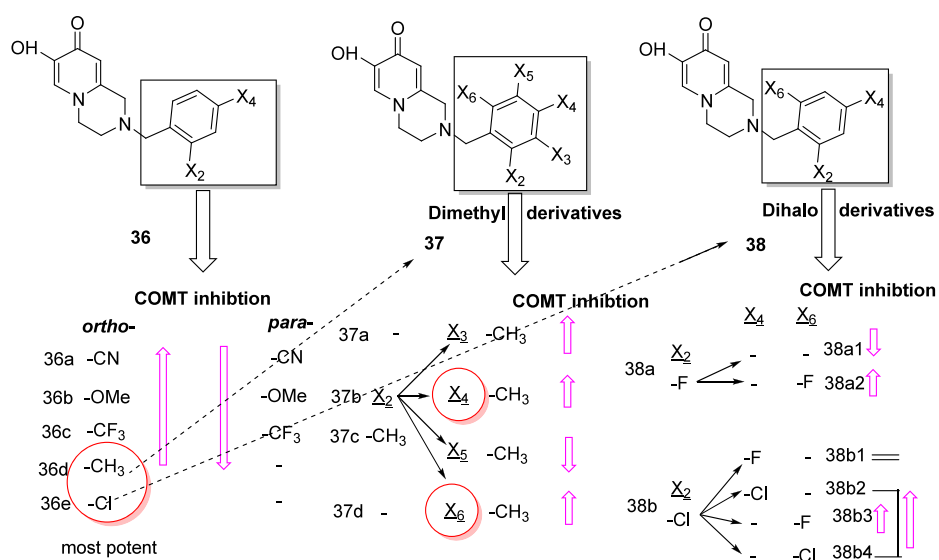


Figure 11. SAR of pyridinone derivatives.

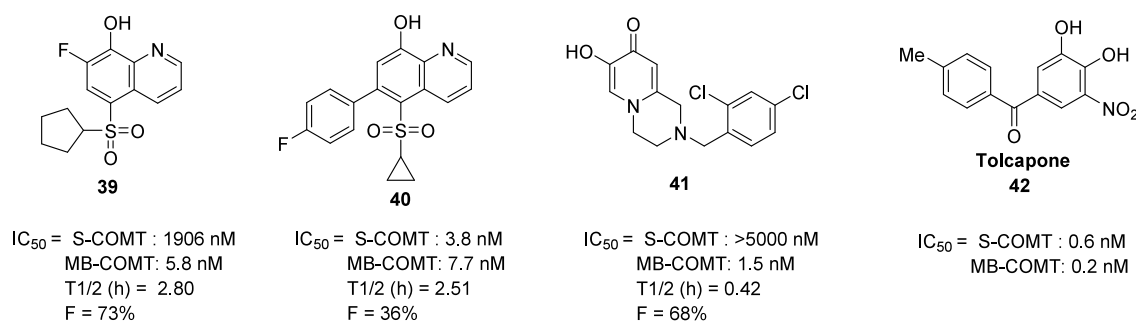


Figure 12. Selected potent inhibitors for *in vivo* COMT evaluations.

The $-CH_3$ and $-Cl$ substituents were further taken to explore the inhibition effect at different positions of the benzyl group by keeping one at the *ortho*-position and another at all available positions. (Figure 11). Among the di $-CH_3$ derivatives 37, methyl groups at the 2,4- (37b) and 2,6-positions (37d) exhibited excellent inhibitory effects with IC_{50} values of 13 and 6.3 nM, respectively.

Similarly, halogen substituents, $-F$ and $-Cl$ (38), were explored for COMT inhibition and 2-F 38a had a slight loss in potency compared to 2-Cl 38b. Among the dihalo derivatives, 2,4-dichloro- (38b2) and 2,6-dichloro- (38b4) exhibited a further increase in potency, with an IC_{50} value of 10 nM, compared with chloro and fluoro substituted derivatives (Figure 11).

Along with potent compounds, such as 2-chloro 36e, 2,4-dichloro 38b2, and 2,6-dimethyl derivative 37d, the parent and 2-chloro-6-fluoro derivative 38b3 were examined for physicochemical and drug metabolism and pharmacokinetics (DMPK) properties, both *in vitro* and *in vivo*. Interestingly, these potent compounds have extraordinarily low levels of protein and tissue binding, which increase the availability of circulating compounds to interact with the target of interest.

The same group subsequently published the *in vivo* activity of novel COMT inhibitors to identify a COMT inhibitor with good oral bioavailability and significant effects on neurochemical biomarkers and behavior in rats along with tolcapone as a reference drug.²³ The selected previously described potent inhibitors (Figure 12) exhibited good *in vivo* COMT inhibitory activity by significantly decreasing homovanillic acid (HVA) and increasing 3,4-dihydroxyphenylacetic acid (DOPAC) at the tested time points. Then the cognitive function was measured as performance in the ASST. These novel inhibitors produced qualitatively similar effects compared to tolcapone.

2.4. Natural Pentacyclic Triterpene Derivatives. Wang et al. (2021) isolated and determined the structure–activity relationship of various series of natural pentacyclic triterpenes such as oleananes, lupanes, friedelanes, and ursanes as COMT inhibitors.²⁴ A two-photon fluorescent probe was used to determine the COMT inhibitory activity. The reaction was carried out in an incubation mixture containing recombinant human S-COMT (2.0 $\mu\text{g}/\text{mL}$), MgCl_2 (5 mM), DTT (1 mM), SAM (200 μM), 3-BTD (2 μM), and varying concentrations (from 0.125 to 20 μM) of tested compounds in a final volume of 200 μL in PBS buffer (50 mM, pH 7.4) in order to determine the IC_{50} values of COMT inhibition. In a series of oleanane derivatives, compound 43a was found as the most active COMT inhibitor with an IC_{50} value of 4.7 μM . The substituent $-COOH$ group at R_1 and $-CH_3$ group at R_3 and R_4 positions was responsible for the COMT inhibitory activity of compound 43a. On replacement of the $-CH_3$ group

at the R_4 position of compound 43a with the CH_2OH group, the derivative retains the COMT inhibitory activity. The substituent $-COOH$ group at R_1 , $-CH_3$ group at R_3 and R_4 positions, and $-OH$ group at R_5 position effect a decrease in COMT inhibitory activity with respect to standard drug epicatechin having an IC_{50} value of 9.5 μM . The substituents $-CH_3$ at R_1 and R_3 positions and $-COOH$ at R_4 position effect a further decrease in COMT inhibitory activity. The substituent $-CH_3$ at R_1 , R_3 , and R_4 positions led to the least potent molecule of this series, 43e, with an IC_{50} value of 114.9 μM . The substituents $-COOH$ at R_1 , $-OH$ at R_2 , $-CH_2OH$ at R_3 , $-CH_3$ at R_4 , and $-OH$ at R_5 position (43f) led to loss of COMT inhibitory activity (Figure 13).

