

Supplemental File

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SUPPLEMENTAL FIGURES AND TABLES

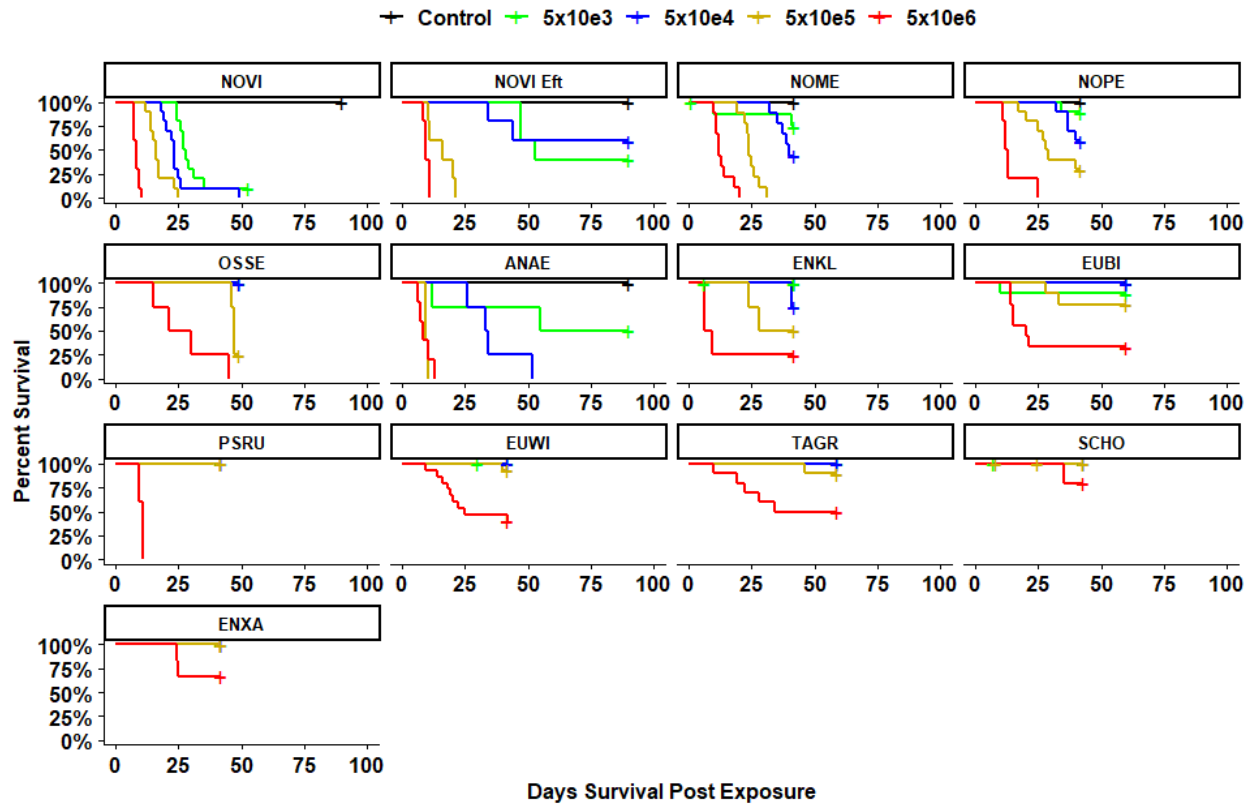


Figure 1. Survival curves among four zoospore exposure doses for species that experienced mortality due to *Batrachochytrium salamandrivorans* (*Bsal*) chytridiomycosis. Curves not shown = 100% survival. ANAE= *Aneides aeneus*, ENKL= *Ensatina eschscholtzii klauberi*, ENXA= *Ensatina eschscholtzii xanthoptica*, EUBI= *Eurycea bislineata*, EUWI= *Eurycea wilderae*, NOME = *Notophthalmus meridionalis*, NOPE= *Notophthalmus perstriatus*, NOVI = *Notophthalmus viridescens*, OSSE= *Osteopilus septentrionalis*, PSRU= *Pseudotriton ruber*, SCHO= *Scaphiopus holbrookii*, and TAGR= *Taricha granulosa*. All species listed here were adults except for NOVI eft = juvenile.

