Genome-Wide Association Analysis Identified ANXA1 Associated with Shoulder Impingement Syndrome in UK Biobank Samples

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ABSTRACT Shoulder impingement syndrome (SIS) is a common shoulder disorder with unclear genetic mechanism. In this study, Genome-wide Association Study (GWAS) was conducted to identify the candidate loci associated with SIS by using the UK Biobank samples (including 3,626 SIS patients and 3,626 control subjects). Based on the GWAS results, gene set enrichment analysis was further performed to detect the candidate gene ontology and pathways associated with SIS. We identified multiple risk loci associated with SIS, such as rs750968 ($P = 4.82 \times 10^{-8}$), rs754832 ($P = 4.83 \times 10^{-8}$) and rs1873119 ($P = 6.39 \times 10^{-8}$) of ANXA1 gene. Some candidate pathways have been identified related to SIS, including those linked to infection response and hypoxia, "ZHOU_INFLAMMATORY_RESPONSE_FIMA_DN" (P = 0.012) and "MANALO_HYPOXIA_UP" ($P = 5.00 \times 10^{-5}$). Our results provide novel clues for understanding the genetic mechanism of SIS.

KEYWORDS

Shoulder impingement syndrome (SIS) Genome-wide association study (GWAS) PLINK Gene set enrichment analysis (GSEA)

Shoulder impingement syndrome (SIS) is a common shoulder disorder which has been considered as the most frequent cause of shoulder pain (Falla *et al.* 2003). The estimated prevalence of shoulder complaints is 7–34% in the general population. SIS is the most commonly reported, accounting for 44–65% of all shoulder pain complaints (De Yang Tien and Tan 2014). It is commonly characterized as recurrent and chronic, with various pathogenic factors including acromial morphology, muscle imbalance, capsular laxity or tightness, rotator cuff muscle weakness, degeneration and inflammation of the tendons or bursa, dysfunctional glenohumeral and scapulothoracic kinematics (Windt *et al.* 1996; Conroy and Hayes 1998). SIS has serious influences on the economy due to its high treatment costs and to losses incurred through absence from work (Brox *et al.* 1993).

Previously, the researches of SIS mainly focused on clinical aspects including surgical treatment and kinesitherapy (Henkus *et al.* 2009; Haahr and Andersen 2006). On the pathophysiological areas, SIS has multiple functions, degradation and mechanical causes (Garving *et al.* 2017). The SIS impingent hypothesis postulated a pathophysiological mechanism, different shoulder joint structures producing mechanical conflicts (Garving *et al.* 2017). A Genome-wide Association Study (GWAS) was recently performed to identify the genetic variants associated with neck or shoulder pain in the UK Biobank cohort, and three genetic loci were identified associated with neck or shoulder pain, such as rs2049604 in *FOXP2* gene and rs62053992 in *LINC01572* gene (Meng *et al.* 2020). However, the molecular mechanisms of SIS are rarely studied and the genetic pathogenesis of SIS is still unclear.

In recent years, increasingly attention is focused on GWAS, an effective tool for studying the relationship between complex diseases and genetic susceptibility (Manolio *et al.* 2009). This technique applies



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millions of single nucleotide polymorphisms (SNPs) from the genome to perform case-control association analysis as markers and screens the SNPs in the whole genome to determine the susceptible sites and regions associated with diseases (Nicolae *et al.* 2010). Traditional GWAS focuses on single SNPs/genes that have a strong statistical association with phenotypic traits. Considering the potential biological interactions of the tested genes, a number of tools for downstream gene set enrichment analysis of GWAS results are widely used, such as GSEA (Wang *et al.* 2007) and FUMA (Watanabe and Taskesen 2017).

Gene set enrichment analysis (GSEA), also known as functional enrichment analysis, has been developed to identify functionally related genes that are significantly enriched in concentrations of genes that are signaly associated with complex diseases or traits (Du *et al.* 2018). Since GSEA integrates the joint genetic effects of multiple genes and prior information about biological processes (such as pathways), it shows a powerful function of biological interactions for pathogenetic research, especially for complex diseases or traits determined by a set of genes with moderate or weak genetic effects (Wang *et al.* 2007; Zhang *et al.* 2014).

