

Short Communication Animal Genetics

Inferring the historical demography of southern African cheetahs (*Acinonyx jubatus*) using Bayesian analyses of molecular genetic data

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Abstract

The contemporary genetic diversity of the cheetah (Acinonyx jubatus) has been the focus of several studies, which have revealed very low levels of variation. Different hypotheses have been proposed to explain this pattern of low diversity, and require additional scrutiny. Here, we used published microsatellite data and coalescence-based analytical methods to explore the historical demography of the largest free-ranging cheetah population, aiming to assess whether present-day diversity may have been impacted by a historical demographic decline. Our results support the hypothesis of a historical (and most likely gradual) demographic decline over the past $\sim 10,000$ years, leading to a present-day $N_{\rm e}$ ranging from 700 to 1,600 individuals. This decline was likely induced by climate-driven vegetational shifts affecting habitat suitability and possibly also interspecies interactions with prey and competitors. These results help clarify the demographic history of cheetahs in southern Africa and its impact on the current genetic diversity of this population.

Keywords: Coalescent modelling, Felidae, population decline, Namibia, microsatellites.

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The historical demography of a species reflects the changes in its population size over time. It plays a crucial role in the survival of species, as reductions in population size negatively affect levels of genetic diversity, and low levels of genetic diversity are often linked to reduced fitness. Moreover, understanding past fluctuations, and their relationships with present-day genetic diversity, is relevant in the context of understanding present-day dynamics and predicting likely outcomes of future scenarios.

Reductions in population size are often caused by external factors such as climate change, habitat degradation, prey-predator dynamics, and disease outbreaks. Throughout the Quaternary period (2.5 million years ago to the present), the climate was highly heterogeneous in Africa, with that of western and eastern Africa being relatively unstable compared to southern Africa (Stokes *et al.*, 1997; Maslin *et al.*, 2012). This variation in climate was accompanied by changes in species-specific habitat suitability and has likely affected the contemporary genetic diversity of many species (Hewitt 2004; Chase *et al.*, 2010). Comparative phylogeography across taxa indicates that southern Africa was a refugium from which populations recolonized more northerly regions (Lorenzen

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et al., 2010; Bertola et al., 2011; Schwab et al., 2011). But even in southern Africa, notable oscillations between wet and dry periods have been reported in the Holocene (Hewitt 2000). Since responses can be species- or population-specific (Lorenzen et al., 2011; Kim et al., 2016; Nadachowska-Brzyska et al., 2016), assessing comparative historical patterns should benefit from a growing body of studies targeting different taxa.

One species that seems to have a particularly interesting demographic history is the cheetah (Acinonyx jubatus), for which remarkably low levels of genetic diversity were first identified in the early 1980s (O'Brien et al., 1983, 1985) and more recently confirmed by studies employing genome-wide data (Dobrynin et al., 2015; Prost et al., 2022). While it is widely recognized that the origin of the cheetah's low extant genetic diversity is predominantly the result of events predating modern civilization, different hypotheses have been proposed to account for this phenomenon. Early genetic studies using various molecular markers (allozymes, mtDNA restriction fragment length polymorphisms, minisatellite fingerprinting) and samples from southern and eastern Africa, hypothesized that the low diversity was likely a consequence of one or more bottlenecks at the end of the Pleistocene (12,000 - 10,000 years ago [ya]) (O'Brien et al., 1983, 1987; Menotti-Raymond and O'Brien, 1993). Two alternative hypotheses to a reduction in population size as cause for the low diversity were subsequently proposed. First, that the low diversity could have been due to the persistence of the species at low effective population size (N), induced by the high reproductive variance observed in species with a polygynous mating system (Pimm et al., 1989). Second, that it could be due to a continuous cycle of

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extinction of subpopulations followed by re-colonization of the areas, following a metapopulation dynamics (Pimm et al., 1989; Gilpin, 1991; Hedrick, 1996). In these two alternative hypotheses, the low diversity could have been maintained over a long period without implying a detectable historical population reduction. Subsequent genetic studies using microsatellite and MHC loci were in agreement with the inference of an ancestral bottleneck (e.g., Driscoll et al., 2002; Castro-Prieto et al., 2011).

