

S-nitroso-N-acetylcysteine attenuates liver fibrosis in experimental nonalcoholic steatohepatitis

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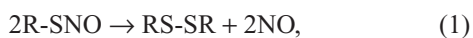
Abstract: S-Nitroso-N-acetylcysteine (SNAC) is a water soluble primary S-nitrosothiol capable of transferring and releasing nitric oxide and inducing several biochemical activities, including modulation of hepatic stellate cell activation. In this study, we evaluated the antifibrotic activity of SNAC in an animal model of nonalcoholic steatohepatitis (NASH) induced in Sprague-Dawley rats fed with a choline-deficient, high trans fat diet and exposed to diethylnitrosamine for 8 weeks. The rats were divided into three groups: SNAC, which received oral SNAC solution daily; NASH, which received the vehicle; and control, which received standard diet and vehicle. Genes related to fibrosis (matrix metalloproteinases [MMP]-13, -9, and -2), transforming growth factor β -1 [TGF β -1], collagen-1 α , and tissue inhibitors of metalloproteinase [TIMP-1 and -2] and oxidative stress (heat-shock proteins [HSP]-60 and -90) were evaluated. SNAC led to a 34.4% reduction in the collagen occupied area associated with upregulation of MMP-13 and -9 and downregulation of HSP-60, TIMP-2, TGF β -1, and collagen-1 α . These results indicate that oral SNAC administration may represent a potential antifibrotic treatment for NASH.

Keywords: nitric oxide, S-nitroso-N-acetylcysteine, fibrogenesis, NASH, diethylnitrosamine

Introduction

Liver fibrosis is one of the main outcomes of chronic nonalcoholic fatty liver disease (NAFLD), which usually precedes hepatocarcinogenesis.¹ The combined pathophysiological features of NAFLD and fibrosis associated hepatocarcinoma have stimulated studies for understanding the basic molecular and cellular mechanisms associated with chronic liver inflammation. Among the central inflammatory signaling pathways driving the transition from chronic liver injury to fibrosis and hepatocarcinoma, oxidative stress is considered to be involved in the onset and progression of NAFLD.^{2,3} Reactive oxygen species are known to activate hepatic stellate cells (HSCs), which leads to hepatic collagen deposition and fibrosis.⁴ In addition, changes in the extracellular matrix composition are accompanied by an imbalance between the rates of matrix synthesis and degradation which, in healthy livers, is maintained by a permanent turnover driven by matrix metalloproteinases (MMPs) and their corresponding tissue inhibitors of metalloproteinases (TIMPs).^{4,5} Chronic liver injury and activation of HSCs lead to upregulation of TIMP-1 and transforming growth factor β -1 (TGF β -1) expression, with inhibition of MMP activity, stimulation of collagen I synthesis, and accumulation of matrix proteins in the extracellular space.^{6,7} Therefore, evaluating the changes in the expression of genes involved in fibrogenesis with antioxidant based treatments in vivo may bring fundamental information regarding the role of oxidative injury in nonalcoholic steatohepatitis (NASH) and liver fibrosis.

Among its many biological actions, endogenous nitric oxide (NO) may be a protective species in inflammatory diseases associated with oxidative injury. Under oxidative stress conditions promoted by superoxide ($O_2^{\cdot-}$) production, NO is capable of exerting a protective action if it is in molar excess relative to $O_2^{\cdot-}$.⁸ One of the strategies for restoring the balance between NO and $O_2^{\cdot-}$ in favor of NO under inflammatory conditions is to supplement NO from an exogenous source. Several NO donor drugs have already been used for this purpose in experimental studies, including metal-nitrosyl complexes, organic nitrites and nitrates, and S-nitrosothiols (RSNOs), which possess the –S–NO moiety.⁹ One of the main synthetic RSNOs described in the literature as an exogenous NO source is S-nitroso-N-acetylpenicillamine (SNAP), which is a tertiary RSNO (where the S–NO moiety is bound to a tertiary carbon atom). SNAP offers the advantage of increased stability¹⁰ due to steric hindrance exerted by the substituent groups of the tertiary carbon atom, complicating the approach of two molecules to form a dimer in the NO release reaction. However, SNAP has low solubility in water, limiting its potential pharmaceutical application as an NO donor in humans. Conversely, several primary RSNOs (where the S–NO moiety is bound to a primary carbon atom) such as S-nitrosoglutathione (GSNO) and S-nitrosoalbumin are water soluble and are endogenously found in mammals with low toxicity. A simpler analog of GSNO is S-nitroso-N-acetylcysteine (SNAC), whose precursor molecule is N-acetylcysteine (NAC), a drug amply used as a mucolytic agent.¹¹ SNAC can be readily obtained from the S-nitrosation of NAC in aqueous media and can spontaneously release NO through homolytic S–N bond cleavage followed by dimerization through the reaction between thiyl radicals (Equation 1).



where $R = C_5H_6NO_3$. SNAC has been shown to exert hypotensive action following intravenous administration^{10,12} and to be protective against liver injury in different animal models of NAFLD and NASH.^{13–18} Therefore, we chose SNAC as a potential antifibrinogenic NO donor drug for treating fibrosis in an animal model of NASH. We used diethylnitrosamine (DEN), a well known alkylating agent with carcinogenic and mutagenic properties, in combination with a choline-deficient high fat diet, to induce NASH and liver fibrosis in Sprague-Dawley rats. The results obtained indicate that oral administration of SNAC may be a potential new antifibrotic therapy for patients with NASH.

