ORIGINAL RESEARCH

Food Science & Nutrition

WILEY

Monitoring of polycyclic aromatic hydrocarbons and probabilistic health risk assessment in yogurt and butter in Iran

Amin Kiani¹ | Mahsa Ahmadloo² | Mojtaba Moazzen³ | Nabi Shariatifar³ | | Saeed Shahsavari⁴ | Majid Arabameri⁵ | Mohammad Mahdi Hasani⁶ | Ali Azari⁷ | Mosaad A. Abdel-Wahhab⁸ | |

¹Department of Public Health, School of Public Health, Fasa University of Medical Sciences, Fasa, Iran

²Department of Food Safety and Hygiene, School of Public Health, Qazvin University of Medical Sciences, Qazvin, Iran

³Department of Environmental Health Engineering, School of Public Health, Tehran University of Medical Sciences, Tehran, Iran

⁴Health Products Safety Research Center, Qazvin University of Medical Sciences, Qazvin, Iran

⁵Food Safety Research Center (salt), Semnan University of Medical Sciences, Semnan, Iran

⁶Department of Environmental Health Engineering, Faculty of Health, Tehran Medical Sciences, Islamic Azad University, Tehran, Iran

⁷Department of Environmental Health Engineering, Faculty of Health, Kashan University of Medical Sciences, Kashan, Iran

⁸Food Toxicology & Contaminants Department, National Research Center, Dokki, Egypt

Correspondence

Nabi Shariatifar, Department of Environmental Health Engineering, School of Public Health, Tehran University of Medical Sciences, Tehran, Iran. Email: nshariati@tums.ac.ir

Mosaad A. Abdel-Wahhab, Food Toxicology & Contaminants Dept., National Research Center, Dokki, Cairo, Egypt. Email: mosaad_abdelwahhab@yahoo.com

Funding information

Fasa University of Medical Sciences, Grant/ Award Number: 96253

Abstract

This study was conducted to determine the polycyclic aromatic hydrocarbons (PAHs) levels and health risk of yogurt and butter samples collected from Tehran using MSPE/ GC-MS (magnetic solid-phase extraction/gas chromatography-mass spectrometry). The results revealed that the limit of detection (LOD) and limit of quantification (LOQ) were ranged from 0.040 to 0.060 and 0.121 to 0.181 μ g/kg, respectively; with recoveries ranged from 86.1% to 100.3%. The highest mean of total PAHs was higher in butter (6.87 \pm 1.21 µg/kg) than in yogurt (3.82 \pm 0.54 µg/kg). The level of benzo (a)pyrene in all samples was lower than of standard levels of the European Union (EU). The highest value of all PAHs in samples was recorded in the winter season and also in the expiration date. The percentile 95% of the total hazard quotient (THQ) due to the consumption of yogurt and butter recorded 1.33E-02 and 3.69E-04 in adults and 6.12E-02 and 1.75E-03 in children, respectively. The percentile of 95% incremental lifetime of cancer risk (ILCR) due to the ingestion of yogurt and butter recorded 1.17E-06 and 2.02E-08 for adults and 5.51E-06 and 9.46E-08 for children, respectively. The rank order of 7 PAHs in adult and children based on P95% Hazard Quotient (HQ) in all samples was benzo(a)anthracene (BaA) > pyrene (P) > fluorene (F) > fluoranthene (FI) > acenaphthylene (Ace) > anthracene (A) > naphthalene (NA). According to the Monte Carlo Simulation (MCS) method, health-risk assessment showed that children and adults are not at significant health risk.

KEYWORDS

butter, gas chromatography-mass spectrometry (GC/MS), health risk assessment, occurrence, polycyclic aromatic hydrocarbons, yogurt

This is an open access article under the terms of the Creative Commons Attribution License, which permits use, distribution and reproduction in any medium, provided the original work is properly cited.

 $\ensuremath{\mathbb{C}}$ 2021 The Authors. Food Science & Nutrition published by Wiley Periodicals LLC

1 | INTRODUCTION

Nowadays, several dangerous and carcinogenic compounds are detected in food and environment, one of which is PAH which are found in air, water, and food (Alharbi et al., 2018; Ali & Aboul-Enein, 2005; Ali et al., 2003, 2005, 2011, 2018; Basheer, 2018). In some industrial areas, the high levels of PAH in unprocessed foods suggest a contamination due to the transport of contaminated particles and natural emissions from forest and volcanic fires or the burning of oil reservoirs (Orecchio et al., 2009; Rey-Salgueiro et al., 2009). There is a positive correlation between the exposure to PAHs and the occurrence of several diseases such as stomach cancer, alteration in the lung including DNA adduct, biochemical and cytogenetic alterations (Rawash et al., 2018; Santonicola et al., 2017).

The United States Environmental Protection Agency (USEPA) has involved 16 of these persistent compounds in its list of persistent organic pollutants (Orecchio et al., 2009; Suchanová et al., 2008). The International Agency for Research on Cancer (LARC) classified BaP as category 2A (probably carcinogenic) to humans (N Aguinaga et al., 2008). Therefore, the EU established a limit of less than 1 μ g/kg of BaP for the processed cereal-based foods prepared for young children, infants, and baby foods (EC 2005).

People can be exposed to PAHs through three main routes including respiratory, skin, and foods (Chung et al., 2010; Cirillo et al., 2004; Shariatifar et al., 2020), and they were reported to be accumulated in various types of food matrices including dairy products (milk, yogurt, butter), etc. (Chung et al., 2010; Cirillo et al., 2004; Shariatifar et al., 2020). These PAHs are persistent in various food products during cooking procedures, thermal processes (such as baking, smoking, grilling, frying), processing, and packaging (Farhadian et al., 2010; Gorji et al., 2016).

Milk and milk products are essential for infants, children and adults, due to their high content of macro and micronutrients (Han et al., 2014; Iwegbue et al., 2014), and yogurt and butter are playing an important role in the human diet (Lee et al., 2015; Martorell et al., 2010). Therefore, milk and dairy products may contain PAHs due to the lipophilic property of PAHs even after exposure to various heat treatments (Lin et al., 2016).

Several methods were developed for the determination of PAHs in milk and dairy product including Liquid–liquid extraction (LLE), Soxhlet apparatus, Solid-phase extraction (SPE), and Pressurized liquid extraction (PLE). The most widely used methods for the extraction of PAHs from an intricutionate sol matrix includes silica cartridges, LLE after saponification with caustic soda, straight immersion solid-phase microextraction (SPME), columns with gel beads, and 2 phase hollow liquid phase microextraction (HF-LPME) (Basheer & Ali, 2018; Moazzen, Khaneghah, et al., 2018; Moazzen, Mahvi, et al., 2018). However, due to the fat content and other materials, alcoholic or caustic solutions are used for the saponification to avoid difficulties from lipids followed by the use of SPE or LLP for the extraction (Moazzen et al., 2013; Naccari et al., 2011). Moreover, several methods were established for the determination of PAHs in milk and dairy products including gas chromatography with flame ionization detection (GC/FID) or gas chromatography with mass detection (GC/MS), high-performance liquid chromatography (HPLC) equipped with ultraviolet and fluorescence detection (UV/FL) and fluorescence detector (FLD) (Ali et al., 2019; Grova et al., 2002; Kamankesh et al., 2015).

For the toxicological evaluation of chemical mixes, toxic equivalency (TEQ) is a concept to indicate the whole toxicity of a mixture, like PAHs. The TEQ was designed to estimate the toxicity and hazards of toxic chemicals related to conventional toxic compounds. However, the toxic equivalency factor (TEF) of each compound shows the level of toxicity compared to the alternative composition, which gives the reference value 1. This TEF method is a common technique to evaluate the risk of PAHs and it was used by USEPA and WHO (1998). The final toxic point and the toxicity grade are the main factors for the TEQ method and TEFs of PAHs, which were proposed based on the growth in the incidence of tumors reported in animal study; however, TEFs for each PAHs compounds were based on their cancer outcomes of various carcinogenic researches (Yoon et al., 2007).

Considering the relatively high consumption of milk products in the Iranian basket, severe air pollution and partly water pollution in a densely populated city like Tehran, and also lack of coherent research about PAHs in dairy product with risk assessment in Iran, it is important to evaluate the amount of PAHs in milk products. Therefore, the goals of the present study are the determination of PAHs in some milk products (butter and yogurt) with the MSPE-GC/ MS method and assessment of the potential health risk produced by PAHs intake using the cancer potency of BaP as a reference member.

2 | MATERIALS AND METHODS

2.1 | Sampling

In this study, 96 samples of milk products (48 yogurt and 48 butter samples) were purchased from local markets in different regions of Tehran, Iran in May, August, December 2018, and February 2019. Three different brand sources were purchased for each type of milk products. All samples were completely homogenized and then five grams of each sample were kept in airtight dishes at -20°C until analysis.

2.2 | Standards and reagents

Standards PAH mix, including 16 mentioned PAHs (benzo(a)pyrene [BaP], acenaphthylene [Ace], naphthalene [NA], fluorene [F], acenaphthene [Ac], phenanthrene [Pa], fluoranthene [Fl], anthracene [A], benzo(a)anthracene [BaA], pyrene [P], benzo(b) fluoranthene [BbF], chrysene [Ch], benzo(k)fluoranthene [BkF], dibenzo[a,h]anthracene [DhA], indeno[1,2,3-cd]pyrene [IP] and benzo[g,h,i]perylene [BgP]) was purchased from Supelco WII FY_Food Science & Nutrition __

(Bellefonte, PA, U. S.). The standard solutions were prepared in dichloromethane, with all mentioned PAHs concentration of 0.1 mg/ml. Stock standard solutions were mixed with methanol-dichloromethane (50:50, v/v) every week for the preparation of a working mixed solution (1 μ g/ml for each mentioned PAHs) which was used to measure the extraction performance under various conditions. The working and stock solutions were preserved at 4°C and biphenyl was used as an internal standard at a concentration of 0.05 μ g/ml in methanol. Multi-walled carbon nanotubes were purchased from Hanwha Nanotech (MWCNT CM-95, Korea) and the adsorbent of MWCNT-MNP was prepared as described previously (Rastkari & Ahmadkhaniha, 2013). All other chemicals and solvents were of analytical grade.

