



SHORT REPORT

A MYLK variant regulates asthmatic inflammation via alterations in mRNA secondary structure

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Myosin light-chain kinase (*MYLK*) is a gene known to be significantly associated with severe asthma in African Americans. Here we further examine the molecular function of a single-nucleotide polymorphism (SNP), located in the non-muscle myosin light-chain kinase isoform (nmMLCK), in asthma susceptibility and pathobiology. We identified nmMLCK variant (reference SNP: rs9840993, NM_053025: 721C>T, c.439C>T) with a distinct mRNA secondary structure from the other variants. The nmMLCK variant (721C) secondary structure exhibits increased stability with an elongated half-life in the human endothelial cell, and greater efficiency in protein translation initiation owing to an increased accessibility to translation start site. Finally, nmMLCK expression of 721C- and 721T-containing *MYLK* transgenes were compared in nmMLCK^{-/-} mice and confirmed deleterious effects of nmMLCK expression on asthmatic indices and implicated the augmented influence of *MYLK* 721C>T (c.439C>T) SNP on asthma severity. The confirmation of the novel mechanism of the regulation of asthmatic inflammation by a *MYLK* advances knowledge of the genetic basis for asthma disparities, and further suggests the potential of nmMLCK as a therapeutic target. Our study suggests that in addition to altering protein structure and function, non-synonymous SNPs may also lead to phenotypic disparity by altering protein expression. *European Journal of Human Genetics* (2015) 23, 874–876; doi:10.1038/ejhg.2014.201; published online 1 October 2014

INTRODUCTION

Within a rising US and world-wide prevalence of asthma, there is strong evidence for ethnic disparities in asthma susceptibility and severity with greater mortality, more severe obstruction and greater number of severe attacks occurring in asthmatics of African descent (AD).2 We recently explored MYLK, a gene encoding the Ca2+/calmodulin-dependent myosin light-chain kinase (MLCK), as a candidate gene for asthma susceptibility³ taking advantage of extensive gene resequencing in samples from Europeans and African Americans.4 We identified the significant association of an African-specific nonsynonymous variant in MYLK (reference single-nucleotide polymorphism (SNP): rs9840993, NM_053025: 721C>T, p.Pro147Ser or c.439C>T) with severe asthma.³ The human MYLK gene encodes three isoforms including non-muscle MLCK isoform (nmMLCK), smooth muscle isoform (smMLCK) and telokin (KRP), a small myosin filament-binding protein. Both smMLCK and nmMLCK phosphorylate myosin light chains to regulate cellular contraction and relaxation.⁵ smMLCK has been well studied in the pathogenesis of asthma as a key contributor to airway smooth muscle contractile function remodeling, characteristic of the asthmatic phenotype.⁶ In contrast, limited information is known about the role of the nmMLCK isoform in asthma pathobiology. The identified asthma susceptibility MYLK variant (NM 053025: 721C>T, p.Pro147Ser or c.439C>T) within the unique N terminus of nmMLCK, residing at significant distance from the smMLCK start site at 922aa.3 Consistent with a potential role for nmMLCK in asthma pathobiology, our structure/function studies in non-muscle tissues, such as gastrointestinal epithelium and lung vascular endothelium, have underscored a key role for nmMLCK in inflammatory responses wherein nmMLCK regulates vascular integrity (via interplay of cell contractile forces and cell-cell/ cell-matrix contacts) and leukocyte influx into lung tissues.^{7,8} We

recently reported that protein levels of nmMLCK correlate with experimental asthma susceptibility and severity. In this report, we analyzed the functionality of the SNP 721C>T (c.439C>T), which is strongly associated with severe asthma in African Americans, and identified a novel aspect of mRNA secondary structure.

MATERIALS AND METHODS

We calculated RNA folding energies (ΔG) using the RNAfold^{10,11} and mfold program.¹² The recombinant nmMLCK1 constructs (721T or 721C) were transfected into human endothelial cells for mRNA decay and protein expression analysis. In addition, these plasmids were used to transfect the mouse lung in a model of OVA-induced experimental asthma. Asthmatic inflammatory parameters were analyzed as described previously.⁹ Detailed methods are in online methods session.