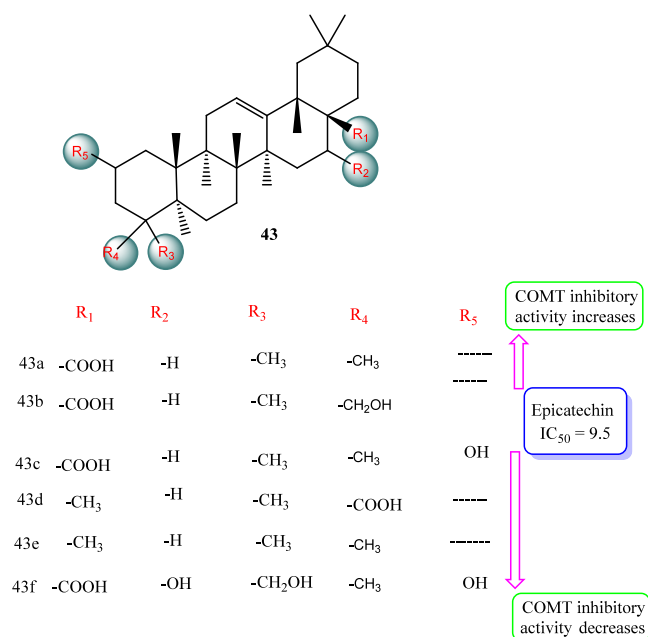


Figure 13. SAR of oleanane derivatives.

In the series of ursane derivatives, all compounds show less activity as compared to the standard drug epicatechin having an IC_{50} value of 9.5 μM . Among all these derivatives, compound 44a was found as the most effective COMT inhibitor. The substituents $-COOH$ at R_2 , $-OH$ at R_4 , and $-CH_3$ at R_3 , R_7 , and R_8 positions were responsible for the highest COMT inhibition of compound 44a. The substituents $-COOH$ at R_1 , $-OH$ at R_4 , and CH_3 at R_3 , R_7 , and R_8 positions led to a decrease in COMT inhibitory activity. The substituents $-COOH$ at R_1 , $-CH_2OH$ at R_3 , $-OH$ at R_4 and R_5 positions, and $-CH_3$ at R_2 , and R_8 position led to the least

potent molecule of this series with IC_{50} value $43.1 \mu M$. With substituents $-COOH$ at R_2 , $-OH$ at R_4 , $=O$ at R_6 , and $-CH_3$ at R_3 and R_8 positions, **44d** exhibited loss of COMT inhibitory activity (Figure 14).

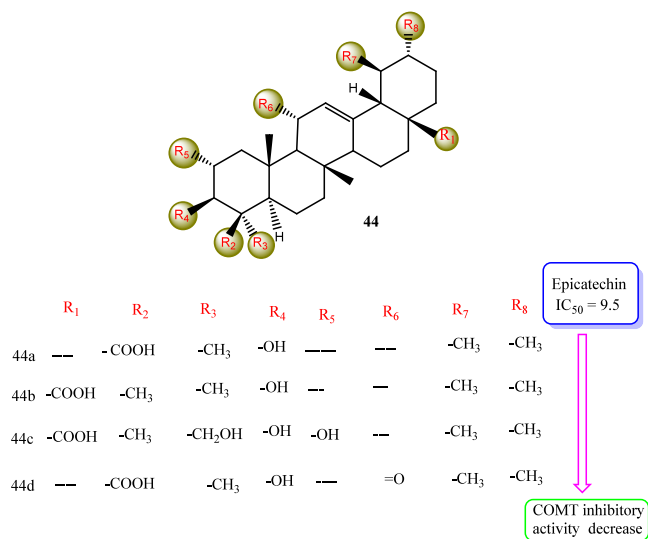


Figure 14. SAR of the ursane derivatives.

In the series of lupane derivatives, the substituent $-COOH$ at R_1 , $-CH_3$ at R_2 and R_3 positions, and $-OH$ at R_4 position conferred the highest efficacy for COMT inhibition within this series, with an IC_{50} value of $5.07 \mu M$. The substituents $-CH_2OH$ at R_1 , $-CH_3$ at R_2 and R_3 positions, and $-OH$ at R_4 position produced a slight increase in COMT inhibitory activity with respect to standard drug epicatechin. The substituents $-CH_3$ at R_1 , R_2 , and R_3 positions and $-OH$ at R_4 position further increased COMT inhibitory activity. The substituents $-C=O$ at R_1 , $-CH_3$ at R_2 and R_3 positions, and $-OH$ at the R_4 position slightly decreased COMT inhibitory activity. Substituents $-COOH$ at R_1 , $-CH_3$ at R_2 and R_3 positions, and $=O$ at R_4 position further decreased COMT inhibitory activity. Substituents $-CH_3$ and $COOH$ at R_1 , $-CH_2OH$ at R_2 , $-CH_3$ at R_3 , and $-OH$ at R_4 produced the least potent molecule (**45f**) of this series with an IC_{50} value of $31.42 \mu M$ (Figure 15).

In the series of friedelane derivatives, they found three derivatives, namely, known as celastrol, hydrocelastrol, and friedelin. Among these compounds, celastrol demonstrates the most significant inhibitory efficacy against COMT, evidenced by an IC_{50} value of $3.89 \mu M$; conversely, hydrocelastrol exhibits a comparatively diminished COMT inhibitory efficacy relative to the reference compound epicatechin, which has an IC_{50} value of $9.5 \mu M$, whereas friedelin manifests no observable COMT inhibitory activity (Figure 16).

2.5. Flavonoid Derivatives. Zhao et al. (2021) isolated and characterized naturally occurring potent COMT inhibitors from the medicinally active plant *Scutellariae radix*.²⁵ A Triple TOF 5600 mass spectrometer (AB SCIEX) running in negative ion mode was coupled to a UPLC system (Shimadzu) for the analysis of the samples, and mixed standards were used to determine the composition. To determine the COMT inhibitory activity, 3-(benzo[*d*]thiazol-2-yl)-7,8-dihydroxy-2*H*-chromen-2-one (3-BTD), an extremely selective optical probe substrate specifically designed for measuring the residual activities of human catechol-*O*-methyltransferase (hCOMT)

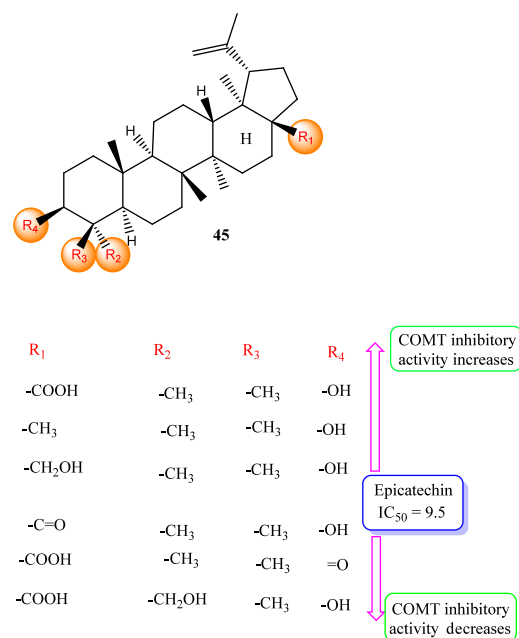


Figure 15. SAR of the lupane derivatives.

