

Received: 2019.09.06

Accepted: 2019.09.29

Published: 2019.10.14

Anti-Colorectal Cancer Mechanisms of Formononetin Identified by Network Pharmacological Approach

Authors' Contribution:

Study Design A

Data Collection B

Statistical Analysis C

Data Interpretation D

Manuscript Preparation E

Literature Search F

Funds Collection G

BCD **Lei Zhang**

BD **Yizhen Gong**

CD **Shuai Wang**

AEG **Feng Gao**

Department of Colorectal and Anal Surgery, The First Affiliated Hospital of Guangxi Medical University, Nanning, Guangxi, P.R. China

Corresponding Author: Feng Gao, e-mail: gxmu_doctorgf@163.com

Source of support: This study is funded by the 2018 Innovation Project of Guangxi Graduate Education (No. YCBZ2018036)

Background: The network pharmacological approach was used to identify the anti-colorectal cancer (CRC) targets of formononetin (FN) and the molecular mechanisms of FN against CRC.

Material/Methods: A tool of the DisGeNET database was used for collection of CRC-based targets. Other tools of SuperPred, herbal ingredients target (HIT), and SwissTargetPrediction databases were applied in prediction of pharmacological targets of FN against cancer. A protein-protein interaction (PPI) network of FN against CRC was obtained by using a STRING database. All top biological functional processes and signaling pathways of FN against CRC were identified by using Database for Annotation, Visualization and Integrated Discovery (DAVID) software and Omicshare cloud platform.

Results: The most key anti-CRC targets of FN were identified as tumor protein p53 (TP53), cytochrome P450 3A4 (CYP3A4), ATP binding cassette subfamily G member 2 (ABCG2), tumor necrosis factor (TNF), epidermal growth factor receptor (EGFR), Erb-B2 receptor tyrosine kinase 2 (ERBB2), and cytochrome P450 1A1 (CYP1A1). In further assays, the treatment of CRC by FN was mainly involved in biological functional processes of reactive oxygen species metabolic process, positive regulation of transcription, DNA-templated, positive regulation of nucleic acid-templated transcription, and positive regulation of RNA metabolic process. anti-CRC by FN of signaling pathways were associated with amyotrophic lateral sclerosis (ALS), allograft rejection, cytokine-cytokine receptor interaction, asthma, mitogen-activated protein kinase (MAPK) signaling pathways, and others.

Conclusions: The anti-CRC molecular mechanisms of FN are implicated in suppression of cellular proliferation and regulation of cancer-related metabolic pathways. Interestingly, 8 optimal biological targets may be used as potential molecular markers for predicting and treating CRC.

MeSH Keywords: **Colorectal Neoplasms • Molecular Mechanisms of Pharmacological Action • Pharmacology**

Full-text PDF: <https://www.medscimonit.com/abstract/index/idArt/919935>



1265



5



23



Background

Globally, epidemiological data indicate that CRC is one of the most common cancer-related causes of death [1]. In clinical practice, most of CRC cases are found with advanced stage, limiting opportunities for medical chemotherapy [2]. Further development of an alternative chemotherapy with high efficiency and low toxicity is a promising anti-CRC research topic. Formononetin (FN), a pharmacologically active compound isolated from *Astragalus* root, reportedly has potent anti-tumor activities [3]. Notably, growing evidence indicates that FN has pharmacological effects against CRC cells, and the underlying mechanism is involved in induction of apoptosis-related pathways [4,5]. However, more detailed anti-CRC mechanisms by FN remain unrevealed, and further study is needed. Interestingly, use of a tool of network pharmacology is a newly emerging strategy for revealing biological targets, functional processes, and molecular pathways of existing treatments for against clinical disease [6]. In addition, the predictive findings may function as possible pharmacological biotargets of chemotherapy against disease prior to in-depth experimental validation [7,8]. At present, there are still limited studies focusing on the biological target and molecular mechanism of FN against CRC. In the present study, a network pharmacological approach was used for identification and visualization of biological targets, functional processes, and signaling pathways of FN against CRC before clinical and experimental validations.

Material and Methods

Experimental design

Figure 1 presents a flow chart showing the experimental design.