RESULTS AND DISCUSSION

Within the four identified coding SNPs in nmMLCK in our previous study (Supplementary Table 1), 3 rs9840993 (NM_053025: 721C>T, c.439C>T) is the only SNP significantly associated with severe asthma (the corresponding variant information has been deposited in the LOVD 3.0 database: http://databases.lovd.nl/shared/variants/0000040191). This is also an AD-specific SNP³ with very low minor allelic frequencies in European decent individuals. To analyze the global and local mRNA stability of each variant, we calculated RNA folding energies (ΔG) using both the RNAfold program in the Vienna package 10,11 and mfold program 12 based on NCBI reference sequence NM_053025 (ancestral allele of each SNP was used to define the wild-type (WT) mRNA sequence). In silico prediction by RNAfold demonstrated that compared with other MYLK variants, the 721T variant greatly changes the global mRNA secondary structure of minimum free energy (MFE; Figure 1a and Supplementary Figure S1).

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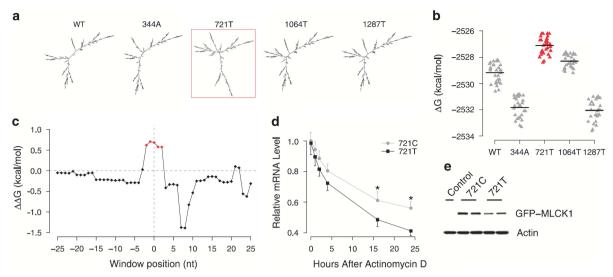


Figure 1 mRNA secondary structure affects MYLK gene translation efficiency. (a) mRNA secondary structures of MFE for wild-type (WT) MLCK gene and its variants (344A, 721T, 1064T and 1287T). (b) Free energy of the top 30 optimal/suboptimal mRNA secondary structures for each MYLK variants. The solid line depicts the mean of 30 values. (c) Landscape of the local accessibility gap ($\Delta\Delta G$) between the secondary structures of 721T and 721C around the start codon. '0' on x axis indicates the position of the start codon. The local accessibility was calculated using a sliding window of three nucleotides in length and one nucleotide in step. The windows with $\Delta\Delta G > 0.5$ kcal/mol were highlighted in red. (d) Relative mRNA decay curve of MLCK1-GFP (721T or 721C) in HLMVECs after actinomycin D exposure (5 µg/ml). *P<0.05. (e) Expression levels of MLCK1-GFP (721T or 721C) protein in human lung ECs 48 h after transfection.

The MFE of 721T is higher than all other variants (WT, 344A, 1064T and 1287T; Supplementary Figure S2). As expected, a similar trend was obtained when the *mfold* program is applied. We used *mfold* to predict the top 30 optimal and suboptimal foldings of each variant. The free energy of the optimal and suboptimal foldings of 721T is significantly higher than that of all the other variants (t-test: $P < 10^{-10}$) (Figure 1b and Supplementary Figure S3). In comparison with MFE structure, RNA centroid structure of the ensemble was reported to produce 30% fewer prediction errors. Here we also computed the centroid structures of WT and 721T by RNAfold13 and found that the folding energy of the centroid structure of 721T is higher than that of WT. The free energy gap $(\Delta \Delta G)$ between 721T and WT is >4 kcal/mol. These results suggest that the global structure of 721T variant is the most unstable compared with other MYLK variants and thus is more likely to be degraded. 14-16

Increasing evidences suggest that reduced mRNA local stability near the translation initiation region may lead to increased translation efficiency. 17-19 Therefore, we further analyzed the mRNA local accessibility (free energy required to open local structure) around the translation start codon using a sliding window of three nucleotides in length and one nucleotide in step. Interestingly, we found that there are five clustered windows with lower accessibility in 721T comparing with 721C (Figure 1c). The $\Delta\Delta G$ between 721T and 721C is >0.5 kcal/mol for each of the five continuous windows, which can be rarely observed along the nmMLCK mRNA (Supplementary Figure S4). We also checked the probability of observing five windows with the sum of $\Delta\Delta G$ larger than that of the five clustered windows around start codon by randomly selecting the nucleotides within nmMLCK mRNA. Among the 10 000 time randomization, we failed to observe such a pattern (Supplementary Figure S5), indicating that the clustered windows with decreased accessibility around start codon in 721T do not reflect random chance but rather suggest that the translation efficiency of 721C is potentially higher than that of 721T.