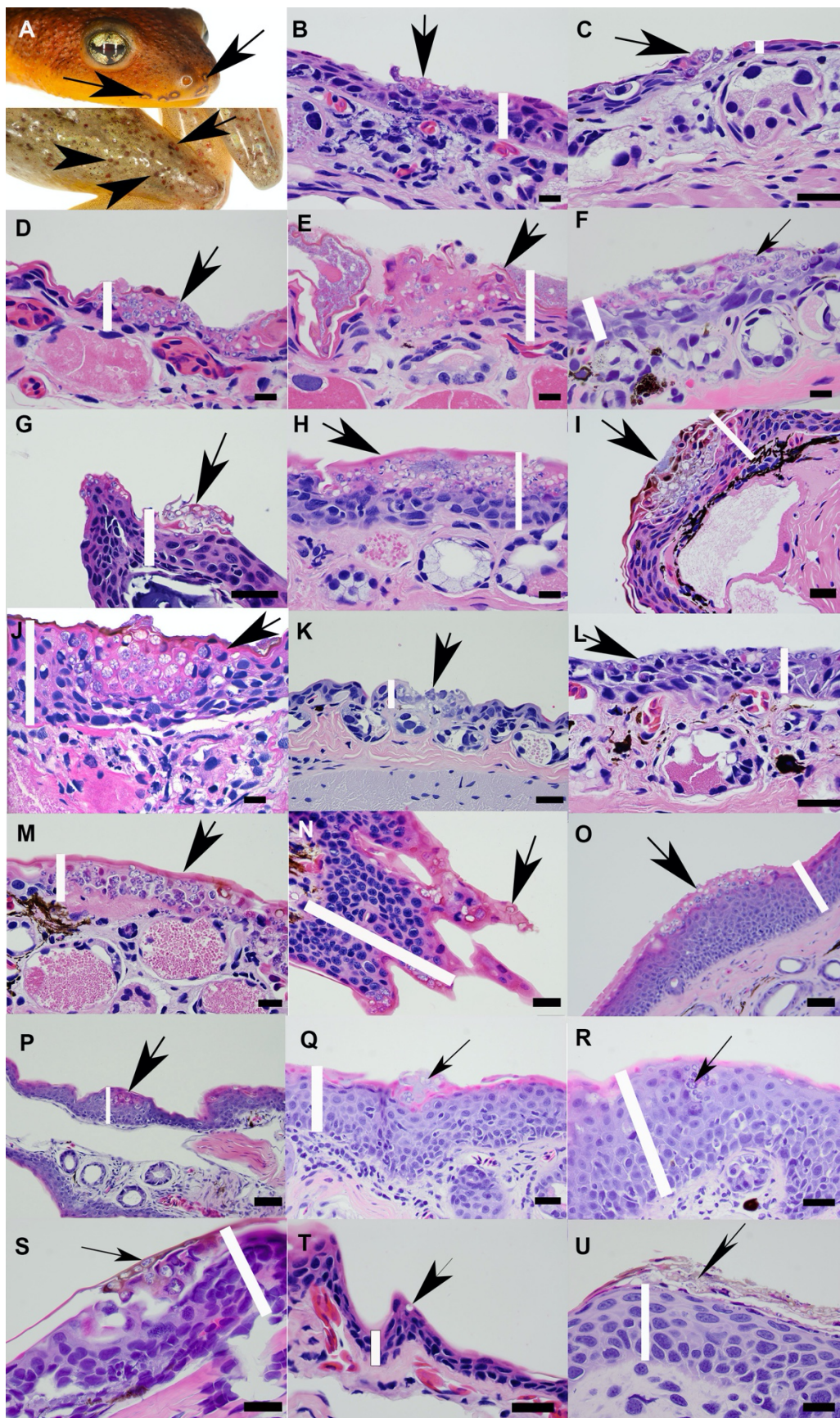


Figure 2. *Batrachochytrium salamandrivorans* (Bsal) chytridiomycosis in experimentally exposed salamanders and anurans. Gross lesions included classic target lesions in salamanders (A top arrows) and solid areas of necrosis as well as occasional target lesions in anurans (A bottom arrowheads and arrow, respectively). Hematoxylin and eosin stained skin sections of the gross lesions with epidermal destruction by *Bsal* fungi (arrows) are shown in the following species: adult *Notophthalmus viridescens* (B) juvenile *N. viridescens* (C), *N. perstriatus* (D), *N. meridionalis* (E), adult *Eurycea bislineata* (F), larval *E. bislineata* (G), *E. wilderae* (H), *Taricha granulosa* (I), *T. torosa* (J), *Aneides aeneus* (K), *Pseudotriton ruber* (L), *Desmognathus escholtzii klauberi* (M), *E. escholtzii xanthoptica* (N), *Osteopilus septentrionalis* (O)*, and *Scaphiopus holbrookii* (P)*. Species with positive qPCR results that had no obvious gross lesions where *Bsal* was detected using histology included: *Anaxyrus americanus* (Q)*, *Hyla chrysoscelis* (R)*, *Plethodon metcalfei* (S), *Eurycea lucifuga* (T), and *Siren lacertina* (U). At least 5 individuals per species were inspected to verify *Bsal* infection and chytridiomycosis using histology (see Supplemental Table 1). Epidermal layer noted by white bars; * = anurans; all others = salamanders; black scale bars = 20 μ m.

