

REVIEW

**OPEN ACCESS**  
Full open access to this and  
thousands of other papers at  
<http://www.la-press.com>.

## Alzheimers Disease: Review of Emerging Treatment Role for Intravenous Immunoglobulins

Rakez Kayed<sup>1-3</sup>, George R. Jackson<sup>1-3</sup>, D. Mark Estes<sup>3,4</sup>, and Alan D.T. Barrett<sup>3,4</sup>

<sup>1</sup>Mitchell Center for Neurodegenerative Diseases, University of Texas Medical Branch, Galveston, TX, USA.

<sup>2</sup>Department of Neurology, University of Texas Medical Branch, Galveston, TX, USA. <sup>3</sup>Sealy Center for Vaccine Development, University of Texas Medical Branch, Galveston, TX, USA. <sup>4</sup>Department of Pathology, University of Texas Medical Branch, Galveston, TX, USA. Corresponding author email: [rakayed@utmb.edu](mailto:rakayed@utmb.edu)

---

**Abstract:** Alzheimer's disease (AD) is the most common neurodegenerative disorder. Currently available therapies are symptomatic but do not alter underlying disease progression. Immunotherapeutic approaches such as anti A $\beta$  peptide active vaccination trials have had limited success to date. Intravenous immunoglobulin (IVIg) is widely used in immune-mediated neurological disorders such as myasthenia gravis and Guillain-Barre syndrome. These preparations have been obtained from the pooled plasma of healthy human donors and contain natural anti-amyloid antibodies and are well tolerated. A small pilot study of passive immunotherapy using IVIg has suggested cognitive improvement. A multicenter phase III trial is ongoing and will determine whether or not this treatment can ameliorate cognitive deficits in mild-to-moderate AD. Here, we briefly review the pathogenic role of amyloid and tau in AD, as well as immunotherapeutic efforts to date. We also summarize what is known about naturally occurring anti-A $\beta$  and tau antibodies in IVIg with a view toward explaining potential mechanisms underlying their therapeutic effects.

**Keywords:** Alzheimer's, immunotherapy, conformation antibodies, tau oligomers, amyloid oligomers.

---

*Journal of Central Nervous System Disease* 2011:3 67–73

doi: [10.4137/JCNSD.S5018](https://doi.org/10.4137/JCNSD.S5018)

This article is available from <http://www.la-press.com>.

© the author(s), publisher and licensee Libertas Academica Ltd.

This is an open access article. Unrestricted non-commercial use is permitted provided the original work is properly cited.



## Introduction

Alzheimer's disease (AD) is the most common neurodegenerative disorder, with devastating personal and financial costs. In the absence of clear prevention strategies or disease modifying therapies, it is expected that the number of people affected by AD worldwide will exceed 100 million in 2050. With improved survival from acute diseases and the increasing lifespan of populations in developed and middle income countries, dramatic increases in the incidence of Alzheimer's disease (AD) are predicted, with dire consequences for the economic and social fabric of many nations.<sup>1</sup> Hence, development of effective disease-modifying therapies for AD is an urgent priority for research in both academia and pharmaceutical companies. AD is a complex disease with two principle hallmark events: 1) the misfolding, aggregation and brain deposition of amyloid- $\beta$  (A $\beta$ ) peptide in amyloid plaques, and 2) the deposition of misfolded tau protein in neurofibrillary tangles (NFT).<sup>2</sup> The A $\beta$  peptide is generated from the cleavage of amyloid precursor protein (APP) by  $\beta$  and  $\gamma$  secretases.<sup>2</sup> Given the preeminence of the amyloid hypothesis<sup>3</sup> in the AD field, extensive efforts have targeted various forms of A $\beta$  aggregates for drug development; these include reduction and alteration of APP processing, prevention of A $\beta$  misfolding and aggregation, minimization or elimination of its neurotoxicity, acceleration of its clearance and degradation,<sup>4-9</sup> as well as active (ie, stimulation of an immune response following administration of an immunogen), and passive (ie, provision of short term protection against infection or clinical condition by administration of antibodies) vaccination strategies to remove amyloid deposits.<sup>10</sup>

An ever-increasing body of evidence implicating tau in neurodegenerative diseases<sup>11,12</sup> supports tau as a potential target for the development of disease-modifying therapeutics.<sup>13,14</sup> Tau-based therapeutic approaches have historically lagged behind anti A $\beta$  approaches. Recently, however, tau-based approaches have been the subject of renewed interest;<sup>13</sup> potential therapeutics may manipulate tau via inhibition of phosphorylation,<sup>15,16</sup> activation of proteolytic or proteasomal degradation pathways,<sup>17,18</sup> microtubule-binding drugs (eg, paclitaxel) for stabilization of microtubule networks,<sup>19,20</sup> inhibition of aggregation by small molecules,<sup>21,22</sup> or clearance by immunotherapy.<sup>23-26</sup>