in the presence or absence of inhibitors, was used. Among the isolated derivatives, compound **49a** was found to be the most active molecule against COMT having an IC_{50} value of $0.018 \mu M$. The substituent $-OCH_3$ at R_2 position was responsible for the highest significant COMT inhibitory activity of compound **49a**. Substituents (1*S*,2*R*,3*S*,4*S*,5*R*)-2,3,4-trihydroxy-5-(λ -oxidaneryl)cyclohexane-1-carboxylic acid at Ar_2 and $-OCH_3$ at R_5 position (**51a**) increased COMT inhibitory activity with respect to entacapone, having IC_{50} value $0.23 \mu M$. The substituent $-OH$ group at the R_1 and R_2 positions conferred a further increase in COMT inhibitory activity. Substituent $-OH$ at the R_2 position slightly decreased the COMT inhibitory activity. Furthermore, the substitution $-OCH_3$ at the R_3 position resulted in a decrease in the COMT inhibitory activity. The substituent (1*S*,2*R*,3*S*,4*S*,5*R*)-2,3,4-trihydroxy-5-(λ -oxidaneryl)cyclohexane-1-carboxylic acid at Ar_2 position produced the least active compound (**51b**) against COMT with IC_{50} value greater than $100 \mu M$ (Figure 17).

Zhao et al. further isolated and characterized flavonoid derivatives from vine tea as COMT inhibitors in 2021.²⁶ A Triple TOF 5600 Mass Spectrometer system (AB SCIEX, Foster City, USA) in conjunction with a UPLC system (Agilent, California, USA) was used to carefully analyze the main ingredients of the bioactive fractions of vine tea. A highly selective optical probe substrate called 3-BTD was utilized to measure the COMT inhibitory activity in the presence or absence of inhibitors. For comparison of the COMT activity, quercetin was used as a standard. Among the isolated flavonoid derivatives, compound **53a** exhibited the highest level of potency with an IC_{50} value of $0.96 \mu M$. Substituents benzene-1,2,3-triol at the Ar_2 position and $-OH$ group at R_1 position were responsible for the highest COMT inhibitory activity of compound **53a**. Substituents benzene-1,2,3-triol at the Ar_2 position and (2*S*,3*R*,4*S*,5*S*,6*R*)-2-methyl-6-(oxidaneryl) tetrahydro-2*H*-pyran-3,4,5-triol at R_1 position slightly decreased COMT inhibitory activity with respect to standard molecule quercetin. Removal of the double bond from a core molecule of flavonoid at the second position of ring A results in a

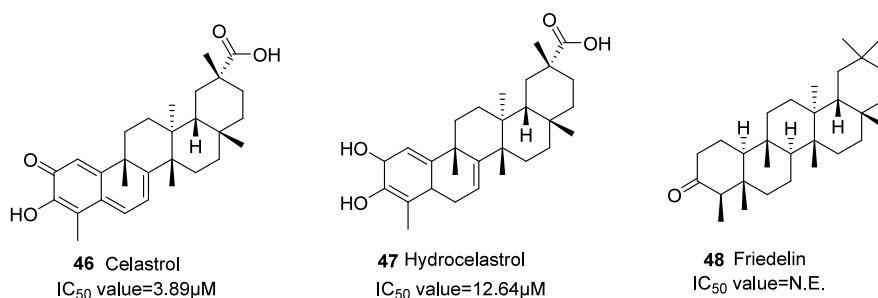


Figure 16. SAR of friedelane derivatives.

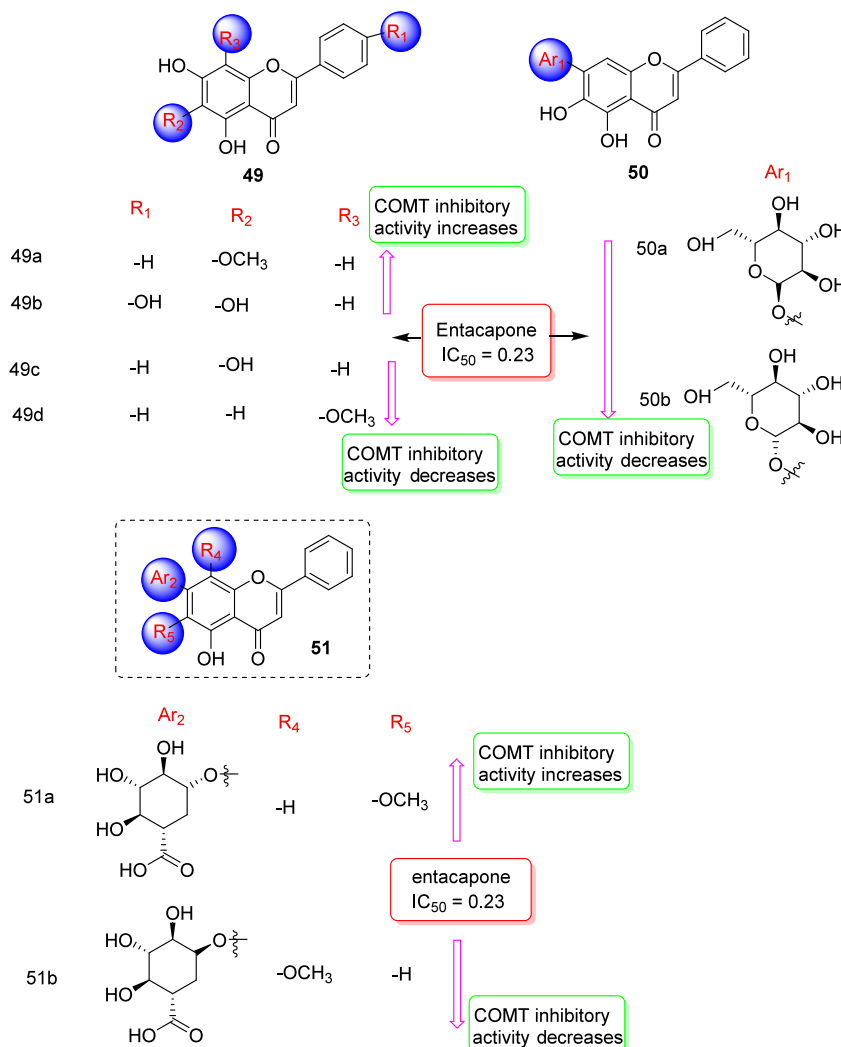


Figure 17. SAR of flavonoid derivatives by Zhao et al. (2021).²⁵

decrease in COMT inhibitory activity. Substitution of benzene-1,2,3-triol at the Ar₁ position of saturated ring A of the flavonoid molecule results in a further decrease in COMT inhibitory activity. Substitution of 4-methylbenzene-1,2-diol at the Ar₁ position of saturated ring A of the flavonoid molecule resulted in the least active molecule against COMT with an IC₅₀ value of 42 μM (Figure 18).