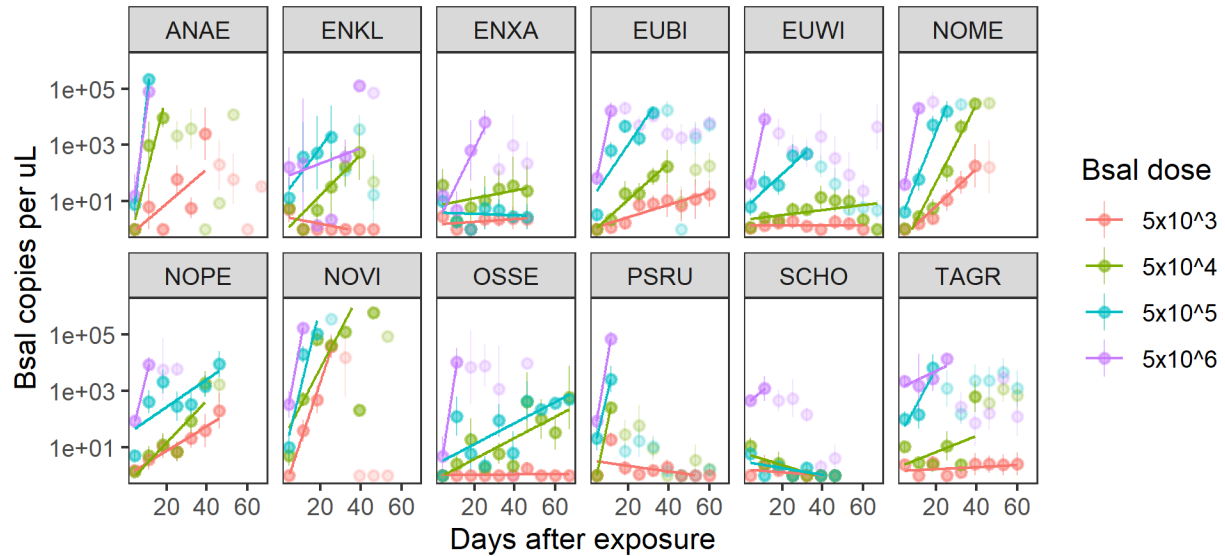


Figure 3. *Bsal* growth rate varies with exposure dose. *Bsal* load (mean and standard error for each sampling period) over the course of the exposure experiments for the species that developed mean loads $> 10^3$ for at least one of the *Bsal* doses (see Supplementary Text for details). Darker symbols represent the linear portion of the growth curve on a log scale (see Supplementary Text for details), lighter symbols represent data collected after this period. Lines represent linear models fit to the log-transformed raw data for the linear portion of the growth curve. According to the ZINB mixed model described in the text, across all species there was a difference in the rate of *Bsal* increase between treatments ($\chi^2=33.35$, $df=6$, $P=9.0 \times 10^{-6}$). This was largely driven by more rapid development of *Bsal* infection in the 5×10^6 treatment compared to the other treatments – between the first week and second weeks after exposure model-predicted *Bsal* load increased 526% in the 5×10^6 treatment and only 165-320% in the other treatments. All hypothesis tests were two-sided. ANAE= *Aneides aeneus*, ENKL= *Ensatina eschscholtzii klauberi*, ENXA= *Ensatina eschscholtzii xanthoptica*, EUBI= *Eurycea bislineata*, EUWI= *Eurycea wilderae*, NOME = *Notophthalmus meridionalis*, NOPE= *Notophthalmus perstriatus*, NOVI = *Notophthalmus viridescens*, OSSE= *Osteopilus septentrionalis*, PSRU= *Pseudotriton ruber*, SCHO= *Scaphiopus holbrookii*, and TAGR= *Taricha granulosa*.

Table 1. Host susceptibility and conservation risk for 35 North American amphibian species exposed to *Batrachochytrium salamandrivorans* (Bsal) at 15°C

