

## Genome Sequence of *Aspergillus aculeatinus* IC\_8, Isolated from an Indoor Air Sample of an Urban Housing Complex in Abidjan, Ivory Coast

**Microbiology**<sup>®</sup>

**Resource Announcements** 

Shu Zhao,<sup>a,b</sup> David Koffi,<sup>c</sup> Jean-Paul Latge,<sup>d</sup> Karidia Sylla,<sup>c</sup> <sup>(D)</sup>John G. Gibbons<sup>a,b,e</sup>

AMERICAN SOCIETY FOR

MICROBIOLOGY

<sup>a</sup>Department of Food Science, University of Massachusetts, Amherst, Massachusetts, USA <sup>b</sup>Molecular and Cellular Biology Graduate Program, University of Massachusetts, Amherst, Massachusetts, USA <sup>c</sup>Parasitology and Mycology Department, Institut Pasteur de Côte d'Ivoire, Abidjan, Ivory Coast <sup>d</sup>Aspergillus Unit, Institut Pasteur, Paris, France

eOrganismic & Evolutionary Biology Graduate Program, University of Massachusetts, Amherst, Massachusetts, USA

**ABSTRACT** Aspergillus aculeatinus is an industrially important species of Aspergillus section *Nigri* capable of producing bioactive, antibiotic, and antitumor compounds. We sequenced the genome of a strain of *A. aculeatinus* that was isolated from the interior of a housing complex in Abidjan, Ivory Coast.

A spergillus section Nigri (the black aspergilli) consists of species that cause food spoilage, cause plant disease, and produce industrially relevant compounds like lipases, amylase, citric acid, and gluconic acid (1). Aspergillus aculeatinus is a member of the black aspergilli and closely related to Aspergillus aculeatus (2). A. aculeatinus has the potential for industrial application, as it produces the bioactive compound neoxaline, the antifungal compound aculeacin, and the antitumor compound paclitaxel (originally named Taxol [Bristol-Myers Squibb]) (2, 3). To date, only one A. aculeatinus genome has been sequenced (4).

To provide additional genomic resources for *A. aculeatinus*, we sequenced the genome of *A. aculeatinus* IC\_8 after isolating it from an indoor air sample of a 23-story urban housing complex in Abidjan, lvory Coast, that houses ~2,000 residents. Specifically, petri dishes with Sabouraud chloramphenicol agar were left open for 24 hours and then incubated at 25°C for 3 days. We used the hyphal tipping approach followed by incubation and single spore isolation to retrieve pure culture. DNA extraction was carried out as previously described (5). Briefly, spores were plated onto potato dextrose agar (PDA) and incubated at 37°C for 96 hours. Spores were collected and directly used for DNA extraction using the MasterPure yeast DNA purification kit following the manufacturer's instructions, with several minor modifications.

Next, 150-bp paired-end libraries were constructed and sequenced on an Illumina NovaSeq 6000 sequencer by Novogene. Raw reads were first deduplicated using Tally v15-065 with the "--with-quality" and "--pair-by-offset" options (6). Trim\_Galore v0.4.2 was then used to remove residual adaptor sequences and to trim low-quality sequences using the parameters "--paired," "--stringency 1," "--quality 30," and "--length 50" (http://www.bioinformatics.babraham.ac.uk/projects/trim\_galore/) (7). The deduplicated and trimmed data set contained 14,017,719 paired reads with a total of 4.07 billion bp. Next, the data were error corrected, and the genome was assembled *de novo* using SPAdes v3.13.1 with the "--careful" mode and a *k*-mer range of 55, 77, and 99 (8).

The assembly consisted of 441 scaffolds, a cumulative assembly size of 36.47 Mb (nearly identical to that of the *A. aculeatinus* CBS 121060 genome [4]), an  $N_{50}$  value

Citation Zhao S, Koffi D, Latge J-P, Sylla K, Gibbons JG. 2021. Genome sequence of *Aspergillus aculeatinus* IC\_8, isolated from an indoor air sample of an urban housing complex in Abidjan, Ivory Coast. Microbiol Resour Announc 10:e00096-21. https://doi.org/ 10.1128/MRA.00096-21.