To the best of our knowledge, no effort has paid to explore the genetic mechanism of SIS. Therefore a GWAS of SIS in UK Biobank was performed in this study. The GWAS results were further performed with GSEA to detect the candidate gene ontology and pathways associated with SIS. Our results may provide a new clue for understanding the molecular genetic mechanisms in the development of SIS.

MATERIALS AND METHODS

UK BioBank samples

For the GWAS, we used the UK Biobank resource, a prospective cohort study including approximately 500,000 candidates from the UK, aged between 40 and 69 years, who have had whole-genome genotyping undertaken and have allowed the linkage of these data with their patient records (Bycroft et al. 2018). Briefly, participants provided selfreported information and collected blood samples for biochemical tests, genotyping, and physical measurements, as described previously (Bycroft et al. 2018). Individuals were linked retrospectively and prospectively to the National Health Service's Hospital Episode Statistics database. The 9th and 10th revisions of the International Classification of Diseases (ICD) were used to define cases based on inpatient Hospital Episode Statistics data. Specific for this study, 3,626 SIS cases were identified according to the ICD10 diagnoses (Data Field: 41202) from the UK Biobank. Among the remaining samples, 3,626 samples were randomly selected as normal controls after excluding osteoarticular disorders according to the ICD 10th. Finally, a total of 7,252 participants from the UK Biobank cohort were included in this study (Table 1).

Genotyping

Genotyping details for UK Biobank participants have been reported previously (Bycroft *et al.* 2018). Briefly, 49,950 candidates were genotyped using the UK BiLEVE Axiom Array (807,411 markers) and 438,427 candidates were genotyped using UK Biobank Axiom Array (825,927 markers); the two arrays share about 95% of their marker content. Genotypes were called from the array intensity data and 106 batches of approximately 4700 samples were using a custom genotype-calling pipeline.

Genome-wide association study of shoulder impingement syndrome

The GWAS of SIS were performed using 7,254 participants of European ancestry in the UK Biobank. All the subjects are unrelated

Table 1 Summary characteristics of the study populations

		Case	Control
Gender	Male/Female	1977/1649	1916/1710
Age (years)	Mean (SD)	57.94 (7.40)	56.90 (7.96)

self-reported White. The unrelated subjects were generated with KING software (http://people.virginia.edu/~wc9c/KING/) (An et al. 2003). The kinship coefficients estimated by KING, and the option "-min" to apply the same cut-off for 3^{rd} degree $(2 \times 1/2^{(9/2)})$ 0.088). The subjects who had a self-reported sex inconsistent with the genetic gender, who were genotyped but not imputed or who withdrew their consents were removed. A logistic regression model (implemented in PLINK2) assuming additive genetic effects was used for association analysis, adjusting for age, sex, PC1, PC2 and PC3 as covariates (Purcell et al. 2007). For quality control, we removed the SNPs with a call rate < 90%, SNP-level missingness (the number of individuals lacking information on a specific SNP is missing in the sample) > 0.1, individual-level missingness (the number of SNPs that is missing for a specific individual) > 0.1, minor allele frequency (MAF) < 0.01 and Hardy-Weinberg equilibrium (HWE) *P* value < 1.0×10^{-3} . The Manhattan plot of GWAS results was drawn by R (https://www.r-project.org/). The quantile-quantile (QQ) plot was made by FUMA (Watanabe and Taskesen 2017).

Linking GWAS findings to eQTL loci

GTEx eQTL dataset were used to identify genes with eQTL association for further investigation in the effect of gene variation on gene expression (GTEx Consortium *et al.* 2017). The identified GWAS SNPs were linked to eQTL loci from GTEx databases and then further evaluated for SIS related gene expression in our GWAS findings.

