

OPEN

Mesothelin expression and its prognostic role according to microsatellite instability status in colorectal adenocarcinoma

Hyunsung Kim, MD, Yumin Chung, MD, Seung Sam Paik, MD, PhD, Kiseok Jang, MD, PhD, Su-Jin Shin, MD, PhD

Abstract

The cell-surface glycoprotein, mesothelin, is normally present on mesothelial cells. Overexpression of mesothelin has been reported in many tumors and is correlated with poor outcome. We investigated the clinicopathologic significance of mesothelin expression in colorectal adenocarcinoma with microsatellites instability (MSI) status.

Mesothelin expression was evaluated immunohistochemically in tissue microarray blocks from 390 colorectal adenocarcinoma samples. Mesothelin expression was interpreted according to the intensity and extent. A score of 2 was considered high expression. We analyzed the correlation between mesothelin expression and clinicopathologic characteristics.

High mesothelin expression was observed in 177 (45.4%) out of 390 colorectal adenocarcinoma samples and was significantly associated with high histologic grade (P=.037), lymphatic invasion (P=.028), lymph node metastasis (P=.028), and high AJCC stage (P=.026). Kaplan–Meier survival curves revealed no significant difference between patients with high mesothelin expression and patients with low mesothelin expression in both recurrence-free survival (RFS) and cancer-specific survival (P=.609 and P=.167, respectively). In subgroup survival analyses, high mesothelin expression was associated with poor RFS in the MSI-High group of colorectal adenocarcinoma (P=.004).

High mesothelin expression was significantly associated with aggressive phenotypes and poor patient outcome in MSI-High colorectal adenocarcinoma.

Abbreviations: CSS = cancer-specific survival, H&E = hematoxylin and eosin, HR = hazard ratio, MSI = microsatellites instability, MSS = microsatellite stable, PD-1 = programmed cell death 1, RFS = recurrence-free survival, SD = standard deviation.

Keywords: colorectal adenocarcinoma, immunohistochemistry, mesothelin, prognosis

1. Introduction

Colorectal adenocarcinoma is one of the most common cancers in the world with over 1.8 million new colorectal adenocarcinomas estimated to have occurred in 2018. [1] Colorectal adenocarcinoma is the second most frequently diagnosed cancer and the fourth leading cause of cancer death in Korea. [2] Surgical excision is the first choice for curing colorectal adenocarcinoma, and adjuvant systemic therapy including immunotherapy can improve survival. [3,4] Although the mortality is decreasing, the prognosis of some colorectal adenocarcinoma is still poor. New prognostic factors that can stratify patients into different risk groups are needed. [4]

Editor: Yan Li.

This work was supported by the research fund of Hanyang University (HY-2016). The authors report no conflicts of interest.

Supplemental Digital Content is available for this article.

Department of Pathology, College of Medicine, Hanyang University, Seoul, Republic of Korea.

* Correspondence: Su-Jin Shin, Department of Pathology, College of Medicine, Hanyang University, 222, Wangsimni-Ro, Seongdong-Gu, Seoul 04763, Republic of Korea (e-mail: charm@hanyang.ac.kr).

Copyright © 2019 the Author(s). Published by Wolters Kluwer Health, Inc. This is an open access article distributed under the terms of the Creative Commons Attribution-Non Commercial-No Derivatives License 4.0 (CCBY-NC-ND), where it is permissible to download and share the work provided it is properly cited. The work cannot be changed in any way or used commercially without permission from the journal.

Medicine (2019) 98:26(e16207)

Received: 30 January 2019 / Received in final form: 15 May 2019 / Accepted: 3 June 2019

http://dx.doi.org/10.1097/MD.000000000016207

Mesothelin is a cell-surface glycoprotein that is normally present on the mesothelial cells lining human body cavities. [5] Many studies have investigated the role of mesothelin in tumor biology and suggested its role in proliferation, local invasion, and metastasis. Overexpression of mesothelin has been reported in many solid tumors including mesothelioma, pancreatic cancer, gastric cancer, ovarian cancer, non-small cell lung cancer, and triple-negative breast cancer. [6–11] Due to its limited expression rate in normal tissue and high expression rate in cancer, therapeutic agents targeting mesothelin have been developed and are currently undergoing clinical trials. [12] Several studies have shown that mesothelin expression is correlated with poor prognosis in stomach cancer and esophageal adenocarcinoma. [8,13–16]

In colorectal adenocarcinoma, mesothelin expression was first described in 2003, and its overexpression was reported in about 50% of colorectal adenocarcinoma cases. [17–19] Still, studies on the prognostic value of mesothelin in colorectal adenocarcinoma are limited. Kawamata et al showed that luminal staining of mesothelin correlated with lymphatic invasion and lymph node metastasis. [19] Moreover, Shiraishi et al reported that non-luminal staining of mesothelin was associated with shorter cancer-specific survival (CSS) in stage II/III, while luminal staining was not associated with CSS. [20] Molecular heterogeneity in colorectal adenocarcinoma influences patient outcome and response to treatment, especially depending on the state of the microsatellite instability (MSI). [21] However, there are no studies on the prognostic value of mesothelin in colorectal cancer according to MSI status.