2.6. Tetralone and Indanone Derivatives. de Beer et al. (2021) designed and synthesized a series of novel tetralone and indanone derivatives as COMT inhibitors.²⁷ The designed compounds were synthesized from 5-nitrovanillin (54), treated with aluminum trichloride (AlCl₃) in the presence of pyridine

to give intermediate 3,4-dihydroxy-5-nitrobenzaldehyde (55), which was reacted in acidic medium with appropriate 1-indanone (56) and 1-tetralone (57) derivatives to give the corresponding indanone derivatives (58) and tetralone derivatives (59) (Figure 19).

The soluble fractions from homogenates of rat liver tissue were used as an enzyme source for the COMT inhibition investigations.²⁸ Among the synthesized compounds, indanone derivatives were found to be more potent inhibitors as compared to tetralone derivatives. In a series of indanone derivatives, the one with a substituent -CH₂O- group at the R₁ position (61a) showed the highest COMT inhibitory

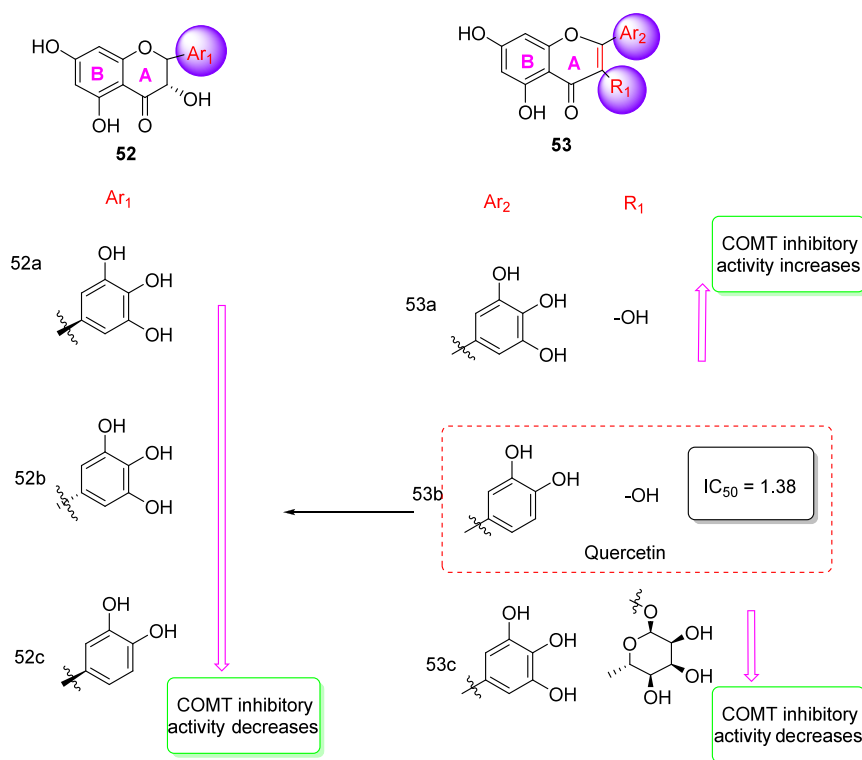


Figure 18. SAR of flavonoid derivatives.

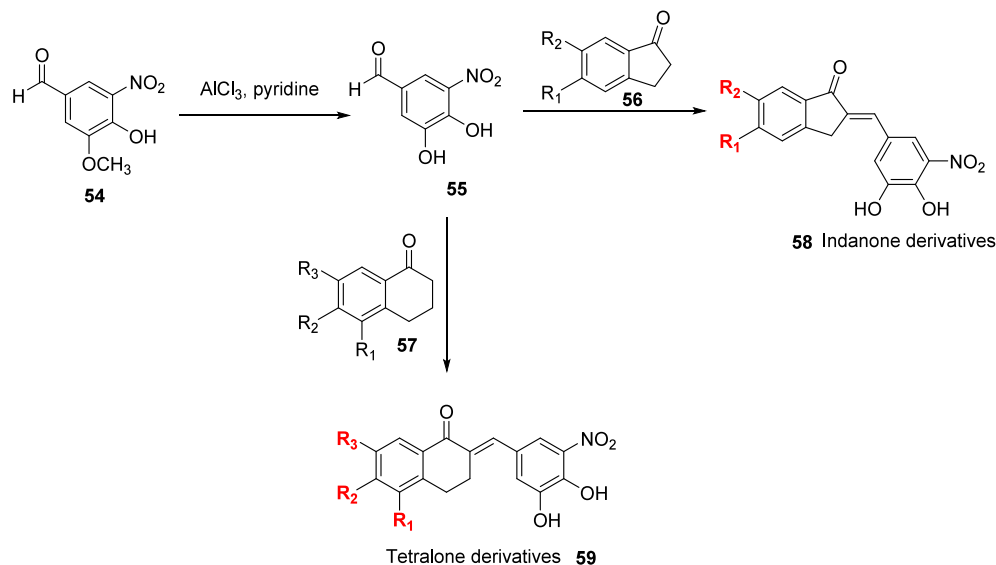


Figure 19. Synthesis of tetralone and indanone derivatives.

activity with IC_{50} value of $0.16 \mu\text{M}$. The substituent $-\text{OCH}_3$ at the R_1 position slightly increased COMT inhibitory activity as compared to the standard molecule entacapone ($IC_{50} = 0.23 \mu\text{M}$). Substitution of hydrogen and $-\text{OCH}_3$ at R_1 and R_2 positions simultaneously results in a decrease in COMT inhibitory activity. Substitution of the hydroxy ($-\text{OH}$) group at the R_1 position, **61e**, results in the least potent COMT inhibitor of this series with an IC_{50} value of $0.38 \mu\text{M}$.

In the series of tetralone derivatives, all molecules were found to be less potent COMT inhibitors compared to entacapone. In this series, the substitution of electron donating groups ($-\text{NH}_2$, $-\text{OCH}_3$) and hydrogen resulted in a decrease in COMT inhibitory activity as compared to entacapone.

Substitution of the $-\text{NH}_2$ group at the R_2 position (**60a**) resulted in the highest COMT inhibition in this series with an IC_{50} value of $0.24 \mu\text{M}$. The substituents $-\text{OCH}_3$ at R_2 and R_3 positions slightly decreased COMT inhibitory activity. The substituent hydrogen at the R_1 , R_2 , and R_3 positions decreased the COMT inhibitory activity. The substituent $-\text{OCH}_3$ at the R_1 or R_3 position further decreased COMT inhibitory activity. The substitution of $-\text{OCH}_3$ at the R_2 position (**60f**) resulted in the least potent COMT inhibitor of this series with an IC_{50} value of $0.79 \mu\text{M}$. Overall indanone derivatives showed better COMT inhibitory activity than tetralone derivatives (Figure 20).