Gene set enrichment analysis

The obtained GWAS summary data of SIS SNP were further subjected to gene set enrichment analysis (Wang et al. 2007; Subramanian et al. 2005). The detailed analytical procedures can be found in a published study (Liu et al. 2010). Briefly, four public pathway databases were utilized to generate a collection of gene ontology terms and pathways for testing in this study, which include BioCarta pathway database (http://www.biocarta.com/genes), Kyoto Encyclopedia of Genes and Genomes (KEGG) pathway database (http://www.genome.ad.jp/kegg/pathway.html), Ambion GeneAssist Pathway Atlas (http://www.ambion.com/tools/pathway), and Gene Ontology (GO) database (http://www.geneontology.org). A total of 2,850 gene ontology terms and pathways were analyzed in this study. 5,000 permutations were conducted to obtain the empirical distribution for the statistical tests. The enrichment scores and the empirical distribution were normalized to account for the different sizes of gene sets for each analyzed pathway databases. For each pathway gene sets, the empirical P value was calculated based on the observed normalized enrichment scores and the empirical distribution (Wang et al. 2007). Significant pathways were identified at empirical *P* values < 0.05.

Data availability

The GWAS data (Category 100314) used in this study are available in http://biobank.ctsu.ox.ac.uk/crystal/label.cgi?id=100314. The UK Biobank resource can be found in http://www.ukbiobank.ac.uk/ scientists-3/genetic-data/. Identification of SIS cases were followed to the ICD10 diagnoses (Data Field: 41202) from the UK Biobank in http://biobank.ctsu.ox.ac.uk/crystal/field.cgi?id=41202. Genome-wide



Figure 1 Manhattan plot for shoulder impingement syndrome in UK. Biobank Genomic coordinates are displayed along the X-axis, with the negative logarithm of the association *P* value for each SNP displayed on the Y-axis, meaning that each dot on the Manhattan plot signifies a SNP. The red line indicates the P-value threshold for genome-wide significance (P < 5 × 10⁻⁸) while the blue line indicates P-value threshold for suggestive significance (P < 5 × 10⁻⁵).

significant loci associated with SIS are summarized in Table S1. Significant pathways associated with TWAS of SIS are summarized in Table S2. Quantile-quantile (QQ) plots of observed against expected *P* values for SIS is shown in Figure S1. All the supplementary materials including the GWAS summary statistics of SIS are available at figshare: https://doi.org/10.25387/g3.12477842.

RESULTS

GWAS results of SIS

After controlling covariates including age, sex, PC1, PC2 and PC3, no statistical difference associations were observed in age, sex, PC1, PC2 and PC3 between SIS cases and controls (P > 0.05). Figure 1 presents the Manhattan plot of GWAS results of SIS. The plot is commonly used in GWAS to display significant SNPs (Gibson 2010). As shown, the strongest associations have the smallest P values; their negative logarithms will be the greatest. Significant associations were detected at the rs750968 ($P = 4.82 \times 10^{-8}$) and rs754832 ($P = 4.83 \times 10^{-8}$) of ANXA1 gene on chromosome nine

(Figure 2). Additionally, 4 SNPs of *ANXA1* showed suggestive association signals, including rs1873119 ($P = 6.39 \times 10^{-8}$), rs2130118 ($P = 6.46 \times 10^{-8}$), rs7029105 ($P = 6.66 \times 10^{-8}$), and rs7874184 ($P = 4.31 \times 10^{-7}$). We identified several candidate locus with suggestive association signals, such as the rs9987487 ($P = 4.63 \times 10^{-5}$) of *PLGRKT* gene on chromosome nine, and rs149598250 ($P = 3.15 \times 10^{-5}$) of *PIK3AP1* gene on chromosome ten (Table 2). The linked genes between identified SNPs of GWAS and eQTL loci of GTEx database are shown in Table S1. For example, *PLGRKT* ($P_{GWAS} = 4.63 \times 10^{-5}$, $P_{eQTL} = 8.90 \times 10^{-7}$), *TEKT3* ($P_{GWAS} = 2.66 \times 10^{-5}$, $P_{eQTL} = 7.30 \times 10^{-7}$) and *LIN28B* ($P_{GWAS} = 2.94 \times 10^{-5}$, $P_{eOTL} = 2.40 \times 10^{-12}$).