In this study, we investigated the clinicopathologic significance of mesothelin expression and its association with survival rates according to MSI molecular subgroups in colorectal adenocarcinoma.

2. Materials and methods

2.1. Patients

We retrospectively collected the data of 390 patients who had undergone curative resection for primary colorectal adenocarcinoma at Hanyang University Hospital from January 2005 to December 2010. Patients who had received neoadjuvant chemotherapy and/or neoadjuvant radiotherapy, or those who had recurrent colorectal cancer were excluded. We reviewed medical records to determine the following clinical characteristics: age, sex, follow-up interval, survival, recurrence status, and adjuvant treatment application status and type. Two surgical pathologists (H. K. and Y. C.) reviewed pathology reports and hematoxylin and eosin (H&E)stained slides to define the clinicopathologic characteristics and confirm the diagnosis. Pathologic parameters including TNM stage (8th AJCC) were determined according to a protocol for examining specimens from patients with primary adenocarcinoma of the colon and rectum. [22,23] Pathologic characteristics included tumor size, histologic grade, lymphatic invasion, vascular invasion, perineural invasion, tumor deposit, tumor budding, TNM stage, and MSI status. [22,24] This study was approved by the Institutional Review Board of the Hanyang University Hospital (HYUH 2016-12-030-001), and the requirement for informed consent was waived.

2.2. Tissue microarray construction

We used a manual tissue microarrayer (Unitma, Seoul, Korea) for tissue microarray construction from archival formalin-fixed, paraffin-embedded tissue blocks. Representative tumor area was selected by light microscopy of H&E-stained sections. Tissue cylinders (diameter: 2 mm) were punched from a previously marked lesion on each donor block and transferred to the recipient block (Unitma).

2.3. Microsatellite analysis

Microsatellite analysis was performed as previously described using a panel of five National Cancer Institute workshop-recommended consensus microsatellite markers (BAT25, BAT26, D2S123, D17S250, and D5S346). Cases showing a shifting of microsatellites at 2 or more markers were classified as MSI-High. A shifting of microsatellites at one marker was classified as MSI-Low. Microsatellite stable (MSS) and MSI-L were classified as non-MSI-High.

2.4. Immunohistochemical study and interpretation

The immunohistochemical study for mesothelin expression was performed with 4-µm-thick sections from TMA blocks using the Ventana Benchmark XT automated staining system (Ventana Medical Systems, Tucson, AZ) according to the manufacturer's protocol. The rabbit monoclonal anti-mesothelin (1:50; SP74; Abcam, Cambridge, UK) antibody was used for immunohistochemical staining. We evaluated the mesothelin expression according to the staining intensity and extent of the tumor cells. A luminal staining pattern without membranous and/or cytoplasmic staining was observed only in sections with weakly positive staining intensity. In moderately and strongly positive staining sections, both luminal and non-luminal staining patterns (membranous and/or cytoplasmic) were observed simultaneously. The intensity of staining (any pattern, luminal and non-luminal) was graded as negative, weakly positive, moderately positive, or strongly positive (Fig. 1). The staining was

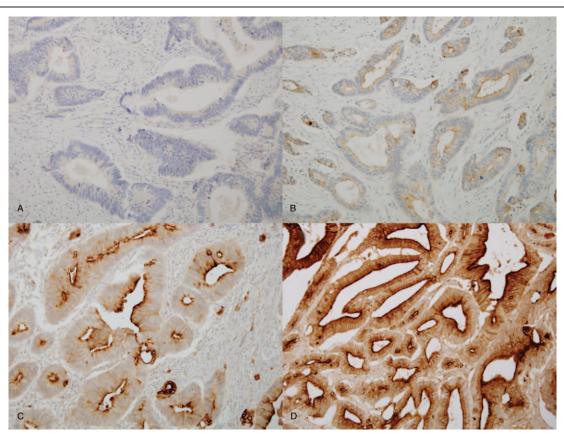


Figure 1. Immunohistochemical staining of mesothelin in colorectal adenocarcinoma. The intensity of membranous and/or cytoplasmic staining was graded as negative (A), weakly positive (B), moderately positive (C), or strongly positive (D) (A–D, ×200).

scored as follows: 0, no expression or any intensity expression less than 5% of tumor cells; 1, weak expression in \geq 5% of tumor cells or moderate to strong expression in \geq 5% and <30% of tumor cells; and 2, moderate to strong expression in \geq 30% of tumor cells. The cases were subdivided into low expression (score 0–1) and high expression groups (score 2).

