Research Article

Evaluating the Risk of Tumors Diseases Based on Measurement of Urinary and Serumal Antioxidants Using the New Agar Diffusion Methods

Ying Zhou,¹ Jing Chen,^{1,2} Zhen Wang,¹ and Hui Liu¹

¹College of Medical Laboratory, Dalian Medical University, Dalian 116044, China
 ²School of Chemistry and Chemical Engineering, Shandong University, Jinan 250100, China

Correspondence should be addressed to Hui Liu; liuhui60@sina.com

Received 4 November 2016; Accepted 22 February 2017; Published 28 March 2017

Academic Editor: Marina Sokovic

Copyright © 2017 Ying Zhou et al. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Objectives. To discuss the characteristics of the amount of urinary total antioxidants in tumor diseases and the possibility of utilizing the changing regulation of urinary antioxidants to diagnose tumor diseases. *Method.* Urine and serum specimens from 130 healthy people were used to investigate the variation of antioxidant capacity against age. Urine and serum specimens from 44 unselected patients with tumors and 44 healthy people with same age background were used to explore the significance of urinary antioxidant capacity in clinic to diagnose tumor diseases. Potassium permanganate agar method and iodine starch method were used to determine the amount of total antioxidants. *Results.* In healthy people, more antioxidants in urine were measured in older people, while the results were opposite in serum. More antioxidants were found in urine of tumor patients than in healthy people with same age-range. *Conclusions.* According to the results of 130 measurements, the amount of antioxidants in urine varies by age. By using agar methods to measure antioxidants, the effect of age is required to be considered. Antioxidants levels from tumor patients were significantly higher than healthy individuals in urine. The combination of urine and serum to determine total antioxidants can better diagnose tumor diseases based on iodine starch method, with area under the receiver operating characteristics curve at 0.787.

1. Introduction

Biological free radicals are general products of metabolism, mainly containing reactive oxygen species (ROS) and reactive nitrogen species (RNS). In common condition, the free radicals producing and removing are kept balanced, which play an important role in biological system. Once the balance is disturbed, free radicals in body can damage cells, tissues, and organs and further cause aging, cardiac diseases, brain diseases, and cancer [1, 2]. Thus, it is of great importance to know the total antioxidant capacity (TAC) of an organism to evaluate its free radicals producing/removing balance.

Nowadays, many researches are investigating the serum TAC [3], but few are focused on the urinary TAC. Human urine metabolome [4] has pointed out that thousands of compounds are detected in urine, including wide-ranged concentrations of urea, urobilinogen, inorganic salts, creatinine, ammonia, organic acids, and water-soluble toxins. As an

important and easily accessible biological fluid, urine reflects the continuously changing environment of an organism [5]. Nevertheless, it has been proved that the level of urea [6], bilirubin [7, 8], and creatinine [9] in serum is associated with the TAC of an organism. So the measurement of antioxidants in urine is important; the variation of urinary metabolite profiling especially the TAC profiling may reveal specific disease.

In tumor patients, obvious oxidative stress is observed, indicating the balance between oxidants and antioxidants is broken up in each studied kind of tumors. In the development of tumors, there exists superfluous generation of ROS and RNS in organism [10]. ROS and RNS can cause DNA damage, protein damage, and lipid peroxidation [11–14]. ROS causes overexpression of Jun gene in lung cancer [15], while the increase of RNS causes protein damage in liver cancer [16]. Free radicals are involved in the initial, enhancement, and accumulation stage of tumor cells developing [17]. And the antioxidants in vivo can fight against with the surplus of free radicals. Low level of antioxidants bilirubin increases the risk of tumor related with smoking and alcohol [18, 19], while antioxidants uric acid is associated with DNA damage [20]. The amount of all the antioxidants in vivo reflects the capacity to react with the surplus of free radicals. However, it is unable to measure all the antioxidants because how many compounds are included in the antioxidants list is still unclear. Now, any proposed methods only cover a subset of the total antioxidants, which cannot truly reflect the redox state of an organism. A method that can measure the amount of total antioxidants in vivo is necessary.

Total antioxidants were previously investigated based on potassium permanganate agar and iodine starch agar method [21, 22]. In both methods, the diffusion area reflects TAC in an organism, and larger diffusion area represents better TAC. Both methods can better reflect the state of TAC than other methods such as ferric reducing antioxidant power assay [23, 24], cupric ion reducing antioxidant capacity assay [25, 26], and 2,2-diphenyl-1-picrylhydrazyl assay [27, 28].