Gene set enrichment analysis

Among all of the 2,850 pathways analyzed by this study, the "MAN-ALO_HYPOXIA_UP" was the most significant associated signal for SIS in our analysis ($P = 5.00 \times 10^{-5}$, FDR = 0.018). We also found several candidate pathways for SIS, such as "BECKER_TAMOXIFEN_RESISTANCE_DN" ($P = 1.00 \times 10^{-4}$, FDR = 0.018), "KEGG_NEUROACTIVE_LIGAND_RECEPTOR_INTERACTION" ($P = 1.00 \times 10^{-5}$, FDR = 0.015) and "IZADPANAH_STEM_CELL_ADIPOSE_VS_BONE_DN" ($P = 1.49 \times 10^{-5}$, FDR = 0.018). Table 3 summarized the ten top-ranking pathways detected by this study. Detailed results of our analysis can be found in Table S2.

DISCUSSION

In this study, we conducted a GWAS of SIS using a large data set from UK biobank samples. GWAS results were further performed with GSEA to detect candidate gene ontology and SIS related pathways. We detected several candidate genes and gene sets for SIS, such as *ANXA1* and *PLGRKT*.

One vital finding of this study is the two significant risk SNPs rs754832 and rs750968, both of them were near to the *ANXA1* (Annexin A1) gene on chromosome nine. This gene encodes a membrane-localized protein that inhibits phospholipase A2 and has anti-inflammatory activity (Cardin *et al.* 2017). A research identified the regulation of the inflammation by regulating the



Figure 2 LocusZoom plots for shoulder impingement syndrome gene ANXA1. Association results for SNPs as a function of genomic distance (chromosomal position from National Center for Biotechnology Information build hg19) for ANXA1. The display range is chr9: 74766780–76785307. The top line shows genomic coverage at the locus, with each vertical tick representing the imputed SNPs. Purple diamond indicate SNP at the locus with the strongest association evidence. Each point represents a SNP. Bottom panel shows genes at each locus as annotated in the UCSC Genome Browser Annotation Database.

Table 2 List of candidate loci identified by GWAS of shoulder impingement syndrome

SNP	CHR	Gene	EA	OR	SE	Р
rs750968	9	ANXA1	G	0.833	0.033	4.82×10 ⁻⁸
rs754832	9	ANXA1	А	0.833	0.033	4.83×10 ⁻⁸
rs1873119	9	ANXA1	А	0.835	0.033	6.39×10 ⁻⁸
rs2130118	9	ANXA1	Т	0.835	0.033	6.46×10 ⁻⁸
rs7029105	9	ANXA1	Т	0.835	0.033	6.66×10 ⁻⁸
rs9987487	9	PLGRKT	А	1.145	0.033	4.63×10^{-5}
rs149598250	10	PIK3AP1	С	2.185	0.188	3.15×10^{-5}

Note: CHR = chromosome; EA = effect allele; OR = odds ratios; SE = standard error; P = P-value.

clearance of neutrophils by macrophages in mouse bone marrow as a critical regulator role for the anti-inflammatory molecule *ANXA1* (Dali *et al.* 2012). Moreover, *ANXA1* is essential to regulate bone marrow mesenchymal stem cells (BMSCs) proliferation and osteogenic differentiation (Pan *et al.* 2015). Likewise, Liang *et al.* in-vestigated the effects of silencing *ANXA1* gene on proliferation and osteogenic differentiation of rabbit BMSCs, and found that silencing *ANXA1* gene could reduce those capacities of BMSCs, which may be one of mechanisms underlying osteoarticular diseases (Liang *et al.* 2012). Our results suggest that *MIR6130* is another gene near the top SNP rs750968. *MIR6130* (microRNA 6130), alias as hsamir-6130, is one of the microRNAs that coordinately regulated in rheumatic heart disease patients with mitral valve stenosis (Lu *et al.* 2018). However, the relationship between *MIR6130* and SIS remains unclear.