2.5. Statistical analysis

Statistical analysis was performed using SPSS software version 21 (IBM Corp., Armonk, NY). The chi-square and Mann–Whitney *U* tests were used to evaluate the association between mesothelin expression and clinicopathologic characteristics including tumor size, histologic grade, lymphatic invasion, vascular invasion, perineural invasion, tumor deposit, tumor budding, TNM stage, and MSI status. CSS was defined as the time interval between the date of curative resection and the date of death due to colorectal adenocarcinoma. Recurrence-free survival (RFS) was defined as the time interval between curative resection and the date of any recurrence. Kaplan–Meier survival curves, the log-rank test, and the Cox proportional hazard regression model were used for survival analysis. Two-sided *P* values <.05 were considered statistically significant.

3. Results

3.1. Baseline characteristics of patients

The clinicopathologic characteristics of patients are summarized in Supplemental Table 1, http://links.lww.com/MD/D62. Median follow-up period was 109.5 months (range 1–166), mean age at surgery was 63.4 years (range 27–89), and 237 (60.8%) patients were men with a male-to-female ratio of 1.54:1. Pathological evaluation revealed that 29 tumors (7.4%) were histologic grade 1, 183 (46.9%) were grade 2, 157 (40.2%) were grade 3, and 21 (5.4%) were grade 4. According to 8th AJCC staging system, 55 (14.1%) were stage I, 121 (31.0%) were stage II, 185 (47.4%) were stage III, and 29 (7.4%) were stage IV. Among 390 cases, 363 (93.1%) were non-MSI-High and 27 (6.9%) were MSI-High. Distant metastasis was identified in 29 (7.4%) patients at the time of initial diagnosis, 82 (21.0%) had experienced metastasis or relapse during the follow-up period, and 138 (35.4%) had died due to colorectal adenocarcinoma.

For treatment, 207 (53.1%) patients received curative surgical resection only, and 183 (46.9%) patients received curative surgical resection plus adjuvant chemotherapy (158 cases) or adjuvant combined chemo-radiotherapy (25 cases). The detailed adjuvant chemotherapy regimens are summarized in supplemental Table 1, http://links.lww.com/MD/D62.

3.2. Correlations between mesothelin expression and clinicopathologic characteristics

Overall, high mesothelin expression was observed in 177 (45.4%) of 390 colorectal cancers. Mesothelin expression was significantly correlated with high histologic grade (P=.037), lymphatic invasion (P=.028), lymph node metastasis (P=.028), and high AJCC stage (P=.026). Moreover, mesothelin expression was more frequent in cases with high-grade tumor budding (Table 1). There was no statistically significant correlation between mesothelin expression and other clinicopathologic parameters. In subgroup analysis, high mesothelin expression was also more frequently observed in cases with lymphatic

Table 1

Correlation between mesothelin expression and clinicopathologic features of colorectal adenocarcinoma patients (n = 390).

Variables	Low expression (n=213)	High expression (n=177)	<i>P</i> value
Age, yr (Mean \pm SD)	63.58 ± 11.96	63.11 ± 10.74	.492*
Sex			.172
Male	136 (57.4%)	101 (42.6%)	
Female	77 (50.3%)	76 (49.7%)	
Tumor size, cm (Mean \pm SD)	5.03 ± 2.12	4.91 ± 2.19	.411*
Histologic grade			.037
Grade1 and Grade2	126 (59.4%)	86 (40.6%)	
Grade3 and Grade 4	87 (48.9%)	91 (51.1%)	
Lymphatic invasion			.028
Absent	108 (60.7%)	70 (39.3%)	
Present	105 (49.5%)	107 (50.5%)	
Vascular invasion			.970
Absent	170 (54.7%)	141 (45.3%)	
Present	43 (54.4%)	36 (45.6%)	
Perineural invasion			.271
Absent	113 (57.4%)	84 (42.6%)	
Present	100 (51.8%)	93 (48.2%)	
Tumor deposit			.368
Absent	175 (55.7%)	139 (44.3%)	
Present	38 (50.0%)	38 (50.0%)	
Tumor budding			.070
Absent/ low/ intermediate-	136 (58.4%)	97 (41.6%)	
grade (0 \sim 9 buds/x200)			
High-grade (≥10 buds/x200)	77 (49.0%)	80 (51.0%)	
pT category			.163
pT1-2	34 (47.2%)	38 (52.8%)	
pT3–4	179 (56.3%)	139 (43.7%)	
pN category			.028
pN0	108 (60.7%)	70 (39.3%)	
pN1-2	105 (49.5%)	107 (50.5%)	
Distant metastasis			.402
Absent	195 (54.0%)	166 (46.0%)	
Present	18 (62.1%)	11 (37.9%)	
AJCC stage [†]			.026
I and II	107 (60.8%)	69 (39.2%)	
III and IV	106 (49.5%)	108 (50.5%)	
MSI status			.367
Non-MSI-High (MSS, MSI-Low)	196 (54.0%)	167 (46.0%)	
MSI-High	17 (63.0%)	10 (37.0%)	
Treatment			.157
Surgery	120 (58.0%)	87 (42.0%)	
Surgery + Adjuvant therapy	93 (50.8%)	90 (49.2%)	