2.3 | Samples preparation

2.3.1 | Yogurt

Yogurt samples were prepared as described in our previous work (Moazzen et al., 2013). This method includes 3 major parts; (a) sample cleanup: Five grams of the sample was ground to fine particles with a pulverization then mixed with 1 ml internal standard and the mixture was homogenized for 15 min. The extraction solution containing 7.5 ml acetonitrile-methanol (70%: v/v) and 7.5 ml potassium hydroxide (1 M) was added to the homogenized mixture then was mixed and sonicated for 7 min in a bath of ultrasonic at 50°C. The mixture was centrifuged for 10 min at $8,944 \times g$, the fat was removed from the mixture in the lipid freezing filtration method (Ahmadkhaniha et al., 2009) and the pH was adjusted to 6.5 using HCl (1 M); (b) Analytes adsorption: after a rudimentary cleanup, the aqueous phase was transferred to another container, 500 mg NaCl and 10 mg of synthesized multi-walled carbon nanotubes magnetic nanoparticle (MWCNT-MNP) as adsorbent was added to the mixture and vortex-mixed vigorously for 5 min. The magnetic synthesized adsorbent was collected to the side of the vial by an exterior magnet; (c) Analytes desorption from the magnetic adsorbent: dichloromethane (5 ml) was added to the elute of PAHs compounds from the synthesized adsorbent, mixed with strong vortex-mixing for 3 min and the supernatant was discarded. The magnetic adsorbent was collected by an exterior magnet to the side of the vial. This phase was repeated for another one more time, afterward, the solvent was evaporated to dryness at 30°C using a gentle stream of pure nitrogen gas. Then the extract was re-dissolved in 50 μ l acetonitrile-methanol (50:50 v/v), vigorously shaken for 1 min with the vortex-mixer, and 1 µl was injected into the GC/MS device. The outcomes of the optimization experiments showed that the mentioned technique permitted for reproducible, quantitative extraction of polycyclic aromatic hydrocarbons (Moazzen et al., 2013). The blank samples include internal standard and QC (quality control) were examined at the beginning of the analysis, middle, and end of each sample. The mean values of results were used for quantification and all the samples in this research were examined in duplicate.

2.3.2 | Butter

Due to the high-fat content in butter, the samples were homogenized before extraction. Five grams of butter were subjected to saponification with methanolic Potassium hydroxide, cold extracted with cyclohexane-dimethylformamide, and cyclohexane for reextraction. The clean-up technique for the extracted sample was carried out by gel permeation chromatography with Bio-Beads SX-3 column followed by adsorption chromatography on silica gel. The final phase was the reduction of the cleaned extracts to a suitable analytical volume using acetonitrile as a solvent (Falco et al., 2003).

2.4 | Analytical conditions and instrumentals

The analysis was conducted on a GC device of the Agilent 6890 with an MS detector 5973 quadrupoles (Agilent Technologies). The GC was adjusted with a capillary column of DB-5 ms (30 m, 0.25 mm i.d., $0.25 \,\mu m$ film thickness) and the mode of splitless was for the inlet. The temperatures were adjusted as follows: 290°C for injector; the initial temperature of the oven was 70°C, kept for one min and increased to 295°C at a rate of 10°C/min then kept for seven min. The transfer line temperature was kept at 300°C and the constant flow of helium as a carrier gas was 1 ml/min. The quadrupole temperature was kept at 150°C and source temperatures were retained at 230°C. The energy of the electronic beam of the MS was adjusted at 70 eV. The identification was carried out by comparing the obtained mass spectra and times of retention to reference spectra and times of retention that were obtained by injection of the calibration standards in the same conditions of GC/MS. The spike calibration standard approach to overcome the problems caused by the matrix was used. In this approach, calibration standards are ready by the addition of standards solution to samples that are subjected to the same sample preparation procedure which is intended to be used for unknown samples. The PAHs analytes were quantified using SIM mode and each sample was injected in duplicate.

2.5 | Health risk assessment

To evaluate the carcinogenic risk of PAHs, the toxicity equivalency factors (TEFs) of benzo(a)pyrene (TEQ BaP) were used. The estimation of TEQ BaP was carried out using a TEFs (Saito et al., 2014). The TEF fixed as one for BaP and dibenzo[a,h]anthracene (DhA); 0.001 for each of naphthalene (NA), acenaphthylene (Ace), acenaphthene (Ac), fluorine (F), Phenanthrene (Pa), fluoranthene (F) and pyrene (P); 0.01 for each of anthracene (A), chrysene (Ch) and benzo[g,h,i]perylene (BgP) and 0.1 for each of benzo(a)anthracene (BaA), benzo(b) fluoranthene (BbF), benzo(k)fluoranthene (BkF), indeno [1,2,3-cd] pyrene (IP), benzo(a)pyrene (BaP) and dibenzo[a,h]anthracene (DhA). TEQ BaP for dietetic contact to PAHs in milk products was computed matched to the following equation (EPA (2010)

$$TEQ_{BaP} = \sum_{i=1}^{n} C_i \times TEF_i$$
(1)

Where C is a quantitative analysis of PAHs and the TEFi is the toxicity equivalency factor of biotype (i) in milk products.

To estimate the hazard of noncarcinogenic of PAH compounds in milk products, the average daily dose (ADD) was applied and calculated by the following equation:

$$ADD = \frac{C \times IR}{BW}$$
(2)

Due to the carcinogenic hazard, the chronic daily intake (CDI) was calculated according to the following equation:

$$CDI = \frac{C \times IR \times ED \times EF}{BW \times AT}$$
(3)

This study assumed that milk items consumed 365 d/year, C is the PAHs levels in milk (mg/kg); IRi as the daily intake of the milk products was set as 73 g/day and 2 g/day for yogurt and butter, respectively (Abdollahi et al., 2014); ED is the time-period of PAHs exposure for adults and children (70 and 6 years old, respectively) as suggested by (Karami et al., 2020). EF is the frequency exposure of PAHs (350 days/years) (Jahanbakhsh et al., 2019); the average body weight (BW) is 15 kg for children and 70 kg for an adult was used in the calculation of PAHs risk assessment (Samiee et al., 2020) and AT is the average time for an adult is 25,550 days and for the children 2,190 days.

The hazard of noncarcinogenic was calculated based on Equation 4:

$$HQ = \frac{ADD}{RfD}$$
(4)

Where, HQ is the hazard quotient; ADD is the average daily dose (mg/kg/d); RfD is the oral reference dose (mg/kg/d) as suggested by (USEPA, 2017) is NA (0.02), AcP (0.06), FL (0.04), AN (0.3), FLUR (0.04), PY (0.03) and BaA (0.0003). If THQ > 1 value, the exposed population is considered at health risk, but if THQ \leq 1, the health risk is not likely (Ghasemidehkordi et al., 2018). In considering the effects of PAHs on the risk of noncarcinogenic, the total hazard quotient (THQ) was determined by the Equation below:

$$THQ = \sum HQ_1 \cdots HQ_n$$
(5)

However, the ILCR of BaP of the milk products was determined according to the following Equation (Fakhri et al., 2018).

$$ILCR = CDI_{BaP} \times CSF_{BaP}$$
(6)

Where: ILCR is incremental lifetime cancer risk and CDI is the chronic daily intake of BaP (mg/kg/d); CSF is the cancer slope factor of BaP which is equal to 7.3 mg/kg/d (Huang et al., 2016; Samiee et al., 2020; USEPA, 2016). According to USEPA, cancer is negligible when ILCR < 10-6, risk, but if ILCR > 10-4, the risk of cancer is unacceptable and finally, when ILCR is between 10-6 to 10-4, the risk of cancer is tolerable for consumers (Huang et al., 2016; USEPA, 2016).

The incremental lifetime carcinogenic risk (ILCR) was evaluated and noncarcinogenic (HQ) of the milk product content of PAHs was navigated by MCS (Crystal Ball v 11.1.2.4.600 software, Oracle, Decisioneering, Denver, CO, USA). To increase the accuracy of health-risk assessment by considering uncertainties, USEPA presented the MCS method (EPA, 2016). The simulation executed by the presented factors, and the MCS model was shown for 10,000 repetitions. Eventually, the mean and 95th percentile of the ILCR and HQ distribution was selected to assess if the exposed society is at risk or not. For estimation of carcinogenic risk of PAHs in milk products, actual ILCR was calculated according to the following equation:

$$ILCR_{(act)} = P95\% ILCR_{Yo} + P95\% ILCR_{Bu}$$
(7)

Where: ILCR (act) is actual ILCR; ILCR $_{\rm yo}$ is ILCR for yogurt and ILCR $_{\rm bu}$ and is ILCR for butter.

For estimation of actual noncarcinogenic risk of PAHs in milk products, actual THQ was calculated based on the following equation:

$$THQ_{(act)} = P95\% THQ_{Yo} + P95\% THQ_{Bu}$$
 (8)

Where THQ (act) is actual THQ; HQbu, is the HQ for butter and HQyo, is the HQ for yogurt.