## Immunotherapy Against A $\beta$

Arguably the most exciting treatment approach for AD to have evolved recently is anti-A $\beta$  immunotherapy using antibodies to A $\beta$  administered on multiple occasions.<sup>27</sup> First introduced by Schenk and colleagues in 1999, promising results were described initially in animal models.<sup>28</sup> Ten years later, enthusiasm for anti A $\beta$  immunotherapy has been largely replaced by frustration due to adverse effects including meningoencephalitis and leukoencephalopathy.<sup>10,29-31</sup> Results from the ongoing phase II trial of anti A $\beta$  humanized monoclonal antibody (Bapineuzumab) have confirmed removal of amyloid plaques as detected by positron emission tomography (PET) scans using Pittsburgh compound B (<sup>11</sup>C-PiB), but without concomitant cognition improvement.<sup>32</sup> In addition, despite evidence of amyloid plaque removal,<sup>33</sup> *post mortem* analysis of brains from those patients has failed to demonstrate changes in tau pathology, neuropil threads, synaptic dysfunction, or cerebral amyloid angiopathy.<sup>29,34,35</sup> Despite these disappointments, the research community has persevered with alternative regimens of administration in efforts to develop and optimize a more effective passive or active A $\beta$ -based vaccine. These efforts have included the use of IVIg preparations that contain naturally occurring anti A $\beta$  antibodies.<sup>36-38</sup> Recent literature reveals a shift in focus from cure toward understanding mechanisms associated with benefits in animal models and etiology of complications reported in both humans and animal models (reviewed in).<sup>39-44</sup>

## Conformation-Specific Antibodies

Amyloid diseases, including many neurodegenerative disorders, are considered conformational diseases, since amyloid formation is triggered by conformational changes in a specific peptide or protein, resulting in its misfolding and deposition as amyloid.<sup>45-47</sup> Moreover, conformation-specific antibodies that recognize specific amyloid species, eg, fibrils or oligomers, from many types of amyloid proteins have been produced and characterized.<sup>48-50</sup> Conformation-specific antibodies were derived from observations reported more than thirty years ago<sup>51,52</sup> indicating that amyloid antibodies react with conformational epitopes and not with native protein structure, ie, suggesting that amyloid fibrils have a non-native structure.<sup>51,52</sup>



Numerous conformation-specific antibodies have been generated and characterized, including a few that are commercially available. Such antibodies have been used to characterize disease progression and to ameliorate amyloid toxicity (see review by Glabe).<sup>50</sup> Moreover, conformation-specific antibody domains and single chain fragment variable (scFv) constructs with similar specificity have been reported; of note, these can cross the blood-brain barrier more efficiently than antibodies and can be expressed intracellularly.<sup>53–55</sup>

The critical role of soluble amyloid oligomers in neurodegeneration has become more generally accepted for multiple neurodegenerative diseases, including AD.<sup>56–60</sup> Results obtained using oligomeric conformation-specific antibodies<sup>49</sup> indicate that oligomers (protofibrils) have a common, generic structure that is distinct from both fibrils and low molecular weight soluble monomer/dimers. Furthermore, such antibodies recognize soluble oligomers from a variety of different amyloids, including lysozyme, islet amyloid polypeptide (IAPP), synuclein, prion protein, polyglutamine, and insulin. The anti oligomer antibody (A11) that binds specifically to amyloid oligomers<sup>49</sup> has more robust effects as compared to other anti amyloid antibodies when injected intrathecally into the TgCRND8 AD mouse model.<sup>61</sup> Surprisingly, similar conformation-specific antibodies have been detected in humans using peptide microarrays. Britschgi et al demonstrated the presence of sequence-independent, oligomeric conformational antibodies in human plasma and CSF.<sup>62</sup> Although the diversity, abundance, and function of such endogenous conformational antibodies remain largely uncharacterized, these investigators have reported that these antibodies decline with age and advancing AD, suggesting that they may play a role in protection against toxic amyloid oligomers.<sup>62</sup>

### **Tau-Based Immunotherapy**

Tau immunotherapy is a new concept.<sup>23</sup> To date, only three reports of tau immunotherapy in animal models have been published, all using active vaccination.<sup>24,25,63</sup> To date, no reports of passive vaccination have appeared. In the first report, the authors used a tau fragment (379–408) phosphorylated at Ser396 and Ser404 (phosphorylation sites commonly associated with NFT) to vaccinate the P301L mouse model.<sup>64</sup>