In this study, another identified candidate genes for SIS was *PLGRKT* (Plasminogen Receptor with A C-Terminal Lysine) which involved in regulation of inflammatory response; regulated monocyte chemotactic migration and matrix metalloproteinase activation, such as of *MMP2* and *MMP9* (Lighvani *et al.* 2011). It has been demonstrated that Plasminogen (PLG) and the plasminogen receptor, *PLGRKT*, play an important role in macrophage recruitment in the inflammatory response, and regulated inflammation via the interplay between PLG and its receptors (Miles *et al.* 2014). Miles *et al.* evaluated the effect of *PLGRKT* deletion on inflammation, and demonstrated that *PLGRKT* was required for plasminogen binding and macrophage migration *in vivo* (Miles *et al.* 2017). Likewise, *PLGRKT* involved in regulation of inflammatory response; regulates matrix metallproteinase activation, which may damage cells by digesting the ECM (Erlykina *et al.* 2015).

GSEA detected several candidate biological gene sets for SIS, such as "ZHOU_INFLAMMATORY_RESPONSE_FIMA_DN" (genes down-regulated in macrophages by *P.gingivalis FimA* pathogen), which was related to inflammatory response. This gene set up-regulated in macrophages by lipopolysaccharide (LPS) and can lead to an inflammatory condition (Zhou and Amar 2007). Several investigators have suggested that fimbriae of *P.gingivalis* can trigger the production of proinflammatory mediators in human macrophages (Nakagawa *et al.* 2002). Besides, LPS in *P.gingivalis* could promote the antibacterial responses of macrophages, and simultaneously cause detrimental lesion in infected tissues due to enhanced local inflammatory reactions (Mancuso *et al.* 2007). Therefore, this gene set may affect the development of SIS by regulating inflammatory responses, but needs further verification.

"MANALO_HYPOXIA_UP" (genes up-regulated in response to both hypoxia and overexpression of an active form of HIF1A), another candidate biological gene sets associated with SIS, played a vital role in the pathogenesis of osteoarthritis (OA) and rheumatoid arthritis (RA) (Johnson et al. 2000; Brouwer et al. 2009; Pfander et al. 2006). However, the molecular mechanism of hypoxic-induced articular lesions in osteoarthrosis remains controversial. A variety of hypotheses on the role of hypoxia in the pathogenesis of osteoarthrosis, such as abnormal energy metabolism and immune inflammation have been proposed in previous studies (Pfander and Gelse 2007). For example, the molecular mechanism of hypoxiainduced joint injury may be partly due to the increase in production of ROS under hypoxia, which can accelerate protein oxidation, and induce mitochondrial injury and apoptosis (Zhang et al. 2011). Based on these evidences above, we suggested that hypoxia might play an important role in the pathogenesis of SIS.

The gene set enrichment researches are receiving increasing attention in genetic study of human complex diseases. Although this is the first molecular mechanism study in SIS combining GWAS and gene set enrichment analysis, there are still several limitations need to be addressed. First, the sample size of SIS GWAS is not large enough. Additionally, our analysis results need to be verified in experimentation via collecting sufficient SIS samples. Consequently, our study results should be interpreted with caution and further studies are needed to confirm the findings and reveal the potential roles of identified locus and pathways in the development of SIS.

Table 3 Top ten of significant pathway associated with shoulder impingement

P value	FDR q value					
5.00× 10 ⁻⁵	0.018					
5.00× 10 ⁻⁵	0.021					
1.00× 10 ⁻⁴	0.018					
1.00× 10 ⁻⁵	0.002					
1.00× 10 ⁻⁵	0.018					
1.00× 10 ⁻⁵	0.035					
1.00× 10 ⁻⁵	0.018					
1.00× 10 ⁻⁵	0.015					
1.00× 10 ⁻⁵	0.018					
1.49× 10 ⁻⁵	0.018					
	P value 5.00×10^{-5} 5.00×10^{-5} 1.00×10^{-5} 1.49×10^{-5}					

In conclusion, we conducted GWAS for SIS, and then performed a gene set enrichment analysis based on GWAS results. Finally, a group of significant difference genes and gene sets within SIS were identified, which may provide novel clues for revealing the complex genetic mechanism of SIS.