During the past decades, there has been a surge of advances in computational methods exploring the historical demography of modern populations using empirically collected molecular data. Of particular interest is the application of the coalescent and Bayesian approaches in population genetics (e.g., Luikart and England, 1999; Storz and Beaumont, 2002; Beaumont MA, 2010; Lopes and Beaumont, 2010). These methods have now been widely used to model changes in N_a of many different species, including the cheetah (Okello et al., 2008; Phillips et al., 2012; Quéméré et al., 2012; Dobrynin et al., 2015). Results based on the analyses of whole genome data support a historical decline in the cheetah population size as the cause for its low genetic diversity, rather than a persistently low effective population sizes or a metapopulation dynamics (as proposed by the alternative hypotheses mentioned above). At the same time, while population decline was supported, even with complete genomes the mode of decline could not be clearly determined (Dobrynin et al., 2015). Different analytical approaches (Dadi vs. PSMC) supported a sharp or a gradual decline, respectively (Dobrynin et al., 2015), indicating that this issue remains incompletely resolved.

To investigate the support for these hypotheses using an independent dataset, here we assessed the demographic history of the Namibian cheetah population using coalescent-based methods applied to a large, previously published microsatellite data set (Marker et al., 2008). Namibia has one of the largest remaining cheetah populations, estimated at 1,500 adult and adolescent individuals, and connected to the broader southern Africa contiguous adult population of ~3,500-6,800 individuals (Charruau et al., 2011; Durant et al., 2017, Weise et al., 2017). This population was considered appropriate for the study due to its relatively large census size and being panmictic (Marker et al., 2008). Panmixia is crucial, as it reduces the risk of false signals of bottleneck caused by sub-structuring (Sousa et al., 2009; Peter et al., 2010), while larger current sizes reduce the likelihood of the population having experienced high genetic drift in the recent past (Tallmon et al., 2010).

Our data set comprises 89 unrelated individuals (determined based on behavioral data, parentage analyses and estimates of genetic relatedness) and originally contained 38 microsatellite loci (Marker *et al.*, 2008). For this study, we only included the 29 loci that showed modal allelic distribution. We used this dataset to assess historical trends in population size using four coalescent-based approaches, implemented in the programs MSVAR 1.3 (Storz and Beaumont, 2002), LAMARC v2.1.2b (Kuhner, 2006), Migrate-n v.3.6.6 (Beerli and Palczewski, 2010) and VAREff v1.2 (Nikolic and Chevalet, 2014). These approaches employ a coalescent framework based on the Wright-Fisher model (Moyer *et al.*, 2012; Sharma

et al., 2013), an assumption which our study population approximates (Marker et al., 2008).

We initially employed MSVAR1.3 to test whether this dataset supports any inference of changes in N_e in this population and, if so, whether this inference was robust to varying model parameters. We ran six independent analyses varying in chain length $(2 \times 10^9 \text{ or } 4 \times 10^9 \text{ steps})$ and spacing among sampled states $(1 \times 10^5 \text{ or } 2 \times 10^5, \text{ respectively})$, in every case yielding 10,000 samples after a 50% burn-in. We also varied the assumed mutation rate $(10^{-6} \text{ or } 10^{-3})$ (Li *et al.* 2002), the assumed demographic trend in the prior model (stable or expanding size), and in case of demographic change, whether it was linear or exponential.

Subsequently, we used LAMARC to estimate trends in this population's long-term N_a and demographic exponential growth rates (g). Two independent runs, each of 2×10^9 steps were performed, with 20,000 recorded parameters sampled every 10⁵ steps, with 10% additional steps employed as burnin. Priors for the population diversity parameter 'theta' (θ) and g were drawn from uniform distributions, and ranged from 1×10^{-5} to 10 and -500 to 500, respectively. Convergence of the runs was assessed using the program Tracer v1.4 (Rambaut and Drummond, 2007). Long-term effective population size was determined by solving the formula $\theta = 4N_e\mu$, where θ is the estimated population diversity parameter, N_e is the effective population size and μ is the mean dinucleotide microsatellite mutation rate per locus per generation (5.6×10^{-4} ; Weber and Wong,1993). Generation time was assumed to be 6 years (Marker and O'Brien, 1989).