Editor Antonis Rokas, Vanderbilt University

**Copyright** © 2021 Zhao et al. This is an openaccess article distributed under the terms of the Creative Commons Attribution 4.0 International license.

Address correspondence to John G. Gibbons, jggibbons@umass.edu.

Received 27 January 2021 Accepted 16 February 2021 Published 11 March 2021



FIG 1 Phylogenetic relationship of 25 Aspergillus section Nigri genomes, including IC\_8. The phylogeny was inferred by the approximately maximum-likelihood approach in FastTree (8) from a concatenated protein alignment of 4,680 sequences. All bootstrap branch support values were 100%. IC\_8 is monophyletic with *A. aculeatinus* CBS 121060, and both taxa have short branch lengths, indicating that the species identity of IC\_8 is *A. aculeatinus* CBS 121060, and both taxa have short branch lengths, indicating that the species identity of IC\_8 is *A. aculeatinus* CBS 121060, and both taxa have short branch lengths, indicating that the species identity of IC\_8 is *A. aculeatinus* CBS 121060, and both taxa have short branch lengths, indicating that the species identity of IC\_8 is *A. aculeatinus* (PSTE00000000), *A. aculeatus* (GCA\_001890905.1), *A. brunneoviolaceus* (PSTC00000000), *A. costaricaensis* (PSTH00000000), *A. ellipticus* (PSSY0000000), *A. eucalypticola* (MSFU00000000), *A. fijensis* (PSTG0000000), *A. heteromorphus* (MSFL00000000), *A. homomorphus* (PSTJ00000000), *A. ibericus* (PSTF00000000), *A. niger* ATCC 13157 (*A. phoenicis*) (QUR00000000), *A. niger* ATCC 13496 (QQZP0000000), *A. sacharolyticus* (MSFR00000000), *A. sacharolyticus* (MSFS00000000), *A. violaceofuscus* (PSTA0000000), *A. welwitschiae* (QQZQ0000000), *A. vadensis* (MSFS00000000), *A. violaceofuscus* (PSTA00000000), and *A. welwitschiae* (QQZQ0000000).

of 649,318 bp, and a GC content of 50.48%. Genome completeness was evaluated with BUSCO v3.1.0 using the "ascomycota\_odb9" gene set (9). A total of 98.9% of BUSCO genes were recovered from the IC\_8 genome, indicating a high-quality genome assembly.

To verify the species of IC\_8, we conducted a phylogenetic analysis of IC\_8 and 24 genomes from 22 *Aspergillus* section *Nigri* species, including *A. aculeatinus* CBS 121060 (4). For all genomes, we used the Funannotate v1.7.0 (10) pipeline to predict gene models. Next, we used Orthofinder v2.3.3 to identify orthologous genes across the 25 genomes (11). A concatenated amino acid sequence alignment was generated from 4,680 translated genes. FastTree v2.1.10 was used to infer the phylogenetic relationship of isolates from the concatenated sequence alignment, using the MLACC=3 and nearest-neighbor interchange (NNI) options, with 100 bootstraps (12, 13). IC\_8 is monophyletic with *A. aculeatinus* CBS 121060, and both taxa have short branch lengths (Fig. 1), providing clear evidence that the species identity of IC\_8 is *A. aculeatinus*.

**Data availability.** The whole-genome shotgun project for *A. aculeatinus* IC\_8 has been deposited in GenBank under the accession number JADPID000000000. Raw Illumina data have been deposited to the NCBI Sequence Read Archive under the BioProject accession number PRJNA675076.

## **ACKNOWLEDGMENTS**

This work was supported by the National Institutes of Health and National Institutes of Allergy and Infectious Diseases (R21AI137485).