Materials and methods

Materials

N-acetyl-L-cysteine (2-acetamido-3-sulfanylpropanoic acid, NAC), sodium nitrite, hydrochloric acid, sodium chloride, phosphate buffered saline (PBS) solution (pH 7.4), and DEN were purchased from Sigma-Aldrich Chemical Co (Milwaukee, WI, USA). Chloroform, isopropanol, and ethanol were obtained from Merck (Darmstadt, Germany). Reagents were used as received.

SNAC synthesis

SNAC was synthesized through S-nitrosation of NAC (40 mM) in an equimolar acidified sodium nitrite solution at pH 4.5. SNAC formation was monitored by ultraviolet-visible (UV-Vis) spectrophotometry based on the formation of its characteristic absorption bands with maxima at 336 and 545 nm using a diode array spectrophotometer (Hewlett-Packard, Model 8453, Palo Alto, CA, USA) referenced against air. A kinetic plot of the absorption at 545 nm in time intervals of 1 minute at 25°C was used for evaluating the time necessary for a complete reaction under stoichiometric conditions. Stock SNAC solutions were prepared at a concentration of 40 mM, and then further diluted to 1 mM in PBS (pH 7.4) for oral administration to the animals. Only freshly prepared SNAC solutions were used in this study.

Thermal decomposition of SNAC solution in simulated gastric fluid

Simulated gastric fluid (SGF) was prepared by dissolving of 2 grams of sodium chloride and 7 mL of 37% hydrochloric acid in a final volume of 1 L of distilled water. The pH of this solution was 1.5. Decomposition of 1.0 mM SNAC incubated in SGF at 37°C was spectrophotometrically characterized over 8 hours by monitoring the spectral changes in the range 220–1100 nm at time intervals of 15 minutes in the dark as described above. Kinetic curves of the decomposition of 1 mM SNAC in SGF were obtained from the absorption changes at 336 nm.

Animals

Adult Sprague-Dawley rats, each weighing 250–300 g, were housed in temperature and humidity controlled rooms under 12 hour light/dark cycles with food and water ad libitum. All procedures for animal experimentation were followed in accordance with the local Animal Experimentation Ethics Committee Guidelines for the Care and Use of Laboratory Animals at the School of Medicine of the University of São Paulo.

NASH induction

NASH with fibrosis was induced in animals with a choline-deficient, high fat diet (35% total fat, 54% trans fatty enriched; Rhostrer Ltda, Brazil) and the administration of 13–15 mg/day of DEN added in the drinking water at a concentration of 135 mg/L for 8 weeks.¹⁹ The animals were randomized and divided into three groups: NASH (n = 10), SNAC (n = 10), and control (n = 3). The NASH and SNAC groups received the choline-deficient, high fat diet and DEN; the SNAC group received SNAC daily by gavage (8.0 μ mol/kg = 1.4 mg/kg) and NASH animals received the vehicle (Ringer's solution). Control animals were fed ad libitum with a standard diet (Nuvilab® Nutrientes Ltda, Colombo, Brazil) and received the vehicle through gavage. After 8 weeks, the animals were anesthetized with intraperitoneal ketamine (0.1 mL/kg) and sacrificed (Figure 1). Samples of liver tissue were obtained for collagen morphometry, nitrotyrosine immunohistochemistry, and gene expression analysis. The association of DEN with a choline-deficient, high fat diet was intended to promote advanced fibrosis within a relative short time in the context of experimental steatohepatitis, as previously published.¹⁹

Collagen morphometry

Collagen was quantified in the histological sections using the picrosirius red staining method with sections examined using an optical microscope (Nikon E-800, Shinagawa-Ku, Tokyo, Japan). Quantification was performed in ten different histological fields of greater collagen deposition of the same histological section considering the heterogeneous distribution of injury in the samples and the different damage patterns.²⁰ This criterion was standardized for all animals in a blinded manner. The collagen fiber area was quantified using the Image Pro-Plus 4.5 software (Media Cybernetics

Inc, Warrendale, PA, USA), and the means and standard deviations were calculated. The results were expressed as the percentage of the area occupied by collagen in the liver parenchyma of each animal, relative to the total field.

Nitrotyrosine immunohistochemistry

Nitrotyrosine immunohistochemistry was performed to assess whether SNAC administration increased peroxynitrite levels in the liver, since NO released by SNAC, in combination with superoxide might lead to peroxynitrite formation. Liver slides were subjected to the antigen retrieval process in citric buffer (10 mM, pH 6.0) in a pressure cooker for 3.5 minutes. After blockade with 6% H₂O₂ solution, slides were incubated in a humidifying chamber overnight at 4°C with mouse anti-nitrotyrosine monoclonal antibody (A-21285; Life Technologies, Carlsbad, CA, USA; 1:200). Samples were then incubated with SuperPicture Poly HRP conjugate (Zymed, San Francisco, CA, USA) for 30 minutes at 37°C, developed in diaminobenzidine (DAB) solution (Sigma, St Louis, MO, USA), and counterstained with Harris hematoxylin. Evaluation consisted of counting positive nitrotyrosine immunostaining hepatocytes per mm² of tissue. The intensity of immunostaining was scored on a 4-tiered scale (0 to 3+), with 0 indicating no staining, 1+ indicating weak staining, 2+ indicating moderate staining, and 3+ indicating strong staining.