We then used the Bayesian inference implemented in Migrate-n to estimate the present-day θ and the demographic history (via skyline plot) of this cheetah population. The main parameters used were: 8 million generations sampled every 200 steps, resulting in 40,000 recorded steps. Burn-in was 500,000 steps. The prior distribution of θ was set to uniform between 0 and 15.

Finally, we used VarEff to estimate past changes of effective population size using approximate likelihoods in a Markov Chain Monte Carlo approach. We employed the command 'Theta' with the following parameters: Theta (NBLOC = 30, JMAX = 5, MODEL = 'G 0.15', MUTAT = 0.00056, NBAR = 2000, VARP1 = 4, RHOCORN = 0, GBAR = 10000, VARP2 = 3, DMAXPLUS = 8, Diagonale = 0.5, NumberBatch = 10000, LengthBatch = 20, SpaceBatch = 20, Burnin = 50000, AccRate=0.25). Mutation rate and generation time were assumed as described above.

All MSVAR runs supported a scenario of population decline, irrespective of the assessed variations in model parameters and demographic assumptions (Figure 1). This included runs in which the population was assumed to have expanded in the prior model, and which still yielded an inference of demographic decline.

Consistently, the LAMARC results also supported a decline scenario by yielding a negative estimated growth rate of -0.2 (95% CI: -0.15 - -0.05). Although this rate of decline is small, its confidence interval did not overlap zero, implying that this approach strongly supported the inference that the population size was larger in the past. In addition, the

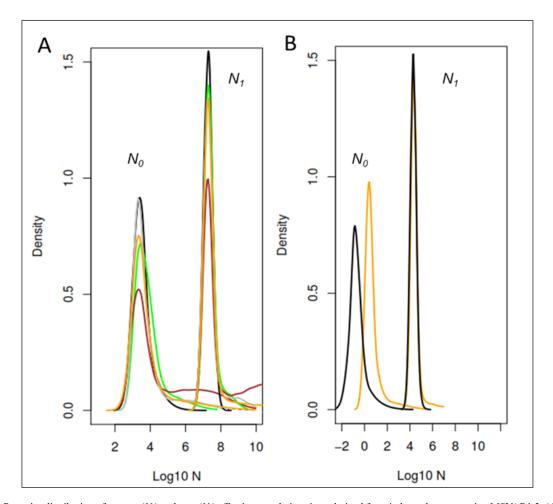


Figure 1 – Posterior distribution of present (N_0) and past (N_1) effective population sizes, derived from independent runs using MSVAR1.3. (A) assumes a microsatellite mutation rate of 10e-6 and exponential changes; black and green lines represent runs that assumed the population to have expanded, while brown and gray to have remained stable; orange represents the combined runs. (B) Results from two independent MSVAR1.3 long chains (4×10^{-9} iterations each) in which the population was modeled as stable but allowed to change exponentially (orange) or linearly (black). N_0 , N_1 = recent, ancestral population sizes, respectively.

LAMARC result yielded a low value for present-day θ (3.44 [95% CI: 3.13 - 3.72]), which translates to an estimated $N_{\rm e}$ of 1,537 (CI: 1.399 - 1.661) individuals.

The Migrate-n skyline plot depicted a continuous declining trend, from an $N_{\rm e}$ of ~40,000 individuals ca. 16,000 ya to ~3,500 individuals ca. 150 ya (Figure 2). Although values older than ca. 6,000 ya are based on small sampling and thus less reliable, the overall trend of decline was robust throughout the whole period, slowing down ca. 1,500 ya. Migrate-n estimated the current θ of the Namibian cheetah population as 3.65 (95% CI = 3.29-4.67) which translates to an $N_{\rm e}$ of 1,629 (1,469-2,085), similar to the results estimated with LAMARC.

The VarEff results also reconstructed a trend of reduction of this cheetah population's N_e in the last 10,000 years (Figure 3), consistent with the Migrate-n and LAMARC results. This trend appears to have been continuous (using both the harmonic mean and median estimates). Prior to ca. 10,000 ya, VarEff showed a population increase that started ca. 30,000 ya, after which the decline ensued. Overall, VarEff suggested smaller population numbers, with a present-day N_e of the Namibian

cheetah population estimated to be of only *ca*. 700-1000 individuals.