Species	Life-stage	% Infection (n) ^a	% Mortality ^a	Avg Copies ^a	Avg Copies SE ^a	Incubation Period ^{a,b,c}	SE ^{a,c}	ID-50 ^c	SE ^{c,d}	LD-50 ^c	SE ^{c,d}	Conservation Risk ^e	Gross Lesions ^f	Histology ^g
<i>Notophthalmus viridescens</i>	Adult	100 (10)	100	128072.1	38263	6.4	0.3	2897	Inf	2897	Inf	Very High	Yes	Yes
<i>Notophthalmus viridescens</i>	Juvenile	100(10)	100	304147.9	159247	4.0	0.0	NE	NE	NE	NE	Very High	Yes	Yes
<i>Notophthalmus perstriatus</i>	Adult	100 (10)	100	20067.27	5970	4.0	0.0	NE	NE	71767	1.8	Very High	Yes	Yes
<i>Eurycea bislineata</i>	Adult	100 (9)	66.67	47998.22	6743	4.0	0.0	2987	Inf	2859708	3.0	High	Yes	Yes
<i>Notophthalmus meridionalis</i>	Adult	100 (9)	100	35909.89	10125	4.7	0.7	3708	Inf	20368	1.5	Very High	Yes	Yes
<i>Taricha granulosa</i>	Adult	100 (10)	50	18077.63	4222	4.0	0.0	5925	Inf	4862047	2.0	Low	Yes	Yes
<i>Osteopilus septentrionalis</i>	Adult	100 (4)	100	57135.48	24901	8.5	1.5	6203	Inf	402625	Inf	High	Yes	Yes
<i>Aneides aeneus</i>	Adult	100 (10)	50	35360	14387	7.0	1.0	9528	2.0	1409309	13.6	Moderate	Yes	Yes
<i>Pseudotriton ruber</i>	Adult	100 (5)	100	86275.63	33811	4.0	0.0	15555	1.8	1570681	Inf	Moderate	Yes	Yes
<i>Ensatina eschscholtzii klauberi</i>	Adult	100 (4)	75	29116.23	14397	4.0	0.0	15921	Inf	973838	2.2	Moderate	Yes	Yes
<i>Eurycea wilderae</i>	Adult	100 (15)	66.67	20969.79	4340	5.7	0.8	42452	1.4	2993608	1.4	Low	Yes	Yes
<i>Eurycea lucifuga</i>	Adult	80 (5)	0	164.6402	124	7.0	1.7	105417	2.5	NE	NE	Low	No	Yes
<i>Ensatina e. xanthoptica</i>	Adult	100 (6)	33	44585.16	23441	9.0	2.4	252249	3.5	5871555	Inf	Low	Yes	Yes
<i>Scaphiopus holbrookii</i>	Juvenile	100 (5)	20	1589.924	546	4.0	0.0	412380	2.8	7063138	Inf	Low	No	Yes
<i>Taricha torosa</i>	Adult	60 (10)	0	80		31.0		450990	NE	NE	NE	Low	Yes	Yes
<i>Rana chiricahuensis</i>	Juvenile	62.5 (8)	0	13.17908	6	4.0	0.0	496096	Inf	NE	NE	Low	No	No
<i>Pseudobranchius striatus</i>	Adult	80 (100)	0	122.9917	37	4.0	0.0	1053259	1.9	NE	NE	Low	No	No
<i>Hyla chrysoscelis</i>	Juvenile	50 (10)	0	14.95589	7	6.4	1.5	5000008	1.1	NE	NE	Low	No	Yes
<i>Plethodon shermani</i>	Adult	42.9 (7)	0	7.06142	6	25.0	15.0	5333243	Inf	NE	NE	Low	No	No
<i>Ambystoma opacum</i>	Adult	10 (10)	0	0.134	0	4.0	0.0	8629525	Inf	NE	NE	Low	No	No
<i>Desmognathus ocoee</i>	Adult	30 (10)	0	1.89775	2	6.0	2.0	15859195	4.1	NE	NE	Low	No	No
<i>Notophthalmus viridescens</i>	Larval	17 (12)	0	116		7.0		25648420	Inf	NE	NE	Low	No	No
<i>Ambystoma mexicanum</i>	Adult	50 (10)	0	0.348	0	17.2	3.5	37417161	130.1	NE	NE	Low	No	No
<i>Plethodon metcalfi</i>	Adult	20 (10)	0	1.13275	1	13.0	3.0	51557075	11.4	NE	NE	Low	No	Yes
<i>Siren lacertina</i>	Adult	20 (5)	0	1.313978	1	45.0	0.0	52891331	35.3	NE	NE	Low	No	Yes
<i>Cryptobranchius alleganiensis</i>	Juvenile	1 (7)	0	0.019	0	40.0	0.0	NE	NE	NE	NE	Low	No	No
<i>Anaxyrus americanus</i>	Juvenile	100 (5)	0	552	369	4.0	0.0	NE	NE	NE	NE	Low	No	Yes
<i>Eurycea bislineata</i>	Larval	28 (7)	0	11160.06	2379	NE	NE	NE	NE	NE	NE	Low	No	Yes
<i>Ambystoma maculatum</i>	Juvenile	0 (12)	0	0	0	NE	NE	NE	NE	NE	NE	Low	No	No
<i>Ambystoma laterale</i>	Adult	0 (5)	0	0	0	NE	NE	NE	NE	NE	NE	Resistant	No	No
<i>Ambystoma maculatum</i>	Larval	0 (10)	0	0	0	NE	NE	NE	NE	NE	NE	Resistant	No	No
<i>Desmognathus aeneus</i>	Adult	0 (10)	0	0	0	NE	NE	NE	NE	NE	NE	Resistant	No	No
<i>Desmognathus monticola</i>	Adult	0 (10)	0	0	0	NE	NE	NE	NE	NE	NE	Resistant	No	No
<i>Hemidactylium scutatum</i>	Adult	0 (5)	0	0	0	NE	NE	NE	NE	NE	NE	Resistant	No	No
<i>Necturus maculosus</i>	Adult	0 (5)	0	0	0	NE	NE	NE	NE	NE	NE	Resistant	No	No
<i>Atelopus zeteki</i>	Larval	0 (10)	0	0	0	NE	NE	NE	NE	NE	NE	Resistant	No	No
<i>Rana catesbeiana</i>	Juvenile	0 (6)	0	0	0	NE	NE	NE	NE	NE	NE	Resistant	No	No
<i>Rana catesbeiana</i>	Larval	0 (30)	0	0	0	NE	NE	NE	NE	NE	NE	Resistant	No	No
<i>Rana pipiens</i>	Adult	0 (5)	0	0	0	NE	NE	NE	NE	NE	NE	Resistant	No	No
<i>Rana sylvatica</i>	Juvenile	0 (5)	0	0	0	NE	NE	NE	NE	NE	NE	Resistant	No	No
<i>Rana pipiens</i>	Larval	0 (20)	0	0	0	NE	NE	NE	NE	NE	NE	Resistant	No	No