Behavioral analysis showed improved performance after immunization as compared to controls. These data demonstrated that antibodies against this immunogen were able to cross the blood-brain barrier and bind to phosphorylated tau.<sup>24</sup> The Rosenmann group used phosphorylated tau with Freund's adjuvant and pertussis toxin adjuvants; these investigators reported a 40% reduction in NFTs and 20% increase in microglia.<sup>25</sup> In 2006, the Rosenmann group also reported that full-length tau was encephalitogenic, triggering a severe autoimmune response.<sup>63</sup> Mice vaccinated with soluble tau developed NFT-like structures, axonal damage, gliosis, mononuclear infiltrates, and motor phenotypes. These data demonstrate the potential dangers of using soluble tau as immunogen, or of antibodies recognizing epitopes of full-length tau for passive vaccination. Although the use of phosphorylated tau antigens seems promising for vaccination studies (ie, presenting specific phosphoepitopes to the immune system), such an approach has significant potential risks, as these phosphorylation sites are mainly associated with NFT.<sup>65,66</sup> An optimal vaccine should target pre-filament tau species (tau oligomers), which form at early stages of NFT development rather than mature, meta-stable NFT.<sup>26</sup> Pre-filament specific phosphorylation sites have yet to be conclusively identified due to the complexity of tau aggregation, the overlap between the three stages of NFTs development with regard to tau phosphorylation sites,<sup>65</sup> and the fact that tau phosphorylation is a physiological process that is essential for tau normal function and reversible.<sup>67</sup> *In vitro* assembled tau paired helical filaments (PHF) similar to those found in AD have  $\beta$ -structure similar to other amyloid fibrils.<sup>68,69</sup> Moreover, tau conformation-specific antibodies (eg, Alz50 and MC-1) recognize conformational tau epitopes associated with PHF.<sup>70,71</sup> Naturally occurring antibodies have been detected in the blood of normal and AD patients, including antibodies against both unphosphorylated and phosphorylated tau.<sup>72</sup> Both IgG and IgM anti-tau antibodies have been identified in serum collected from AD patients and controls; very few tau antibodies are found in CSF. Higher anti-phosphorylated-tau IgM antibodies have been found in AD patients relative to controls. In this work of Steinitz and coworkers, all subjects (both AD and controls) were greater than 70 years of age;<sup>72</sup> it is reasonable to assume that higher levels



of anti-phosphotau IgM would be present in healthy young controls, as reported for A $\beta$  antibodies. If so, it seems likely that larger studies will confirm these findings and identify higher concentrations of neuroprotective tau antibodies in young healthy people.

## IVIg Immunotherapy

Intravenous immunoglobulin (IVIg) is an antibody product obtained from human plasma from thousands of donors. For more than thirty years, IVIg has been used for the treatment of post-exposure to infectious diseases, immune disorders and the management of patients with neurological conditions.<sup>73–76</sup> IVIg treatment is used routinely for some immune-mediated neurological disorders such as Guillain-Barre syndrome, patients treated with IVIg have reduced risk of developing Alzheimer's disease and recently IVIg has been investigated for the treatment of neurodegenerative disorders.<sup>77</sup> Commercially available IVIg has been used in small pilot trial in AD.<sup>36</sup> In this open label study, 8 mildly affected patients received IVIg for a total of 15 months over two intervals (6 months, discontinued for 3 months, then followed by 9 months further treatment). Infusions were generally well-tolerated; of note, both anti-A $\beta$  antibodies and A $\beta$  levels in the serum increased after each infusion in proportion to IVIg dose, whereas A $\beta$  levels in CSF decreased significantly at 6 months, returned to baseline after washout, and decreased again after IVIg was re-administered for the additional 9 months. Mini-mental state scores increased an average of 2.5 points after 6 months, returned to baseline during washout, and remained stable during subsequent 9 months treatment.<sup>36</sup> A smaller study previously had shown positive effects in five AD patients, each of whom received one dose of IVIg per month for 6 months.<sup>38</sup> Taken together, these findings justified a large-scale randomized phase III clinical trial that has enrolled more than 360 AD patients.<sup>76,78</sup>

Naturally occurring antibodies that can detect and block the toxicity of special conformations displayed by misfolded proteins, including amyloid- $\beta$ ,  $\alpha$ -synuclein, and prion protein, are detectable in human serum and CSF. Although such antibodies are detectable in both healthy individuals and patients, it is clear that their levels are reduced in the latter.<sup>62,78–82</sup> However, an assay for precise quantification of endogenous human A $\beta$  antibodies

is not yet available. Efforts are underway to identify and overcome factors contributing to wide disparities among reported measurements.<sup>83–85</sup> Such efforts seem likely to improve the quality of IVIg preparations for AD treatment, and they may lead to the development of methods to isolate specific anti-amyloid antibody populations that can then be tested for their potential to treat AD and other neurodegenerative diseases by passive administration.<sup>80,83–85</sup> Humanized monoclonal antibodies have applications but they recognize only a single epitope whereas naturally occurring antibodies are polyclonal and will recognize multiple epitopes, and more likely to have stronger therapeutic effects.

## Which Antibodies Should We be Looking for in IVIg Preparations?

In addition to the anti A $\beta$  oligomer antibodies reported in IVIg,<sup>82,83</sup> these preparations may contain neuroprotective anti tau oligomer antibodies that may account, at least in part, for the positive results derived from IVIg treatment in AD. Recently, tau oligomers have emerged as a likely pathogenic entity in tauopathies. Although their formation and role in neurodegeneration has yet to be fully elucidated, an increasing literature argues that they play a crucial role in AD and other tauopathies. Stereological analysis of AD demonstrates that neuronal loss actually precedes NFT formation.<sup>86,87</sup> This observation, as well as data emerging from biochemical, cell-based and transgenic mouse studies, suggests that soluble tau aggregates may be the most toxic and pathologically significant tau species.<sup>57,88–92</sup> Studies using different animal models suggest that tau oligomers play a key role in impaired synaptic function, hippocampal synapse loss, and microgliosis leading to neurodegeneration and behavioral impairments. All of these phenomena are concurrent with accumulation of soluble aggregated tau species and dissociated from the accumulation of NFT.<sup>57,93,94</sup> Tau oligomers have been characterized biochemically in a conditional mouse model (rTg4510) expressing the P301L human tau mutant; surprisingly, the accumulation of oligomeric tau (but not NFT) correlated with neuronal loss and behavioral deficits in this model.<sup>95,96</sup> Tau oligomers have been characterized in human brain; a correlation between disease progression and the accumulation of granular tau oligomers in the brains of AD