2.6 | Statistical examination

The results were presented as mean \pm SD and the statistical significance was carried out by SPSS (version 24.0) software. Data for PAHs levels in milk products (yogurt and butter) were evaluated for normality (Kolmogorov-Smirnov test). For normally distributed data, comparisons among yogurt and butter were assessed using an independent *t*-test also if non-normally distributed data, comparisons among yogurt and butter were assessed using Mann-Whitney as shown in Table 1. Values were considered statistically significant when *p* <.05. In cases, when PAH analytes were ND (not detected), 1/2 of the LOD was used to calculate the mean concentration.

3 | RESULTS AND DISCUSSIONS

3.1 | Performance evaluation of the analytical method

The extracted PAHs were then determined using a GC-MS technique. Toward the recognition goal, the full scan mass spectrum, 4 characteristic ions ratios, and the RTT of \pm 0.5% tolerance criteria were applied for the quantification purpose compared to the standard, and the most intense ions were used for each compound and these compounds were then quantified using SIM (selected

TABLE 1	Linear range ($\mu g/kg$), Limit of detection (LOD; $\mu g/kg$), limit of quantification (LOQ; $\mu g/kg$), Coefficient of estimation (r2),
repeatabilit	y relative standard deviation (RSDr; $n = 6$), and reproducibility relative standard deviation (RSDR; $n = 6$)

Target compound	Linear range (µg/kg)	Limit of detection (LOD) (µg/kg)	Limit of quantification (LOQ) (µg/kg)	Coefficient of estimation (r2)	Recoveries (%)	Repeatability (RSDr) (%)	Reproducibility (RSDR) (%)
NA	0.050-1.000	0.040	0.121	0.988	91.2	9	6, 10, 14
Ace	0.050-1.000	0.040	0.121	0.981	93.4	10.1	9, 12, 17
Ac	0.050-1.000	0.040	0.121	0.990	86.1	3.2	5, 10,12
F	0.050-1.000	0.040	0.121	0.992	96.1	6.4	14, 8, 16
Pa	0.050-1.000	0.050	0.150	0.991	99.7	7	13, 11, 8
А	0.050-1.000	0.040	0.121	0.989	89.9	10	7, 12, 10
FI	0.050-1.000	0.040	0.121	0.983	100.1	9	10, 13, 7
Р	0.050-1.000	0.055	0.165	0.986	93.5	9.1	11, 9, 18
BaA	0.050-1.000	0.040	0.121	0.984	94.7	5	6, 7, 12
Ch	0.050-1.000	0.040	0.121	0.991	98.1	4.5	9, 13, 11
BbF	0.050-1.000	0.060	0.181	0.992	99.9	8	12, 7, 10
BkF	0.050-1.000	0.040	0.121	0.986	100.3	9	14, 11, 8
BaP	0.050-1.000	0.040	0.121	0.988	94.5	8.6	7, 15, 11
IP	0.050-1.000	0.040	0.121	0.989	89.9	6	5, 9, 12
DhA	0.050-1.000	0.040	0.121	0.992	97.1	7	14, 6, 10
BgP	0.050-1.000	0.040	0.121	0.991	97.8	9.2	7, 5, 12

Note: RSDR of 1 μ g/Kg, 5 μ g/Kg, and 10 μ g/Kg standard value (n = 6).

TABLE 2 Mean \pm SD of PAH factors in yogurt and butter in Iran (μ g/kg)

	Type of	f dairy produc	ts						
	Yogurt				Butter	Butter			
	N	Min	Max	Mean ± SD	N	Min	Max	Mean \pm SD	
NA	48	0.02	0.02	0.02 ± 0.00	48	0.02	0.02	0.02 ± 0.00	_
Ace	48	0.30	1.17	0.80 ± 0.27	48	1.10	2.30	1.62 ± 0.32	p <.001
Ac	48	0.02	0.02	0.02 ± 0.00	48	0.02	0.02	0.02 ± 0.00	-
F	48	0.02	0.02	0.02 ± 0.00	48	0.02	0.02	0.02 ± 0.00	-
Pa	48	1.06	1.42	1.21 ± 0.12	48	1.40	2.80	2.01 ± 0.38	p <.001
А	48	0.02	0.02	0.02 ± 0.00	48	0.02	0.02	0.02 ± 0.00	-
FI	48	0.02	0.02	0.02 ± 0.00	48	0.02	0.02	0.02 ± 0.00	-
Р	48	1.28	1.64	1.39 ± 0.10	48	1.90	3.20	2.45 ± 0.38	p <.001
BaA	48	0.02	0.02	0.02 ± 0.00	48	0.02	0.02	0.02 ± 0.00	-
Ch	48	0.02	0.12	0.05 ± 0.03	48	0.02	0.12	0.05 ± 0.03	1.000
BbF	48	0.08	0.23	0.15 ± 0.05	48	0.37	0.75	0.51 ± 0.11	p <.001
BkF	48	0.02	0.02	0.02 ± 0.00	48	0.02	0.02	0.02 ± 0.00	-
BaP	48	0.02	0.02	0.02 ± 0.00	48	0.02	0.02	0.02 ± 0.00	.737
IP	48	0.02	0.02	0.02 ± 0.00	48	0.02	0.02	0.02 ± 0.00	_
DhA	48	0.02	0.02	0.02 ± 0.00	48	0.02	0.02	0.02 ± 0.00	-
BgP	48	0.02	0.06	0.03 ± 0.01	48	0.02	0.08	0.03 ± 0.02	.087
Total	48	3.01	4.86	3.82 ± 0.54	48	5.01	9.47	6.87 ± 1.21	p <.001
PAH4	48	0.14	0.41	0.23 ± 0.08	48	0.43	0.93	0.60 ± 0.15	p <.001

 TABLE 3
 Comparison of PAHs contamination levels in different study

-WILEY

Location of study	Analyte	Sample type	Amount	Detection method	References
France	8 PAHs	Fresh milk	Σ PAHs (26.7 \pm 10.8 ng/g)	GC-MS	(Grova et al., 2002)
Spain	16 PAHs	Infant formula	ND	GC-MS	(Aguinaga et al., 2007)
		Half-fat milk	ND		
		Skimmed milk	ND		
		Full fat milk	Σ FLUR and PY (2.16 \pm 0.43 $\mu g/L$)		
Spain	10 PAHs	Infant formula	ND	GC-MS	(Aguinaga et al., 2008)
		Soya milk	ND		
		Full-fat milk	ND		
		Skimmed milk	Σ AcPY, FL, PHEN, AN, FLUR and PY (449.4 \pm 31.51 ng/L)		
Italy	16 PAHs	Milk	ND	GC-MS	(Bianchi et al., 2008)
Taiwan	16 PAHs	Milk	NaP, AcP, Flu, Pa and in some samples Bap	GC-MS	(Chung et al., 2010)
Nigeria	16 PAHs	Infant formula:	ΣPAHs	GC-MS	(Iwegbue et al., 2014)
		0–6 months	0.1 to 1.98		
		6–12 months	0.05 to 1.98		
		1–3 years	0.02 to 2.54		
		0–12 months	0.51 to 0.7 μg/kg		
Egypt	15 PAHs		ΣPAHs	GC-MS	(Abou-Arab et al., 2014)
		Farm raw milk	1.01 µg/kg		
		Commercial raw milk	0.37 μg/kg		
		Pasteurized milk	0.2 μg/kg		
Mexico	16 PAHs	Milk	ΣPAHs	GC-FID	(Gutiérrez et al., 2015)
		2008	2.06 ± 2.83		
		2009	1.65 to 2.14		
		2010	$1.24 \pm 1.28 \ \mu g/g$		
Korea	8 PAHs		ΣPAHs	GC-MS	(Lee et al., 2015)
		Dairy products	1.52 μg/kg		
		Sea foods	1.06 μg/kg		
Romania	15 PAHs	Milk powders	Σ PAHs (0.47 ± 0.04 to 1.4 ± 0.17 µg/kg)	GC-MS	(Dobrinas et al., 2016)
China	6 PAHs	Milk	Σ PHE, ANT, FLA and PYR (6.0 ng/g)	GC-MS	(Lin et al., 2016)
Canada	15 PAHs		ΣPAHs	HPLC-FID	(Lawrence & Weber, 1984)
		Skim Milk	0-2.7 μg/kg		
		Infant formula	8.1 µg/kg		
Japan	12 PAHs		ΣPAHs	HPLC-FID	(Kishikawa et al., 2003)
		Commercial milk	$0.99\pm0.37\mu\text{g/kg}$		
		Infant formula	$2.01\pm0.30\mu\text{g/kg}$		
		Human milk	$0.75 \pm 0.47 \ \mu g/kg$		
Poland	16 PAHs	Milk powder	Σ BaA, Ch, BbF, BkF, BaP and BP(1.83 $\mu g/kg)$	HPLC-FID	(Wegrzyn et al., 2006)
Czech republic	15 PAHs	Industrial smoked cheese	Σ PAHs (0.11 μ g/kg)	HPLC-FID	(Suchanová et al., 2008)
Spain	11 PAHs	Infant food	BkF (0.1-0.3 μg/kg)	HPLC-FID	(Rey-Salgueiro et al., 2009)

TABLE 3 (Continued)