^a Highest exposure dose results (see Supplemental Table 2); percent infection and mortality were calculated using the total sample size (n) in this dose; copies refer to *Bsal* fungal DNA copies per microliter.

^b Incubation period was calculated by averaging the number of days it took for each individual to test positive for *Bsal* DNA using quantitative-PCR for each species.

^c NE= Not estimable, because either no or all animals became infected or died.

^d Inf= Infinite standard error estimation due to limits of model estimation based on available data.

^e Conservation risk (also referred to as mortality risk, Figure 3) was estimated as the product of the median infectious dose (ID-50) and lethal dose (LD-50) estimates, and ranked according to the 25th percentile quartiles. All species that became infected but did not die also were classified as low susceptibility and risk. Species with 100% infection and mortality at all treatment groups were classified as very high conservation risk. Note: NOVI larvae infection only occurred in metamorphosing individuals.

^f Gross lesions were noted during each observation event. A “yes” in this column indicates that the species had gross lesions typically consistent with *Bsal* chytridiomycosis^{1,2}. According to the case definition of *Bsal*, gross signs should not be used to diagnosis *Bsal* chytridiomycosis¹; histology in combination with another diagnostic methods (e.g., qPCR, culture) are recommended².

^g Histological examinations were performed on each species which had *Bsal* detected via qPCR. A “yes” in this column indicates that *Bsal* infections were confirmed through histological observations (see Supplemental Figure 2).

Table 2. Collecting location and experimental conditions for *Batrachochytrium salamandrivorans* (*Bsal*) exposure experiments. Asterisk (*) indicates collected as eggs from the wild.

Species	Lifestage	Order	Family	IUCN Conservation Status	Source	Location	Start Date (Mo-Yr)	Housing Conditions	Exposure Method	Treatments	n	qPCR Processing Location
<i>Ambystoma laterale</i>	Adult	Caudata	Ambystomatidae	Least Concern	Wild Caught	ME, USA	May-16	Terrestrial	Water Bath	Control 5x10e3 5x10e4 5x10e5 5x10e6	4 5 5 5	UT Knoxville
<i>Ambystoma maculatum</i>	Larval	Caudata	Ambystomatidae	Least Concern	Wild Caught	MA, USA	Aug-16	Aquatic	Water Bath	Control 5x10e3 5x10e4 5x10e5 5x10e6	 10	UMass Boston
<i>Ambystoma maculatum</i>	Juvenile	Caudata	Ambystomatidae	Least Concern	Wild Caught	MA, USA	Dec-16	Terrestrial	Water Bath	Control 5x10e3 5x10e4 5x10e5 5x10e6	 10	UMass Boston
<i>Ambystoma mexicanum</i>	Adult	Caudata	Ambystomatidae	Critically Endangered	Captive	MA, USA	Apr-17	Aquatic	Water Bath	Control 5x10e3 5x10e4 5x10e5 5x10e6	7 10 10 10	UT Knoxville
<i>Ambystoma opacum</i>	Adult	Caudata	Ambystomatidae	Least Concern	Wild Caught	TN, USA	Jan-16	Terrestrial	Water Bath	Control 5x10e3 5x10e4 5x10e5 5x10e6	10 10 10 10	UT Knoxville
<i>Anaxyrus americanus</i>	Juvenile	Anura	Bufonidae	Least Concern	Captive*	TN, USA	Oct-17	Terrestrial	Water Bath	Control 5x10e3 5x10e4 5x10e5 5x10e6	2 5	UT Knoxville
<i>Aneides aeneus</i>	Adult	Caudata	Plethodontidae	Near Threatened	Captive	Indoor Ecosystems LLC	Feb-17	Terrestrial	Water Bath	Control 5x10e3 5x10e4 5x10e5 5x10e6	7 10 10 10	UT Knoxville
<i>Atelopus zeteki</i>	Larval	Anura	Bufonidae	Critically Endangered	Captive	Maryland Zoo	Mar-19	Aquatic	Water Bath	Control 5x10e3 5x10e4 5x10e5 5x10e6	 10	UMass Boston
<i>Cryptobranchus alleganiensis</i>	Juvenile	Caudata	Cryptobranchidae	Near Threatened	Captive	Nashville Zoo	Oct-16	Aquatic	Pipette Exposure	Control 5x10e3 5x10e4 5x10e5 5x10e6	3 6 7 7	UT Knoxville
<i>Desmognathus aeneus</i>	Adult	Caudata	Plethodontidae	Near Threatened	Captive	Indoor Ecosystems LLC	Jul-16	Terrestrial	Water Bath	Control 5x10e3 5x10e4 5x10e5 5x10e6	5 10 10 10	UT Knoxville
<i>Desmognathus monticola</i>	Adult	Caudata	Plethodontidae	Least Concern	Wild Caught	GA, USA	Jul-16	Terrestrial	Water Bath	Control 5x10e3 5x10e4 5x10e5 5x10e6	8 10 10 10	UT Knoxville
<i>Desmognathus ocoee</i>	Adult	Caudata	Plethodontidae	Least Concern	Wild Caught	NC, USA	May-16	Terrestrial	Water Bath	Control 5x10e3 5x10e4 5x10e5 5x10e6	5 10 10 10	UT Knoxville
<i>Ensatina eschscholtzii klauberi</i>	Adult	Caudata	Plethodontidae	Least Concern	Captive	Indoor Ecosystems LLC	Mar-18	Terrestrial	Water Bath	Control 5x10e3 5x10e4 5x10e5 5x10e6	2 4 4 4	UT Knoxville