Tissue RNA extraction

Liver tissue samples, received in liquid nitrogen, were fragmented in a tissue pulverizer (Mikro-Dismembrenator II, B. Braun, Melsungen, Germany). After the pulverization, 1.0 mL of TRIzol® was added (Life Technologies) to the mixture, which was incubated for 5 minutes at room

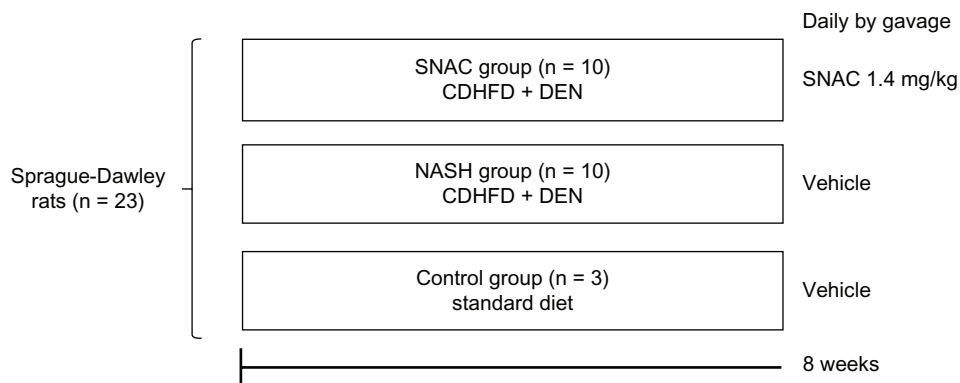


Figure 1 Scheme of the study design describing the three groups of animals treated over eight weeks with: standard diet and oral vehicle (control group); choline-deficient high fat diet plus diethylnitrosamine (CDHFD + DEN) and oral vehicle (nonalcoholic steatohepatitis group); and CDHFD + DEN and oral S-nitroso-N-acetylcysteine (SNAC) solution (1.4 mg/kg) (SNAC group).

Abbreviations: CDHFD, choline-deficient high fat diet; DEN, diethylnitrosamine; NASH, nonalcoholic steatohepatitis; SNAC, S-nitroso-N-acetylcysteine.

temperature. Two hundred microliters of chloroform was added to the solution, which was stirred vigorously for 15 seconds, and then incubated for 5 minutes at room temperature. The solution was then centrifuged for 15 minutes at 12,000 rpm at 4°C (Eppendorf 5417-R, Hamburg, Germany). The supernatant was transferred to a new 1.5 mL sterile tube, and RNA was precipitated with 500 μ L of isopropanol. The mixture was allowed to stand for 10 minutes and centrifuged for 15 minutes at 12,000 rpm at 4°C. The supernatant was removed, and the RNA at the bottom was washed with 1.0 mL of ethanol (70%), centrifuged again, and resuspended in an appropriate volume of sterile water (UltraPure™ DNase/RNase-Free Distilled Water [Life Technologies]).

Real time quantitative reverse transcription polymerase chain reaction (RT-qPCR)

The RT-qPCR reactions for TIMP-1 and -2, MMP-9, and HSP-60 and -90 were performed using the Rotor-Gene RG-3000 (Corbett Research, Sydney, Australia) and the reagent set of the SuperScript™ III Platinum® One-Step Quantitative RT-PCR System (Life Technologies) as recommended by the manufacturer. Gene expression intensities were obtained using threshold cycle (CT) values, in which the signal increase associated with the fragment amplification exponential phase begins to be detected. The reaction for each analyzed gene was carefully standardized to avoid nonspecific amplification. The annealing temperature for amplification of all genes was 60°C. All RT-qPCR reactions were performed in duplicate for each sample of liver tissue for both the target gene and for the control, β -actin. The primers for the genes of TIMP-1, TIMP-2, MMP-9, HSP-60, HSP-90, and β -actin were designed using the Primer3 Input program (<http://frodo.wi.mit.edu/>). The β -actin gene was used as an endogenous control. The sequences of primers for each gene were as follows: – HSP-60 forward: AGC AAA GGG GCT AAT CCA GT; HSP-60 reverse: TGA CAC CCT TTC TTC CAA CC; – HSP-9 forward: GAT TGA CAT CAT CCC CAA CC; HSP-90 reverse: CTG CTC ATC ATC GTT GTG CT; – TIMP-1 forward: TCC CCA GAA ATC ATC GAG AC; TIMP-1 reverse: TCA GAT TAT GCC AGG GAA CC; – TIMP-2 forward: GCA TCA CCC AGA AGA AGA GC; TIMP-2 reverse: GGG TCC TCG ATG TCA AGA AA; – MMP-9 forward: CAA ACC CTG CGT ATT TCC AT; MMP-9 reverse: AGA GTA CTG CTT GCC CAG GA.

For MMP-2, MMP-13, TGF β -1, and collagen-1 α , total RNA was reverse transcribed to yield cDNA and preamplified using the TaqMan® PreAmp Master Mix Kit (Applied Biosystems, Foster City, CA, USA). Profibrogenic and fibrolytic related genes were analyzed by RT-qPCR (ABI PRISM® 7000, Applied Biosystems). α -Actin gene expression was used as a housekeeping endogenous control. All primers and probes were purchased from Applied Biosystems (Assay ID: β -actin: 4331348; collagen-1 α : Rn00801649_q1; TGF β -1: Rn00572010_m1; MMP-2: Rn01538167_m1; TIMP-1: Rn00587558_m1 and MMP-13: Rn01448191_g1). Analysis of relative gene expression data was performed according to the 2(-Delta Delta C[T]) method²¹ and was expressed as fold increase over values obtained from control rats.