Location of study	Analyte	Sample type	Amount	Detection method	References
Poland	19 PAHs	Infant formula	Σ PAHs (0.28–7.45 $\mu g/kg)$	HPLC-FID/DAD	(Ciecierska & Obiedziński, 2010)
Calabria	16 PAHs		ΣPAHs	HPLC-FID	(Naccari et al., 2011)
		Raw milk	5.44 ng/g		
		Pasteurized milk	6.52 ng/g		
		Semi-skimmed milk	5.94 ng/g		
		Whole milk	7.75 ng/g		
Italy	16 PAHs		ΣPAHs	HPLC-UV	(Cirillo et al., 2004)
		Non-smoked Cheese	59.11-160.05 μg/kg		
		Smoked cheese	67.49-399.90 μg/kg		
Korea	7 PAHs		ΣPAHs	HPLC-FID	(Cho & Shin, 2012)
		Infant formula	0.43 µg/kg		
		Mixed milk powders	0.46 μg/kg		
Argentina, Brazil	15 PAHs	Milk powders	Σ PAHs (11.8-78.4 µg/kg)	HPLC-FID	(Garcia Londoño et al., 2013)
Korea	8 PAHs	Infant formula	Σ PAHs (0.09–0.18 $\mu g/kg)$	HPLC-FID	(Han et al., 2014)
Iran	15 PAHs	Kebab (grilled meat)	Σ PAHs (7.37–17.94 $\mu g/kg)$	GC-MS	(Gorji et al., 2016)
Iran	3 PAHs	Grilled meat	Σ PAHs (3.51-132 ng/g)	HPLC-FD	(Farhadian et al., 2010)
Sweden	1 PAHs	Swedish smoked meat	Σ BaP (6.6-36.9 μg/kg)	HRGC-MS	(Wretling et al., 2010)
Estonia	6 PAHs	smoked fish	Σ PAHs (12.37 $\mu g/kg)$	GC-MS	(Yurchenko & Mölder, 2005)
Spain	16 PAHs		ΣPAHs	HRGC- HRMS	(Martorell et al., 2010)
		Meat and meat products	38.99 μg/kg		
		Fish and shellfish	2.87 μg/kg		
		Vegetables	1.22 μg/kg		
		Fruits	0.81 µg/kg		
		Milk	0.47 μg/kg		
		Oils and fats	18.75 μg/kg		
Italy	22 PAHs	Coffee	Σ PAHs 0.52 to 1.8 μg/L	GC-MS	(Orecchio et al., 2009)

ion monitoring) mode. According to Moazzen et al. (2013), one quantitation and two qualifier ions were controlled for each compound. The optimum conditions for the analysis were used for the establishment of the calibration curves ($0.050-1.000 \mu g/kg$) considering the coefficient of correlation of 0.981-0.992. The LOQs and LODs of PAHs compounds were $0.121-0.181 \mu g/kg$ and $0.040-0.060 \mu g/kg$, respectively. The accuracy of the technique was assessed according to interday precision via the QC analysis for samples ready at four three repeated days. Additionally, the values of interday precision for all PAHs compounds were less than 8.7% and the recorded values were 3.2% to 10.1% for repeatability and 5%-18% for reproducibility with estimated recoveries of 86.1%-100.3% (Table 1). The feasibility and reliability of this method were also confirmed by the measurement PAHs in the milk, doogh, and milk powder (Kiani et al., 2019).

3.2 | Comparing the mean values of PAHs in the tested milk products

The mean values of PAHs compounds based on the type of dairy products are presented in Table 2 and showed that the highest levels of all PAHs compounds were found in the butter. The recorded means of total PAHs in yogurt and butter were $3.82 \pm 0.54 \mu g/kg$ and $6.87 \pm 1.21 \mu g/kg$, respectively (Table 2). The results also indicated that the higher the amount of fat content and the lower the amount of water content, the more contaminant of PAHs was observed. In this study, P recorded the maximum mean value in yogurt ($1.39 \pm 0.10 \mu g/kg$) and butter ($2.45 \pm 0.38 \mu g/kg$). Among the groups suggested in EC Regulation Number 835/ 2011 (2011), a maximum value of PAH4 of $1 \mu g/kg$ was reported only in milk powder which corresponding to the infant products. The mean levels of

TABLE 4 Mean \pm SD of PAH factors in yogurt and butter in Iran based on seasons of year (μ g/kg)

	Yogurt				Butter				
	Season				Season	Season			
	Spring	Summer	Autumn	Winter	Spring	Summer	Autumn	Winter	
Analyte	Mean ± SD								
NA	0.02 ± 0.00	0.02 ± 0.00	0.02 ± 0.00	0.02 ± 0.00	0.02 ± 0.00	0.02 ± 0.00	0.02 ± 0.00	0.02 ± 0.00	
Ace	0.52 ± 0.21	0.72 ± 0.17	0.91 ± 0.19	1.07 ± 0.08	1.33 ± 0.16	1.43 ± 0.19	1.80 ± 0.23	1.90 ± 0.28	
Ac	0.02 ± 0.00	0.02 ± 0.00	0.02 ± 0.00	0.02 ± 0.00	0.02 ± 0.00	0.02 ± 0.00	0.02 ± 0.00	0.02 ± 0.00	
F	0.02 ± 0.00	0.02 ± 0.00	0.02 ± 0.00						
Pa	1.10 ± 0.03	1.14 ± 0.05	1.28 ± 0.10	1.32 ± 0.09	1.60 ± 0.13	1.87 ± 0.10	2.17 ± 0.29	2.40 ± 0.32	
А	0.02 ± 0.00	0.02 ± 0.00	0.02 ± 0.00						
FI	0.02 ± 0.00	0.02 ± 0.00	0.02 ± 0.00	0.02 ± 0.00	0.02 ± 0.00	0.02 ± 0.00	0.02 ± 0.00	0.02 ± 0.00	
Р	1.32 ± 0.07	1.34 ± 0.04	1.42 ± 0.09	1.49 ± 0.10	2.03 ± 0.12	2.23 ± 0.12	2.68 ± 0.19	2.87 ± 0.26	
BaA	0.02 ± 0.00	0.02 ± 0.00	0.02 ± 0.00	0.02 ± 0.00	0.02 ± 0.00	0.02 ± 0.00	0.02 ± 0.00	0.02 ± 0.00	
Ch	0.02 ± 0.01	0.04 ± 0.02	0.06 ± 0.04	0.07 ± 0.04	0.02 ± 0.01	0.04 ± 0.02	0.06 ± 0.04	0.07 ± 0.04	
BbF	0.09 ± 0.01	0.12 ± 0.01	0.17 ± 0.03	0.20 ± 0.02	0.41 ± 0.04	0.45 ± 0.05	0.54 ± 0.09	0.63 ± 0.10	
BkF	0.02 ± 0.00	0.02 ± 0.00	0.02 ± 0.00	0.02 ± 0.00	0.02 ± 0.00	0.02 ± 0.00	0.02 ± 0.00	0.02 ± 0.00	
BaP	0.02 ± 0.00								
IP	0.02 ± 0.00								
DhA	0.02 ± 0.00								
BgP	0.02 ± 0.00	0.02 ± 0.00	0.03 ± 0.01	0.04 ± 0.01	0.02 ± 0.00	0.02 ± 0.00	0.04 ± 0.02	0.06 ± 0.02	
total	3.27 ± 0.22	3.57 ± 0.29	4.05 ± 0.45	4.40 ± 0.34	5.62 ± 0.41	6.24 ± 0.43	7.48 ± 0.82	8.13 ± 1.02	
PAH4	0.16 ± 0.01	0.20 ± 0.03	0.27 ± 0.07	0.32 ± 0.06	0.41 ± 0.12	0.45 ± 0.07	0.47 ± 0.09	0.55 ± 0.09	

TABLE 5Mean \pm SD of PAH factorsin yogurt and butter in Iran based onproduction date ($\mu g/kg$)

	Yogurt		Butter			
	Production date					
	Production date	Expire date	Production date	Expire date		
Analyte	Mean ± SD	Mean \pm SD	Mean ± SD	Mean \pm SD		
NA	0.02 ± 0.00	0.02 ± 0.00	0.02 ± 0.00	0.02 ± 0.00		
Ace	0.66 ± 0.26	0.95 ± 0.19	1.44 ± 0.22	1.79 ± 0.31		
Ac	0.02 ± 0.00	0.02 ± 0.00	0.02 ± 0.00	0.02 ± 0.00		
F	0.02 ± 0.00	0.02 ± 0.00	0.02 ± 0.00	0.02 ± 0.00		
Pa	1.15 ± 0.07	1.27 ± 0.12	1.87 ± 0.25	2.15 ± 0.44		
А	0.02 ± 0.00	0.02 ± 0.00	0.02 ± 0.00	0.02 ± 0.00		
FI	0.02 ± 0.00	0.02 ± 0.00	0.02 ± 0.00	0.02 ± 0.00		
Р	1.35 ± 0.06	1.44 ± 0.12	2.35 ± 0.30	2.56 ± 0.44		
BaA	0.02 ± 0.00	0.02 ± 0.00	0.02 ± 0.00	0.02 ± 0.00		
Ch	0.03 ± 0.01	0.07 ± 0.03	0.03 ± 0.01	0.07 ± 0.03		
BbF	0.13 ± 0.04	0.16 ± 0.05	0.45 ± 0.06	0.57 ± 0.12		
BkF	0.02 ± 0.00	0.02 ± 0.00	0.02 ± 0.00	0.02 ± 0.00		
BaP	0.02 ± 0.00	0.02 ± 0.00	0.02 ± 0.00	0.02 ± 0.00		
IP	0.02 ± 0.00	0.02 ± 0.00	0.02 ± 0.00	0.02 ± 0.00		
DhA	0.02 ± 0.00	0.02 ± 0.00	0.02 ± 0.00	0.02 ± 0.00		
BgP	0.02 ± 0.00	0.03 ± 0.01	0.02 ± 0.01	0.04 ± 0.02		
total	3.54 ± 0.40	4.11 ± 0.53	6.35 ± 0.82	7.38 ± 1.35		
PAH4	0.20 ± 0.05	0.27 ± 0.09	0.51 ± 0.07	0.68 ± 0.15		