patients has been reported.<sup>97,98</sup> Moreover, increased levels of tau oligomers are detected in the frontal cortex at very early stage of the disease (Braak stage I), before clinical manifestations of AD and NFT are believed to develop.<sup>97,98</sup> Given the heterogeneity of tau species, ranging from monomer to oligomer, to PHF/NFT, it seems prudent that future efforts should focus on complete characterization of all tau antibodies present in CSF, rather than on any particular species. Meticulous analysis of both anti tau and anti amyloid antibodies present in IVIg preparations is critical for a better understanding of mechanisms underlying potential benefits of such preparations in AD.

## Conclusions

The results from the pilot clinical trial justify further investigation of IVIg for the treatment of AD. Further examination of endogenous neuroprotective antibodies should help us to better understand their mechanism of action. Given that anti-A $\beta$  immunotherapy trials were disappointing, it seems unlikely that naturally occurring A $\beta$  antibodies are likely to account for observed clinical benefits of IVIg; further, such IVIg preparations may be effective because they contain both anti-tau and anti-A $\beta$  natural antibodies. Apart from the presence of these antibodies, IVIg may be useful for AD treatment due to anti-inflammatory effects at high doses mediated in part via Fc receptor uptake of IgG by inflammatory cells.<sup>73,74</sup> IVIg is a very expensive treatment, and despite its safety record in the treatment of multiple neurological disorders, potential complications include headache, dermatitis, infection, pulmonary edema, allergic/anaphylactic reactions, acute renal failure, venous thrombosis, and aseptic meningitis.<sup>99,100</sup> These side effects may hinder the use of IVIg in AD, which has a long asymptomatic phase and survival period once symptomatic. Therefore, critical analysis of IVIg preparations and further examination of endogenous neuroprotective antibodies are necessary to better understand their mechanism of action and optimize their role as a disease modifying therapeutic in AD and other neurodegenerative diseases. Finally, significant advances have been made in the use of humanized monoclonal antibodies for passive vaccination against infectious diseases that will do doubt have applications.

## Disclosure

This manuscript has been read and approved by all authors. This paper is unique and is not under consideration by any other publication and has not been published elsewhere. The authors and peer reviewers of this paper report no conflicts of interest. The authors confirm that they have permission to reproduce any copyrighted material.

## References

1. Editorial. How much is dementia care worth? *The Lancet Neurology*. 2010; 9:1037.
2. Selkoe DJ. Alzheimer's disease: Genes, proteins, and therapy. *Physiol Rev*. 2001;81:741–66.
3. Hardy J, Selkoe DJ. The amyloid hypothesis of Alzheimer's disease: progress and problems on the road to therapeutics. *Science*. 2002;297:353–6.
4. Roberson ED, Mucke L. 100 years and counting: prospects for defeating Alzheimer's disease. *Science*. 2006;314:781–4.
5. Shah RS, Lee HG, Xiongwei Z, Perry G, Smith MA, Castellani RJ. Current approaches in the treatment of Alzheimer's disease. *Biomed Pharmacother*. 2008;62:199–207.
6. Stains CI, Mondal K, Ghosh I. Molecules that target beta-amyloid. *Chem Med Chem*. 2007;2:1674–92.
7. Klaver DW, Wilce MC, Cui H, Hung AC, Gasperini R, Foa L, et al. Is BACE1 a suitable therapeutic target for the treatment of Alzheimer's disease? Current strategies and future directions. *Biol Chem*. 2010;391:849–59.
8. Kovacs T. Therapy of Alzheimer disease. *Neuropsychopharmacol Hung*. 2009;11:27–33.
9. Wolfe MS. Inhibition and modulation of gamma-secretase for Alzheimer's disease. *Neurotherapeutics*. 2008;5:391–8.
10. Wisniewski T, Boutajangout A. Vaccination as a therapeutic approach to alzheimer's disease. *Mt Sinai J Med*. 2010;77:17–31.
11. Ballatore C, Lee VM, Trojanowski JQ. Tau-mediated neurodegeneration in Alzheimer's disease and related disorders. *Nat Rev Neurosci*. 2007;8: 663–72.
12. Haroutunian V, Davies P, Vianna C, Buxbaum JD, Purohit DP. Tau protein abnormalities associated with the progression of alzheimer disease type dementia. *Neurobiol Aging*. 2007;28:1–7.
13. Schneider A, Mandelkow E. Tau-based treatment strategies in neurodegenerative diseases. *Neurotherapeutics*. 2008;5:443–57.
14. Iqbal K, Liu F, Gong CX, Alonso AD, Grundke-Iqbal I. Mechanisms of tau-induced neurodegeneration. *Acta Neuropathol*. 2009;118:53–69.
15. Iqbal K, Grundke-Iqbal I. Tau phosphatase activity as a therapeutic target for AD. *Drug News Perspect*. 1998;11:10–4.
16. Iqbal K, Grundke-Iqbal I. Inhibition of neurofibrillary degeneration: a promising approach to Alzheimer's disease and other tauopathies. *Curr Drug Targets*. 2004;5:495–502.
17. Johnson GV, Jope RS, Binder LI. Proteolysis of tau by calpain. *Biochem Biophys Res Commun*. 1989;163:1505–11.
18. Sengupta S, Horowitz PM, Karsten SL, Jackson GR, Geschwind DH, Fu Y, et al. Degradation of tau protein by puromycin-sensitive aminopeptidase in vitro. *Biochemistry*. 2006;45:15111–9.
19. Trojanowski JQ, Smith AB, Huryn D, Lee VM. Microtubule-stabilising drugs for therapy of Alzheimer's disease and other neurodegenerative disorders with axonal transport impairments. *Expert Opin Pharmacother*. 2005;6:683–6.
20. Zhang B, Maiti A, Shively S, Lakhani F, McDonald-Jones G, Bruce J, et al. Microtubule-binding drugs offset tau sequestration by stabilizing microtubules and reversing fast axonal transport deficits in a tauopathy model. *Proc Natl Acad Sci U S A*. 2005;102:227–31.
21. Bulic B, Pickhardt M, Schmidt B, Mandelkow EM, Waldmann H, Mandelkow E. Development of tau aggregation inhibitors for Alzheimer's disease. *Angew Chem Int Ed Engl*. 2009;48:1740–52.