^{*} Mann-Whitney test.

invasion (P=.057), high-grade tumor budding (P=.081), lymph node metastasis (P=.057), and high AJCC stage (P=.055) in the non-MSI-High group. In the MSI-High group, mesothelin expression was significantly correlated with high histologic grade (P=.018) and tumor deposit (P=.047; Table 2).

3.3. Prognostic significance of mesothelin expression

There were no significant differences between the high mesothelin expression and low mesothelin expression groups for both RFS and CSS (P=.609 and P=.167, respectively; Fig. 2A–B). In subgroup analyses, there were no significant differences in both the RFS and CSS in the non-MSI-High group (P=.870 and P=.156, respectively; Fig. 2 C–D); however, high mesothelin expression was associated with shorter RFS in the MSI-High

[†] AJCC 8th edition.

Kim et al. Medicine (2019) 98:26

Table 2

Correlation between mesothelin expression and clinicopathologic features of non-MSI-High (n=363) and MSI-High (n=27) colorectal adenocarcinoma patients.

1101	Non-MSI-High (n=363)		MSI-High (n=27)		
Low expression Variables (n = 196)	High expression (n = 167)	P value	Low expression $(n=17)$	High expression $(n=10)$	P value
63.88 ± 11.70	63.24 ± 10.49	.364*	60.06 ± 14.54	61.00 ± 14.94	.980*
		.306			.219
123 (56.2%)	96 (43.8%)		13 (72.2%)	5 (27.8%)	
73 (50.7%)	71 (49.3%)		4 (44.4%)	5 (55.6%)	
4.83 ± 1.95	4.73 ± 2.05	.429*	7.24 ± 2.82	7.80 ± 2.62	.711*
		.134			.018
114 (57.6%)	84 (42.4%)		12 (85.7%)	2 (14.3%)	
			5 (38.5%)	, ,	
,	,	.057	,	,	.212
97 (59.5%)	66 (40.5%)		11 (73.3%)	4 (26.7%)	
\ /	\ /			'	
((/	.896	((.561
155 (53.8%)	133 (46,2%)		15 (65.2%)	8 (34.8%)	
, ,	, ,		, ,	, ,	
(= ,=)	- ((.360	_ (******)	_ (************************************	.573
101 (56.4%)	78 (43.6%)		12 (66.7%)	6 (33.3%)	
, ,	, ,			, ,	
00 (011070)	00 (101170)	.723	0 (00.070)	. (/ 0)	.047
159 (54.5%)	133 (45.5%)		16 (72.7%)	6 (27.3%)	
'	, ,		, ,	'	
01 (021170)	0 1 (11 10 70)	.081	. (20.070)	1 (001070)	.406
129 (57 6%)	95 (42 4%)		7 (77 8%)	2 (22 2%)	1.00
120 (01.070)	00 (42.470)		7 (17.070)	2 (22.270)	
67 (48 2%)	72 (51.8%)		10 (55 6%)	8 (44 4%)	
01 (40.270)	12 (01.070)	122	10 (00.070)	0 (44.470)	.260
32 (45 7%)	38 (54 3%)		2 (100 0%)	0 (0 0%)	.200
, ,	, ,			, ,	
101 (00.070)	120 (11.070)	057	10 (00.070)	10 (10.070)	.257
97 (59 5%)	66 (40 5%)	.007	11 (73 3%)	4 (26.7%)	.201
\ /			'	'	
33 (1 3.370)	101 (50.570)	363	0 (30.070)	0 (30.070)	
178 (53 3%)	156 (46 7%)	.505	17 (63 0%)	10 (37 0%)	
, ,	, ,		17 (03.070)	10 (07.070)	
10 (02.170)	11 (37.370)	055			.257
96 (59 6%)	65 (40 4%)	.000	11 (73 3%)	4 (26.7%)	.201
	, ,		' '	, ,	
100 (43.570)	102 (30.370)	53/	0 (30.070)	0 (30.070)	.002
105 (55 60/)	QA (AA A0/.)	.554	15 (82 20/.)	2 (16 70/)	.002