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LITERATURE CITED

- An, D. S., Y. Xie, S. H. Mao, K. Morizono, S. K. Kung *et al.*, 2003 Efficient lentiviral vectors for short hairpin RNA delivery into human cells. Hum. Gene Ther. 14: 1207–1212. https://doi.org/10.1089/104303403322168037
- GTEx Consortium; Battle, A., C. D. Brown, B. E. Engelhardt, S. B. Montgomery *et al.*, 2017 Genetic effects on gene expression across human tissues. Nature 550: 204–213. https://doi.org/10.1038/nature24277
- Brouwer, E., A. S. Gouw, M. D. Posthumus, M. A. van Leeuwen, A. L. Boerboom *et al.*, 2009 Hypoxia inducible factor-1-alpha (HIF-1alpha) is related to both angiogenesis and inflammation in rheumatoid arthritis. Clin. Exp. Rheumatol. 27: 945–951.
- Brox, J. I., P. H. Staff, A. E. Ljunggren, and J. I. Brevik, 1993 Arthroscopic surgery compared with supervised exercises in patients with rotator cuff disease (stage II impingement syndrome). BMJ 307: 1269. https://doi.org/ 10.1136/bmj.307.6909.899
- Bycroft, C., C. Freeman, D. Petkova, G. Band, L. T. Elliott *et al.*, 2018 The UK Biobank resource with deep phenotyping and genomic data. Nature 562: 203–209. https://doi.org/10.1038/s41586-018-0579-z
- Cardin, L. T., N. M. Sonehara, K. K. O. Mimura, A. R. D. D. Santos, W. A. D. S. Junior *et al.*, 2017 ANXA1 Ac2–26 peptide, a possible therapeutic approach in inflammatory ocular diseases. Gene 614: 26–36. https:// doi.org/10.1016/j.gene.2017.02.032
- Conroy, D. E., and K. W. Hayes, 1998 The effect of joint mobilization as a component of comprehensive treatment for primary shoulder impingement syndrome. J. Orthop. Sports Phys. Ther. 28: 3–14. https://doi.org/ 10.2519/jospt.1998.28.1.3
- Nicolae D. L., E. Gamazon, W. Zhang, S. Duan, M. E. Dolan, N. J. Cox, 2010 Trait-associated SNPs are more likely to be eQTLs: annotation to enhance discovery from GWAS. PLoS Genet. 6: e1000888. https://doi.org/ 10.1371/journal.pgen.1000888
- De Yang Tien, J., and A. H. C. Tan, 2014 Shoulder Impingement Syndrome, a Common Affliction of the Shoulder: A Comprehensive Review. Proceedings of Singapore Healthcare. 23: 297–305. https://doi.org/10.1177/ 201010581402300406
- Du, Y., Y. Ning, Y. Wen, L. Liu, X. Liang et al., 2018 A genome-wide pathway enrichment analysis identifies brain region related biological pathways associated with intelligence. Psychiatry Res. 268: 238–242. https://doi.org/ 10.1016/j.psychres.2018.07.029
- Erlykina, E. I., L. M. Obukhova, A. B. Yazykova, V. F. Rossokhin, T. N. Gorshkova *et al.*, 2015 Molecular Mechanisms of Relationship Between Blood Plasma Beta-globulins and Protein Markers of Renal and Bladder Cancer. Bionanoscience 5: 84–90. https://doi.org/10.1007/s12668-015-0165-x
- Falla, D. L., S. Hess, and C. Richardson, 2003 Evaluation of shoulder internal rotator muscle strength in baseball players with physical signs of glenohumeral joint instability. Br. J. Sports Med. 37: 430–432. https://doi.org/ 10.1136/bjsm.37.5.430
- Garving, C., S. Jakob, I. Bauer, R. Nadjar, and U. H. Brunner,
- 2017 Impingement Syndrome of the Shoulder. Dtsch. Arztebl. Int. 114: 765–776.
- Gibson, G., 2010 Hints of hidden heritability in GWAS. Nat. Genet. 42: 558– 560. https://doi.org/10.1038/ng0710-558