Collection and identification of anti-CRC targets of FN

The SuperPred database was used to obtain known targets of FN, and the SwissTargetPrediction database was employed in identification of predictive targets of FN. The CRC-diseased targets were identified by using the DisGeNET database. Subsequently, the combined targets of FN and CRC were further used to produce the anti-CRC targets by FN through the UniProt database.

Construction of PPI network of FN against CRC and topological analysis

The mapped anti-CRC targets of FN were used to obtain the function-related proteins by using a STING database, and protein interaction with confidence score greater than 0.9 was set to exclude targets. The collective targets were imported into Cytoscape software to construct a PPI network of FN against

CRC. The Network-Analyzer in Cytoscape was used to further analyze the average degree of freedom and degree of freedom, and thus the core targets were screened and identified according to degree value. The upper limit of the screening range was the large degree value in the topological data, and the lower limit was twice the average degrees of freedom [9].

Cluster analysis

A multivariate conditional outlier detection (MCODE) algorithm was used to cluster the PPI network of FN-target-CRC, as reported previously [10].

Biological function and pathway enrichment analyses of core targets

The Database for Annotation, Visualization and Integrated Discovery (DAVID) database was used to obtain biological functions and molecular pathways of core targets in FN against CRC before being visualized through the Omicshare cloud platform. Advanced bubble diagrams of signaling pathways and biological processes were plotted according to *P* values. The Kyoto Encyclopedia of Genes and Genomes (KEGG) signaling pathway enrichment analyses of all identifiable targets were performed to further reveal detailed pharmacological mechanisms of FN in the treatment of CRC.

Results

Target information

All 6229 genes found using the DisGeNET database were used for pathogenetic targets. The top 200 targets with *P* values greater than 0.05 were selected as the optimal targets of CRC. Ten anti-cancer targets were obtained through the HIT database, 23 known targets and 20 predictive targets were screened through the SuperPred database, and 15 identifiable targets of FN were identified through the SwissTargetPrediction database. After mapping, 19 intersecting targets of FN against CRC were obtained, followed by 86 interacted and correlated edges (Figure 2). In addition, the most key anti-CRC targets of FN were identified as TP53, CYP3A4, ABCG2, TNF, EGFR, ERBB2, and CYP1A1 (Figure 3).

Cluster analysis and enrichment assay

The subnetworks clustered by the MCODE algorithm using Cytoscape are shown in Figure 4. The predictive targets of CYP2C19, CYP2D6, CYP3A4, and CYP2C9 were clustered as a group. IL4, PPARG, TNF, TP53, ESR1, ESR2, EGFR, BRCA1, ERBB2, ABCG2 targets were clustered and grouped. The Cluego plugin and GraphPad Prism software were used to achieve visualization