WILEY

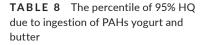
TABLE 6 Mean \pm SD of PAH factors in yogurt and butter in Iran based on brand type (μ g/kg)

Type of dairy products							
	Yogurt			Butter			
	Brand type			Brand type			
	A	В	с	A	В	с	
Analyte	Mean ± SD						
NA	0.02 ± 0.00						
Ace	0.80 ± 0.28	0.85 ± 0.26	0.76 ± 0.29	1.63 ± 0.33	1.70 ± 0.33	1.53 ± 0.32	
Ac	0.02 ± 0.00	0.02 ± 0.00	0.02 ± 0.00	0.02 ± 0.00	0.02 ± 0.00	0.02 ± 0.00	
F	0.02 ± 0.00	0.02 ± 0.00	0.02 ± 0.00	0.02 ± 0.00	0.02 ± 0.00	0.02 ± 0.00	
Pa	1.21 ± 0.12	1.22 ± 0.12	1.20 ± 0.12	2.04 ± 0.36	2.13 ± 0.39	1.86 ± 0.38	
A	0.02 ± 0.00	0.02 ± 0.00	0.02 ± 0.00	0.02 ± 0.00	0.02 ± 0.00	0.02 ± 0.00	
FI	0.02 ± 0.00	0.02 ± 0.00	0.02 ± 0.00	0.02 ± 0.00	0.02 ± 0.00	0.02 ± 0.00	
Р	1.38 ± 0.10	1.40 ± 0.12	1.39 ± 0.09	2.45 ± 0.40	2.58 ± 0.40	2.34 ± 0.36	
BaA	0.02 ± 0.00	0.02 ± 0.00	0.02 ± 0.00	0.02 ± 0.00	0.02 ± 0.00	0.02 ± 0.00	
Ch	0.05 ± 0.03	0.05 ± 0.04	0.04 ± 0.03	0.05 ± 0.03	0.05 ± 0.04	0.04 ± 0.03	
BbF	0.15 ± 0.05	0.16 ± 0.05	0.14 ± 0.05	0.51 ± 0.12	0.53 ± 0.12	0.49 ± 0.11	
BkF	0.02 ± 0.00	0.02 ± 0.00	0.02 ± 0.00	0.02 ± 0.00	0.02 ± 0.00	0.02 ± 0.00	
BaP	0.02 ± 0.00	0.02 ± 0.00	0.02 ± 0.00	0.02 ± 0.00	0.02 ± 0.00	0.02 ± 0.00	
IP	0.02 ± 0.00	0.02 ± 0.00	0.02 ± 0.00	0.02 ± 0.00	0.02 ± 0.00	0.02 ± 0.00	
DhA	0.02 ± 0.00	0.02 ± 0.00	0.02 ± 0.00	0.02 ± 0.00	0.02 ± 0.00	0.02 ± 0.00	
BgP	0.03 ± 0.01	0.03 ± 0.01	0.02 ± 0.00	0.03 ± 0.02	0.04 ± 0.02	0.03 ± 0.02	
Total	3.81 ± 0.57	3.91 ± 0.58	3.75 ± 0.54	6.90 ± 1.23	7.22 ± 1.28	6.49 ± 1.18	
PAH4	0.23 ± 0.08	0.25 ± 0.09	0.22 ± 0.08	0.59 ± 0.15	0.62 ± 0.16	0.57 ± 0.14	

TABLE 7 Toxic equivalency factor (TEFs) and toxicity equivalency quotient TEQ BaP (µg/kg) concentrations in yogurt and butter

TEQBaP (μg/kg)							
Analytes	TEFs	Yogurt	Butter				
NA	0.00100	0.00002	0.00002				
Ace	0.00100	0.00162	0.00080				
Ac	0.00100	0.00002	0.00002				
F	0.00100	0.00002	0.00002				
Pa	0.00100	0.00201	0.00121				
А	0.01000	0.00020	0.00020				
FI	0.00100	0.00002	0.00002				
Р	0.00100	0.00245	0.00139				
BaA	0.10000	0.00200	0.00200				
Ch	0.01000	0.00046	0.00046				
BbF	0.10000	0.05075	0.01463				
BkF	0.10000	0.00200	0.00200				
BaP	1.00000	0.02167	0.02167				
IP	0.10000	0.00200	0.00200				
DhA	1.00000	0.02000	0.02000				
BgP	0.01000	0.00033	0.00025				

PAH4 observed in the present study were below the standard limits since the recorded values were 0.23 \pm 0.08 and 0.60 \pm 0.15 $\mu g/$ kg for yogurt and butter, respectively. The EC set $1 \mu g/kg$ as a maximum level for BaP in the baby foods for infant and young children, processed cereal-based foods, infant milk, and infant formulae. The current results showed that the levels of BaP were also below the standard levels (between 0.02–0.08 μ g/kg) in all samples and the recorded means were 0.03 \pm 0.02 and 0.03 \pm 0.01 $\mu g/kg$ in butter and yogurt, respectively. In a previous study, we reported that the means of total PAHs, PAH4, and BaP in doogh samples were $1.96 \pm 0.39, 0.15 \pm 0.07$, and $0.03 \pm 0.01 \, \mu g/kg$, respectively (Kiani et al., 2019). In this concern, Falco et al. (2003) reported a mean concentration of 6.636 μ g/kg for total PAHs in the dairy product (yogurt and cheese) and it was reported that the detection range of the sum of PAHs in conventional and organic yogurt ranged between 38.28 and 306.59 µg/kg fat (Rodríguez-Hernández et al., 2015). Recently, Kacmaz (2019) reported that the highest concentrations for the sum of the 4PAHs (benz[a]anthracene, chrysene, benzo[b] fluoranthene and benzo[a]pyrene) were 0.33 \pm 0.18 µg/kg of retail Turkish yogurt, 0.59 μ g/kg and 0.95 μ g/kg in yogurt with low-fat and high-fat content, respectively (Kacmaz 2019). Additionally, previous studies revealed that the concentrations of total PAHs were $3.695 \pm 0.43 - 5.443 \pm 0.66 \,\mu$ g/g in milk and dairy-based products



		Adults	Adults		
Analyte	RfD	Yogurt	Butter	Yogurt	Butter
NA	0.02	5.18E-07	1.43E-08	2.43E-06	6.66E-08
AcP	0.06	5.05E-04	1.42E-05	2.44E-03	6.59E-05
FL	0.04	7.66E-04	2.08E-05	3.65E-03	9.95E-05
AN	0.30	1.02E-04	2.84E-06	4.66E-04	1.29E-05
FLUR	0.04	7.67E-04	2.08E-05	3.57E-03	9.98E-05
PY	0.03	1.01E-03	2.76E-05	4.77E-03	1.34E-04
BaA	0.0003	1.02E-02	2.83E-04	4.63E-02	1.33E-03
Total	0.00	1.33E-02	3.69E-04	6.12E-02	1.75E-03

ood Science & Nutrition

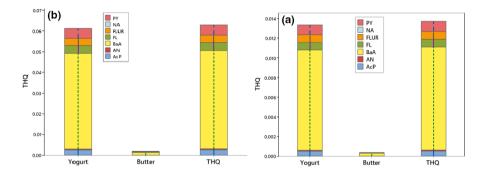


FIGURE 1 THQ value and percentage share due to content of 7 PAHs in the adult (a) and children (b) in milk products. PAHs, polycyclic aromatic hydrocarbons; THQ, total hazard quotient

(E.-S. A. Rawash et al., 2018) and 72.8 μ g/kg in butter samples collected from Egypt (Loutfy et al., 2007), 2.4 to 4 μ g/kg in Finnish butter (Hopia et al., 1986), 0.65 and 18.7 μ g/kg in fresh Spanish yogurt and butter, respectively (Martorell et al., 2010) and 0.61 and 18.3 μ g/kg in fresh yogurt and butter from Northwest Spain, respectively (Martí-Cid et al., 2008).

Due to this concern, many studies have been conducted on other food samples in different countries (Table 3) revealed shown that the results are varied due to different types of food. The reasons for these variations can be briefly summarized as follow: the low level may be due to the remoteness of a farm from the industrial environment, cities, roads, and do not consume or reduce the use of fossil fuels in livestock buildings to heat the environment, the use of healthy food and non-polluting water for animals, less food fats, the use of correct methods in food processing such as using less heat, steaming and not contacting foods with very hot surfaces and direct flame. However, the high levels can be due to the failure to comply with the above which increased the PAHs contaminant in food types (Kishikawa et al., 2003; Lawrence & Weber, 1984; Lin et al., 2016; Rawash et al., 2018; Rey-Salgueiro et al., 2009; Santonicola et al., 2017; Yurchenko & Mölder, 2005).

The means of the PAHs based on the type of milk product in different seasons are presented in Table 4 and indicated that the means of all PAH compounds recorded the highest value in the winter samples. The maximum means of total PAHs and PAH4 were 8.13 ± 1.02 and $0.55 \pm 0.09 \ \mu g/kg$ in butter, 4.40 ± 0.34 and $0.32 \pm 0.06 \ \mu g/kg$ in yogurt. The current results also showed that the maximum BaP mean in all of the brands was below the standard levels of the EU.