Statistical analysis

All data were expressed as the mean or median depending on the variable distribution, with standard deviation (SD) values. For Gaussian distribution variables, *t*-test, one-way ANOVA, and Newman–Keuls Multiple Comparison Posttest were used. For non Gaussian distribution variables, Mann–Whitney, Kruskal–Wallis, and Dunn’s Multiple Comparison Posttests were used. A *P*-value less than 0.05 was deemed statistically significant. All calculations and graphics were made using SPSS 17.0 (SPSS Inc, Chicago, IL, USA) and GraphPad Prism v5.0 (GraphPad Software Inc, La Jolla, CA, USA).

Results

Kinetics of SNAC formation and decomposition

Figure 2A and B show the UV-Vis absorption spectra of SNAC obtained in the S-nitrosation reaction of NAC. The absorption bands at 336 and 545 nm are assigned to the $\pi \rightarrow \pi$ and $n_N \rightarrow \pi$ electronic transitions of the SNO moiety of the molecule, respectively,²² confirming SNAC formation. Figure 2C shows a representative kinetic curve of SNAC formation during its synthesis from NAC at pH 4.5 and 25°C, based on monitoring of the absorption band at 545 nm. The reaction was observed to be relatively fast under this condition, with a yield of approximately 90% after 3 minutes.

Figure 2D shows the kinetic curve of 1 mM SNAC decomposition in SGF (pH 1.5) at 37°C over 4 hours, based on its absorption band at 336 nm. The disappearance of this band gives a real time measurement of the SNAC decomposition in this medium. The intensity of the band was

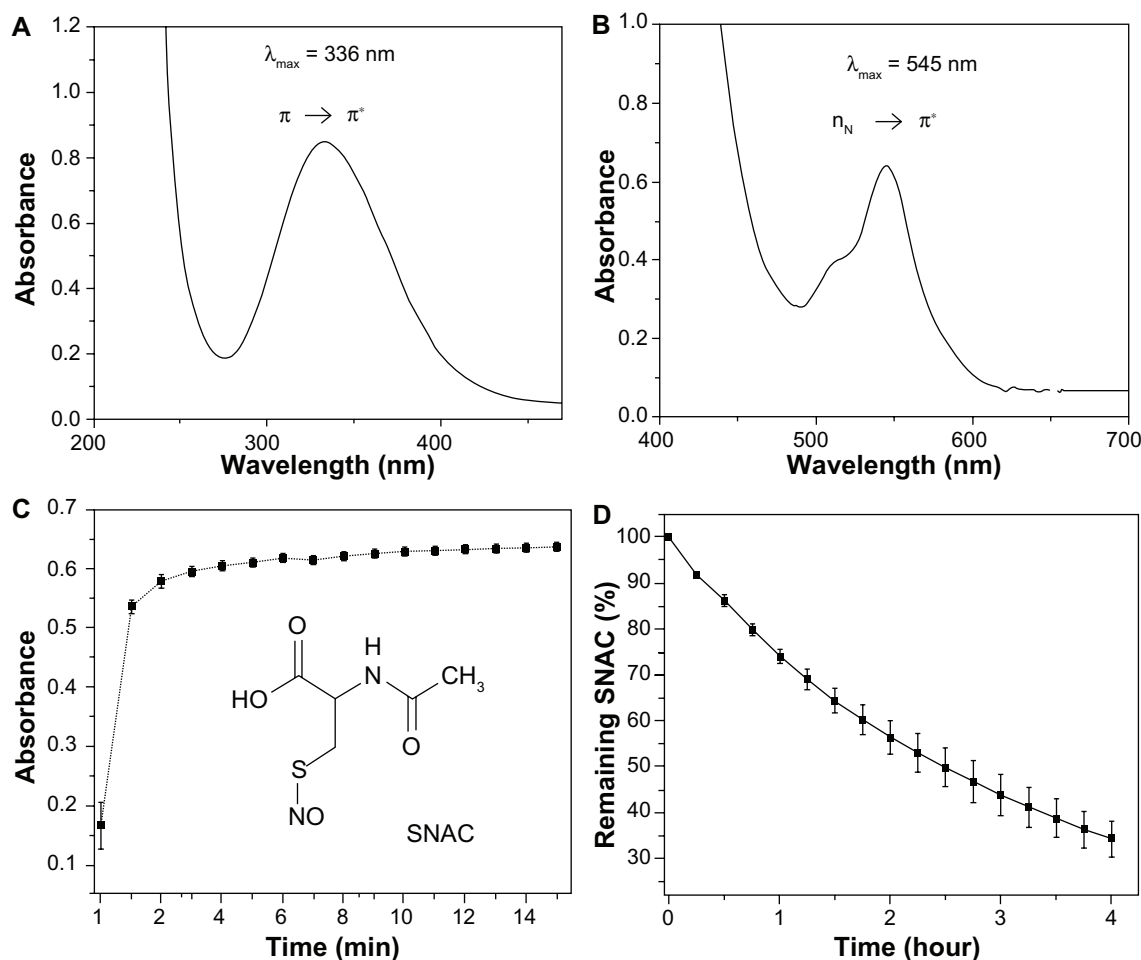


Figure 2 (A and B) Ultraviolet-visible spectral changes associated with the formation of S-nitroso-N-acetylcysteine (SNAC) through the S-nitrosation of N-acetylcysteine (NAC) with nitrous acid showing the two absorption bands characteristic of primary S-nitrosothiols with maxima at 336 and 545 nm. The concentration of the SNAC solution is 1 and 40 mM, respectively. **(C)** Kinetic curve of SNAC formation during the S-nitrosation of NAC based on the absorption band of SNAC at 545 nm. Inset: SNAC structure. **(D)** Kinetic curve of SNAC decomposition in simulated gastric fluid at a concentration of 1 mM, pH 1.5 and 37°C over 4 hours based on the absorption band of SNAC at 336 nm.