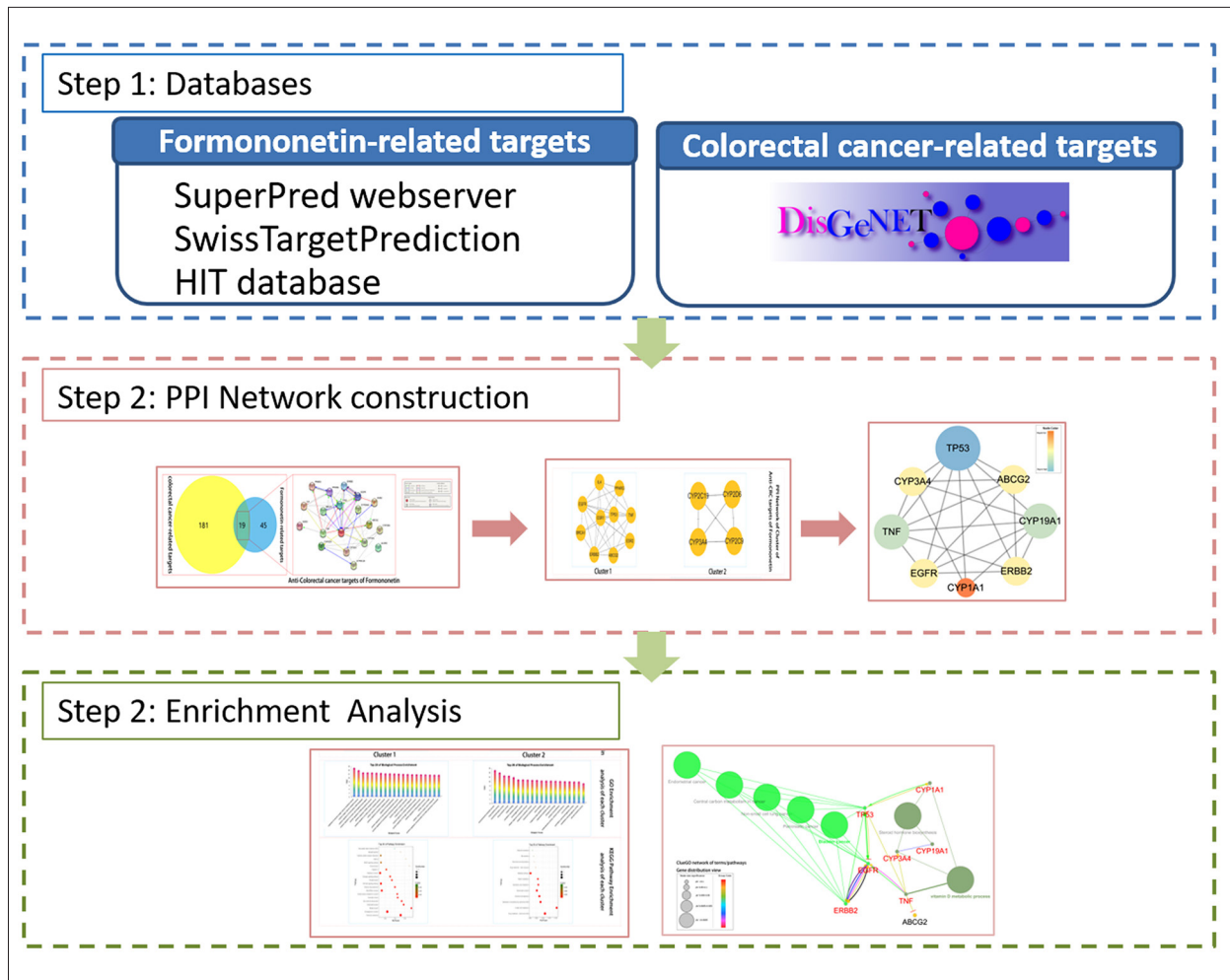


Figure 1. Flow chart of FN against CRC by using a strategy of network pharmacological approach.

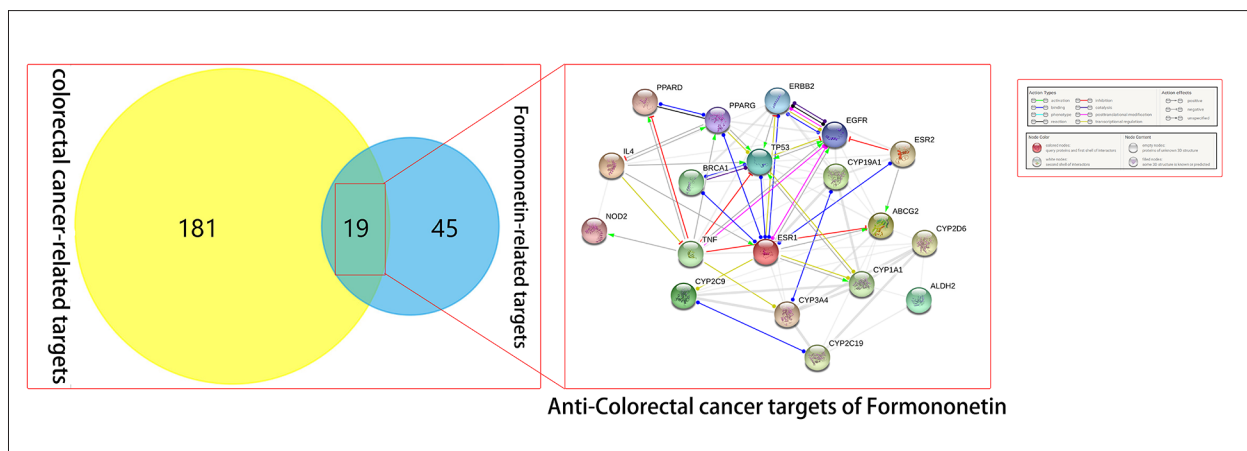


Figure 2. Mapper targets of FN and CRC for construction of a PPI network.