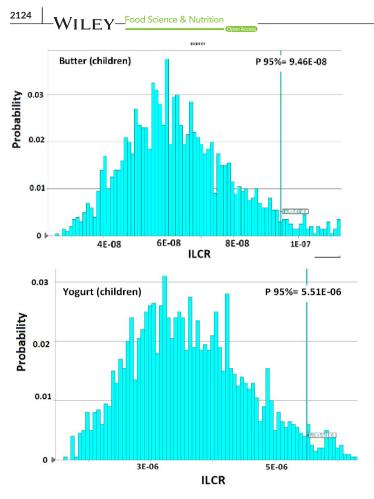
The lowest mean recorded for total PAHs and PAH4 were in spring, which reflects the high pollutants in winter due to the use of fossil fuels in livestock, or water, air, and livestock feed due to the lack of fresh fodder in this season.

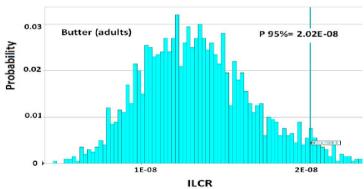
The means of PAHs compound based on the type of milk product, production date, and expiration are presented in Table 5. The results revealed that the means of all PAHs compounds were higher in expire date than the production date for the two tested milk products. In the expiration date, the maximum mean of both total PAHs and PAH4 were 7.38 \pm 1.35 and 0.68 \pm 0.15 µg/kg in butter and 4.11 \pm 0.53 and 0.27 \pm 0.09 µg/kg in yogurt. Additionally, the maximum mean of BaP in all fat status was below the standard limits of the EU. These compounds may be higher at the date of expiration for several reasons such as binding of these compounds together, turning from other substances to these compounds, and freeing them from containers (2020), but in general, the difference between production time and expiration time was not significant.

The means of PAHs analysts based on the type of brands are presented in Table 6 and indicated that the mean of all compounds in B and A brand in the samples showed the highest and lowest value, respectively. So that in brand B, the maximum mean of total PAHs and PAH4 were $7.22 \pm 1.28 \ \mu\text{g/kg}$ and $0.62 \pm 0.16 \ \mu\text{g/kg}$ in butter, and $3.91 \pm 0.58 \ \mu\text{g/kg}$ and $0.25 \pm 0.09 \ \mu\text{g/kg}$ in yogurt. On the other hand, the maximum mean of BaP in the tested brands was low compared with the standard levels of the EU. The possible reasons for this high level of PAHs in brand B may be due to the proximity of the produced animals to industrial environments, the pollution of water, air, or livestock feeds in this area.

2123

WILEY





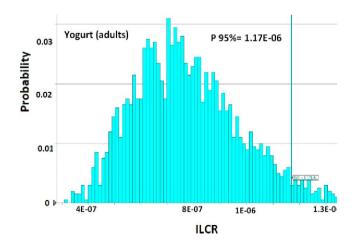


FIGURE 2 ILCR in the consumers due to ingestion yogurt and butter contents. ILCR, incremental lifetime of cancer risk

3.3 | Risk assessment of PAHs in milk products

The toxicity equivalency quotient (TEQ) of benzo(a)pyrene was evaluated to assess the recommended health risk. Our outcome revealed that yogurts had the highest TEQBaP (0.105 µg/kg), while butter had less TEQBaP (0.066 μ g/kg) as shown in Table 7. In a previous study, the level of TEQ in milk was reported as 0.21 μ g/kg by Yoon et al. (2007) who examined the risk assessment of analytes of PAHs in milk sold in Korea. The non-carcinogenic risk due to consumption of 7 PAHs in yogurt and butter showed that the rank order of HQ for adults and children was BaA > PY> F > FL>AcP > A > NA (Table 8). Because of the low oral reference dose (0.0003 mg kg⁻ ¹ day⁻¹), HQ of BaA was higher than other PAHs in milk products and the recorded values in these products were lower than 1 suggesting that subjects are not considerable at carcinogenic risk. The percentile of 95% THQ of 7 PAHs in the adult owing to ingestion of yogurt and butter was measured as 1.33E-02 and 3.69E-04, respectively. Furthermore, percentile 95% THQ in the children owing to ingestion of yogurt and butter was determined as 6.12E-02 and 1.75E-03, respectively. Moreover, THQ for adults and children was equal to 1.39E-01 and 6.37E-01, respectively. The comparison between THQ of PAHs in the samples showed that no carcinogenic risk due to the consumption of milk products for the adult and children since the recorded THQ of PAHs values were lower than 1. The share of the parameter in health risk of noncarcinogenic in the consumers due to ingestion of yogurt and butter obtained by the MCS (Figure 1a,b) concluded that BaA with 76.3% and 75.8% have a higher effect on THQ act in the adult and children, respectively. The results of carcinogenic risk of PAHs content due to the consumption of milk products (Figure 2) showed that ILCR index in adults owing to the consumption of yogurt and butter were 2.20E-07 and 5.98E-09, respectively. However, the percentile ILCR index in the children owing to ingestion of yogurt and butter were 1.02E-06 and 2.89E-8, respectively (Figure 2). Percentile 95% ILCR, PAHs in the yogurt and butter was between 1E-4 to 1E-9, hence, the carcinogenic risk for consumers is negligible (Ahmadkhaniha et al., 2009; USEPA, 2016). Furthermore, ILCR act in the adult and children was equal to 2.53E-07 and 1.18E-06, respectively; hence, the consumers are not at carcinogenic risk owing to the consumption of these milk products.

4 | CONCLUSIONS

This research provides a reliable, useful, and sensitive MSPE/GC-MS technique for measurement and risk assessment of PAHs in yogurt and butter samples by using a simple technique for the preparation of samples. The results of our study showed that the BaP in all samples was below the EU standard level for milk products. The mean of total PAHs in butter samples was higher than yogurt samples. Based on HQ no public health risk was detected and ILCR revealed no significant potential for carcinogenic risk in both children and adult consumers.

The analytical methods used in the current study are accurate and practical. It is recommended to be used on the industrial scale to the checking of PAHs in other milk products, such as cream, ice cream, cheese, etc. for toxicological and epidemiological studies. Moreover, it will help to ensure the safety of dairy industries.

ACKNOWLEDGMENTS

This research was supported by the Department of Public Health, Fasa University of Medical Sciences, Fasa, Iran (Grant no. 96253).

CONFLICT OF INTEREST

The authors are fully aware that there is no competing interest in this study.

ETHICAL REVIEW

This study does not involve any human or animal testing.

DATA AVAILABILITY STATEMENT

The required data will be available upon request.

ORCID

Nabi Shariatifar D https://orcid.org/0000-0002-2861-5758 Majid Arabameri https://orcid.org/0000-0003-0422-037X Mosaad A. Abdel-Wahhab D https://orcid.org/0000-0002-7174-3341

REFERENCES

- Abdollahi, M., Mohammadi-Nasrabadi, F., Houshiarrad, A., Ghaffarpur, M., Ghodsi, D., & Kalantari, N. (2014). Socio-economic differences in dietary intakes: The comprehensive study on household food consumption patterns and nutritional status of IR Iran. Nutrition and Food Sciences Research, 1(1), 19–26.
- Abou-Arab, A., Abou-Donia, M., El-Dars, F., Ali, O., & Hossam, A. (2014). Detection of polycyclic aromatic hydrocarbons levels in Egyptian meat and milk after heat treatment by Gas chromatography-mass spectrometry. International Journal of Current Microbiology and Applied Sciences, 3(7), 294–305.
- Aguinaga, N., Campillo, N., Vinas, P., & Hernández-Córdoba, M. (2007). Determination of 16 polycyclic aromatic hydrocarbons in milk and related products using solid-phase microextraction coupled to gas chromatography-mass spectrometry. *Analytica Chimica Acta*, 596(2), 285-290. https://doi.org/10.1016/j.aca.2007.06.005
- Aguinaga, N., Campillo, N., Viñas, P., & Hernández-Córdoba, M. (2008). A headspace solid-phase microextraction procedure coupled with gas chromatography-mass spectrometry for the analysis of volatile polycyclic aromatic hydrocarbons in milk samples. *Analytical and Bioanalytical Chemistry*, 391(3), 753–758. https://doi.org/10.1007/ s00216-008-1841-2
- Ahmadkhaniha, R., Shafiee, A., Rastkari, N., & Kobarfard, F. (2009). Accurate quantification of endogenous androgenic steroids in cattle's meat by gas chromatography mass spectrometry using a surrogate analyte approach. Analytica Chimica Acta, 631(1), 80-86. https://doi.org/10.1016/j.aca.2008.10.011
- Alharbi, O. M., Khattab, R. A., & Ali, I. (2018). Health and environmental effects of persistent organic pollutants. *Journal of Molecular Liquids*, 263, 442–453. https://doi.org/10.1016/j.molliq.2018.05.029
- Ali, I., & Aboul-Enein, H. Y. (2005). Chiral pollutants: Distribution, toxicity and analysis by chromatography and capillary electrophoresis: John Wiley & Sons.