Note: *Excited state of a molecular orbital.

shown to be decreased by approximately 40% after 2 hours, which represents the time estimated for gastric emptying of approximately 50% of the ingested food.²³

Dietary ingestion and weight gain

NASH animals showed a lower median dietary intake when compared with SNAC animals (10.83 mg \pm 0.56 versus 11.99 mg \pm 1.27, respectively; $P = 0.0002$), and there was no difference in weight gain ($P = 0.393$).

Collagen morphometry

The percentages of the collagen occupied area relative to the total field were 0.33 ± 0.07 for the control group, 1.75 ± 0.27 for the SNAC group, and 2.48 ± 0.34 for the NASH group. The differences in collagen quantification were highly significant between all groups ($P < 0.0001$), with SNAC treated animals

showing a reduction of 34.4% in the collagen occupied area when compared with the NASH group (Figure 3).

Nitrotyrosine immunohistochemistry

No immunostaining reaction for nitrotyrosine was observed in normal liver sections. In liver sections of the NASH and SNAC groups, nitrotyrosine produced weak to moderate diffuse cytoplasmic immunostaining reactions for nitrotyrosine in all hepatocytes. However, there were no differences between the NASH and SNAC groups (Figure 4).

Expression of genes involved in fibrogenesis

The median expression of MMP-13 was significantly higher in the SNAC group when compared with the NASH group [1.70 (1.15–2.82) versus 0.98 (0.71–1.3), respectively; $P = 0.0005$],

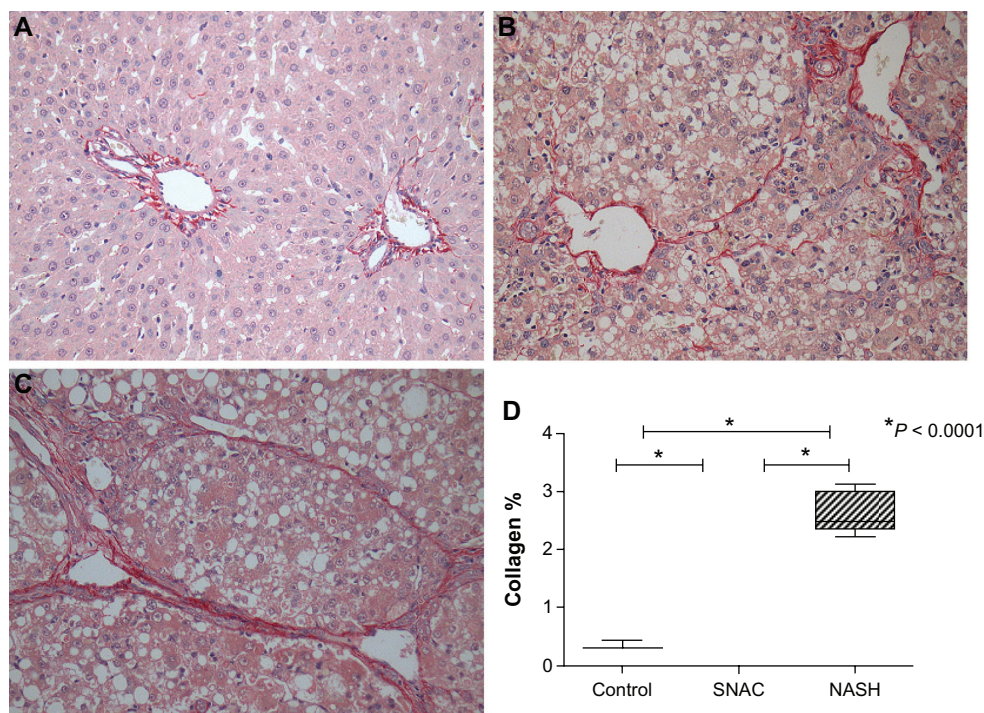


Figure 3 Representative picosirius red staining images of hepatic histology of (A) control, (B) nonalcoholic steatohepatitis (NASH), and (C) S-nitroso-N-acetylcysteine (SNAC) treated animals. Magnification is $\times 200$. (D) Collagen quantification of the control, SNAC, and NASH groups.

and the same was noted regarding MMP-9 expression [1.90 (1.156–2.23) versus 0.81 (0.52–1.05), respectively; $P < 0.0001$]. In addition, the expression of TIMP-2 was lower in the SNAC group than in the NASH group [0.68 (0.48–0.88) versus 1.04 (0.91–3.78), respectively; $P < 0.0001$]. No differences were found in the expression of TIMP-1 and MMP-2 between the two groups ($P = 0.842$ and $P = 0.123$, respectively). There was a marked reduction in TGF β -1 expression in SNAC treated animals [0.61 (0.35–0.80) versus 0.92 (0.41–1.60), respectively; $P = 0.0089$], and the same was noted regarding collagen-1 α expression [0.59 (0.28–0.90) versus 0.93 (0.52–1.55), respectively; $P = 0.0207$] (Figures 5 and 6).