Overall, our results were very consistent in the reconstruction of the demographic history of Namibian cheetahs, which represent the broader southern African population considering their connectivity across this region (Charruau *et al.*, 2011). We initially focused on the question of whether an independent genetic dataset (relative to the genome analyses reported by Dobrynin et al. 2015) supports the hypothesis of a historical demographic decline in this species. While there was some variation among the estimates derived from our four methods (MSVAR, LAMARC, Migrate-n, VarEff), all analyses supported a historical decline in this cheetah population. Two of the methods (Migrate-n and VarEff) allowed a reconstruction of its demographic trajectory, with concordant indication of a decline beginning at least ~10,000 years ago (see Figures 2 and 3).

The magnitude and rate of decline varied among the analyses. While LAMARC estimated a mild but significant rate of decline (-0.2), Migrate-n and VarEff retrieved a 4 to 10-fold decrease in N_e over a 10,000-year period (i.e., ~30,000 to ~3,500; ~3,000 to ~700 individuals, respectively). Although

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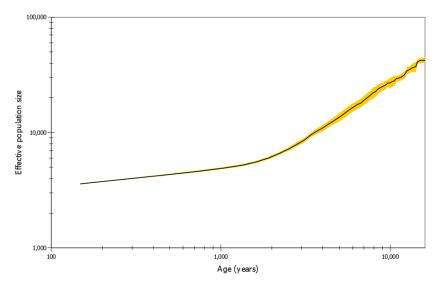


Figure 2 – Skyline plot generated with Migrate-n, presenting estimated N_e values between 16,000 and 100 ya in log scale. The black line is the median estimate and the yellow band shows the 95% highest posterior density intervals.

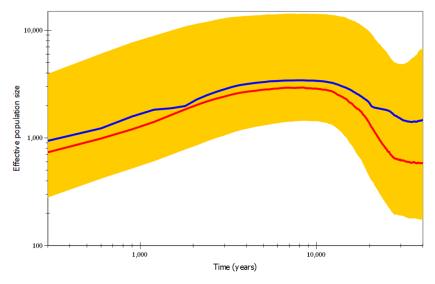


Figure 3 – VarEff skyline plot presenting estimated N_e values between 40,000 and 100 ya in log scale. The red and blue lines are the harmonic mean and the median estimates, respectively, and the yellow band shows the 95% highest posterior density intervals.

the estimated absolute timing of those demographic changes would be affected by the assumed mutation rate, microsatellite mutation rates are species- and locus-specific (Ellegren, 2004), and no specific rates have been directly ascertained for cheetahs. While assuming different rates would change the timeframe of inferred shifts in population size, the conclusion regarding the declining trend would still hold.

Given the consistent support for a historical decline, our second focus was to assess whether this decline was gradual or abrupt. Our findings consistently support a gradual decline, in agreement with the results of a PSMC analysis using whole genome data (Dobrynin *et al.*, 2015). We also observed a recent attenuation of the rate of population decrease in the Migrate-n skyline plot (Figure 1), suggesting that the process of demographic decline may have occurred at different paces over time.

Overall, our results indicate that the low levels of genetic diversity in present-day cheetahs (at least in southern African populations) can be explained by the inferred historical demographic decline. Accordingly, we observed no evidence for the long-term maintenance of $N_{\rm e}$ at low levels, as would be expected from the alternative hypotheses that imply either high reproductive variance due to the cheetah's polygynous mating system, or a continuous metapopulation dynamics (Pimm *et al.*, 1989; Gilpin, 1991; Hedrick, 1996).

An additional inference based on our analyses pertains to the estimates of present-day N_e in this cheetah population (which likely represents the genetically continuous southern African population; Weise *et al.*, 2017). The estimates were consistently low across all methods, ranging from 700-1000 individuals for VarEff to 1,537 for LAMARC and 1,629 for Migrate-n. It is frequently accepted that N_e tends to be 5-10

times smaller than the census size of a population (Frankham $et\,al,2014$), which would suggest that cheetah census numbers in the region (southern Africa) range from 3,500 to 16,000 individuals. These values are comparable with the overall estimated numbers for cheetahs in this area, whose latest estimate for adult individuals alone was 3,500-4,000 (Durant $et\,al.,2017$). Weise $et\,al.,2017$). The congruence between this census estimate and our $N_{\rm e}$ -based estimate indicates robustness of the genetic assessments of present-day diversity and previous demographic trends leading up to this inferred scenario. In addition, this congruence highlights the usefulness of available genetic data, including those obtained with microsatellite markers, to estimate demographic parameters from present-day populations.