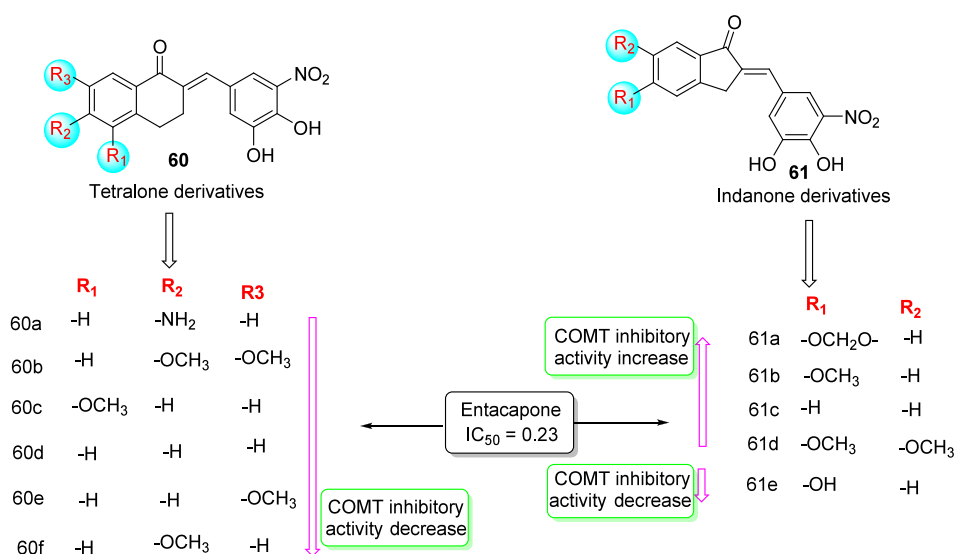


Figure 20. SAR of tetralone and indanone derivatives.

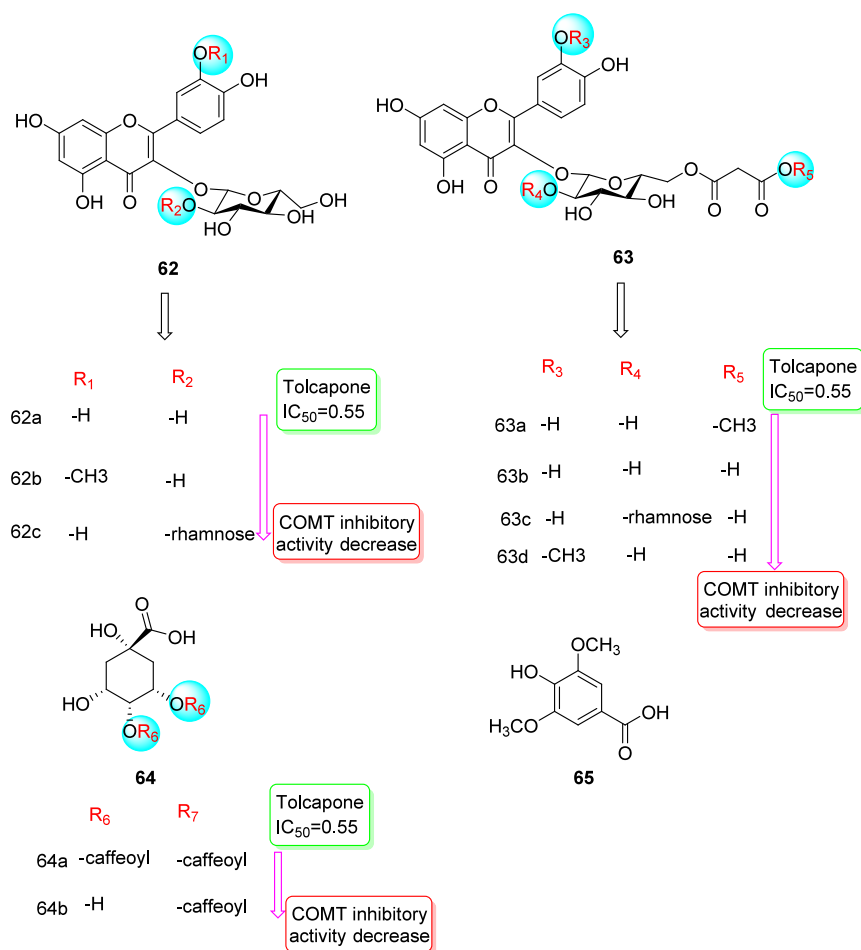


Figure 21. SAR of various COMT inhibitors from *Calendula officinalis* leaf by Kadowaki et al.²⁹

2.7. Miscellaneous. Kadowaki et al. (2023) isolated various COMT inhibitors from *Calendula officinalis* leaf.²⁹ The primary constituents of *Calendula* leaves were determined by isolating ten specific compounds through a variety of chromatographic methods. The isolated compounds were identified by mass spectroscopy (MS) and nuclear magnetic resonance (NMR) spectroscopy to be quercetin 3-*O*-

glucoside (62a), isorhamnetin 3-*O*- β -glucoside (62b), quercetin 3-*O*- β -neohesperidoside (62c), quercetin 3-*O*-(6''-*O*-methylmalonyl)- β -glucoside (63a), quercetin 3-*O*-(6''-*O*-malonyl)- β -glucoside (63b), quercetin 3-*O*-(2''-*O*- α -rhamnosyl-6''-*O*-malonyl)- β -glucoside (63c), isorhamnetin 3-*O*-(6''-*O*-malonyl)- β -glucoside (63d), isorhamnetin 3-*O*-(6''-*O*-malonyl)- β -glucoside (63d), chlorogenic acid (64a), 3,4-dicaffeoylquinic acid (64b), and syringic acid (65). For determining the

COMT inhibitory 3-(benzo[*d*]thiazol-2-yl)-7,8-dihydroxy-2*H*-chromen-2-one (3-BTD) was used.³⁰ For comparison of the COMT inhibitory activity, tolcapone was used as a standard drug. All of the isolated compounds showed less COMT inhibitory activity than the standard drug tolcapone, having an IC₅₀ value of 0.55 μM. Among all isolated derivatives, compound **64a** was found to be the most active COMT inhibitor with an IC₅₀ value of 21 μM. The substituent caffeoyl at R₆ and R₇ positions was responsible for the COMT inhibitory activity of compound **64a**. The substituent caffeoyl at the R₇ position resulted in a decrease in COMT inhibitory activity. The substituent hydrogen at R₁ and R₂ positions maintained the COMT inhibitory activity with IC₅₀ value 50 μM. Substituents –CH₃ at the R₁ position and rhamnose at the R₂ position simultaneously decreased the COMT inhibitory. The substituents hydrogen at the R₃ and R₄ positions and –CH₃ at the R₅ position produced promising COMT inhibitory activity with an IC₅₀ value of 42 μM. Substituent hydrogen at R₃, R₄ and R₅ positions decreased COMT inhibitory activity. The substituents hydrogen at the R₃ and R₅ positions and rhamnose at the R₄ position further decreased in the COMT inhibitory activity. Compound **63d** with substituents –CH₃ at R₃ position and hydrogen at R₄ and R₅ positions and compound **65** showed the least potent COMT inhibition with an IC₅₀ value greater than 100 μM (Figure 21).