To further validate our findings, we assayed and compared mRNA stability and translational efficiency of variants of 721C and 721T. EGFP-labeled nmMLCK constructs with 721C or 721T were ectopically transfected and expressed in human endothelial cells to compare the rate of mRNA decay. Twenty four hours after the transfection, transcription was inhibited with actinomycin D (5 µg/ml), and the levels of nmMLCK-GFP transcripts at 0-24 h were determined by realtime PCR. Actinomycin D significantly reduced mRNA levels by both constructs, while the relative mRNA level by 721C variant is significantly higher than that of 721T (Figure 1d). These observations indicate that 721C variant mediates an increased mRNA stability and reduced intracellular decay. Protein levels of nmMLCK-GFP were determined in these ECs without actinomycin D treatment (Figure 1e). Our results demonstrated that mutating nmMLCK at position 721 from T to C recapitulates higher expression levels of protein, indicating that protein translation is more efficient with the variant 721C.

To verify the functional effects of 721C variant (147Pro) in contributing to asthmatic susceptibility and severity, we reversely expressed human nmMLCK1 in the pulmonary endothelium of nmMLCK^{-/-} mice utilizing the ACE antibody-tagged liposome delivery system used to deliver MLCK expressing plasmids. This gene delivery system is consistently targeting pulmonary endothelium with to overexpress nmMLCK1 in mouse lung endothelium (Supplementary Figure S6). nmMLCK1 overexpression in mouse lung tissues with the nmMLCK variant 721C (147Pro) exhibited higher expression efficiency than that of the 721T variant (147Ser or WT; Figure 2a and b), consistent with the findings in computation analysis and endothelial cell in vitro assays. As expected, nmMLCK1 overexpression augmented asthmatic inflammatory parameters including airway hyper-responsiveness (Figure 2c), with the disease-associated 721C variant eliciting stronger allergic inflammation than the MYLK variant 721T (Figure 2c). Similar effects were observed in other inflammatory parameters such as airway inflammatory leukocyte infiltration (Figure 2d-f) and BAL protein leakage (Figure 2g). These data are consistent with findings of upregulation of nmMLCK transcripts in both human or murine asthmatic subjects (Supplementary Figure S7). These studies represent the first characterization of a biological function of a MYLK SNP in vivo.

In summary, our studies underscore the contributory role of MYLK genetic variants to asthma susceptibility in populations of African



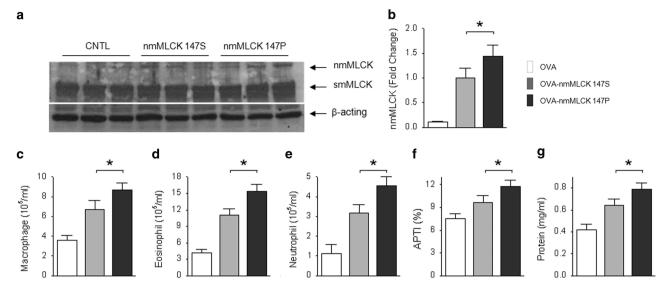


Figure 2 Effect of overexpressed human nmMLCK1 variants (147P or 147S) on asthmatic inflammation in a murine model. OVA-challenged nmMLCK-/-mice received nmMLCK1 express transgene to overexpress nmMLCK1 (147P or 147S). (a) Representative nmMLCK1 overexpression in mouse lung tissues after transfection. (b) Quantified relative levels of expressed nmMLCK variants. (c-e) Effect of nmMLCK1 variant overexpression on inflammatory leukocyte infiltration (macrophage, eosinophil and neutrophil). (f) Effects of nmMLCK1 variant overexpression on airway hyper-reactivity reflected by acetylcholine-induced APTI. (g) Effect of nmMLCK1 variant overexpression on BAL protein levels. *P<0.05 between groups of nmMLCK1 147P and 147S. n=5-6.

ancestry, and highlight the role of nmMLCK expression, especially in lung endothelium, to asthma pathogenesis. Although the link between MYLK variants, expression of specific MLCK isoforms and asthma appears to be certain, additional studies are required to establish a mechanistic relationship between genetic variants and functional alterations of this interesting gene product. Moreover, although MYLK variants may account for severe symptoms in African American asthmatics, further association studies, analyzing additional nearby SNPs and independent samples are required.