Expression of genes involved in oxidative stress

As shown in Figure 6, HSP-60 was significantly less expressed in the SNAC group [0.78 (0.75–0.81)] when compared with the NASH group [median: 0.98 (0.95–1.04); $P < 0.0001$]. However, there was no difference in HSP-90 expression between these groups [1.00 (0.97–1.07) versus 0.99 (0.94–1.02), respectively; $P = 0.326$].

Discussion

SNAC formation from the S-nitrosation of NAC in aqueous acidic sodium nitrite solution is a fast reaction, allowing the

easy preparation of SNAC immediately before use, and this is an important point for therapeutic purposes. The reaction can be written as follows:



where R = C₅H₆NO₃, and nitrous acid (HONO) is formed from the protonation of nitrite according to Equation 3:



In the SNAC structure, an acetyl group is attached to the nitrogen atom of an S-nitrosated cysteine molecule (Figure 2C, inset). Calculations described elsewhere¹³ have shown that SNAC may establish an intramolecular hydrogen bond between the hydrogen atom of the amino group and the oxygen atom of the NO group in a seven-membered ring, favored by a second hydrogen bond between the hydrogen of the carboxyl group and the carbonyl oxygen of the acetate group. In this optimized geometry, the SNO group remains sterically unprotected and may undergo oxidation and dimerization reactions. In the last case, spontaneous S-N bond cleavage is favored by the low dissociation energy of the S-N bond (152.9 kJ mol⁻¹)¹³ and this reaction can be considered the main pathway of NO release by this molecule (Equation 1).

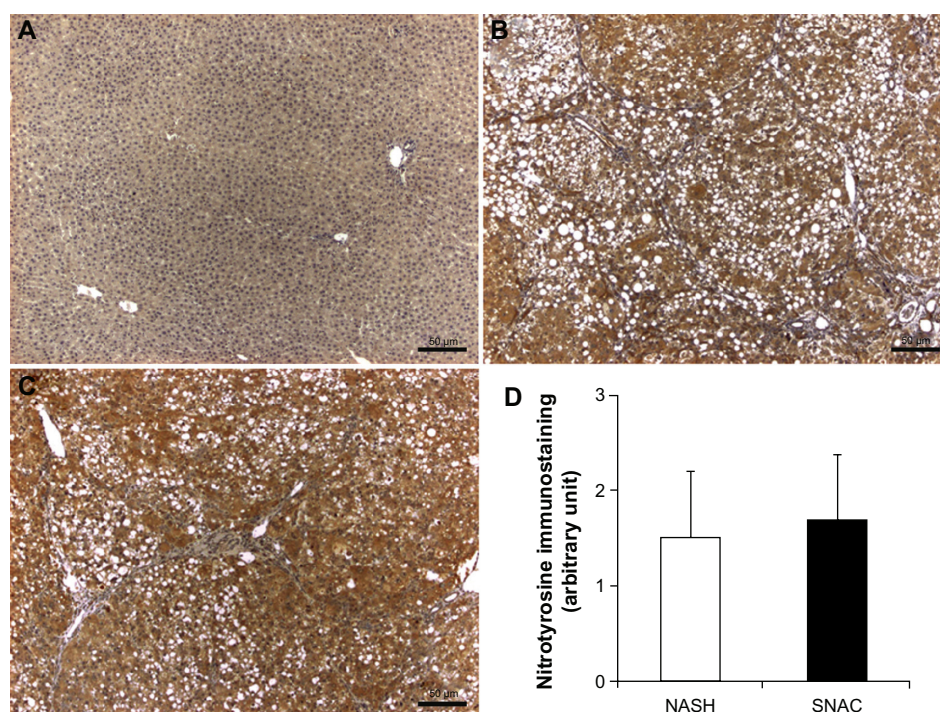


Figure 4 Representative nitrotyrosine immunostaining of liver sections.

Notes: Absent nitrotyrosine staining or hepatocyte injury is seen in the control group (A). Intense positivity is seen in both nonalcoholic steatohepatitis (NASH) (B) and S-nitroso-N-acetylcysteine (SNAC) (C) groups. (D) Nitrotyrosine immunostaining quantification of the NASH and SNAC groups.

In the particular condition of the stomach medium, SNAC decomposition is expected to lead to the formation of HONO and NAC (a free thiol, RSH):



Both intact SNAC and HONO formed in the acid catalyzed decomposition reaction are nitrosating agents, capable of modifying enzymes through transnitrosation reactions.²⁴ SNAC is also expected to undergo homolytic bond cleavage with release of free NO (Equation 1), which may cross the gastric epithelium easily to exert a gastric vasodilator effect. Finally, NAC may also exert antioxidant actions.¹¹ Therefore, oral administration of SNAC leads to a mixture of species capable of exerting nitrosating, antioxidant, and vasodilator actions in the gastric mucosa. The decomposition of approximately 40% of SNAC in SGF after 2 hours means that most of its biochemical actions are expected to be maintained during its residence time in the stomach (Figure 2D). Diffusion of intact SNAC through the gastrointestinal mucosa may thus propagate these actions systemically, reaching the liver, where a local reduction of oxidative stress may be achieved.