Table 2. Continued.

Species	Lifestage	Order	Family	IUCN Conservation Status	Source	Location	Start Date (Mo-Yr)	Housing Conditions	Exposure Method	Treatments	n	qPCR Processing Location
<i>Ensatina e. xanthoptica</i>	Adult	Caudata	Plethodontidae	Least Concern	Wild Caught	CA, USA	Mar-18	Terrestrial	Water Bath	Control 5x10e3 5x10e4 5x10e5 5x10e6	6 6 6 6 6	UT Knoxville
<i>Eurycea bislineata</i>	Larval	Caudata	Plethodontidae	Least Concern	Wild Caught	MA, USA	Aug-19 Oct-17	Aquatic	Water Bath	Control 5x10e3 5x10e4 5x10e5 5x10e6	3 5 5 5 7	UTK UMass Boston
<i>Eurycea bislineata</i>	Adult	Caudata	Plethodontidae	Least Concern	Wild Caught	MA, USA	Aug-19	Terrestrial	Water Bath	Control 5x10e3 5x10e4 5x10e5 5x10e6	8 9 9 9 9	UT Knoxville
<i>Eurycea lucifuga</i>	Adult	Caudata	Plethodontidae	Least Concern	Captive	Indoor Ecosystems LLC	Nov-17	Terrestrial	Water Bath	Control 5x10e3 5x10e4 5x10e5 5x10e6	2 5 5 5 5	UT Knoxville
<i>Eurycea wilderae</i>	Adult	Caudata	Plethodontidae	Least Concern	Wild / Captive	NC, VA, Indoor Ecosystems LLC	Oct-16 Oct-17 Nov-17	Terrestrial	Water Bath	Control 5x10e3 5x10e4 5x10e5 5x10e6	15 11 15 15 15	UT Knoxville
<i>Hemidactylium scutatum</i>	Adult	Caudata	Plethodontidae	Least Concern	Captive	Indoor Ecosystems LLC	Feb-17	Terrestrial	Water Bath	Control 5x10e3 5x10e4 5x10e5 5x10e6	5 5 5 5 5	UT Knoxville
<i>Hyla chrysoscelis</i>	Adult	Anura	Hylidae	Least Concern	Captive*	TN, USA	May-16	Terrestrial	Water Bath	Control 5x10e3 5x10e4 5x10e5 5x10e6	10 10 10 10 10	UT Knoxville
<i>Necturus maculosus</i>	Adult	Caudata	Proteidae	Least Concern	Wild Caught	OH, USA	Apr-16	Aquatic	Pipette Exposure	Control 5x10e3 5x10e4.5 5x10e6	2 5 4 5	UT Knoxville
<i>Notophthalmus meridionalis</i>	Adult	Caudata	Salamandridae	Endangered	Captive	Fort Worth Zoo	Apr-17	Terrestrial	Water Bath	Control 5x10e3 5x10e4 5x10e5 5x10e6	4 9 9 9 9	UT Knoxville
<i>Notophthalmus perstriatus</i>	Adult	Caudata	Salamandridae	Near Threatened	Captive	Omaha Zoo	Apr-17	Terrestrial	Water Bath	Control 5x10e3 5x10e4 5x10e5 5x10e6	10 10 10 10 10	UT Knoxville
<i>Notophthalmus viridescens</i>	Adult	Caudata	Salamandridae	Least Concern	Wild Caught	TN, USA	Oct-18	Aquatic	Water Bath	Control 5x10e3 5x10e4 5x10e5 5x10e6	5 10 10 10 10	UT Knoxville
<i>Notophthalmus viridescens</i>	Eft	Caudata	Salamandridae	Least Concern	Wild Caught	PA & MA, USA	Aug-19	Terrestrial	Water Bath	Control 5x10e3 5x10e4 5x10e5 5x10e6	5 5 5 5	UT Knoxville
<i>Notophthalmus viridescens</i>	Larval	Caudata	Salamandridae	Least Concern	Wild Caught	MA, USA	Aug-18	Aquatic	Water Bath	Control 5x10e3 5x10e4 5x10e5 5x10e6	12 . 12 12 12	UMass Boston
<i>Osteopilus septentrionalis</i>	Adult	Anura	Hylidae	Least Concern	Wild Caught	FL, USA	Jan-19	Terrestrial	Water Bath	Control 5x10e3 5x10e4 5x10e5 5x10e6	2 4 4 4 4	UT Knoxville