In this work, potassium permanganate agar method and iodine starch agar method were used to determine the antioxidants in urine and serum, with the aim of applying antioxidant capacity to distinguish healthy people and tumor patients and to evaluate the risk of tumor diseases in clinic in a simpler way.

2. Materials and Method

2.1. The Variation of Total Antioxidants Measured in Urine and Serum in Healthy People

2.1.1. Specimens. Urine specimens from 130 healthy people were collected from the first hospital affiliated with Dalian Medical University and the second hospital affiliated with Dalian Medical University. The urine specimens were divided into 13 groups equally for every five years; each group contained 5 males and 5 females; the ages of subjects ranged from 20 to 85 years. Serum specimens were also collected from the above. Urine and serum specimens were stored at -20° C after being collected and melted in room temperature before experiment.

2.1.2. Method. The amount of total antioxidants in urine was determined by potassium permanganate agar method [21] and iodine starch agar method [22], respectively. Serum specimens were prepared and measured in the same way.

2.2. The Clinical Significance in Measuring the Amount of Total Antioxidants in Urine and Serum

2.2.1. Specimens. Urine specimens were collected from the first hospital affiliated with Dalian Medical University and the second hospital affiliated with Dalian Medical University. Forty-four unselected patients with tumors were regarded as experimental group, including 19 males and 25 females, mean age at 57.84 \pm 10.94 years. Forty-four healthy people were collected as control group, including 31 males and 13 females, mean age at 57.80 \pm 11.45 years. The diagnoses of experimental group include gastric cancer, intestinal cancer,

lung cancer, and breast cancer. Serum specimens were also collected from the above. Urine and serum specimens were stored at -20° C after being collected and restored in room temperature before experiment.

2.2.2. Method. This section was the same as Section 2.1.2.

2.3. Statistical Analysis. Nonparametric correction was used to analyze the variation of antioxidant capacity in urine as well as serum. Nonparametric test was used to compare the results of urine diffusion. *T*-test was used to compare the results of serum diffusion. Receiver operating characteristics (ROC) curves were constructed to assess sensitivity, specificity, and respective areas under the curves (AUCs) with 95% confidence interval (CI). A value of p < 0.05 (two tailed) was considered significant. Statistical software package SPSS 13.0 was used to evaluate the results.

2.4. *Ethical Approval.* This article does not contain any studies with human participants or animals performed. The protocol has been approved by the Ethical Committee of Dalian Medical University.

3. Results and Discussion

In our previous publications, potassium permanganate agar method and iodine starch agar method were used to determine TAC in urine and serum [21, 22]. The diffusion area was dependent on the amount of antioxidants in urine and serum, in which larger diffusion area represents the fact that more antioxidants were measured. So, the amount of antioxidants reflects the antioxidant capacity in urine and serum; more antioxidants represent better antioxidant capacity. Both methods have good linearity and precision and can better reflect the state of TAC than reported methods in urine [22-26]. Neutral environment required in our methods is closer to physiological pH, high standard electrode potential of MnO₄^{-/}MnO₂ can oxidize most antioxidants, and indicator starch-iodine is highly sensitive to judge the end point and the covering of liquid paraffin on the surface of the agar can exclude the interference of external O_2 .

3.1. The Variation of Total Antioxidants Measured in Healthy People by Potassium Permanganate Agar Method and Iodine Starch Agar Method. In Table 1, the amount of antioxidants was measured in both urine and serum of 130 healthy people. In urine, positive correction coefficients (p = 0.025 in potassium permanganate agar method and p = 0.016 in iodine starch agar method) between age and urine diffusion area represent the fact that more urinary antioxidants exist in older people than the younger age. Contrastingly, in serum, negative correction coefficients (p < 0.001 in potassium permanganate agar and p = 0.014 in iodine starch agar method) represent the fact that less antioxidants exist in sera from older people. Similar results were observed in both methods, which ensure the accuracy of the results that older people has more antioxidants in urine and less antioxidant in serum comparing with the younger ones.