The inferred demographic decline of southern African cheetahs has likely been driven by one or more ecological processes, such as climate-induced vegetational shifts and changes in inter-species dynamics with prey and/or competitors (including humans). Climate-driven habitat shifts in this time frame have been pervasive in Africa (Stokes et al., 1997; Hewitt 2000; Hewitt 2004; Chase et al., 2010; Lorenzen et al., 2010; Bertola et al., 2011; Schwab et al., 2011; Maslin et al., 2012), and may have affected cheetah densities directly. Furthermore, it is plausible that interspecific competition with lions (Panthera leo), leopards (Panthera pardus), and spotted hyenas (Crocuta crocuta), the cheetah's main competitors (Durant, 2000; Walker et al., 2022), were exacerbated during times of reduced habitat and prey availability. In addition to the historical declines detected in this study, cheetahs have undergone severe recent population declines, largely attributed to anthropogenic threats and habitat reduction (Marker-Kraus et al., 1996; Durant et al., 2017). Overall, the results from this study provide useful information for improving our understanding of the cheetah's long-term demography, especially for populations in southern Africa, and highlight the importance of genetic research on this complex and threatened species.

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Conflict of Interest

The authors declare that there is no conflict of interest that could be perceived as harmful to the impartiality of the reported research.

Author Contributions

ECF and EE conceived and designed the study; ASK, LM and EE supervised study implementation; ECF and SLB analyzed the data; ASK and EE supervised data interpretation; ECF wrote the first draft of the manuscript; ASK and EE wrote additional sections of the manuscript; SLB, SJO and LM edited the manuscript; all authors read and approved the final version.

References

- Beaumont MA (2010) Approximate Bayesian computation in evolution and ecology. Annu Rev Ecol Evol Syst 41:379-406.
- Beerli P and Palczewski M (2010) Unified framework to evaluate panmixia and migration direction among multiple sampling locations. Genetics 185:313-26.
- Bertola LD, van Hooft WF, Vrieling K, Uit de Weerd DR, York DS, Bauer H, Prins HHT, Funston PJ, Udo de Haes HA, Leirs H *et al.* (2011) Genetic diversity, evolutionary history and implications for conservation of the lion (*Panthera leo*) in West and Central Africa. J Biogeogr 38:1356-1367.
- Castro-Prieto A, Wachter B and Sommer S (2011) Cheetah paradigm revisited: MHC diversity in the world's largest free-ranging population. Mol Biol Evol 28:1455-1468.
- Charruau P, Fernandes C, Orozco-Terwengel P, Peters J, Hunter L, Ziaie H, Jourabchian A, Jowkar H, Schaller G, Ostrowski S *et al.* (2011) Phylogeography, genetic structure and population divergence time of cheetahs in Africa and Asia: Evidence for long-term geographic isolates. Mol Ecol 20:706-724.
- Chase BM, Meadows ME, Carr AS and Reimer PJ (2010) Evidence for progressive Holocene aridification in southern Africa recorded in Namibian hyrax middens: Implications for African Monsoon dynamics and the "African Humid Period". Quatern Res 74:36-45.
- Dobrynin P, Liu S, Tamazian G, Xiong Z, Yurchenko AA, Krasheninnikova K, Kliver S, Schmidt-Küntzel A, Koepfli KP, Johnson W *et al.* (2015) Genomic legacy of the African cheetah, *Acinonyx jubatus*. Genome Biol 16:277.
- Driscoll CA, Menotti-Raymond M, Nelson G, Goldstein D and O'Brien SJ (2002) Genomic microsatellites as evolutionary chronometers: A test in wild cats. Genome Res 12:414-423.
- Durant SM (2000) Living with the enemy: Avoidance of hyenas and lions by cheetahs in the Serengeti. Behav Ecol 11:624-632.
- Durant SM, Mitchell N, Groom R, Pettorelli N, Ipavec A, Jacobson AP, Woodroffe R, Böhm M, Hunter LTB, Becker MS et al. (2017) The global decline of cheetah Acinonyx jubatus and what it means for conservation. Proc Natl Acad Sci U S A 114:528-533.
- Ellegren H (2004) Microsatellites: Simple sequences with complex evolution. Nat Rev Genet 5:435-445.
- Frankham R, Bradshaw CJA and Brook BW (2014) Genetics in conservation management: Revised recommendations for the 50/500 rules, Red List criteria and population viability analyses. Biol Conserv 170:56-63.
- Gilpin M (1991) The genetic effective size. Biol J Linn Soc 42:165-175.
- Hedrick PW (1996) Bottleneck(s) in cheetahs or metapopulation. Conserv Biol 10:897-899.
- Hewitt G (2000) The genetic legacy of the Quaternary ice ages. Nature 405:907-913.
- Hewitt GM (2004) Genetic consequences of climatic oscillations in the Quaternary. Philos Trans R Soc Lond B Biol Sci 359:183-195.
- Kim S, Cho YS, Kim H-K, Chung O, Kim H, Jho S, Seomun H, Kim J, Bang WY, Kim C et al. (2016) Comparison of carnivore, omnivore, and herbivore mammalian genomes with a new leopard assembly. Gen Biol 17:211.
- Kuhner M (2006) LAMARC 2.0: Maximum likelihood and Bayesian estimation of population parameters. Bioinformatics 22:768-770.
- Li Y-C, Korol AB, Fahima T, Beiles A and Nevo R (2002) Microsatellites: Genomic distribution, putative functions and mutational mechanisms: a review. Mol Ecol 11: 2453-2465.