CONFLICT OF INTEREST

The authors declare no conflict of interest.

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- 1 Masoli M, Fabian D, Holt S, Beasley R: The global burden of asthma: executive summary of the GINA Dissemination Committee report. Allergy 2004; 59: 469-478.
- 2 El-Ekiaby A, Brianas L, Skowronski ME et al. Impact of race on the severity of acute episodes of asthma and adrenergic responsiveness. Am J Respir Crit Care Med 2006; 174: 508–513.
- 3 Flores C, Ma SF, Maresso K, Ober C, Garcia JG: A variant of the myosin light chain kinase gene is associated with severe asthma in African Americans. *Genet Epidemiol* 2007; 31: 296–305.
- 4 Gao L, Grant A, Halder I et al: Novel polymorphisms in the myosin light chain kinase gene confer risk for acute lung injury. Am J Respir Cell Mol Biol 2006; 34: 487–495
- 5 Dudek SM, Garcia JG: Cytoskeletal regulation of pulmonary vascular permeability. J Appl Physiol 2001; 91: 1487–1500.
- 6 Benayoun L, Druilhe A, Dombret MC, Aubier M, Pretolani M: Airway structural alterations selectively associated with severe asthma. Am J Respir Crit Care Med 2003; 167: 1360–1368.

- 7 Clayburgh DR, Rosen S, Witkowski ED et al: A differentiation-dependent splice variant of myosin light chain kinase, MLCK1, regulates epithelial tight junction permeability. J Biol Chem 2004; 279: 55506–55513.
- 8 Garcia JG, Verin AD, Herenyiova M, English D: Adherent neutrophils activate endothelial myosin light chain kinase: role in transendothelial migration. *J Appl Physiol* 1998; **84**: 1817–1821.
- 9 Wang T, Moreno-Vinasco L, Ma SF et al: Nonmuscle myosin light chain kinase regulates murine asthmatic inflammation. Am J Respir Cell Mol Biol 2014; 50: 1129–1135.
- 10 Hofacker IL, Stadler PF: Memory efficient folding algorithms for circular RNA secondary structures. *Bioinformatics* 2006; 22: 1172–1176.
- 11 Hofacker IL, Fontana W, Stadler PF, Bonhoeffer LS, Tacker M, Schuster P: Fast folding and comparison of RNA secondary structures. *Monatshefte für Chemie* 1994; 125: 167–188.
- 12 Zuker M, David H, Mathews DH, Turner DH. Algorithms and Thermodynamics for RNA Secondary Structure Prediction: a Practical Guide. RNA Biochemistry and Biotechnology. Dordrecht, NL, USA: Kluwer Academic Publishers, 1999, pp 11–43.
- 13 Ding Y, Chan CY, Lawrence CE: RNA secondary structure prediction by centroids in a Boltzmann weighted ensemble. RNA 2005; 11: 1157–1166.
- 14 Nackley AG, Shabalina SA, Tchivileva IE et al: Human catechol-O-methyltransferase haplotypes modulate protein expression by altering mRNA secondary structure. Science 2006; 314: 1930–1933.
- 15 Chamary JV, Hurst LD: Evidence for selection on synonymous mutations affecting stability of mRNA secondary structure in mammals. Genome Biol 2005; 6: R75.
- 16 Qiu L, Moreira A, Kaplan G et al: Degradation of hammerhead ribozymes by human ribonucleases. Mol Gen Genet 1998; 258: 352–362.
- 17 Gingold H, Pilpel Y: Determinants of translation efficiency and accuracy. Mol Syst Biol 2011; 7: 481.
- 18 Kudla G, Murray AW, Tollervey D, Plotkin JB: Coding-sequence determinants of gene expression in Escherichia coli. Science 2009; 324: 255–258.
- 19 Gu W, Zhou T, Wilke CO: A universal trend of reduced mRNA stability near the translation-initiation site in prokaryotes and eukaryotes. *PLoS Comput Biol* 2010; 6: e1000664.

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