## REFERENCES

- Varga J, Frisvad JC, Kocsubé S, Brankovics B, Tóth B, Szigeti G, Samson RA. 2011. New and revisited species in Aspergillus section Nigri. Stud Mycol 69:1–17. https://doi.org/10.3114/sim.2011.69.01.
- Noonim P, Mahakarnchanakul W, Varga J, Frisvad JC, Samson RA. 2008. Two novel species of Aspergillus section Nigri from Thai coffee beans. Int J Syst Evol Microbiol 58:1727–1734. https://doi.org/10.1099/ijs.0.65694-0.
- Qiao W, Tang T, Ling F. 2020. Comparative transcriptome analysis of a taxol-producing endophytic fungus, Aspergillus aculeatinus Tax-6, and its mutant strain. Sci Rep 10:10558. https://doi.org/10.1038/s41598-020 -67614-1.
- 4. Vesth TC, Nybo JL, Theobald S, Frisvad JC, Larsen TO, Nielsen KF, Hoof JB, Brandl J, Salamov A, Riley R, Gladden JM, Phatale P, Nielsen MT, Lyhne EK, Kogle ME, Strasser K, McDonnell E, Barry K, Clum A, Chen C, LaButti K, Haridas S, Nolan M, Sandor L, Kuo A, Lipzen A, Hainaut M, Drula E, Tsang A, Magnuson JK, Henrissat B, Wiebenga A, Simmons BA, Mäkelä MR, de Vries RP, Grigoriev IV, Mortensen UH, Baker SE, Andersen MR. 2018. Investigation of inter- and intraspecies variation through genome sequencing of Aspergillus section Nigri. Nat Genet 50:1688–1695. https://doi.org/10 .1038/s41588-018-0246-1.
- Zhao S, Latgé JP, Gibbons JG. 2019. Genome sequences of two strains of the food spoilage mold Aspergillus fischeri. Microbiol Resour Announc 8: e01328-19. https://doi.org/10.1128/MRA.01328-19.
- Davis MP, van Dongen S, Abreu-Goodger C, Bartonicek N, Enright AJ. 2013. Kraken: a set of tools for quality control and analysis of high-

- Martin M. 2011. Cutadapt removes adapter sequences from high-throughput sequencing reads. EMBnet J 17:10–12. https://doi.org/10.14806/ej.17.1.200.
- Bankevich A, Nurk S, Antipov D, Gurevich AA, Dvorkin M, Kulikov AS, Lesin VM, Nikolenko SI, Pham S, Prjibelski AD, Pyshkin AV, Sirotkin AV, Vyahhi N, Tesler G, Alekseyev MA, Pevzner PA. 2012. SPAdes: a new genome assembly algorithm and its applications to single-cell sequencing. J Comput Biol 19:455–477. https://doi.org/10.1089/cmb.2012.0021.
- Seppey M, Manni M, Zdobnov EM. 2019. BUSCO: assessing genome assembly and annotation completeness. Methods Mol Biol 1962:227–245. https://doi.org/10.1007/978-1-4939-9173-0\_14.
- 10. Palmer J, Stajich J. 2017. Funannotate: eukaryotic genome annotation pipeline. Zenodo. https://doi.org/10.5281/zenodo.3548120.
- Price MN, Dehal PS, Arkin AP. 2010. FastTree 2—approximately maximum-likelihood trees for large alignments. PLoS One 5:e9490. https://doi .org/10.1371/journal.pone.0009490.
- Emms DM, Kelly S. 2015. OrthoFinder: solving fundamental biases in whole genome comparisons dramatically improves orthogroup inference accuracy. Genome Biol 16:157. https://doi.org/10.1186/s13059-015-0721-2.
- Emms DM, Kelly S. 2019. OrthoFinder: phylogenetic orthology inference for comparative genomics. Genome Biol 20:238. https://doi.org/10.1186/ s13059-019-1832-y.