Previous reports have demonstrated that SNAC can induce the differentiation of myofibroblasts into a lipocyte phenotype

in a murine hepatic stellate cell line, with concomitant reduction of profibrogenic molecules and upregulation of MMP-13.²⁵ In the present study, we demonstrate in vivo for the first time that SNAC displays antifibrotic activity in an animal model of DEN induced NASH and that this action is associated with downregulation of profibrogenic molecules and upregulation of MMP-13, the latter of which plays a key role in the degradation of interstitial collagen in liver fibrosis. The smaller dietary intake of animals not receiving SNAC, compared with those receiving SNAC, can be attributed to a worse health condition that developed in NASH during the treatment, considering that these animals had more hepatic collagen deposition at the end of the study.

Collagen quantification by collagen fiber morphometry, which shows a good correlation between the amounts of tissue bound picrosirius red staining quantification,²⁶ demonstrated the beneficial effects of SNAC over fibrogenesis because SNAC treated animals showed a 34.4% reduction in the relative volume occupied by collagen when compared with the NASH group ($P < 0.0001$, Figure 3D).

The attenuation of fibrosis promoted by SNAC in this particular experimental model is in accordance with that in previous studies,^{14,16,18} which showed that SNAC could positively interfere with known triggers of liver fibrosis in NASH/NAFLD pathogenesis.^{3,6} The beneficial effects of

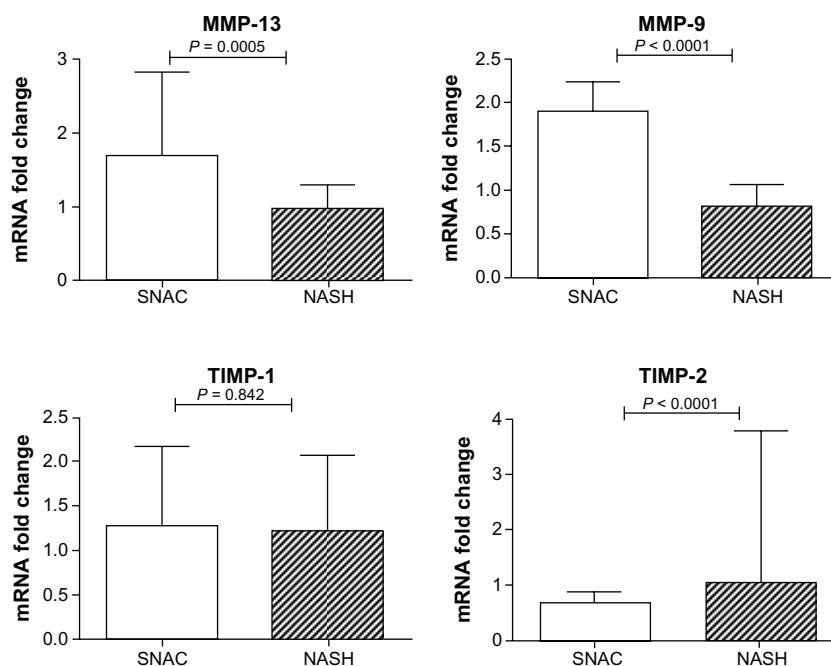


Figure 5 Fold change in matrix metalloproteinase (MMP)-13 and -9, and tissue inhibitor of metalloproteinases (TIMP)-1 and -2 mRNA expression in the nonalcoholic steatohepatitis groups against S-nitroso-N-acetylcysteine groups.

Abbreviations: NASH, nonalcoholic steatohepatitis; SNAC, S-nitroso-N-acetylcysteine.

SNAC observed in the present work are also in agreement with results already reported in an experimental biliary cirrhosis model, where the authors demonstrated that SNAC treated animals displayed lower accumulation of histologically detectable fibrosis, accompanied by downregulation of

collagen-1 α and decreased expression of smooth muscle specific α actin (α SMA).²⁷ More recently, other reports have shown that SNAC can induce differentiation of myofibroblasts into a lipocyte phenotype in the GRX cell line and decrease α SMA gene expression.²⁵ It must be

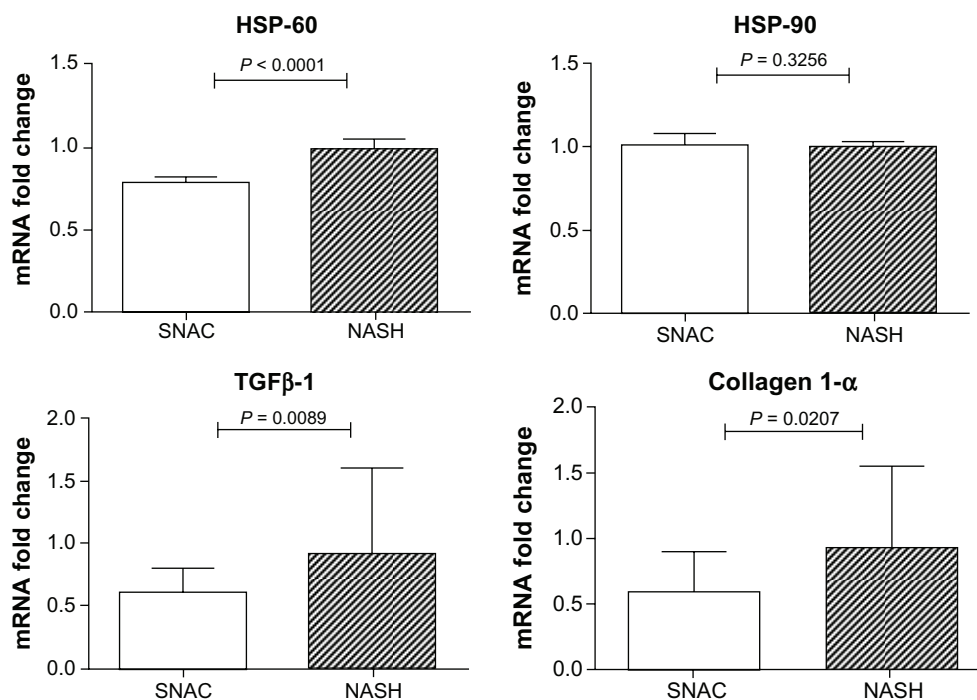


Figure 6 Fold change in heat-shock protein (HSP)-60 and -90, transforming growth factor β -1, and collagen 1 α mRNA expression in the nonalcoholic steatohepatitis groups against S-nitroso-N-acetylcysteine groups.