- Haahr, J. P., and J. H. Andersen, 2006 Exercises may be as efficient as subacromial decompression in patients with subacromial stage II impingement: 4–8-years\" follow-up in a prospective, randomized study. Scand. J. Rheumatol. 35: 224–228. https://doi.org/10.1080/ 03009740600556167
- Henkus, H. E., P. B. Witte, and R. G. H. H. Nelissen, R. Brand, E. R. A. van Arkel, 2009 Bursectomy compared with acromioplasty in the management of subacromial impingement syndrome: a prospective randomised study. J. Bone Joint Surg. Br. 91: 504–510. https://doi.org/10.1302/0301-620X.91B4.21442
- Dalli J., C. P. Jones, D. M. Cavalcanti, S. H. Farsky, M. Perretti, S. M. Rankin, 2012 Annexin A1 regulates neutrophil clearance by macrophages in the mouse bone marrow. FASEB J. 26: 387–396. https://doi.org/10.1096/fj.11-182089
- Johnson, K., A. Jung, A. Murphy, A. Andreyev, J. Dykens et al., 2000 Mitochondrial oxidative phosphorylation is a downstream regulator of nitric oxide effects on chondrocyte matrix synthesis and mineralization. Arthritis Rheum. 43: 1560–1570. https://doi.org/ 10.1002/1529-0131(200007)43:7<1560::AID-ANR21>3.0.CO;2-S
- Liang, B. W., X. Y. Pan, J. M. Zhao, and G. Q. Yin, 2012 Effects of silencing annexin A1 gene on proliferation and osteogenic differentiation of rabbit bone marrow mesenchymal stem cells. Chinese Journal of Tissue Engineering Research. 16: 3421.
- Liu, Y. J., Y. F. Guo, L. S. Zhang, Y. F. Pei, N. Yu et al., 2010 Biological pathway-based genome-wide association analysis identified the vasoactive intestinal peptide (VIP) pathway important for obesity. Obesity (Silver Spring) 18: 2339–2346. https://doi.org/10.1038/oby.2010.83
- Lu, Q., Y. Sun, Y. Duan, B. Li, J. Xia *et al.*, 2018 Comprehensive microRNA profiling reveals potential augmentation of the IL1 pathway in rheumatic heart valve disease. BMC Cardiovasc. Disord. 18: 53. https://doi.org/ 10.1186/s12872-018-0788-2
- Mancuso, G., A. Midiri, C. Biondo, C. Beninati, S. Zummo et al., 2007 Type I IFN signaling is crucial for host resistance against different species of pathogenic bacteria. J. Immunol. 178: 3126–3133. https://doi.org/10.4049/ jimmunol.178.5.3126
- Manolio, T. A., F. S. Collins, N. J. Cox, D. B. Goldstein, L. A. Hindorff et al., 2009 Finding the missing heritability of complex diseases. Nature 461: 747–753. https://doi.org/10.1038/nature08494
- Meng, W., B. W. Chan, C. Harris, M. B. Freidin, H. L. Hebert *et al.*, 2020 A genome-wide association study finds genetic variants associated with neck or shoulder pain in UK Biobank. Hum. Mol. Genet. 29: 1396–1404. https:// doi.org/10.1093/hmg/ddaa058
- Miles, L. A., N. Baik, S. Lighvani, S. Khaldoyanidi, N. M. Varki *et al.*,
 2017 Deficiency of plasminogen receptor, Plg-RKT, causes defects in plasminogen binding and inflammatory macrophage recruitment in vivo.
 J. Thromb. Haemost. 15: 155–162. https://doi.org/10.1111/jth.13532
- Miles, L. A., S. Lighvani, N. Baik, C. M. Parmer, S. Khaldoyanidi *et al.*, 2014 New Insights into the Role of Plg-R KT in Macrophage Recruitment. Int. Rev. Cell Mol. Biol. 309: 259–302. https://doi.org/10.1016/B978-0-12-800255-1.00005-3
- Nakagawa, I., A. Amano, M. Kuboniwa, T. Nakamura, S. Kawabata *et al.*, 2002 Functional differences among FimA variants of Porphyromonas gingivalis and their effects on adhesion to and invasion of human epithelial cells. Infect. Immun. 70: 277–285. https://doi.org/10.1128/IAI.70.1.277-285.2002
- Pan, X., L. Peng, and G. Yin, 2015 Downregulation of Annexin A1 by short hairpin RNA inhibits the osteogenic differentiation of rat bone marrowderived mesenchymal stem cells. Int. J. Mol. Med. 36: 406–414. https:// doi.org/10.3892/ijmm.2015.2243
- Pfander, D., and K. Gelse, 2007 Hypoxia and osteoarthritis: how chondrocytes survive hypoxic environments. Curr. Opin. Rheumatol. 19: 457– 462. https://doi.org/10.1097/BOR.0b013e3282ba5693
- Pfander, D., B. Swoboda, and T. Cramer, 2006 The role of HIF-1alpha in maintaining cartilage homeostasis and during the pathogenesis of osteoarthritis. Arthritis Res. Ther. 8: 104. https://doi.org/10.1186/ar1894
- Lighvani S., H. Baik, J. E. Diggs, S. Khaldoyanidi, R. J. Parmer, L. A. Miles, 2011 Regulation of macrophage migration by a novel plasminogen