3. CONCLUSION

The SAR of COMT inhibitors has emerged as a vital tool in the pursuit of novel therapeutics for Parkinson's disease, depression, and anxiety disorders. In this review, we discussed the synthesis and SAR of various catechol, nitrocatechol, pyridine, pyridinone, flavonoid, tetralone, indanone, and natural pentacyclic triterpene derivatives which have been recently discovered (2019–2024). By elucidating the intricate relationships between chemical structure and biological activity, this review will provide a roadmap for researchers for the design and optimization of next-generation COMT inhibitors. As the field continues to evolve, the insights gleaned from SAR will be crucial in unlocking the full potential of COMT inhibitors and enabling the development of safer, more effective, and personalized treatments for patients in need. Through leveraging the potential of SAR, researchers and clinicians can collaborate to influence the future trajectory of neurology and psychiatry, enhancing lives and improving outcomes.

AUTHOR INFORMATION

Corresponding Authors

Haya Khader Ahmad Yasin – Department of Pharmaceutical Sciences, College of Pharmacy and Health Sciences, Ajman University, Ajman 346, United Arab Emirates; Center of Medical and Bio-allied Health Sciences Research, Ajman University, Ajman 346, United Arab Emirates;
Email: haya.yasin@gmail.com, h.yasin@ajman.ac.ae

T. M. Rangarajan – Department of Chemistry, Sri Venkateswara College, University of Delhi, New Delhi 110021, India; orcid.org/0000-0002-5972-1879;
Email: rangarajan93150@gmail.com

Bijo Mathew – Department of Pharmaceutical Chemistry, Amrita School of Pharmacy, Amrita Vishwa Vidyapeetham, Kochi 682041, India; orcid.org/0000-0002-6658-4497;
Email: bijomathew@aims.amrita.edu

Authors

Sandeep Bindra – Department of Pharmaceutical Chemistry, Amrita School of Pharmacy, Amrita Vishwa Vidyapeetham, Kochi 682041, India

Ananya Datta – Department of Biochemistry, Sri Venkateswara College, University of Delhi, New Delhi 110021, India

Riya Rachel Thomas – Department of Pharmaceutical Chemistry, Amrita School of Pharmacy, Amrita Vishwa Vidyapeetham, Kochi 682041, India

Shailesh Verma – Department of Biochemistry, Sri Venkateswara College, University of Delhi, New Delhi 110021, India

Ankita Patel – Department of Biochemistry, Sri Venkateswara College, University of Delhi, New Delhi 110021, India

Della Grace Thomas Parambi – Department of Pharmaceutical Chemistry, College of Pharmacy, Jouf University, Sakaka, Aljouf 72341, Saudi Arabia

Suraj N. Mali – School of Pharmacy, D.Y. Patil University (Deemed to be University), 400706 Navi Mumbai, India; orcid.org/0000-0003-1995-136X

Complete contact information is available at:
<https://pubs.acs.org/10.1021/acsomega.4c06190>

Notes

The authors declare no competing financial interest.

Biographies

Riya Rachel Thomas is a dedicated researcher with a strong academic background in pharmaceutical sciences. She completed her Bachelor of Pharmacy (B.Pharm) at Amrita School of Pharmacy in Kochi and is currently advancing her studies with a Master of Pharmacy (M.Pharm) in Pharmaceutical Chemistry from the same institution under the guidance of Prof. Dr. Bijo Mathew. Her research interests lie in the synthesis of small molecules and their potential applications in treating neurodegenerative disorders. Driven by a passion for innovative therapeutic development, she aims to contribute to the field of pharmaceutical chemistry by discovering new compounds that could offer hope for patients with neurological diseases.

Rangarajan T. M. is an Assistant Professor of Chemistry at Sri Venkateswara College, University of Delhi, India. He obtained his Ph.D. in Organic Chemistry in 2015 from the University of Delhi. His research interests include organic synthesis, fluoroorganic chemistry, electrochemical perfluorination reactions, and drug development for neurodegenerative diseases and antimalarial treatment.

Ananya Datta is a third-year B.Sc. (H) Biochemistry student at Sri Venkateswara College, University of Delhi, India. She is a recipient of the prestigious INSPIRE Scholarship by the Department of Science and Technology, Government of India. She is one of the Editorial board members of the “Biochemical Society-CATALYSIS” of Sri Venkateswara College. Her research interests include psychoneuroimmunology, oncology, and neurodegenerative disorders and related drug development.

Shailesh Verma is currently in the third year of B.Sc. (H) Biochemistry at Sri Venkateswara College, University of Delhi, India. His major area of research interest is Genomics and Neuroscience.

Ankita Patel is a third year Undergraduate Biochemistry student at Sri Venkateswara College, University of Delhi, India. She is one of the members of the “Biochemical Society- CATALYSIS” of Sri Venkateswara College. Her research interests include molecular biology and bioinformatics.

Sandeep Bindra is a dedicated postgraduate student pursuing an M. Pharm. in Pharmaceutical Chemistry at Amrita School of Pharmacy, Amrita Vishwa Vidyapeetham, Coimbatore, India. With a strong passion for medicinal chemistry, particularly synthetic chemistry, he is committed to deepening his knowledge and making significant contributions to the academic community. He earned his Bachelor's degree from Dr. Bhim Rao Ambedkar University in Agra, India. Currently, as a postgraduate student, he is actively engaged in the design and synthesis of thio/semicarbazide-acetophenone based carbonic anhydrase inhibitors. His research focuses on developing novel inhibitors to treat neurodegenerative diseases.

Haya Yasin is currently a faculty member at the Department of Pharmaceutical Sciences, College of Pharmacy and Health Sciences, Ajman University, United Arab Emirates. She earned her Master of Science in Pharmaceutical Technology from Jordan University of Science and Technology, Irbid/Amman, in August 2011, following her Bachelor of Pharmacy degree from the same institution in June 2009. Her research focuses on areas such as drug delivery and targeting, 3D printing of pharmaceutical formulations, and pharmacy practice, contributing to advancements in pharmaceutical education and innovation. Haya Yasin has received several prestigious awards and honors in her field. At the 26th Dubai International Pharmaceutical and Technology Conference and Exhibition (DUPHAT 2021), she earned the First Best Pharmacy Poster under the "Thematic" category and the Fourth Best Pharmacy Poster under the "Quality" category. Additionally, she achieved second and third place in the poster competition at the International Conference of Pharmacy and Medicine (ICPM), demonstrating her excellence in research and contribution to pharmaceutical science. Currently, she has 25 publications of repute as per Scopus Database.