- Ali, I., Aboul-Enein, H. Y., & Ghanem, A. (2005). Enantioselective toxicity and carcinogenesis. Current Pharmaceutical Analysis, 1(1), 109–125.
- Ali, I., Alharbi, O. M. L., ALOthman, Z. A., Al-Mohaimeed, A. M., & Alwarthan, A. (2019). Modeling of fenuron pesticide adsorption on CNTs for mechanistic insight and removal in water. *Environmental Research*, 170, 389–397. https://doi.org/10.1016/j. envres.2018.12.066
- Ali, I., Alharbi, O. M., Tkachev, A., Galunin, E., Burakov, A., & Grachev, V. A. (2018). Water treatment by new-generation graphene materials: Hope for bright future. *Environmental Science and Pollution Research*, 25(8), 7315–7329. https://doi.org/10.1007/ s11356-018-1315-9
- Ali, I., Gupta, V., & Aboul-Enein, H. Y. (2003). Chirality: A challenge for the environmental scientists. *Current Science-Bangalore*, 84(2), 152–156.
- Ali, I., Wani, W. A., & Saleem, K. (2011). Cancer scenario in India with future perspectives. *Cancer Therapy*, 8(8), 56–70.
- Basheer, A. A. (2018). Chemical chiral pollution: Impact on the society and science and need of the regulations in the 21st century. *Chirality*, 30(4), 402–406.
- Basheer, A. A., & Ali, I. (2018). Stereoselective uptake and degradation of (±)-o, p-DDD pesticide stereomers in water-sediment system. *Chirality*, 30(9), 1088–1095.
- Bianchi, F., Careri, M., Mangia, A., Mattarozzi, M., & Musci, M. (2008). Experimental design for the optimization of the extraction conditions of polycyclic aromatic hydrocarbons in milk with a novel diethoxydiphenylsilane solid-phase microextraction fiber. *Journal* of Chromatography A, 1196, 41-45. https://doi.org/10.1016/j. chroma.2008.04.018
- Cho, H.-K., & Shin, H.-S. (2012). Evaluation of polycyclic aromatic hydrocarbon contents and risk assessment for infant formula in Korea. *Food Science and Biotechnology*, 21(5), 1329–1334. https://doi. org/10.1007/s10068-012-0175-1
- Chung, T.-L., Liao, C.-J., & Chen, M.-F. (2010). Comparison of liquidliquid extraction and solid-phase extraction for the determination of polycyclic aromatic hydrocarbons in the milk of Taiwan. Journal of the Taiwan Institute of Chemical Engineers, 41(2), 178–183. https://doi. org/10.1016/j.jtice.2009.07.003
- Ciecierska, M., & Obiedziński, M. (2010). Polycyclic aromatic hydrocarbons in infant formulae, follow-on formulae and baby foods available in the Polish market. *Food Control*, 21(8), 1166–1172. https://doi. org/10.1016/j.foodcont.2010.01.013
- Cirillo, T., Milano, N., & Cocchieri, R. A. (2004). Polycyclic aromatic hydrocarbons (PAHs) in traditional smoked dairy products from Campania (Italy). *Italian Journal of Public Health*, 1(1–2), e6143.
- Dobrinas, S., Soceanu, A., Popescu, V., & Coatu, V. (2016). Polycyclic aromatic hydrocarbons and pesticides in milk powder. *Journal of Dairy Research*, 83(2), 261–265. https://doi.org/10.1017/S002202991 6000169
- EPA. (2010). Application of the Margin of Exposure (MoE) approach to substances in food that are genotoxic and carcinogenic example: Benzo[a] pyrene and polycyclic aromatic hydrocarbons, https://cfpub.epa.gov/ si/si_public_record_report.cfm?Lab=NCCT&dirEntryId=218290. In Appendix A to: US Environmental Protection Agency Washington, DC.
- EPA. (2016). Use of Monte Carlo simulation in risk assessments EPA. https://www.epa.gov/risk/use-monte-carlo-simulation-risk-asses sments
- European Commission. (2011). Amending Regulation (EC) No. 1881/2006 as regards maximum levels for polycyclic aromatic hydrocarbons in foodstuffs. Official Journal of the European Union, 835/2011(215), 4-8. https://eur-lex.europa.eu/LexUriServ/LexUriServ.do?uri=OJ:L: 2011:215:0004:0008:EN:PDF
- Fakhri, Y., Bjørklund, G., Bandpei, A. M., Chirumbolo, S., Keramati, H., Pouya, R. H., Asadi, A., Amanidaz, N., Sarafraz, M., Sheikhmohammad, A., Alipour, M., Baninameh, Z., Mohseni, S. M.,

& Sarkhosh, M., & Sheikhmohammad, A. (2018). Concentrations of arsenic and lead in rice (*Oryza sativa* L.) in Iran: A systematic review and carcinogenic risk assessment. *Food and Chemical Toxicology*, 113, 267–277.

- Falco, G., Domingo, J. L., Llobet, J. M., Teixido, A., Casas, C., & Müller, L. (2003). Polycyclic aromatic hydrocarbons in foods: Human exposure through the diet in Catalonia. *Spain. Journal of Food Protection*, 66(12), 2325–2331.
- Farhadian, A., Jinap, S., Abas, F., & Sakar, Z. I. (2010). Determination of polycyclic aromatic hydrocarbons in grilled meat. *Food Control*, 21(5), 606–610.
- Garcia Londoño, V. A., Garcia, L. P., Scussel, V. M., & Resnik, S. (2013). Polycyclic aromatic hydrocarbons in milk powders marketed in Argentina and Brazil. *Food Additives & Contaminants: Part A*, 30(9), 1573–1580.
- Ghasemidehkordi, B., Malekirad, A. A., Nazem, H., Fazilati, M., Salavati, H., Shariatifar, N., Rezaei, M., Fakhri, Y., & Khaneghah, A. M. (2018). Concentration of lead and mercury in collected vegetables and herbs from Markazi province, Iran: Non-carcinogenic risk assessment. *Food and Chemical Toxicology*, 113, 204–210.
- Gorji, M. E. H., Ahmadkhaniha, R., Moazzen, M., Yunesian, M., Azari, A., & Rastkari, N. (2016). Polycyclic aromatic hydrocarbons in Iranian Kebabs. Food Control, 60, 57–63.
- Grova, N., Feidt, C., Crépineau, C., Laurent, C., Lafargue, P. E., Hachimi, A., & Rychen, G. (2002). Detection of polycyclic aromatic hydrocarbon levels in milk collected near potential contamination sources. *Journal of Agricultural and Food Chemistry*, 50(16), 4640–4642.
- Gutiérrez, R., Vega, S., Ortiz, R., Pérez, J. J., & Schettino, B. (2015). Presence of PAHs in milk of industrial farms from Tizayuca, Hidalgo, Mexico. Journal of Environmental Science and Health, Part B, 50(5), 317-321.
- Han, J.-H., Kim, M.-J., & Shin, H.-S. (2014). Evaluation of polycyclic aromatic hydrocarbon contents and risk assessment for infant formula in Korea. *Journal of the Korean Society for Applied Biological Chemistry*, 57(2), 173–179. https://doi.org/10.1007/s13765-013-4219-x
- Hopia, A., Pyysalo, H., & Wickström, K. (1986). Margarines, butter and vegetable oils as sources of polycyclic aromatic hydrocarbons. *Journal of the American Oil Chemists' Society*, 63(7), 889–893. https:// doi.org/10.1007/BF02540921
- Huang, C.-L., Bao, L.-J., Luo, P., Wang, Z.-Y., Li, S.-M., & Zeng, E. Y. (2016). Potential health risk for residents around a typical e-waste recycling zone via inhalation of size-fractionated particle-bound heavy metals. *Journal of Hazardous Materials*, 317, 449–456. https://doi. org/10.1016/j.jhazmat.2016.05.081
- Iwegbue, C. M., Edeme, J. N., Tesi, G. O., Bassey, F. I., Martincigh, B. S., & Nwajei, G. E. (2014). Polycyclic aromatic hydrocarbon concentrations in commercially available infant formulae in Nigeria: Estimation of dietary intakes and risk assessment. *Food and Chemical Toxicology*, 72, 221–227. https://doi.org/10.1016/j.fct.2014.06.026
- Jahanbakhsh, M., Afshar, A., Momeni Feeli, S., Pabast, M., Ebrahimi, T., Mirzaei, M., Akbari-Adergani, B., & Farid, M. & Arabameri, M. (2019). Probabilistic health risk assessment (Monte Carlo simulation method) and prevalence of aflatoxin B1 in wheat flours of Iran. International Journal of Environmental Analytical Chemistry, 1–12. https://doi. org/10.1080/03067319.03062019.01676421
- Kamankesh, M., Mohammadi, A., Hosseini, H., & Tehrani, Z. M. (2015). Rapid determination of polycyclic aromatic hydrocarbons in grilled meat using microwave-assisted extraction and dispersive liquid– liquid microextraction coupled to gas chromatography-mass spectrometry. *Meat Science*, 103, 61–67. https://doi.org/10.1016/j.meats ci.2015.01.001
- Karami, H., Shariatifar, N., Nazmara, S., Moazzen, M., Mahmoodi, B., & Mousavi Khaneghah, A. (2020). The Concentration and probabilistic health risk of potentially toxic elements (PTEs) in Edible Mushrooms (Wild and Cultivated) samples collected from different cities of

Iran. Biological Trace Element Research, 199(1), 389-400. https://doi. org/10.1007/s12011-020-02130-x