Table 2. Continued.

Species	Lifestage	Order	Family	IUCN Conservation Status	Source	Location	Start Date (Mo-Yr)	Housing Conditions	Exposure Method	Treatments	n	qPCR Processing Location
<i>Plethodon metcalfi</i>	Adult	Caudata	Plethodontidae	Least Concern	Wild Caught	NC, USA	Jul-16	Terrestrial	Water Bath	Control 5x10e3 5x10e4 5x10e5 5x10e6	5 10 10 10 10	UT Knoxville
<i>Plethodon shermani</i>	Adult	Caudata	Plethodontidae	Vulnerable	Wild Caught	NC, USA	Jan-16	Terrestrial	Water Bath	Control 5x10e3 5x10e4 5x10e5 5x10e6	7 7 7 7 7	UT Knoxville
<i>Pseudobranchius striatus</i>	Adult	Caudata	Sirenidae	Least Concern	Wild Caught	FL, USA	Jun-19	Aquatic	Water Bath	Control 5x10e3 5x10e4 5x10e5 5x10e6	5 5 5 5 5	UT Knoxville
<i>Pseudotriton ruber</i>	Adult	Caudata	Plethodontidae	Least Concern	Captive	Indoor Ecosystems LLC	Nov-17	Terrestrial	Water Bath	Control 5x10e3 5x10e4 5x10e5 5x10e6	5 5 5 5 5	UT Knoxville
<i>Rana catesbeiana</i>	Juvenile	Anura	Ranidae	Least Concern	Wild Caught	NC, USA	Mar-16	Aquatic	Water Bath	Control 5x10e3 5x10e4 5x10e5 5x10e6	6 6 6 6 6	UT Knoxville
<i>Rana catesbeiana</i>	Larval	Anura	Ranidae	Least Concern	Captive*	TN, USA	Nov-18	Terrestrial	Water Bath	Control 5x10e3 5x10e4 5x10e5 5x10e6	10 . 10 10 30	UMass Boston
<i>Rana chiricahuensis</i>	Juvenile	Anura	Ranidae	Vulnerable	Captive *	Fort Worth Zoo	Mar-16	Terrestrial	Water Bath	Control 5x10e3 5x10e4 5x10e5 5x10e6	8 8 8 8 8	UT Knoxville
<i>Rana pipiens</i>	Larval	Anura	Ranidae	Least Concern	Captive*	UMass Boston	Jul-19	Aquatic	Water Bath	Control 5x10e3 5x10e4 5x10e5 5x10e6	15 15 15 15 5	UMass Boston
<i>Rana pipiens</i>	Juvenile	Anura	Ranidae	Least Concern	Captive *	UMass Boston	Aug-19	Terrestrial	Water Bath	Control 5x10e3 5x10e4 5x10e5 5x10e6	 5	UMass Boston
<i>Rana sylvatica</i>	Juvenile	Anura	Ranidae	Least Concern	Captive*	TN, USA	Mar-16	Terrestrial	Water Bath	Control 5x10e3 5x10e4 5x10e5 5x10e6	5 5 5 5 5	UT Knoxville
<i>Scaphiopus holbrookii</i>	Juvenile	Anura	Scaphiopodidae	Least Concern	Captive*	TN, USA	Oct-17	Terrestrial	Water Bath	Control 5x10e3 5x10e4 5x10e5 5x10e6	5 5 5 5 5	UT Knoxville
<i>Siren lacertina</i>	Adult	Caudata	Sirenidae	Least Concern	Wild Caught	FL, USA	Jun-19	Aquatic	Pipette Exposure	Control 5x10e3 5x10e4 5x10e5 5x10e6	5 5 5 5 5	UT Knoxville
<i>Taricha granulosa</i>	Adult	Caudata	Salamandridae	Least Concern	Wild Caught	CA, USA	Sep-17	Terrestrial	Water Bath	Control 5x10e3 5x10e4 5x10e5 5x10e6	8 10 10 10 10	UT Knoxville
<i>Taricha torosa</i>	Adult	Caudata	Salamandridae	Least Concern	Wild Caught	CA, USA	Mar-18	Terrestrial	Water Bath	Control 1x10e3 1x10e4 1x10e5 1x10e6	10 10 10 10 10	Washington State University