22. Duff K, Kuret J, Congdon EE. Disaggregation of Tau as a Therapeutic Approach to Tauopathies. *Curr Alzheimer Res.* 2010;7:235–40.
23. Sigurdsson EM. Tau-focused immunotherapy for Alzheimer's disease and related tauopathies. *Curr Alzheimer Res.* 2009;6:446–50.
24. Asuni AA, Boutajangout A, Quartermain D, Sigurdsson EM. Immunotherapy targeting pathological tau conformers in a tangle mouse model reduces brain pathology with associated functional improvements. *J Neurosci.* 2007;27:9115–29.
25. Boimel M, Grigoriadis N, Loubopoulos A, Haber E, Abramsky O, Rosenmann H. Efficacy and safety of immunization with phosphorylated tau against neurofibrillary tangles in mice. *Exp Neurol.* 2010;224:472–85.
26. Kayed R, Jackson GR. Prefilament tau species as potential targets for immunotherapy for Alzheimer disease and related disorders. *Curr Opin Immunol.* 2009;21:359–63.
27. Wisniewski T, Boutajangout A. Immunotherapeutic approaches for Alzheimer's disease in transgenic mouse models. *Brain Struct Funct.* 2010;214:201–18.
28. Schenk D, Barbour R, Dunn W, Gordon G, Grajeda H, Guido T, et al. Immunization with amyloid-beta attenuates Alzheimer-disease-like pathology in the PDAPP mouse. *Nature.* 1999;400:173–7.
29. Holmes C, Boche D, Wilkinson D, Yadegarfar G, Hopkins V, Bayer A, et al. Long-term effects of Abeta42 immunisation in Alzheimer's disease: follow-up of a randomised, placebo-controlled phase I trial. *Lancet.* 2008;372:216–23.
30. Nicoll JA, Barton E, Boche D, Neal JW, Ferrer I, Thompson P, et al. Abeta species removal after abeta42 immunization. *J Neuropathol Exp Neurol.* 2006;65:1040–8.
31. Weiner HL, Frenkel D. Immunology and immunotherapy of Alzheimer's disease. *Nat Rev Immunol.* 2006;6:404–16.
32. Rinne JO, Brooks DJ, Rossor MN, Fox NC, Bullock R, Klunk WE, et al. 11C-PiB PET assessment of change in fibrillar amyloid-beta load in patients with Alzheimer's disease treated with bapineuzumab: a phase 2, double-blind, placebo-controlled, ascending-dose study. *Lancet Neurol.* 2010;9:363–72.
33. Schenk D. Amyloid-beta immunotherapy for Alzheimer's disease: the end of the beginning. *Nat Rev Neurosci.* 2002;3:824–8.
34. Nicoll JA, Wilkinson D, Holmes C, Steart P, Markham H, Weller RO. Neuropathology of human Alzheimer disease after immunization with amyloid-beta peptide: a case report. *Nat Med.* 2003;9:448–52.
35. Boche D, Zotova E, Weller RO, Love S, Neal JW, Pickering RM, et al. Consequence of Abeta immunization on the vasculature of human Alzheimer's disease brain. *Brain.* 2008;131:3299–310.
36. Relkin NR, Szabo P, Adamiak B, Burgut T, Monthe C, Lent RW, et al. 18-Month study of intravenous immunoglobulin for treatment of mild Alzheimer disease. *Neurobiol Aging.* 2009;30:1728–36.
37. Weksler ME, Pawelec G, Franceschi C. Immune therapy for age-related diseases. *Trends Immunol.* 2009;30:344–50.
38. Dodel RC, Du Y, Depboylu C, Hampel H, Frolich L, Haag A, et al. Intravenous immunoglobulins containing antibodies against beta-amyloid for the treatment of Alzheimer's disease. *J Neurol Neurosurg Psychiatry.* 2004;75:1472–4.
39. Woodhouse A, Dickson TC, Vickers JC. Vaccination strategies for Alzheimer's disease: A new hope? *Drugs Aging.* 2007;24:107–19.
40. Wilcock DM, Colton CA. Anti-amyloid-beta immunotherapy in Alzheimer's disease: relevance of transgenic mouse studies to clinical trials. *J Alzheimers Dis.* 2008;15:555–69.
41. Brendza RP, Holtzman DM. Amyloid-beta immunotherapies in mice and men. *Alzheimer Dis Assoc Disord.* 2006;20:118–23.
42. Brody DL, Holtzman DM. Active and passive immunotherapy for neurodegenerative disorders. *Annu Rev Neurosci.* 2008;31:175–93.
43. Lichtlen P, Mohajeri MH. Antibody-based approaches in Alzheimer's research: safety, pharmacokinetics, metabolism, and analytical tools. *J Neurochem.* 2008;104:859–74.
44. Schenk DB, Seubert P, Grundman M, Black R. A beta immunotherapy: Lessons learned for potential treatment of Alzheimer's disease. *Neurodegener Dis.* 2005;2:255–60.