receptor Plg-R KT. Blood 118: 5622-5630. https://doi.org/10.1182/blood-2011-03-344242

- Purcell S., B. Neale, K. Todd-Brown, L. Thomas, M. A. R. Ferreira *et al.*, 2007 PLINK: a tool set for whole-genome association and populationbased linkage analyses. Am. J. Hum. Genet. 81: 559–575. https://doi.org/ 10.1086/519795
- Subramanian, A., P. Tamayo, V. K. Mootha, S. Mukherjee, B. L. Ebert *et al.*, 2005 Gene set enrichment analysis: a knowledge-based approach for interpreting genome-wide expression profiles. Proc. Natl. Acad. Sci. USA 102: 15545–15550. https://doi.org/10.1073/pnas.0506580102
- Wang, K., M. Li, and M. Bucan, 2007 Pathway-based approaches for analysis of genomewide association studies. Am. J. Hum. Genet. 81: 1278–1283. https://doi.org/10.1086/522374
- Watanabe, K., and E. Taskesen, 2017 Functional mapping and annotation of genetic associations with FUMA. Nat. Commun. 8: 1826. https://doi.org/ 10.1038/s41467-017-01261-5

- Windt, D. A. V., B.W. Koes, A.J. Boeke, W. Devillé, B.A. Jong *et al.*, 1996 Shoulder disorders in general practice: prognostic indicators of outcome. Br. J. Gen. Pract. 46: 519–523.
- Zhang, F., X. Guo, W. Wang, H. Yan, and C. Li, 2011 Genome-wide gene expression analysis suggests an important role of hypoxia in the pathogenesis of endemic osteochondropathy Kashin-Beck disease. PLoS One 6: e22983. https://doi.org/10.1371/journal.pone.0022983
- Zhang, F., Y. Wen, X. Guo, T. Yang, H. Shen *et al.*, 2014 Trans-omics pathway analysis suggests that eQTLs contribute to chondrocyte apoptosis of Kashin-Beck disease through regulating apoptosis pathway expression. Gene 553: 166–169. https://doi.org/10.1016/j.gene.2014.10.018
- Zhou, Q., and S. Amar, 2007 Identification of signaling pathways in macrophage exposed to Porphyromonas gingivalis or to its purified cell wall components.
 J. Immunol. 179: 7777–7790. https://doi.org/10.4049/jimmunol.179.11.7777

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