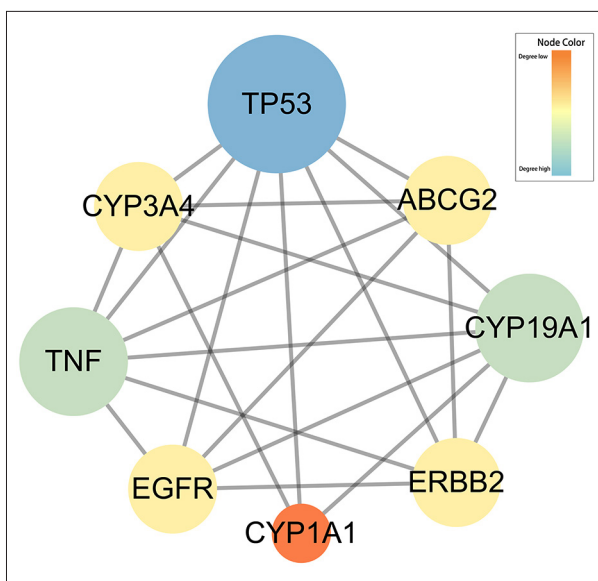


Figure 3. Hub targets of the most important molecules of FN and CRC.

of molecular functions and biological pathways of the most key anti-CRC targets of FN, as revealed in Figure 5.

Discussion

CRC is the third most life-threatening cancer in the world, with a high mortality rate. The lethality rate of CRC ranks the second of fatal malignancies, just after lung carcinoma and breast cancer [11]. Moreover, most CRC patients are diagnosed with advanced stage because of inapparent early symptoms and insufficient clinical inspection [12]. In recent medical regimens, therapeutic bioactive compounds against CRC have been developed as a substitute for traditional cancer treatment methods such as surgery, radiotherapy and chemotherapy. Growing evidence indicates that marked TP53 mutations are found in CRC cells and human samples. Abnormally mutated changes of the TP53 gene play a key role in advanced CRC [13]. Compared with non-cancer tissues, the methylation levels of microRNA-34a in TP53 PIN A1A1 and TP53 MSP GG genotypes in CRC tumors