Age groups	Urine		Serum		
	Potassium permanganate agar	Iodine starch agar	Potassium permanganate agar	Iodine starch agar	
20~	3.35 ± 0.93	1.21 ± 0.25	2.52 ± 0.18	3.48 ± 0.51	
25~	1.99 ± 1.42	1.19 ± 0.48	2.34 ± 0.15	3.15 ± 0.42	
30~	2.24 ± 1.57	1.48 ± 0.51	2.41 ± 0.16	3.41 ± 0.39	
35~	2.14 ± 0.96	1.22 ± 0.47	2.45 ± 0.16	3.36 ± 0.42	
40~	2.15 ± 1.59	1.27 ± 0.32	2.39 ± 0.18	3.08 ± 0.30	
45~	2.04 ± 1.17	1.10 ± 0.31	2.37 ± 0.21	3.37 ± 0.60	
50~	2.33 ± 1.43	1.25 ± 0.44	2.37 ± 0.14	3.25 ± 0.47	
55~	1.72 ± 0.98	1.38 ± 0.53	2.39 ± 0.13	3.29 ± 0.33	
60~	3.10 ± 0.88	1.52 ± 0.50	2.26 ± 0.21	3.33 ± 0.48	
65~	2.47 ± 1.29	1.57 ± 0.49	2.35 ± 0.19	3.44 ± 0.46	
70~	2.28 ± 1.07	1.30 ± 0.51	2.28 ± 0.12	3.04 ± 0.27	
75~	2.71 ± 1.73	1.22 ± 0.37	2.27 ± 0.33	3.24 ± 0.49	
80~85	3.85 ± 1.27	1.47 ± 0.23	2.39 ± 0.22	3.03 ± 0.39	
Correlation coefficient	0.139	0.150	-0.239	-0.152	
p	0.025	0.016	<0.001	0.014	

TABLE 1: The diffusion area (cm^2) of urine and serum in healthy people in different groups (mean $\pm S$).

TABLE 2: Urine and serum diffusion area (cm²) of tumor patients and healthy people in the two methods.

Diffusion area (cm ²)		Urine			Serum		
		Median	Z	P	Mean \pm SD	t	Р
KMnO ₄	Tumor patients	1.83	-2.076	0.038	2.02 ± 0.17	-1.424	0.158
	Healthy people	1.54			1.97 ± 0.17		
<i>I</i> ₂	Tumor patients	1.54	-2.399	0.016	3.20 ± 0.40	-2.867	0.005
	Healthy people	1.13			2.94 ± 0.46		

Because urinary diffusion areas in both methods were in nonnormal distribution, nonparametric test was used to compare the results in the two groups. While the serumal diffusion areas in both methods were in normal distribution, *t*-test was used to compare the results in the two groups. SD, standard deviation.

The contrasting variations of antioxidants in urine and serum might be caused by the different antioxidant mechanisms. Future work is deserved to compare antioxidants profiling and fluid pathway in serum and urine. The question is whether the antioxidants in urine are derived from blood through glomerular filtration and tubular secretion.

The reason why we consider the age-background factor of healthy people is that, in previous reports, the serumal antioxidant capacity of the elderly will decline, but it is still unclear whether it is the same situation in urine. Thus, we measured the urinary antioxidants of healthy people to eliminate the age influence. Because there is age dependent trend observed between TAC and both urine and serum, the ages of tumor patients group and healthy group were designed to be matched to eliminate the influence.

3.2. The Total Antioxidants in Urine and Serum Were Measured in Unselected Patients with Tumors by Potassium Permanganate Agar Method and Iodine Starch Agar Method. Obvious oxidative stress is observed in patients with tumor. The balance between oxidants and antioxidants was broken up, regardless of the kinds of tumor. Here, clinical specimens of urine and serum were collected from patients with tumors without specifying the kind of tumors, which were labeled as "unselected patients with tumors" group.

In Table 2, because the urinary diffusion areas were not normally distributed in both methods, median diffusion areas were compared between the two groups. In potassium permanganate agar, the median diffusion area in tumor patients was 1.83 cm², while it was 1.54 cm² in healthy group; more antioxidants were measured in tumor patients, where p = 0.038; in iodine starch agar method, the median diffusion area in tumor patients was 1.54 cm², while it was 1.13 cm^2 in healthy group; more antioxidants were measured in tumor patients, where p = 0.016. While in serum the serum diffusion areas were normally distributed in both methods, average diffusion areas were compared between the two groups. In potassium permanganate agar, the average diffusion area in tumor patients was $2.02 \pm 0.17 \text{ cm}^2$, while it was 1.97 ± 0.17 cm² in healthy group, because p = 0.158; no evidence showed that more antioxidants were measured in tumor patients; in iodine starch agar method, the average diffusion area in tumor patients was 3.20 ± 0.40 cm², while it was 2.94 ± 0.46 cm² in healthy group; more antioxidants were measured in tumor patients, where p = 0.005.