Table 3. Parameter estimates for analysis of *Bsal* growth rate across species.*Random effects*

Group	Negative binomial Standard deviation	Correlation	Zero-inflated Standard deviation	Correlation
Species (intercept)	1.94		4.25	
Species (slope – Day of swab)	0.01	-0.36	0.05	-0.88
Individual (intercept)	2.73		2.18	

Fixed effects

Parameter	Negative binomial	Zero-inflated
(Intercept)	2.07	1.92
Day_of_Swab	0.12	-0.05
Treatment5x10 ⁴	0.72	-1.14
Treatment5x10 ⁵	4.27	-3.12
Treatment5x10 ⁶	5.29	0.34
Day_of_Swab:Treatment5x10 ⁴	0.06	-0.05
Day_of_Swab:Treatment5x10 ⁵	0.04	-0.06
Day_of_Swab:Treatment5x10 ⁶	0.17	-1.19

Table 4. Post-hoc tests for differences in *Bsal* growth rate between exposure doses across species. All tests two-sided with Tukey adjustments for multiple comparisons.

Contrast	Negative binomial			Zero-inflated		
	<i>t</i>	df	<i>P</i>	<i>t</i>	df	<i>P</i>
10 ³ - 10 ⁴	-2.71	1986	0.035	2.53	1986	0.055
10 ³ - 10 ⁵	-1.56	1986	0.40	2.40	1986	0.078
10 ³ - 10 ⁶	-3.15	1986	0.0091	2.35	1986	0.088
10 ⁴ - 10 ⁵	1.18	1986	0.64	0.63	1986	0.92
10 ⁴ - 10 ⁶	-2.08	1986	0.16	2.26	1986	0.11
10 ⁵ - 10 ⁶	-2.47	1986	0.065	2.22	1986	0.11

SUPPLEMENTAL NOTE 1

R Code for Bsal Detection Estimates

Calculation 1

N = 1283 # Total sample

n = 336 # Control subset

true_prevalence = 0.01 # True prevalence/probability of being infected

What is the probability of not detecting Bsal in 336 tested individuals. Assuming all individuals
have an equal probability of having Bsal

prob_of_detection = 1 - (1 - true_prevalence)^n

With true_prevalence = 0.05, the probability of detecting Bsal approx 1 -> you are going to detect it on
one of your control animals

With true_prevalence = 0.01, the probability of detection Bsal = 0.966 -> There is a < 5% change that
you falsely say no Bsal on controls

Check with a simulation

sims = 10000

detected = array(NA, sims)

```
for(i in 1:sims){  
  newts = rbinom(N, 1, true_prevalence)  
  samp_newts = any(sample(newts, n, replace=FALSE))  
  detected[i] = samp_newts  
}
```

#Probability of detecting Bsal

sum(detected) / sims

Same results as above: < 5% chance of failing to detect Bsal if it exists.

Calculation 2

Let's now say each individual has a different probability of having Bsal, but that the
average is 0.01 or 0.05

true_prevalence = 0.01

phi = 1 # Dispersion parameter of beta distribution

alpha = phi * true_prevalence

beta = phi*(1 - true_prevalence)

Check with a simulation

sims = 10000

detected = array(NA, sims)

```
for(i in 1:sims){
```

```

# Beta binomial simulation
probs = rbeta(N, alpha, beta)
newts = rbinom(N, size=1, prob=probs)
samp_newts = any(sample(newts, n, replace=FALSE))
detected[i] = samp_newts
}

```

```

# Probabiliy of detecting Bsal
sum(detected) / sims

```

```

# There is no substantial change -> there is still less than a 5% that you fail to detect
# Bsal given its presence.

```

SUPPLEMENTAL INFORMATION REFERENCES

- 1 White, C. L. *et al.* Amphibian: A case definition for Batrachochytrium salamandrivorans chytridiomycosis. *Herpetological Review* **47**, 207-207 (2016).
- 2 Thomas, V. *et al.* Recommendations on diagnostic tools for Batrachochytrium salamandrivorans. *Transbound Emerg Dis* **65**, e478-e488, doi:10.1111/tbed.12787 (2018).