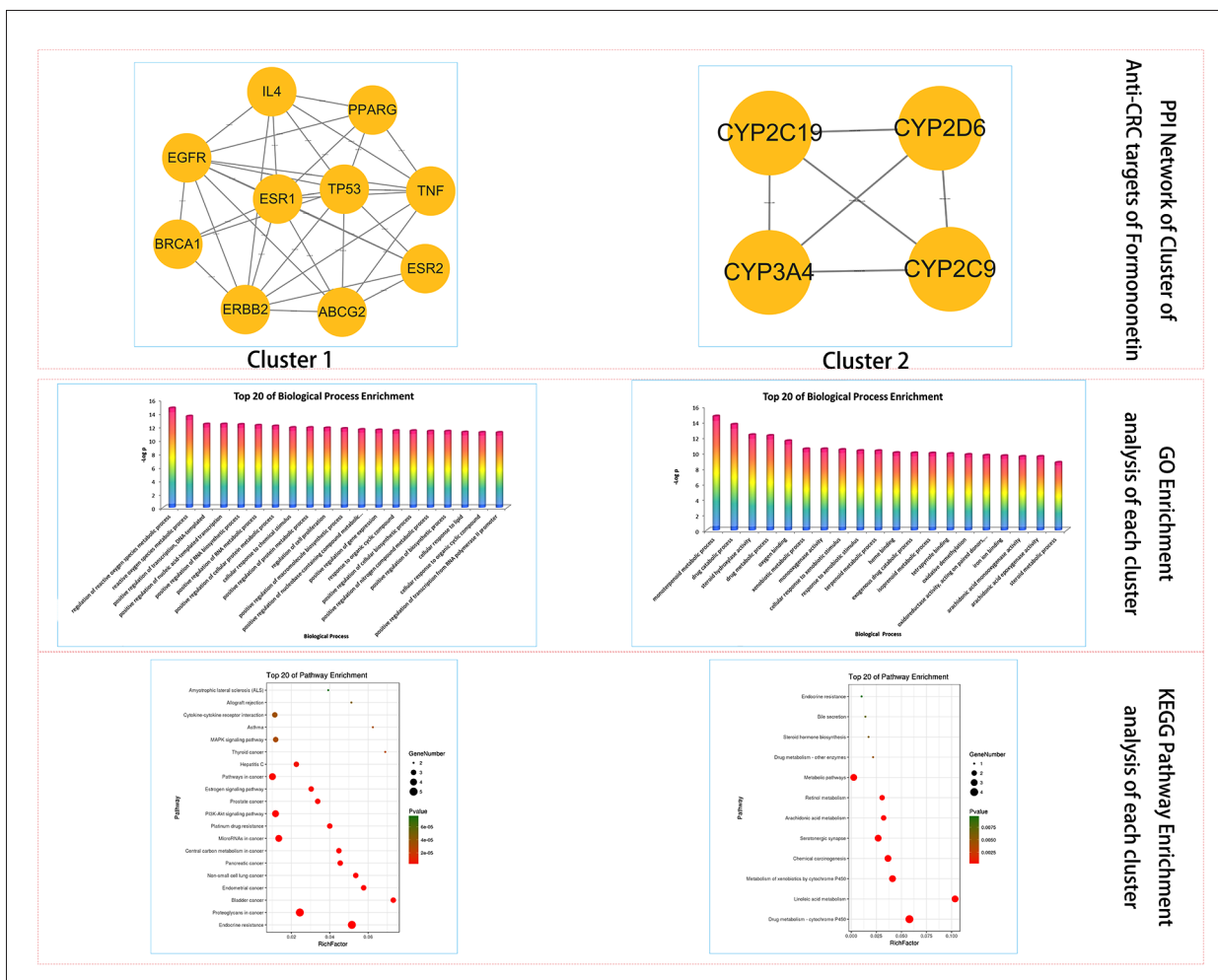


Figure 4. Cluster analysis for correlative association of core biotargets of FN and CRC.