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Lorenzen ED, Masembe C, Arctander P and Siegismund HR (2010) A long-standing Pleistocene refugium in southern Africa and a mosaic of refugia in East Africa: Insights from mtDNA and the common eland antelope. J Biogeogr 37:571-581.

- Lorenzen ED, Nogués-Bravo D, Orlando L, Weinstock J, Binladen J, Marske KA, Ugan A, Borregaard MK, Gilbert MT, Nielsen RRN *et al.* (2011) Species-specific responses of Late Quaternary megafauna to climate and humans. Nature 479:359-364.
- Lopes JS and Beaumont MA (2010) ABC: A useful Bayesian tool for the analysis of population data. Infect Genet Evol 10:826-833.
- Luikart G and England PR (1999) Statistical analysis of microsatellite DNA data. Trends Ecol Evol. 14:253-256.
- Marker L and O'Brien SJ (1989) Captive breeding of the cheetah *Acinonyx jubatus* in North American Zoos (1871-1986). Zoo Biology 8:3-16.
- Marker L, Pearks Wilkerson AJ, Sarno RJ, Martenson J, Breitenmoser-Würsten C, Obrien SJ and Johnson WE (2008) Molecular genetic insights on cheetah (*Acinonyx jubatus*) ecology and conservation in Namibia. J Hered 99:2-13.
- Marker-Kraus L, Kraus D, Barnett D and Hurlbut S (1996) Cheetah Survival on Namibian Farmlands, Cheetah Conservation Fund, Windhoek, Namibia.
- Maslin MA, Pancost RD, Wilson KE, Lewis J and Trauth MH (2012)
 Three and half million year history of moisture availability of
 South West Africa: Evidence from ODP site 1085 biomarker
 records. Palaeogeogr Palaeoclimatol Palaeoecol 317-318:4147.
- Menotti-Raymond MA and O'Brien SJ (1993) Dating the genetic bottleneck of the African cheetah. Proc Natl Acad Sci U S A 90:3172-3176.
- Moyer GR, Sweka JA and Peterson DL (2012) Past and present processes influencing genetic diversity and effective population size in a natural population of Atlantic sturgeon. Trans Am Fish Soc 141:56-67.
- Nadachowska-Brzyska K, Burri R, Smeds L and Ellegren H (2016) PSMC analysis of effective population sizes in molecular ecology and its application to black-and-white Ficedula flycatchers. Mol Ecol 25:1058-1072.
- Nikolic N and Chevalet C (2014) Detecting past changes of effective population size. Evol Appl 7:663-681.
- O'Brien SJ, Wildt DE, Goldman D, Merril CR and Bush M (1983) The cheetah is depauperate in genetic variation. Science 221:459-462.
- O'Brien SJ, Roelke ME, Marker LL, Newman A, Winkler CA, Meltzer D, Colly L, Evermann JF, Bush M and Wildt DE (1985) Genetic basis for species vulnerability in the cheetah. Science 227:1428-1434.
- O'Brien SJ, Wildt DE, Bush M, Caro TM, FitzGibbon C, Aggundey I and Leakey RE (1987) East African cheetahs: Evidence for two population bottlenecks? Proc Natl Acad Sci U S A 84:508-511.
- Okello JB, Wittemyer G, Rasmussen HB, Arctander P, Nyakaana S, Douglas-Hamilton I and Siegismund HR (2008) Effective population size dynamics reveal impacts of historic climatic events and recent anthropogenic pressure in African elephants. Mol Ecol 17:3788-3799.
- Peter BM, Wegmann D and Excoffier L (2010) Distinguishing between population bottleneck and population subdivision by a Bayesian model choice procedure. Mol Ecol 19:4648-4660.
- Phillips CD, Hoffman JI, George JC, Suydam RS, Huebinger RM, Patton JC and Bickham JW (2012) Molecular insights into the