'	, ,			, ,	
	Low expression (n=196) 63.88±11.70 123 (56.2%) 73 (50.7%)	Low expression (n = 196) High expression (n = 167) 63.88 ± 11.70 63.24 ± 10.49 123 (56.2%) 96 (43.8%) 73 (50.7%) 71 (49.3%) 4.83 ± 1.95 4.73 ± 2.05 114 (57.6%) 84 (42.4%) 82 (49.7%) 83 (50.3%) 97 (59.5%) 66 (40.5%) 99 (49.5%) 101 (50.5%) 155 (53.8%) 133 (46.2%) 41 (54.7%) 34 (45.3%) 101 (56.4%) 78 (43.6%) 95 (51.6%) 89 (48.4%) 159 (54.5%) 33 (45.5%) 37 (52.1%) 34 (47.9%) 129 (57.6%) 95 (42.4%) 67 (48.2%) 72 (51.8%) 32 (45.7%) 38 (54.3%) 164 (56.0%) 129 (44.0%) 97 (59.5%) 66 (40.5%) 99 (49.5%) 101 (50.5%) 178 (53.3%) 156 (46.7%) 18 (62.1%) 11 (37.9%) 96 (59.6%) 65 (40.4%) 100 (49.5%) 102 (50.5%) 105 (55.6%) 84 (44.4%)	Low expression (n = 196) High expression (n = 167) P value 63.88 ± 11.70 63.24 ± 10.49 .364* 123 (56.2%) 96 (43.8%) .306 73 (50.7%) 71 (49.3%) .429* 4.83 ± 1.95 4.73 ± 2.05 .429* 114 (57.6%) 84 (42.4%) .82 (49.7%) 82 (49.7%) 83 (50.3%) .057 97 (59.5%) 66 (40.5%) .99 (49.5%) 101 (50.5%) .896 .896 155 (53.8%) 133 (46.2%) .360 41 (54.7%) 34 (45.3%) .360 101 (56.4%) 78 (43.6%) .95 (51.6%) 95 (51.6%) 89 (48.4%) .723 159 (54.5%) 33 (45.5%) .723 159 (54.5%) 34 (47.9%) .081 129 (57.6%) 95 (42.4%) .081 129 (57.6%) 95 (42.4%) .057 97 (59.5%) 38 (54.3%) .057 97 (59.5%) 66 (40.5%) .99 99 (49.5%) 101 (50.5%) .057 96 (59.6%)	Low expression (n=196) High expression (n=167) P value Low expression (n=17) 63.88±11.70 63.24±10.49 .364* 60.06±14.54 .306 13 (72.2%) 306 73 (50.7%) 71 (49.3%) 4 (44.4%) 4.83±1.95 4.73±2.05 .429* 7.24±2.82 .134 12 (85.7%) 82 (49.7%) 83 (50.3%) 5 (38.5%) 82 (49.7%) 83 (50.3%) .057 11 (73.3%) 97 (59.5%) 66 (40.5%) 11 (73.3%) 6 (50.0%) 99 (49.5%) 101 (50.5%) 896 15 (65.2%) 155 (53.8%) 133 (46.2%) 15 (65.2%) 2 (50.0%) 41 (54.7%) 34 (45.3%) 2 (50.0%) 2 (50.0%) 101 (56.4%) 78 (43.6%) 5 (55.6%) 12 (66.7%) 95 (51.6%) 89 (48.4%) 5 (55.6%) 12 (66.7%) 37 (52.1%) 33 (45.5%) 16 (72.7%) 1 (20.0%) 37 (52.1%) 34 (47.9%) 10 (55.6%) 1 (20.0%) 40 (48.2%) 72 (51.8%) 10 (55.6%) 10 (55.6%)	Low expression (n=196) High expression (n=167) P value Low expression (n=17) High expression (n=10) 63.88±11.70 63.24±10.49 .364* 60.06±14.54 61.00±14.94 123 (56.2%) 96 (43.8%) 13 (72.2%) 5 (27.8%) 73 (50.7%) 71 (49.3%) 4 (44.4%) 5 (55.6%) 4.83±1.95 4.73±2.05 .429* 7.24±2.82 7.80±2.62 114 (67.6%) 84 (42.4%) 12 (85.7%) 2 (14.3%) 82 (49.7%) 83 (50.3%) 5 (38.5%) 8 (61.5%) 97 (59.5%) 66 (40.5%) 11 (73.3%) 4 (26.7%) 99 (49.5%) 101 (50.5%) 896 15 (65.2%) 8 (34.8%) 155 (53.8%) 133 (46.2%) 15 (65.2%) 8 (34.8%) 41 (54.7%) 34 (45.3%) 2 (50.0%) 2 (50.0%) 95 (51.6%) 78 (43.6%) 5 (55.6%) 4 (44.4%) 95 (51.6%) 34 (47.9%) 12 (66.7%) 6 (33.3%) 95 (51.6%) 34 (47.9%) 10 (56.6%) 6 (67.3%) 95 (51.6%) 34 (47.9%) 10