Abbreviations: NASH, nonalcoholic steatohepatitis; SNAC, S-nitroso-N-acetylcysteine.

highlighted that SNAC did not negatively interfere with the regulation of TIMP-1 but led to diminished expression of TIMP-2 because a significant reduction in TIMP-2 was also observed in previous reports during fibrosis resolution^{28,29} with an associated increase in the overall collagenase activity and extracellular matrix degradation.^{30,31} Data from chronic toxic liver injury in mice also sets TIMP-2 as an MMP-2 activator, leading to pathological collagen degradation, and consequently, degradation of normal liver matrix.³² Our results are consistent with this concept because SNAC did not upregulate MMP-2 expression and downregulated TIMP-2 expression, leading to an antifibrogenic gene expression profile (Figure 5).

One of the most intriguing findings reflecting SNAC's beneficial effect over liver fibrosis was the upregulation of MMP-13 in SNAC treated animals (Figure 5). MMP-13 has been characterized by its ability to degrade fibrillar collagens, such as collagen type I, the main collagen deposited in liver fibrosis. Previous studies have emphasized the importance of collagen type I initial cleavage for the overall regression of liver fibrosis because mice expressing a mutated, cleavage resistant collagen type I were unable to resolve fibrosis.^{33,34} Experimental studies have also shown that MMP-13 expression is markedly enhanced during the recovery phase of liver fibrosis.⁵ It is also noteworthy that SNAC induced downregulation of TGF- β 1, the most potent stimulus for collagen I production, and downregulated collagen-1 α mRNA expression, two important additional findings of SNAC's ability to attenuate hepatic fibrosis. The absence of TIMP-1 downregulation associated with upregulation of MMP-13 observed in SNAC treated animals may be justified by SNAC induced downregulation of TIMP-2 and suppression of TGF- β . SNAC treated animals also showed increased expression of MMP-9. This MMP itself exhibits no activity against collagen type I. Nevertheless, an indirect involvement of MMP-9 in fibrolysis has been assumed because MMP-9 expression increases at the beginning of fibrolysis.³⁵⁻³⁷ A possible mechanism may involve an indirect contribution of MMP-9 to fibrolysis by accelerating HSC apoptosis.³⁸ In line with its known antioxidant effects, we found that SNAC led to decreased expression of HSP-60, an oxidative stress related gene, suggesting a possible protective effect of SNAC in oxidative stress and corroborating the collagen quantification performed in the present study (Figure 6).^{39,40} Although these findings, based on mRNA expression levels, show a coherent set of changes associated with the attenuation of liver collagen deposition in the SNAC treated animals, it must be taken into account that mRNA

expression levels are not always directly correlated with protein expression levels and that our mRNA level results may be only correlative, instead of causative. Therefore, further studies to measure both mRNA expression levels and protein concentrations are needed to establish the existing correlations between changes in mRNA expression induced by SNAC treatment and protein expression.

The beneficial effects of oral SNAC administration over liver fibrosis may be associated with either direct antioxidant action of SNAC in primary reactions with reactive oxygen species or the secondary reaction of free NO released in a primary enzymatic reduction of SNAC to NAC plus NO (Equation 1). In the second case, the formed NAC may also exert a protective effect as already described in the literature.^{39,40} It must be noted that although NAC is a notorious antioxidant, SNAC is already reported to have a more effective antioxidant action than NAC in *in vitro*¹⁵ and *in vivo*.¹⁴ In the last case, oral SNAC administration in an animal model of NAFLD led to a significant decrease in the concentration of hydroperoxides in the liver homogenate when compared with the control group or with the administration of NAC at a dose five times higher. The protective action of SNAC in the present study may be due, in part, to its ability to act as an exogenous source of NO, leading to an increased local or systemic availability of NO. A common concern in the administration of exogenous NO in oxidative stress conditions is that NO may lead to increased peroxynitrite (OONO⁻) production with a consequent increase in nitrotyrosine formation (a marker of OONO⁻ production). The intense positivity of nitrotyrosine staining for the NASH and SNAC groups observed in Figure 4B and C is expected, since both groups developed NASH with liver injury. The absence of difference in the nitrotyrosine staining between the NASH and SNAC groups (Figure 4D) is evidence that SNAC administration did not lead to a significant increase in OONO⁻ formation due to a possible increase in hepatic NO concentration, as might be expected.

In conclusion, we show for the first time that oral administration of SNAC leads to an antifibrogenic effect in experimental NASH. Because longitudinal outcome studies have shown the prognostic importance of liver fibrosis for the natural history of NAFLD,^{41,42} these findings may represent a possible strategy to attenuate fibrosis in NASH.

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Disclosure

The authors declare no conflict of interest in this work.

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