- historic demography of bowhead whales: Understanding the evolutionary basis of contemporary management practices. Ecol Evol 3:18-37.
- Pimm SL, Gittleman JL, McCracken GF and Gilpin M (1989) Plausible alternatives to bottlenecks to explain reduced genetic diversity. Trends Ecol Evol. 4:176-178.
- Prost S, Machado AP, Zumbroich J, Preier L, Mahtani-Williams S, Meissner R, Guschanski K, Brealey JC, Fernandes CR, Vercammen P et al. (2022) Genomic analyses show extremely perilous conservation status of African and Asiatic cheetahs (*Acinonyx jubatus*). Mol Ecol 31:4208-4223.
- Quéméré E, Amelot X, Pierson J, Crouau-roy B and Chikhi L (2012) Genetic data suggest a natural prehuman origin of open habitats in northern Madagascar and question the deforestation narrative in this region. Proc Natl Acad Sci U S A 109:13028-13033.
- Schwab P, Debes PV, Witt T, Hartl GB, Hmwe SS, Zachos FE and Grobler JP (2011) Genetic structure of the common impala (*Aepyceros melampus melampus*) in South Africa: Phylogeography and implications for conservation. J Zoolog Syst Evol Res 50:76-84.
- Sharma S, Dutta T, Maldonado JE, Wood TC, Panwar HS and Seidensticker J (2013) Forest corridors maintain historical gene flow in a tiger metapopulation in the highlands of central India. Proc R Soc B 280:20131506.
- Sousa VC, Fritz M, Beaumont MA and Chicki L (2009) Approximate Bayesian computation without summary statistics: The case of admixture. Genetics 1519:1507-1519.
- Storz JF and Beaumont MA (2002) Testing for genetic evidence of population expansion and contraction: An empirical analysis of microsatellite DNA variation using a hierarchical Bayesian model. Evolution 56:154-166.
- Stokes S, Thomas DSG and Washington R (1997) Multiple episodes of aridity in southern Africa since the last interglacial period. Nature 388:2-6.
- Tallmon DA, Gregovich D, Waples RS, Baker CS, Taylor BL, Archer E, Martien KK, Allendorf FW and Schwartz MK (2010) When are genetic methods useful for estimating contemporary abundance and detecting population trends? Mol Ecol Notes 10:684-692.
- Walker EH, Verschueren S, Schmidt-Küntzel A and Marker L (2022) Recommendations for the rehabilitation and release of wildborn, captive-raised cheetah: The importance of pre- and postrelease management for optimising survival. Oryx 56:495-504.
- Weber JL and Wong C (1993) Mutation of human short tandem repeats. Hum Mol Genet. 2:1123-1128.
- Weise FJ, Vijay V, Jacobson AP, Schoonover RF, Groom RJ, Horgan J, Keeping D, Klein R, Marnewick K, Maude G *et al.* (2017) The distribution and numbers of cheetah (*Acinonyx jubatus*) in southern Africa. PeerJ 5:e4096.

Internet Resources

Rambaut A and Drummond AJ (2007) Tracer v 1.7.2, https://github.com/beast-dev/tracer/releases/tag/v1.7.2 (accessed 30 May 2024).

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