45. Zerovnik E. Protein conformational pathology in Alzheimer's and other neurodegenerative diseases; new targets for therapy. *Curr Alzheimer Res.* 2010;7:74–83.
46. Reiss C, Ehrlich R, Lesnick T, Parvez S, Parvez H. Conformational diseases: misfolding mechanisms may pave the way to early therapy. *Neurotoxicol Teratol.* 2002;24:ix–xiv.
47. Chiti F, Dobson CM. Protein misfolding, functional amyloid, and human disease. *Annu Rev Biochem.* 2006;75:333–66.
48. Kayed R, Head E, Sarsoza F, Saing T, Cotman CW, Necula M, et al. Fibril specific, conformation dependent antibodies recognize a generic epitope common to amyloid fibrils and fibrillar oligomers that is absent in prefibrillar oligomers. *Mol Neurodegener.* 2007;2:18.
49. Kayed R, Head E, Thompson JL, McIntire TM, Milton SC, Cotman CW, et al. Common structure of soluble amyloid oligomers implies common mechanism of pathogenesis. *Science.* 2003;300:486–9.
50. Glabe CG. Conformation-dependent antibodies target diseases of protein misfolding. *Trends Biochem Sci.* 2004;29:542–7.
51. Franklin EC, Zucker-Franklin D. Antisera specific for human amyloid reactive with conformational antigens. *Proc Soc Exp Biol Med.* 1972;140:565–8.
52. Linke RP, Zucker-Franklin D, Franklin ED. Morphologic, chemical, and immunologic studies of amyloid-like fibrils formed from Bence Jones Proteins by proteolysis. *J Immunol.* 1973;111:10–23.
53. Robert R, Dolezal O, Waddington L, Hattarki MK, Cappai R, Masters CL, et al. Engineered antibody intervention strategies for Alzheimer's disease and related dementias by targeting amyloid and toxic oligomers. *Protein Eng Des Sel.* 2009;22:199–208.
54. Habicht G, Haupt C, Friedrich RP, Hortschansky P, Sachse C, Meinhardt J, et al. Directed selection of a conformational antibody domain that prevents mature amyloid fibril formation by stabilizing Abeta protofibrils. *Proc Natl Acad Sci U S A.* 2007;104:19232–7.
55. Barkhordarian H, Emadi S, Schulz P, Sierks MR. Isolating recombinant antibodies against specific protein morphologies using atomic force microscopy and phage display technologies. *Protein Eng Des Sel.* 2006;19:497–502.
56. Haass C, Selkoe DJ. Soluble protein oligomers in neurodegeneration: lessons from the Alzheimer's amyloid beta-peptide. *Nat Rev Mol Cell Biol.* 2007;8:101–12.
57. Brunden KR, Trojanowski JQ, Lee VM. Evidence that non-fibrillar tau causes pathology linked to neurodegeneration and behavioral impairments. *J Alzheimers Dis.* 2008;14:393–9.
58. Caughey B, Baron GS, Chesebro B, Jeffrey M. Getting a grip on prions: oligomers, amyloids, and pathological membrane interactions. *Annu Rev Biochem.* 2009;78:177–204.
59. Glabe CG. Common mechanisms of amyloid oligomer pathogenesis in degenerative disease. *Neurobiol Aging.* 2006;27:570–5.
60. Glabe CG. Structural classification of toxic amyloid oligomers. *J Biol Chem.* 2008;283:29639–43.
61. Chauhan NB. Intracerebroventricular passive immunization with anti-oligoAbeta antibody in TgCRND8. *J Neurosci Res.* 2007;85:451–63.
62. Britschgi M, Olin CE, Johns HT, Takeda-Uchimura Y, LeMieux MC, Rufibach K, et al. Neuroprotective natural antibodies to assemblies of amyloidogenic peptides decrease with normal aging and advancing Alzheimer's disease. *Proc Natl Acad Sci U S A.* 2009;106:12145–50.
63. Rosenmann H, Grigoriadis N, Karussis D, Boimel M, Touloumi O, Ovadia H, et al. Tauopathy-like abnormalities and neurologic deficits in mice immunized with neuronal tau protein. *Arch Neurol.* 2006;63:1459–67.
64. Lewis J, McGowan E, Rockwood J, Melrose H, Nacharaju P, van Slegtenhorst M, et al. Neurofibrillary tangles, amyotrophy and progressive motor disturbance in mice expressing mutant (P301L) tau protein. *Nat Genet.* 2000;25:402–5.
65. Augustinack JC, Schneider A, Mandelkow EM, Hyman BT. Specific tau phosphorylation sites correlate with severity of neuronal cytopathology in Alzheimer's disease. *Acta Neuropathol.* 2002;103:26–35.
66. Trinczek B, Biernat J, Baumann K, Mandelkow EM, Mandelkow E. Domains of tau protein, differential phosphorylation, and dynamic instability of microtubules. *Mol Biol Cell.* 1995;6:1887–902.