^{*} Mann-Whitney test.

group (P=.004; Fig. 2E). We additionally divided the groups according to treatment. Mesothelin expression had no prognostic implication for RFS and CSS in both colorectal cancer groups with adjuvant therapy (n=183; P=.944 and P=.480, respectively) and without adjuvant therapy (n=207; P=.763 and P=.291, respectively).

4. Discussion

In this study, we revealed that mesothelin expression was correlated with aggressive phenotypes in colorectal adenocarcinoma, such as high histologic grade, frequent lymphatic invasion, high-grade tumor budding, lymph node metastasis, and high AJCC stage. Mesothelin was not a prognostic indicator of CSS and RFS in all patients but served as a

significant prognostic factor of RFS in the MSI-High group of colorectal adenocarcinoma. Although the number of MSI-high cases was small, this finding is interesting. The survival discrepancy between groups might be explained by their distinct genomic background. Recent studies revealed that MSS/MSI-Low and MSI-High groups of colorectal adenocarcinoma exhibit distinct genomic profiles and show different phenotypes and prognoses. The MSI-High group of colorectal adenocarcinoma patients usually show proximal colon predominance, poor histologic differentiation and/or mucin production, and increased tumor-infiltrating lymphocytes. [26] Several studies have shown that MSI-High colorectal adenocarcinoma has a better prognosis than MSS/MSI-Low colorectal adenocarcinoma. [27] A recent study has revealed that advanced stage MSI-High colorectal adenocarcinoma has

[†] AJCC 8th edition.

Kim et al. Medicine (2019) 98:26 www.md-journal.com

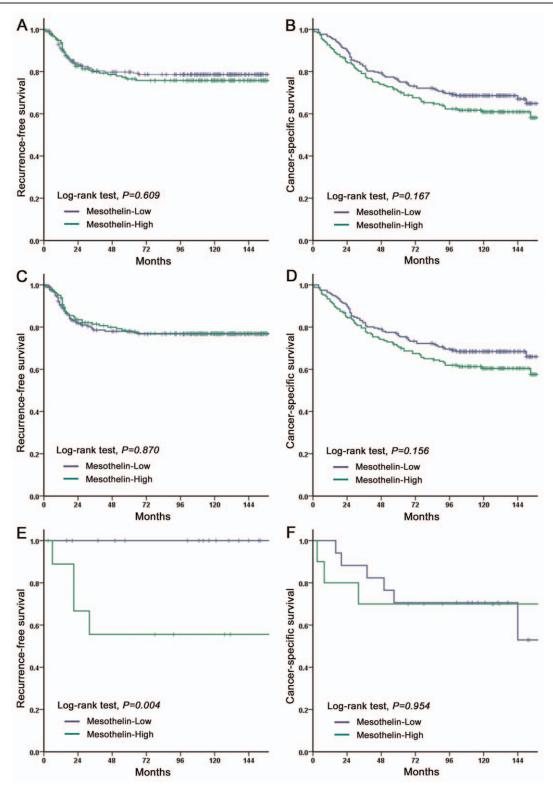


Figure 2. Kaplan-Meier survival analysis of recurrence-free survival (RFS) and cancer-specific survival (CSS) in colorectal adenocarcinoma patients. No significant difference in RFS or CSS was observed in all colorectal adenocarcinoma (A-B) and non-MSI-High subgroup (C-D). High mesothelin expression was associated with shorter RFS (E), and no significant difference in CSS (F) was observed in MSI-High subgroup.

higher rates of treatment response and progression-free survival to pembrolizumab, an immunomodulatory drug targeting programmed cell death 1 (PD-1) protein, than MSS/MSI-Low colorectal adenocarcinoma. [21]

Mesothelin overexpression in malignancies is associated with tumor cell migration and invasion. [6,28] These findings correspond with our results that mesothelin overexpression correlated with frequent lymphatic invasion and high-grade tumor budding.