Suraj Mali holds a Ph.D. in Pharmacy and currently serves as an Assistant Professor of Pharmaceutical Chemistry at the School of Pharmacy, DY Patil University, Navi Mumbai, India. A former analytical scientist at Dr. Reddy's Laboratories in Hyderabad, India, Dr. Mali has an M. Pharm. in Pharmaceutical Science and Technology from the Institute of Chemical Technology, Mumbai. He is a distinguished reviewer for 267 scientific journals and was named a Bentham Science Brand Ambassador for 2019–2020. With over 140 international journal publications and an H-index of 27 (Scopus Citations: 1635), Dr. Mali's research focuses on molecular modeling, synthetic chemistry, phytochemistry, pharmacology, and analytics, with a keen interest in drug design and synthesis. His notable achievements include the Institute of Chemical Technology's Masters Best Thesis Aditya Birla Award in 2019. His work has garnered international recognition, including a publication in *Nature Scientific Reports* on identifying antimycobacterial agents through computational methods. Furthermore, Dr. Mali was ranked among the world's top 2% of scientists by Stanford University in both 2023 and 2024, a testament to his outstanding contributions to pharmaceutical science.

Della Grace Thomas Parambi is presently working as Associate Professor in the Faculty of Pharmacy, Jouf University, Saudi Arabia. She has accomplished her pharmacy Ph.D. in Pharmaceutical analysis and has more than 20 years of teaching experience in Analytical Chemistry, Medicinal Chemistry, Molecular Modeling, and Drug Design. The primary research activities in her research group focus on the design of MAO/ChE inhibitors for neurodegenerative disorders. She has also published more than 70 research articles in peer-reviewed international journals and authored one book, 9 book chapters and one patent to her credit. Dr. Della, despite being one of the best chemists and academicians, has also been able to translate difficult chemistry into general understanding for students.

Bijo Mathew is a Professor in the Pharmaceutical Chemistry department at Amrita School of Pharmacy, Amrita Vishwa Vidyapeetham, Kochi, India. In 2024, Dr. Mathew worked as Visiting Professor in the department of medicinal chemistry at University of Bari, Italy. Bijo Mathew obtained his M.Pharm. in Pharmaceutical Chemistry from The Tamilnadu Dr. M.G.R. Medical University, Tamilnadu, India, in 2008 and Ph.D. in Pharmaceutical Sciences from Jawaharlal Nehru Technological University, Hyderabad, India, in 2016. Mathew's primary focus areas are drug discovery of neurodegenerative disorders and cancer and molecular modeling. His main research interest is in the design of enzyme inhibitors, especially MAOs, ChEs, and BACE1. He received the Top 2% Scientists category from the world Stanford's list in 2020 to 2024. He has published 245 papers [h-index = 44; the number of citations: 6423 in Google Scholar, h-index = 41; the number of citations: 370 in Scopus; Cumulative Impact factor: 790.0: August 2024]. With 16 years of teaching and research experience, Mathew authored one book, edited two books, authored seven book chapters, and gave 37 invited talks at national and international levels. Dr. B. Mathew serves as a chief executive guest editor of the journals *Molecules*, *Current Topics in Medicinal Chemistry*, and *Current Drug Targets*. He serves on the editorial board of journals *Brain Sciences* and *Combinatorial Chemistry and High Throughput Screening*. For the development of MAO inhibitor drugs, Dr. Mathew obtained funds from many government funding organizations in India, including DRDO and ICSSR. He received Amrita Vishwa Vidyapeetham Chancellor's Publication and Merit awards in 2021 and 2024. He received three patents from the Korean and two from the German Intellectual Property office.

ACKNOWLEDGMENTS

H.K.A.Y. would like to thank the Deanship of Graduate Studies and Research, Ajman University, UAE, for their support in providing assistance in article processing charges for this manuscript.

REFERENCES

- (1) Guldberg, H. C.; Marsden, C. A. Catechol-O-Methyl Transferase: Pharmacological Aspects and Physiological Role. *Pharmacol Rev.* **1975**, *27* (2), 135–206.
- (2) Nissinen, E.; Männistö, P. T. Biochemistry and Pharmacology of Catechol-O-Methyltransferase Inhibitors. *International Review of Neurobiology* **2010**, *95*, 73–118.
- (3) Kiss, L. E.; Soares-da-Silva, P. Medicinal Chemistry of Catechol O -Methyltransferase (COMT) Inhibitors and Their Therapeutic Utility. *J. Med. Chem.* **2014**, *57* (21), 8692–8717.
- (4) Borchardt, R. T.; Cheng, C.-F.; Cooke, P. H.; Creveling, C. R. The Purification and Kinetic Properties of Liver Microsomal-Catechol-O-Methyltransferase. *Life Sci.* **1974**, *14* (6), 1089–1100.
- (5) Tunbridge, E. M. Catechol-O-Methyltransferase Inhibition Improves Set-Shifting Performance and Elevates Stimulated Dopamine Release in the Rat Prefrontal Cortex. *J. Neurosci.* **2004**, *24* (23), 5331–5335.
- (6) Grossman, M. H.; Emanuel, B. S.; Budarf, M. L. Chromosomal Mapping of the Human Catechol-O-Methyltransferase Gene to 22q11.1→q11.2. *Genomics* **1992**, *12* (4), 822–825.
- (7) Chen, J.; Lipska, B. K.; Halim, N.; Ma, Q. D.; Matsumoto, M.; Melhem, S.; Kolachana, B. S.; Hyde, T. M.; Herman, M. M.; Apud, J.; Egan, M. F.; Kleinman, J. E.; Weinberger, D. R. Functional Analysis of Genetic Variation in Catechol-O-Methyltransferase (COMT): Effects on mRNA, Protein, and Enzyme Activity in Postmortem Human Brain. *American Journal of Human Genetics* **2004**, *75* (5), 807–821.
- (8) Martin, J. SAM (Dependent) I AM: The S-Adenosylmethionine-Dependent Methyltransferase Fold. *Curr. Opin Struct Biol.* **2002**, *12* (6), 783–793.