67. Su B, Wang X, Drew KL, Perry G, Smith MA, Zhu X. Physiological regulation of tau phosphorylation during hibernation. *J Neurochem*. 2008;105:2098–108.
68. Berriman J, Serpell LC, Oberg KA, Fink AL, Goedert M, Crowther RA. Tau filaments from human brain and from in vitro assembly of recombinant protein show cross-beta structure. *Proc Natl Acad Sci U S A*. 2003;100:9034–8.
69. Barghorn S, Davies P, Mandelkow E. Tau paired helical filaments from Alzheimer's disease brain and assembled in vitro are based on beta-structure in the core domain. *Biochemistry*. 2004;43:1694–703.
70. Weaver CL, Espinoza M, Kress Y, Davies P. Conformational change as one of the earliest alterations of tau in Alzheimer's disease. *Neurobiol Aging*. 2000;21:719–27.
71. Jicha GA, Bowser R, Kazam IG, Davies P. Alz-50 and MC-1, a new monoclonal antibody raised to paired helical filaments, recognize conformational epitopes on recombinant tau. *J Neurosci Res*. 1997;48:128–32.
72. Rosenmann H, Meiner Z, Geylis V, Abramsky O, Steinitz M. Detection of circulating antibodies against tau protein in its unphosphorylated and in its neurofibrillary tangles-related phosphorylated state in Alzheimer's disease and healthy subjects. *Neurosci Lett*. 2006;410:90–3.
73. Jacob S, Rajabally YA. Current proposed mechanisms of action of intravenous immunoglobulins in inflammatory neuropathies. *Curr Neuropharmacol*. 2009;7:337–42.
74. Durandy A, Kaveri SV, Kuijpers TW, Basta M, Miescher S, Ravetch JV, et al. Intravenous immunoglobulins—understanding properties and mechanisms. *Clin Exp Immunol*. 2009;158 Suppl 1:2–13.
75. Kivity S, Katz U, Daniel N, Nussinovitch U, Papageorgiou N, Shoenfeld Y. Evidence for the use of intravenous immunoglobulins—a review of the literature. *Clin Rev Allergy Immunol*. 2010;38:201–69.
76. Hughes RA, Dalakas MC, Comblath DR, Latov N, Weksler ME, Relkin N. Clinical applications of intravenous immunoglobulins in neurology. *Clin Exp Immunol*. 2009;158 Suppl 1:34–42.
77. Fillit H, Hess G, Hill J, Bonnet P, Toso C. IV immunoglobulin is associated with a reduced risk of Alzheimer disease and related disorders. *Neurology*. 2009;73:180–5.
78. Dodel R, Neff F, Noelker C, Pul R, Du Y, Bacher M, et al. Intravenous immunoglobulins as a treatment for Alzheimer's disease: rationale and current evidence. *Drugs*. 2010;70:513–28.
79. Neff F, Wei X, Nolker C, Bacher M, Du Y, Dodel R. Immunotherapy and naturally occurring autoantibodies in neurodegenerative disorders. *Autoimmun Rev*. 2008;7:501–7.
80. Du Y, Wei X, Dodel R, Sommer N, Hampel H, Gao F, et al. Human anti-beta-amyloid antibodies block beta-amyloid fibril formation and prevent beta-amyloid-induced neurotoxicity. *Brain*. 2003;126:1935–9.
81. Patrias LM, Klaver AC, Coffey MP, Loeffler DA. Specific antibodies to soluble alpha-synuclein conformations in intravenous immunoglobulin preparations. *Clin Exp Immunol*. 2010;161:527–35.
82. Szabo P, Relkin N, Weksler ME. Natural human antibodies to amyloid beta peptide. *Autoimmun Rev*. 2008;7:415–20.
83. Klaver AC, Finke JM, Digambaranath J, Balasubramaniam M, Loeffler DA. Antibody concentrations to Abeta1–42 monomer and soluble oligomers in untreated and antibody-antigen-dissociated intravenous immunoglobulin preparations. *Int Immunopharmacol*. 2010;10:115–9.
84. Klaver AC, Patrias LM, Coffey MP, Finke JM, Loeffler DA. Measurement of anti-Abeta1–42 antibodies in intravenous immunoglobulin with indirect ELISA: the problem of nonspecific binding. *J Neurosci Methods*. 2010;187:263–9.