Variables	В	SE	Wald	Sig.
U_I	0.850	0.280	9.192	0.002
S _I	2.029	0.651	9.702	0.002
Constant	-7.691	2.199	12.230	< 0.001

TABLE 3: Variables in binary logistic regression model.

 U_I , urine diffusion area in iodine starch agar; S_I , serum diffusion area in iodine starch agar.

TABLE 4: Results for the measurement of total antioxidants in urine and serum in the diagnosis of tumor diseases.

Variable	AUC	Std. error	Asymptotic Sig.	95% CI
$U_{ m Mn}$	0.626	0.060	0.043	0.509~0.742
U_I	0.629	0.061	0.037	0.509~0.749
S _{Mn}	0.503	0.064	0.963	0.377~0.629
S_I	0.669	0.058	0.006	0.556~0.782
Logistical regression model	0.787	0.047	< 0.001	0.694~0.880

 U_{Mn} , urine diffusion area in potassium permanganate agar; U_I , urine diffusion area in iodine starch agar; S_{Mn} , serum diffusion area in potassium permanganate agar; S_I , serum diffusion area in iodine starch agar; AUC, area under curve; CI, confidence interval.

To ensure the accuracy of the results, only the results were significant in both methods; the amount of antioxidants was considered different between the two groups. Thus, more urinary antioxidants were observed in tumor patients, because the results were significant in both methods. However, there was no difference in the serum antioxidants between the two groups; the result was significant only in iodine starch method. From the data presented in Table 1, it can be seen in older healthy people that less serumal antioxidants were measured. This is opposite to the results presented in Table 2, where no more serumal antioxidants were measured in tumor patients. However, in urine, more antioxidants were measured in the both older group and tumor patients. Thus, it may indicate a fact that the occurrence of tumor diseases is related to not only aging but also other factors. Obviously, the antioxidant mechanism of aging and tumor diseases has its own characteristic, and the mechanism of tumor diseases is still to be explored. Thus, we can conclude that, in tumor diseases, the change of antioxidants in urine was more sensitive than that in serum. Compared with serum, urine is a better specimen to evaluate the risk of tumor diseases. Exploring the diagnosis value of measuring the total antioxidants in urine to evaluate the risk of tumor diseases was deserved.

3.3. Measuring the Urinary Total Antioxidants Can Improve the Diagnosis Efficiency of Tumor Diseases Based on the Measurement of Antioxidants in an Organism to Diagnose Tumor Diseases. In Table 4, four ROC curves for urine and serum in both methods were constructed, but all AUCs at either urine or serum were small. So the combination of them was in consideration. To assess the combined use of the measurement of urine diffusion area in potassium permanganate agar (U_{Mn}) , serum diffusion area in potassium permanganate agar (S_{Mn}) , urine diffusion area in iodine starch agar (U_I) , and serum diffusion area in iodine starch agar (S_I) , binary logistic regression was conducted. In Table 3, only U_I and S_I were significant in the regression model, where

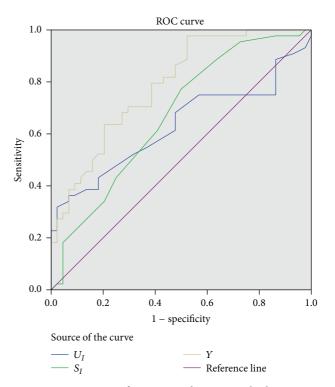


FIGURE 1: ROC curves for urine and serum in both potassium permanganate agar and iodine starch agar method.

the *p* values were both 0.002. $U_{\rm Mn}$ and $S_{\rm Mn}$ were excluded; the logistical regression model was $Y = 0.85 * U_I + 2.029 * S_I - 7.691$. Thus, U_I , S_I , and Y were chosen to build ROC, and it can be seen from Figure 1 that the AUC of Y was 0.787, which is larger than the use of U_I and S_I alone (Figure 1).

The reason why U_{Mn} and S_{Mn} were not selected in the regression model is that the sensitivity of potassium permanganate method is relatively lower than the iodine starch agar method; only few antioxidants that represent the differences between tumor diseases and healthy subjects were measured, but with the use of indicator starch, the sensitivity of iodine starch method was good; it can measure most antioxidants that represent the differences between tumor diseases and healthy subjects. And it also can be inferred that the electric potential of antioxidants in tumor patients was not high.