- (9) Rutherford, K.; Le Trong, I.; Stenkamp, R. E.; Parson, W. W. Crystal Structures of Human 108V and 108M Catechol O-Methyltransferase. *J. Mol. Biol.* **2008**, *380* (1), 120–130.
- (10) Lerner, C.; Ruf, A.; Gramlich, V.; Masjost, B.; Zürcher, G.; Jakob-Roetne, R.; Borroni, E.; Diederich, F. X-Ray Crystal Structure of a Bisubstrate Inhibitor Bound to the Enzyme Catechol-O-Methyltransferase: A Dramatic Effect of Inhibitor Preorganization on Binding Affinity We Thank F. Hoffmann–La Roche for Generous Support of This Work. We Are Grateful to P. Malherbe for the Cloning of COMT, P. Caspers for the Expression of COMT, A. Cesura for Enzyme Purification, B. Wipf for Fermentation, and H. W. Lahm for Sequencing. *Angew. Chem., Int. Ed.* **2001**, *40* (21), 4040.
- (11) Ma, Z.; Liu, H.; Wu, B. Structure-Based Drug Design of Catechol-O-Methyltransferase Inhibitors for CNS Disorders. *Br. J. Clin. Pharmacol.* **2014**, *77* (3), 410–420.
- (12) Lautala, P.; Ulmanen, I.; Taskinen, J. Molecular Mechanisms Controlling the Rate and Specificity of Catechol O-Methylation by Human Soluble Catechol O-Methyltransferase. *Mol. Pharmacol.* **2001**, *59* (2), 393–402.
- (13) Bacq, Z. M.; Gosselin, L.; Dresse, A.; Renson, J. Inhibition of O-Methyltransferase by Catechol and Sensitization to Epinephrine. *Science* (1979) **1959**, *130* (3373), 453–454.
- (14) Wood, A. J. J.; Calne, D. B. Treatment of Parkinson's Disease. *New England Journal of Medicine* **1993**, *329* (14), 1021–1027.
- (15) Akhtar, M. J.; Yar, M. S.; Grover, G.; Nath, R. Neurological and Psychiatric Management Using COMT Inhibitors: A Review. *Bioorg Chem.* **2020**, *94*, No. 103418.
- (16) Katherine Hatstat, A.; Kennedy, G. M.; Squires, T. R.; Xhafkollari, G.; Skyler Cochrane, C.; Cafiero, M.; Peterson, L. W. Synthesis and Analysis of Novel Catecholic Ligands as Inhibitors of Catechol-O-Methyltransferase. *Bioorg. Med. Chem. Lett.* **2023**, *88*, No. 129286.
- (17) Hatstat, A. K.; Morris, M.; Peterson, L. W.; Cafiero, M. Ab Initio Study of Electronic Interaction Energies and Desolvation Energies for Dopaminergic Ligands in the Catechol-O-Methyltransferase Active Site. *Comput. Theor Chem.* **2016**, *1078*, 146–162.
- (18) Hitge, R.; Smit, S.; Petzer, A.; Petzer, J. P. Evaluation of Nitrocatechol Chalcone and Pyrazoline Derivatives as Inhibitors of Catechol-O-Methyltransferase and Monoamine Oxidase. *Bioorg. Med. Chem. Lett.* **2020**, *30* (12), No. 127188.
- (19) Manna, F.; Chimenti, F.; Fioravanti, R.; Bolasco, A.; Secci, D.; Chimenti, P.; Ferlini, C.; Scambia, G. Synthesis of Some Pyrazole Derivatives and Preliminary Investigation of Their Affinity Binding to P-Glycoprotein. *Bioorg. Med. Chem. Lett.* **2005**, *15* (20), 4632–4635.
- (20) Engelbrecht, I.; Petzer, J. P.; Petzer, A. Nitrocatechol Derivatives of Chalcone as Inhibitors of Monoamine Oxidase and Catechol-O-Methyltransferase. *Cent Nerv Syst. Agents Med. Chem.* **2018**, *18* (2), 115–127.
- (21) de Beer, J.; Petzer, J. P.; Lourens, A. C. U.; Petzer, A. Design, Synthesis and Evaluation of 3-Hydroxypyridin-4-Ones as Inhibitors of Catechol-O-Methyltransferase. *Mol. Divers* **2021**, *25* (2), 753–762.
- (22) Ernst, G.; Akuma, D.; Au, V.; Buchler, I. P.; Byers, S.; Carr, G. V.; Defays, S.; de León, P.; Demaude, T.; DePasquale, M.; Durieu, V.; Huang, Y.; Jigorel, E.; Kimos, M.; Kolobova, A.; Montel, F.; Moureau, F.; Poslusney, M.; Swinnen, D.; Vandergeten, M.-C.; Van houtvin, N.; Wei, H.; White, N.; Wood, M.; Barrow, J. C. Synthesis and Evaluation of Bicyclic Hydroxypyridones as Inhibitors of Catechol O-Methyltransferase. *ACS Med. Chem. Lett.* **2019**, *10* (11), 1573–1578.
- (23) Byers, S.; Buchler, I. P.; DePasquale, M.; Rowley, H. L.; Kulkarni, R. S.; Pinder, L.; Kolobova, A.; Li, C.; Au, V.; Akuma, D.; Zhang, G.; Wei, H.; Cheetham, S. C.; Barrow, J. C.; Carr, G. V. Novel, Non-Nitrocatechol Catechol-O-Methyltransferase Inhibitors Modulate Dopamine Neurotransmission in the Frontal Cortex and Improve Cognitive Flexibility. *Psychopharmacology (Berl)* **2020**, *237* (9), 2695–2707.
- (24) Wang, F.-Y.; Wei, G.-L.; Fan, Y.-F.; Zhao, D.-F.; Wang, P.; Zou, L.-W.; Yang, L. Inhibition of Catechol-O-Methyltransferase by Natural Pentacyclic Triterpenes: Structure–Activity Relationships and Kinetic Mechanism. *J. Enzyme Inhib Med. Chem.* **2021**, *36* (1), 1079–1087.
- (25) Zhao, D.-F.; Fan, Y.-F.; Wang, F.-Y.; Hou, F.-B.; Gonzalez, F. J.; Li, S.-Y.; Wang, P.; Xia, Y.-L.; Ge, G.-B.; Yang, L. Discovery and Characterization of Naturally Occurring Potent Inhibitors of Catechol-O-Methyltransferase from Herbal Medicines. *RSC Adv.* **2021**, *11* (17), 10385–10392.
- (26) Zhao, D.-F.; Fan, Y.-F.; Yu, H.-N.; Hou, F.; Xiang, Y.-W.; Wang, P.; Ge, G.-B.; Yang, L.; Xu, J.-G. Discovery and Characterization of Flavonoids in Vine Tea as Catechol-O-Methyltransferase Inhibitors. *Fitoterapia* **2021**, *152*, No. 104913.
- (27) de Beer, A. D.; Legoabe, L. J.; Petzer, A.; Petzer, J. P. The Inhibition of Catechol O-Methyltransferase and Monoamine Oxidase by Tetralone and Indanone Derivatives Substituted with the Nitrocatechol Moiety. *Bioorg Chem.* **2021**, *114*, No. 105130.
- (28) Engelbrecht, I.; Petzer, J. P.; Petzer, A. Nitrocatechol Derivatives of Chalcone as Inhibitors of Monoamine Oxidase and Catechol-O-Methyltransferase. *Cent Nerv Syst. Agents Med. Chem.* **2018**, *18* (2), 115–127.
- (29) Kadowaki, W.; Miyata, R.; Fujinami, M.; Sato, Y.; Kumazawa, S. Catechol-O-Methyltransferase Inhibitors from *Calendula Officinalis* Leaf. *Molecules* **2023**, *28* (3), 1333.
- (30) Juvekar, V.; Lee, H. W.; Kim, H. M. Two-Photon Fluorescent Probes for Detecting Enzyme Activities in Live Tissues. *ACS Appl. Bio Mater.* **2021**, *4* (4), 2957–2973.