A previous study demonstrated that mesothelin overexpression promotes tumor cell motility and invasion via the activation of MMP-7 and MMP-9.[6,29]

In gastrointestinal tumors, mesothelin expression is a poor prognostic factor. [30] Co-expression of mesothelin and CA125 was associated with poor patient prognosis in pancreatic cancer. [7] Luminal membrane expression of mesothelin was one of the poor prognostic factors in gastric cancer.[8] Intracellular mesothelin expression correlated with poor patient prognosis in extrahepatic bile duct cancer. [31] In several studies, the expression pattern of mesothelin is divided into luminal and non-luminal patterns. In a 2014 study, luminal staining of mesothelin was correlated with poor prognostic factors in colorectal adenocarcinoma. [19] In another recent study, a luminal pattern of mesothelin expression was not associated with CSS and poor prognostic factors; however, a non-luminal pattern of mesothelin expression was associated with CSS and poor prognostic factors, such as tumor budding, high histologic grading, and lymph node metastasis. [20] In the present study, a completely luminal pattern of staining was observed only in cases with weak intensity, while both luminal and non-luminal patterns were observed simultaneously in cases with moderate to strong positive staining. A score of 2 (moderate to strong expression in ≥30% of tumor cells) was considered as high expression. As a result, cases with only a luminal pattern were considered to have low expression, and non-luminal pattern was associated with poor prognostic factors. These findings are consistent with a previous study; although not statistically significant, our study also showed shorter CSS in patients with mesothelin overexpression.^[20]

Clinical trials of mesothelin-targeting therapy are currently under way for malignant mesothelioma and pancreatic cancer. [12] Mesothelin expression was found in a large number of cases (45.4%) and was associated with poor prognosis. Mesothelin-targeted therapy may be a new treatment candidate for colorectal adenocarcinoma.

5. Conclusion

We revealed that mesothelin expression was significantly associated with aggressive phenotypes in colorectal adenocarcinoma and shorter RFS in the MSI-High group. Further studies are warranted to assess larger cohorts with a large number of MSI-High cases.

Author contributions

Conceptualization: Su-Jin Shin.

Data curation: Hyunsung Kim, Yumin Chung, Seung Sam Paik,

Kiseok Jang.

Formal analysis: Hyunsung Kim, Su-Jin Shin. Investigation: Hyunsung Kim, Yumin Chung.

Methodology: Kiseok Jang. Supervision: Su-Jin Shin.

Validation: Seung Sam Paik, Kiseok Jang. Writing - original draft: Hyunsung Kim. Writing - review & editing: Su-Jin Shin. Su-Jin Shin orcid: 0000-0001-9114-8438.

References

[1] Bray F, Ferlay J, Soerjomataram I, et al. Global cancer statistics 2018: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. CA Cancer J Clin 2018;68:394-424.