In all, the measurement of the urinary total antioxidants can improve the diagnosis efficiency of tumor diseases.

4. Conclusions

The antioxidant capacity of organism varies by age. More urinary antioxidants were measured in older people, while less antioxidants exist in serum of older people. The influence of age should be taken into consideration when discussing the TAC of an organism.

In patients with tumors, more antioxidants were found in urine with potassium permanganate agar method (p = 0.038) and iodine starch agar method (p = 0.016). More antioxidants were found in serum only in iodine starch agar method (p = 0.005). The change of the total amounts in urine was more sensitive than that in serum in tumor diseases.

The combined measurement of antioxidants in urine and serum could improve the diagnostic ability of tumor diseases, where AUC = 0.787. The measurement of total antioxidants in urine should be applied alone or in combination in clinic to evaluate the risk of tumor diseases.

Conflicts of Interest

No conflicts of interest exist in this manuscript.

References

- H. Kawagishi and T. Finkel, "Unraveling the truth about antioxidants: ROS and disease: finding the right balance," *Nature Medicine*, vol. 20, no. 7, pp. 711–713, 2014.
- [2] R. A. Floyd, R. A. Towner, T. He, K. Hensley, and K. R. Maples, "Translational research involving oxidative stress and diseases of aging," *Free Radical Biology and Medicine*, vol. 51, no. 5, pp. 931–941, 2011.
- [3] A. Buico, C. Cassino, M. Ravera, P.-G. Betta, and D. Osella, "Oxidative stress and total antioxidant capacity in human plasma," *Redox Report*, vol. 14, no. 3, pp. 125–131, 2009.
- [4] S. Bouatra, F. Aziat, R. Mandal et al., "The human urine metabolome," *PLoS ONE*, vol. 8, no. 9, Article ID e73076, 2013.
- [5] B. Kirschbaum, "Total urine antioxidant capacity," *Clinica Chimica Acta*, vol. 305, no. 1-2, pp. 167–173, 2001.
- [6] E. Sofic, A. Rustembegovic, G. Kroyer, and G. Cao, "Serum antioxidant capacity in neurological, psychiatric, renal diseases and cardiomyopathy," *Journal of Neural Transmission*, vol. 109, no. 5-6, pp. 711–719, 2002.
- [7] R. Stocker, Y. Yamamoto, A. F. McDonagh, A. N. Glazer, and B. N. Ames, "Bilirubin is an antioxidant of possible physiological importance," *Science*, vol. 235, no. 4792, pp. 1043–1046, 1987.
- [8] B. N. Ames, R. Cathcart, E. Schwiers, and P. Hochstein, "Uric acid provides an antioxidant defense in humans against oxidant- and radical-caused aging and cancer: a hypothesis,"

Proceedings of the National Academy of Sciences of the United States of America, vol. 78, no. 11, pp. 6858–6862, 1981.