- Kiani, A., Shariatifar, N., Shahsavari, S., Ahmadloo, M., & Moazzen, M. (2019). Investigating the presence of polycyclic aromatic hydrocarbons in Doogh. *Journal of Mazandaran University of Medical Sciences*, 29(178), 10–23.
- Kishikawa, N., Wada, M., Kuroda, N., Akiyama, S., & Nakashima, K. (2003). Determination of polycyclic aromatic hydrocarbons in milk samples by high-performance liquid chromatography with fluorescence detection. *Journal of Chromatography B*, 789(2), 257–264. https://doi. org/10.1016/S1570-0232(03)00066-7
- Lawrence, J. F., & Weber, D. F. (1984). Determination of polycyclic aromatic hydrocarbons in Canadian samples of processed vegetable and dairy products by liquid chromatography with fluorescence detection. Journal of Agricultural and Food Chemistry, 32(4), 794–797. https://doi.org/10.1021/jf00124a023
- Lee, S.-Y., Lee, J.-Y., & Shin, H.-S. (2015). Evaluation of chemical analysis method and determination of polycyclic aromatic hydrocarbons content from seafood and dairy products. *Toxicological Research*, 31(3), 265. https://doi.org/10.5487/TR.2015.31.3.265
- Lin, W., Wei, S., Jiang, R., Zhu, F., & Ouyang, G. (2016). Calibration of the complex matrix effects on the sampling of polycyclic aromatic hydrocarbons in milk samples using solid phase microextraction. *Analytica Chimica Acta*, 933, 117–123. https://doi.org/10.1016/j. aca.2016.05.045
- Loutfy, N., Fuerhacker, M., Tundo, P., Raccanelli, S., & Ahmed, M. T. (2007). Monitoring of polychlorinated dibenzo-p-dioxins and dibenzofurans, dioxin-like PCBs and polycyclic aromatic hydrocarbons in food and feed samples from Ismailia city, Egypt. Chemosphere, 66(10), 1962–1970. https://doi.org/10.1016/j.chemosphere.2006.07.081
- Martí-Cid, R., Llobet, J. M., Castell, V., & Domingo, J. L. (2008). Evolution of the dietary exposure to polycyclic aromatic hydrocarbons in Catalonia. Spain. Food and Chemical Toxicology, 46(9), 3163–3171. https://doi.org/10.1016/j.fct.2008.07.002
- Martorell, I., Perelló, G., Martí-Cid, R., Castell, V., Llobet, J. M., & Domingo, J. L. (2010). Polycyclic aromatic hydrocarbons (PAH) in foods and estimated PAH intake by the population of Catalonia, Spain: Temporal trend. *Environment International*, 36(5), 424–432. https://doi.org/10.1016/j.envint.2010.03.003
- Moazzen, M., Ahmadkhaniha, R., Gorji, M. E., Yunesian, M., & Rastkari, N. (2013). Magnetic solid-phase extraction based on magnetic multiwalled carbon nanotubes for the determination of polycyclic aromatic hydrocarbons in grilled meat samples. *Talanta*, 115, 957–965. https://doi.org/10.1016/j.talanta.2013.07.005
- Moazzen, M., Khaneghah, A. M., Shariatifar, N., Ahmadloo, M., Eş, I., Baghani, A. N., Dobaradaran, S., Rastkari, N., Nazmara, S., Delikhoon, M., & Jahed Khaniki, G. R., (2018). Multi-walled carbon nanotubes modified with iron oxide and silver nanoparticles (MWCNT-Fe3O4/ Ag) as a novel adsorbent for determining PAEs in carbonated soft drinks using magnetic SPE-GC/MS method. Arabian Journal of Chemistry, 12(4), 476–488.
- Moazzen, M., Mahvi, A. H., Shariatifar, N., Jahed Khaniki, G., Nazmara, S., Alimohammadi, M., Ahmadkhaniha, R., Rastkari, N., Ahmadloo, M., Akbarzadeh, A., Dobaradaran, S., & Norouzian Baghani, A. (2018). Determination of phthalate acid esters (PAEs) in carbonated soft drinks with MSPE/GC-MS method. *Toxin Reviews*, 37(4), 319-326. https://doi.org/10.1080/15569543.2017.1378234
- Naccari, C., Cristani, M., Giofrè, F., Ferrante, M., Siracusa, L., & Trombetta, D. (2011). PAHs concentration in heat-treated milk samples. *Food Research International*, 44(3), 716–724. https://doi.org/10.1016/j. foodres.2010.12.029
- Orecchio, S., Ciotti, V. P., & Culotta, L. (2009). Polycyclic aromatic hydrocarbons (PAHs) in coffee brew samples: Analytical method by GC-MS, profile, levels and sources. *Food and Chemical Toxicology*, 47(4), 819–826. https://doi.org/10.1016/j.fct.2009.01.011

- Rastkari, N., & Ahmadkhaniha, R. (2013). Magnetic solid-phase extraction based on magnetic multi-walled carbon nanotubes for the determination of phthalate monoesters in urine samples. *Journal* of Chromatography A, 1286, 22–28. https://doi.org/10.1016/j. chroma.2013.02.070
- Rawash, E.-S., Mohamed, G., Souaya, E., Khalil, L., El-Chaghaby, G., & El-Gammal, M. (2018). Distribution and health hazards of polycyclic aromatic hydrocarbons in Egyptian Milk and dairy-based products. *Beverages*, 4(3), 63–73.
- Rey-Salgueiro, L., Martínez-Carballo, E., García-Falcón, M. S., González-Barreiro, C., & Simal-Gándara, J. (2009). Occurrence of polycyclic aromatic hydrocarbons and their hydroxylated metabolites in infant foods. *Food Chemistry*, 115(3), 814–819. https://doi.org/10.1016/j. foodchem.2008.12.095
- Rodríguez-Hernández, Á., Camacho, M., Boada, L. D., Ruiz-Suarez, N., Almeida-González, M., Henríquez-Hernández, L. A., Zumbado, M., & Luzardo, O. P. (2015). Daily intake of anthropogenic pollutants through yogurt consumption in the Spanish population. *Journal of Applied Animal Research*, 43(4), 373–383. https://doi. org/10.1080/09712119.2014.978777
- Saito, E., Tanaka, N., Miyazaki, A., & Tsuzaki, M. (2014). Concentration and particle size distribution of polycyclic aromatic hydrocarbons formed by thermal cooking. *Food Chemistry*, 153, 285–291. https:// doi.org/10.1016/j.foodchem.2013.12.055
- Samiee, S., Fakhri, Y., Sadighara, P., Arabameri, M., Rezaei, M., Nabizadeh, R., Shariatifar, N., & Mousavi Khaneghah, A. (2020). The concentration of polycyclic aromatic hydrocarbons (PAHs) in the processed meat samples collected from Iran's market: A probabilistic health risk assessment study. *Environmental Science and Pollution Research*, 27(17), 21126–21139. https://doi.org/10.1007/ s11356-020-08413-z
- Santonicola, S., Albrizio, S., Murru, N., Ferrante, M. C., & Mercogliano, R. (2017). Study on the occurrence of polycyclic aromatic hydrocarbons in milk and meat/fish based baby food available in Italy. *Chemosphere*, 184, 467–472. https://doi.org/10.1016/j.chemosphere.2017.06.017
- Sibe, K. (2019). Polycyclic aromatic hydrocarbons in retail Turkish yogurts. Quality Assurance and Safety of Crops & Foods, 11(4), 361–367.
- Shariatifar, N., Dadgar, M., Fakhri, Y., Shahsavari, S., Moazzen, M., Ahmadloo, M., Kiani, A., Aeenehvand, S., Nazmara, S., & Mousavi Khanegah, A. (2020). Levels of polycyclic aromatic hydrocarbons in milk and milk powder samples and their likely risk assessment in Iranian population. *Journal of Food Composition and Analysis*, 85, 103331.-https://doi.org/10.1016/j.jfca.2019.103331
- Suchanová, M., Hajšlová, J., Tomaniová, M., Kocourek, V., & Babička, L. (2008). Polycyclic aromatic hydrocarbons in smoked cheese. *Journal* of the Science of Food and Agriculture, 88(8), 1307–1317. https://doi. org/10.1002/jsfa.3198
- USEPA. (2016). U.S. Environmental Protection Agency, Supplemental guidance for assessing susceptibility from early-life exposure to carcinogens. http://www3.epa.gov/airtoxics/childrenssupplementfinal.pdf. Accessed on January 25, 2016.
- USEPA, (2017). Toxicological Review of Benzo[a]pyrene Executive Summary [CASRN 50-32-8]. Integrated Risk Information System, National Center for Environmental Assessment, Office of Research and Development, U.S. Environmental Protection Agency, Washington, DC.
- Wegrzyn, E., Grzeskiewicz, S., Poplawska, W., & Glod, B. (2006). Modified analytical method for polycyclic aromatic hydrocarbons, using sec for sample preparation and RP-HPLC with fluorescence detection. Application to different food samples. Acta Chromatographica, 17, 233.
- Wretling, S., Eriksson, A., Eskhult, G., & Larsson, B. (2010). Polycyclic aromatic hydrocarbons (PAHs) in Swedish smoked meat and fish. *Journal of Food Composition and Analysis*, 23(3), 264–272. https://doi. org/10.1016/j.jfca.2009.10.003

ILEY_Food Science & Nutrition

- Yoon, E., Park, K., Lee, H., Yang, J.-H., & Lee, C. (2007). Estimation of excess cancer risk on time-weighted lifetime average daily intake of PAHs from food ingestion. *Human and Ecological Risk Assessment*, 13(3), 669–680. https://doi.org/10.1080/10807030701226871
- Yurchenko, S., & Mölder, U. (2005). The determination of polycyclic aromatic hydrocarbons in smoked fish by gas chromatography mass spectrometry with positive-ion chemical ionization. *Journal of Food Composition and Analysis*, 18(8), 857–869. https://doi.org/10.1016/j. jfca.2004.11.004

How to cite this article: Kiani A, Ahmadloo M, Moazzen M, et al. Monitoring of polycyclic aromatic hydrocarbons and probabilistic health risk assessment in yogurt and butter in Iran. *Food Sci Nutr.* 2021;9:2114–2128. <u>https://doi.</u> org/10.1002/fsn3.2180