- [2] Jung KW, Won YJ, Kong HJ, et al. Cancer statistics in Korea: incidence, mortality, survival, and prevalence in 2015. Cancer Res Treat
- [3] Lindskog EB, Gunnarsdottir KA, Derwinger K, et al. A population-based cohort study on adherence to practice guidelines for adjuvant chemotherapy in colorectal cancer. BMC Cancer 2014;14:948.
- Riley JM, Cross AW, Paulos CM, et al. The clinical implications of immunogenomics in colorectal cancer: a path for precision medicine. Cancer 2018;124:1650-9.
- [5] Chang K, Pastan I. Molecular cloning of mesothelin, a differentiation antigen present on mesothelium, mesotheliomas, and ovarian cancers. Proc Natl Acad Sci U S A 1996;93:136-40.
- [6] Servais EL, Colovos C, Rodriguez L, et al. Mesothelin overexpression promotes mesothelioma cell invasion and MMP-9 secretion in an orthotopic mouse model and in epithelioid pleural mesothelioma patients. Clin Cancer Res 2012;18:2478-89.
- [7] Einama T, Kamachi H, Nishihara H, et al. Co-expression of mesothelin and CA125 correlates with unfavorable patient outcome in pancreatic ductal adenocarcinoma. Pancreas 2011;40:1276-82.
- Einama T, Homma S, Kamachi H, et al. Luminal membrane expression of mesothelin is a prominent poor prognostic factor for gastric cancer. Br J Cancer 2012;107:137-42.
- [9] Rump A, Morikawa Y, Tanaka M, et al. Binding of ovarian cancer antigen CA125/MUC16 to mesothelin mediates cell adhesion. I Biol Chem 2004;279:9190-8.
- [10] Ho M, Bera TK, Willingham MC, et al. Mesothelin expression in human lung cancer. Clin Cancer Res 2007;13:1571-5.
- Tozbikian G, Brogi E, Kadota K, et al. Mesothelin expression in triple negative breast carcinomas correlates significantly with basal-like phenotype, distant metastases and decreased survival. PloS One 2014;9:e114900.
- [12] Morello A, Sadelain M, Adusumilli PS. Mesothelin-targeted CARs: driving T cells to solid tumors. Cancer Discov 2016;6:133-46.
- [13] Einama T, Kawamata F, Kamachi H, et al. Mesothelin-specific immune responses and targeted immunotherapy for mesothelin-expressing tumors. EBioMedicine 2017;24:16-7.
- [14] Baba K, Ishigami S, Arigami T, et al. Mesothelin expression correlates with prolonged patient survival in gastric cancer. J Surg Oncol 2012;105:195-9.
- [15] Illei PB, Alewine C, Zahurak M, et al. Mesothelin expression in advanced gastroesophageal cancer represents a novel target for immunotherapy. Appl Immunohistochem Mol Morphol 2016;24:246-52.
- [16] Alvarez H, Rojas PL, Yong KT, et al. Mesothelin is a specific biomarker of invasive cancer in the Barrett-associated adenocarcinoma progression model: translational implications for diagnosis and therapy. Nanomedicine 2008;4:295-301.
- [17] Ordonez NG. Application of mesothelin immunostaining in tumor diagnosis. Am J Surg Pathol 2003;27:1418-28.
- [18] Liebig B, Brabletz T, Staege MS, et al. Forced expression of deltaN-TCF-1B in colon cancer derived cell lines is accompanied by the induction of CEACAM5/6 and mesothelin. Cancer Lett 2005;223:159-67.
- [19] Kawamata F, Homma S, Kamachi H, et al. C-ERC/mesothelin provokes lymphatic invasion of colorectal adenocarcinoma. J Gastroenterol 2014;49:81-92.
- [20] Shiraishi T, Shinto E, Mochizuki S, et al. Mesothelin expression has prognostic value in stage II/III colorectal cancer. Virchows Arch 2019; 474:297-307.
- [21] Le DT, Uram JN, Wang H, et al. PD-1 blockade in tumors with mismatch-repair deficiency. N Engl J Med 2015;372:2509-20.
- [22] Washington MK, Berlin J, Branton P, et al. Protocol for the examination of specimens from patients with primary carcinoma of the colon and rectum. Arch Pathol Lab Med 2009;133:1539-51.
- [23] Amin MB, Jessup JM, Goldberg RM, et al. AJCC Cancer Staging Manual. 8th ed. NY: Springer; 2017.
- [24] Kim H, Rehman A, Chung Y, et al. Clinicopathologic significance of extranodal tumor extension in colorectal adenocarcinoma with regional lymph node metastasis. Gastroenterol Res Pract 2016;2016:5620765.
- [25] Boland CR, Thibodeau SN, Hamilton SR, et al. A National Cancer Institute Workshop on Microsatellite Instability for cancer detection and familial predisposition: development of international criteria for the determination of microsatellite instability in colorectal cancer. Cancer Res 1998;58:5248-57.
- [26] Boland CR, Goel A. Microsatellite instability in colorectal cancer. Gastroenterology 2010;138:2073-87. e2073.
- [27] Ryan E, Sheahan K, Creavin B, et al. The current value of determining the mismatch repair status of colorectal cancer: a rationale for routine testing. Crit Rev Oncol Hematol 2017;116:38-57.

- [28] Li M, Bharadwaj U, Zhang R, et al. Mesothelin is a malignant factor and therapeutic vaccine target for pancreatic cancer. Mol Cancer Ther 2008;7:286–96.
- [29] Chen SH, Hung WC, Wang P, et al. Mesothelin binding to CA125/ MUC16 promotes pancreatic cancer cell motility and invasion via MMP-7 activation. Sci Rep 2013;3:1870.
- [30] Einama T, Kawamata F, Kamachi H, et al. Clinical impacts of mesothelin expression in gastrointestinal carcinomas. World J Gastrointest Pathophysiol 2016;7:218–22.
- [31] Kawamata F, Kamachi H, Einama T, et al. Intracellular localization of mesothelin predicts patient prognosis of extrahepatic bile duct cancer. Int J Oncol 2012;41:2109–18.