85. Szabo P, Mujalli DM, Rotondi ML, Sharma R, Weber A, Schwarz HP, et al. Measurement of anti-beta amyloid antibodies in human blood. *J Neuroimmunol*. 2010;227:167–74.
86. Gomez-Isla T, Hollister R, West H, Mui S, Growdon JH, Petersen RC, et al. Neuronal loss correlates with but exceeds neurofibrillary tangles in Alzheimer's disease. *Ann Neurol*. 1997;41:17–24.
87. Van de Nes JA, Nafe R, Schlote W. Non-tau based neuronal degeneration in Alzheimer's disease—an immunocytochemical and quantitative study in the supragranular layers of the middle temporal neocortex. *Brain Res*. 2008;1213:152–65.
88. Kaye R, Pensalfini A, Margol L, Sokolov Y, Sarsoza F, Head E, et al. Annular protofibrils are a structurally and functionally distinct type of amyloid oligomer. *J Biol Chem*. 2009;284:4230–7.
89. Marx J. Alzheimer's disease. A new take on tau. *Science*. 2007;316:1416–7.
90. Meraz-Rios MA, Lira-De Leon KI, Campos-Pena V, de Anda-Hernandez MA, Mena-Lopez R. Tau oligomers and aggregation in Alzheimer's disease. *J Neurochem*. 2010;112:1353–67.
91. Lasagna-Reeves CA, Roi B, Guerrero-Muñoz MJ, Castillo-Carranza DL, Troncoso JC, Kaye R, et al. Role of Tau oligomers in Parkinson's disease and dementia with Lewy bodies. *Movement Disorders*. 2010;25:S181–565.
92. Dance A, Landhuis E, Strobel G. The Society for Neuroscience 2009 meeting report, part 2. *J Alzheimers Dis*. 2010;19:1409–15.
93. Andorfer C, Acker CM, Kress Y, Hof PR, Duff K, Davies P. Cell-cycle reentry and cell death in transgenic mice expressing nonmutant human tau isoforms. *J Neurosci*. 2005;25:5446–54.
94. Yoshiyama Y, Higuchi M, Zhang B, Huang SM, Iwata N, Saido TC, et al. Synapse loss and microglial activation precede tangles in a P301S tauopathy mouse model. *Neuron*. 2007;53:337–51.
95. Berger Z, Roder H, Hanna A, Carlson A, Rangachari V, Yue M, et al. Accumulation of pathological tau species and memory loss in a conditional model of tauopathy. *J Neurosci*. 2007;27:3650–62.
96. Spire TL, Orne JD, SantaCruz K, Pitstick R, Carlson GA, Ashe KH, et al. Region-specific dissociation of neuronal loss and neurofibrillary pathology in a mouse model of tauopathy. *Am J Pathol*. 2006;168:1598–607.
97. Maeda S, Sahara N, Saito Y, Murayama M, Yoshiike Y, Kim H, et al. Granular tau oligomers as intermediates of tau filaments. *Biochemistry*. 2007;46:3856–61.
98. Maeda S, Sahara N, Saito Y, Murayama S, Ikai A, Takashima A. Increased levels of granular tau oligomers: an early sign of brain aging and Alzheimer's disease. *Neurosci Res*. 2006;54:197–201.
99. Duhem C, Dicato MA, Ries F. Side-effects of intravenous immune globulins. *Clin Exp Immunol*. 1994;97 Suppl 1:79–83.
100. Carbone J. Adverse reactions and pathogen safety of intravenous immunoglobulin. *Curr Drug Saf*. 2007;2:9–18.

**Publish with Libertas Academica and every scientist working in your field can read your article**

*"I would like to say that this is the most author-friendly editing process I have experienced in over 150 publications. Thank you most sincerely."*

*"The communication between your staff and me has been terrific. Whenever progress is made with the manuscript, I receive notice. Quite honestly, I've never had such complete communication with a journal."*

*"LA is different, and hopefully represents a kind of scientific publication machinery that removes the hurdles from free flow of scientific thought."*

**Your paper will be:**

- Available to your entire community free of charge
- Fairly and quickly peer reviewed
- Yours! You retain copyright

<http://www.la-press.com>