- [9] S. Percário, S. P. D. T. Domingues, L. F. M. Teixeira et al., "Effects of creatine supplementation on oxidative stress profile of athletes," *Journal of the International Society of Sports Nutrition*, vol. 9, no. 1, article 56, 2012.
- [10] X. Zhao, S. Zhou, R. Liu et al., "Proteomics analysis of tumor microenvironment: implications of metabolic and oxidative stresses in tumorigenesis," *Mass Spectrometry Reviews*, vol. 32, no. 4, pp. 267–311, 2013.
- [11] A. Valavanidis, T. Vlachogianni, and C. Fiotakis, "8-Hydroxy-21-deoxyguanosine (8-OHdG): a critical biomarker of oxidative stress and carcinogenesis," *Journal of Environmental Science and Health. Part C Environmental Carcinogenesis and Ecotoxicology Reviews*, vol. 27, no. 2, pp. 120–139, 2009.
- [12] T. Liu, A. Stern, L. J. Roberts, and J. D. Morrow, "The isoprostanes: novel prostaglandin-like products of the free radical-catalyzed peroxidation of arachidonic acid," *Journal of Biomedical Science*, vol. 6, no. 4, pp. 226–235, 1999.
- [13] H. A. Headlam and M. J. Davies, "Markers of protein oxidation: different oxidants give rise to variable yields of bound and released carbonyl products," *Free Radical Biology and Medicine*, vol. 36, no. 9, pp. 1175–1184, 2004.
- [14] C.-C. Sung, Y.-C. Hsu, C.-C. Chen, Y.-F. Lin, and C.-C. Wu, "Oxidative stress and nucleic acid oxidation in patients with chronic kidney disease," *Oxidative Medicine and Cellular Longevity*, vol. 2013, Article ID 301982, 15 pages, 2013.
- [15] E. Szabo, M. E. Riffe, S. M. Steinberg, M. J. Birrer, and R. I. Linnoila, "Altered cJun expression: an early event in human lung carcinogenesis," *Cancer Research*, vol. 56, no. 2, pp. 305–315, 1996.
- [16] P. Klatt and S. Lamas, "Regulation of protein function by Sglutathiolation in response to oxidative and nitrosative stress," *European Journal of Biochemistry*, vol. 267, no. 16, pp. 4928– 4944, 2000.
- [17] D. Dreher and A. F. Junod, "Role of oxygen free radicals in cancer development," *European Journal of Cancer*, vol. 32, no. 1, pp. 30–38, 1996.
- [18] J. L. Hu, Z. Y. Li, W. Liu et al., "Polymorphism in heme oxygenase-1 (HO-1) promoter and alcohol are related to the risk of esophageal squamous cell carcinoma on Chinese males," *Neoplasma*, vol. 57, no. 1, pp. 86–92, 2010.
- [19] A. Kikuchi, M. Yamaya, S. Suzuki et al., "Association of susceptibility to the development of lung adenocarcinoma with the heme oxygenase-1 gene promoter polymorphism," *Human Genetics*, vol. 116, no. 5, pp. 354–360, 2005.
- [20] X. Xu, G. S. Rao, V. Groh et al., "Major histocompatibility complex class I-related chain A/B (MICA/B) expression in tumor tissue and serum of pancreatic cancer: role of uric acid accumulation in gemcitabine-induced MICA/B expression," *BMC Cancer*, vol. 11, article no. 1471, 2011.
- [21] Y. Zhou, M. Zhang, and H. Liu, "Total antioxidant capacity of serum determined using the potassium permanganate agar method based on serum diffusion in agar," *Bioinorganic Chemistry and Applications*, vol. 2015, Article ID 406071, 6 pages, 2015.
- [22] Y. Zhou, N. Xu, M. Zhang, and H. Liu, "A new method for measuring total antioxidant capacity in urine using the iodine starch agar based on agar diffusion," *Current Analytical Chemistry*, vol. 12, no. 5, pp. 425–430, 2016.

- [23] I. F. F. Benzie and J. J. Strain, "The ferric reducing ability of plasma (FRAP) as a measure of "antioxidant power": The FRAP assay," *Analytical Biochemistry*, vol. 239, no. 1, pp. 70–76, 1996.
- [24] I. F. F. Benzie and J. J. Strain, "Ferric reducing/antioxidant power assay: direct measure of total antioxidant activity of biological fluids and modified version for simultaneous measurement of total antioxidant power and ascorbic acid concentration," *Methods in Enzymology*, vol. 299, pp. 15–27, 1998.
- [25] R. Apak, K. Güçlü, M. Özyürek, and S. E. Karademir, "Novel total antioxidant capacity index for dietary polyphenols and vitamins C and E, using their cupric ion reducing capability in the presence of neocuproine: CUPRAC method," *Journal of Agricultural and Food Chemistry*, vol. 52, no. 26, pp. 7970–7981, 2004.
- [26] R. Apak, K. Güçlü, M. Özyürek, B. Bektaşoğlu, and M. Bener, "Cupric ion reducing antioxidant capacity assay for antioxidants in human serum and for hydroxyl radical scavengers," *Methods in Molecular Biology*, vol. 594, pp. 215–239, 2010.
- [27] G. Litwinienko and K. U. Ingold, "Abnormal solvent effects on hydrogen atom abstractions. 1. The reactions of phenols with 2,2-diphenyl-1-picrylhydrazyl (dpph*) in alcohols," *Journal of Organic Chemistry*, vol. 68, no. 9, pp. 3433–3438, 2003.
- [28] R. Amorati, S. Menichetti, E. Mileo, G. F. Pedulli, and C. Viglianisi, "Hydrogen-atom transfer reactions from ortho-alkoxysubstituted phenols: an experimental approach," *Chemistry*, vol. 15, no. 17, pp. 4402–4410, 2009.