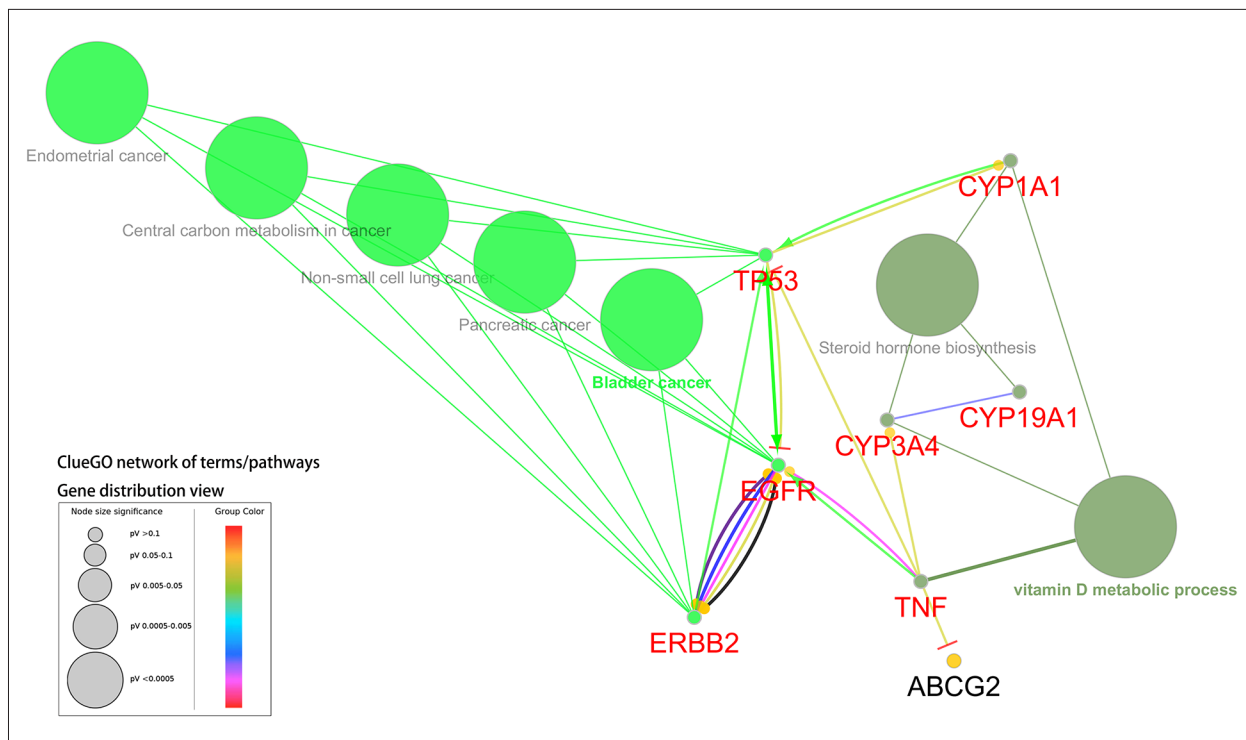


Figure 5. Targets-pathways for anti-CRC exerted by FN, associated with hub target.

are clearly higher [14]. Thus, overexpression of TP53 mutation is a pathological target in CRC progression. Oxidative stress and inflammation play important roles in malignant CRC, and the high expression of ABCG2 in CRC tissue may be a feedback of overoxidation reaction, associated with poor prognosis. A cell culture study showed that downregulation of ABCG2 can inhibit the production of antioxidants, inducing tumor cell growth [15], suggesting that ABCG2 is a potential therapeutic target of CRC. Excessive expression of CYP3A4 is detected in colon tissue [16]. Intestinal CYP3A and P-glycoprotein (P-gp), some important components in the host defense barrier, exert an important effect in the pathogenesis of disease, drug exposure, and environmental stimulus. Clinical and experimental data suggest that the expressions of CYP3A and P-gp in the intestinal tract of patients with inflammatory bowel disease is clearly lower, accompanied with imbalance of the gut microbiome [17]. The steroid-related gene CYP19A1 can affect estrogen metabolic processes, and it can increase cancer risk through inflammatory stress [18]. Cytochrome P450 1A1 (CYP1A1) is a crucial metabolic enzyme involved in the metabolism of many xenobiotic organisms. One of the pathogenetic mechanisms of CYP1A1-mediated carcinogenesis is reported be that CYP1A1 metabolizes polycyclic aromatic hydrocarbons (PAHs) to active epoxy intermediates, and then covalently binds into DNA for inducing tumors [19]. Collectively, cytochrome P450 enzymes (CYP3A4, CYP19A1, and CYP1A1) may function as potent biological and therapeutic targets against CRC. In addition, endogenous TNF/TNF receptor (TNFR) pathways affect survival rates

of patients with cancer [20], suggesting that TNF exerts a key role in CRC development. High expression of EGFR is observed in cancer cells and tissues [21]. EGFR is functionally involved in proliferation, survival, invasion, and immunodeficiency of metastatic CRC [22]. ERBB2 (HER2) amplification is an emerging biomarker in CRC, increasing sensitivity to combination therapy against HER2 [23]. According to the literature, it can be speculated that FN plays an effective anti-cancer role by regulating the expression of predictive targets in CRC cells. Molecular classification and identification of new and emerging biomarker indicates a key role in cancer progression, and it may provide insight into clinical information for prolonging the survival of patients with CRC. In addition, GO annotation analysis and KEGG pathway assay of biological targets of FN against CRC showed most of the enriched top signaling pathways.

Conclusions

The prediction of biotargets and molecular mechanisms by using a tool of network pharmacology is accurate and practical, and this approach sheds light on an attractive scientific foundation for further exploring the pharmacological targets and signaling mechanisms of FN against CRC.

Conflict of interests

None.

References:

1. Thomas J, Ohtsuka M, Pichler M et al: MicroRNAs: Clinical relevance in colorectal cancer. *Int J Mol Sci*, 2015; 16: 28063–76
2. Fakhri MG: Metastatic colorectal cancer: Current state and future directions. *J Clin Oncol*, 2015; 33: 1809–24
3. Tay KC, Tan LT, Chan CK et al: A review of its anticancer potentials and mechanisms. *Front Pharmacol*, 2019; 10: 820
4. Auyeung KK, Law PC, Ko JK: Novel anti-angiogenic effects of formononetin in human colon cancer cells and tumor xenograft. *Oncol Rep*, 2012; 28: 2188–94
5. Wang AL, Li Y, Zhao Q, Fan LQ: Formononetin inhibits colon carcinoma cell growth and invasion by microRNA-149-mediated EphB3 downregulation and inhibition of PI3K/AKT and STAT3 signaling pathways. *Mol Med Rep*, 2018; 17: 7721–29
6. Su M, Guo C, Liu M et al: Therapeutic targets of vitamin C on liver injury and associated biological mechanisms: A study of network pharmacology. *Int Immunopharmacol*, 2019; 66: 383–87
7. Zhou R, Wu K, Su M, Li R: Bioinformatic and experimental data decipher the pharmacological targets and mechanisms of plumbagin against hepatocellular carcinoma. *Environ Toxicol Pharmacol*, 2019; 70: 103200
8. Wu K, Wei P, Liu M et al: To reveal pharmacological targets and molecular mechanisms of curcumin against interstitial cystitis. *J Adv Res*, 2019; 20: 43–50
9. Li J, Guo C, Lu X, Tan W: Anti-colorectal cancer biotargets and biological mechanisms of puerarin: Study of molecular networks. *Eur J Pharmacol*, 2019; 858: 172483
10. Li R, Ma X, Song Y et al: Anti-colorectal cancer targets of resveratrol and biological molecular mechanism: Analyses of network pharmacology, human and experimental data. *J Cell Biochem*, 2019; 120: 11265–73
11. Siegel RL, Miller KD, Jemal A: Cancer statistics, 2019. *Cancer J Clin*, 2019; 69: 7–34
12. Brenner H, Kloor M, Pox CP: Colorectal cancer. *Lancet*, 2014; 383: 1490–502
13. Sikorska A, Flisikowska T, Stachowiak M et al: Elevated expression of p53 in early colon polyps in a pig model of human familial adenomatous polyposis. *J Appl Genet*, 2018; 59: 485–91
14. Jun HH, Kwack K, Lee KH et al: Association between TP53 genetic polymorphisms and the methylation and expression of miR-34a, 34b/c in colorectal cancer tissues. *Oncol Lett*, 2019; 17: 4726–34
15. Nie S, Huang Y, Shi M et al: Protective role of ABCG2 against oxidative stress in colorectal cancer and its potential underlying mechanism. *Oncol Rep*, 2018; 40: 2137–46
16. Bethke L, Webb E, Sellick G et al: Polymorphisms in the cytochrome P450 genes CYP1A2, CYP1B1, CYP3A4, CYP3A5, CYP11A1, CYP17A1, CYP19A1 and colorectal cancer risk. *BMC Cancer*, 2007; 7: 123
17. Gao XJ, Li T, Wei B et al: [Regulatory mechanisms of gut microbiota on intestinal CYP3A and P-glycoprotein in rats with dextran sulfate sodium-induced colitis.] *Yao Xue Xue Bao*, 2017; 52: 34–43 [in Chinese]
18. Slattery ML, Lundgreen A, Herrick JS et al: Variation in the CYP19A1 gene and risk of colon and rectal cancer. *Cancer Causes Control*, 2011; 22: 955–63
19. Zhu X, Wang Z, He J et al: Associations between CYP1A1 rs1048943 A>G and rs4646903 T>C genetic variations and colorectal cancer risk: Proof from 26 case-control studies. *Oncotarget*, 2016; 7: 51365–74
20. Guo Y, Feng Y, Liu H et al: Potentially functional genetic variants in the TNF/TNFR signaling pathway genes predict survival of patients with non-small cell lung cancer in the PLCO cancer screening trial. *Mol Carcinog*, 2019; 58: 1094–104
21. Miyamoto Y, Muguruma N, Fujimoto S et al: EGFR-targeted molecular imaging of colorectal tumors: Detection and treatment evaluation of tumors in animal models. *Cancer Sci*, 2019; 110: 1921–30
22. Mizukami T, Izawa N, Nakajima TE et al: Targeting EGFR and RAS/RAF signaling in the treatment of metastatic colorectal cancer: From current treatment strategies to future perspectives. *Drugs*, 2019; 79: 633–45
23. Siravegna G, Sartore-Bianchi A, Nagy RJ et al: Plasma HER2 (ERBB2) copy number predicts response to HER2-targeted therapy in metastatic colorectal cancer. *Clin Cancer Res*, 2019; 25: 3046–53