



Complete Genome Sequence of a *Rhodococcus* Species Isolated from the Winter Skate *Leucoraja ocellata*

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We report here a genome sequence for *Rhodococcus* sp. isolate UM008 isolated from the renal/interrenal tissue of the winter skate *Leucoraja ocellata*. Genome sequence analysis suggests that *Rhodococcus* bacteria may act in a novel mutualistic relationship with their elasmobranch host, serving as biocatalysts in the steroidogenic pathway of 1 α -hydroxycorticosterone.

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lasmobranch fish (sharks, skates, and rays) possess a unique corticosteroid, 1α -hydroxycorticosterone (1α -OH-B), produced in the anatomically distinct interrenal gland (1, 2). The complete biosynthetic pathway remains to be elucidated, particularly with respect to the mechanism and enzyme(s) responsible for α -hydroxylation at C-1 (2). Microbial conversion may be a contributing factor, as the presence of a C₁₁ hydroxyl group in the β -configuration results in steric interactions favoring this unique α -configuration at C-1 (1, 3). Microbial hydroxylation of C₁₉ steroids at positions C-1 and C-2, as well as microbial 1α hydroxylation, have been demonstrated elsewhere (4, 5). In this view, we have recently isolated bacteria of the genus Rhodococcus from the renal/interrenal tissue of the winter skate Leucoraja ocellata (J. Wiens, R. Ho, A. K. C. Brassinga, C. A. Deck, P. J. Walsh, R. N. Ben, K. McClymont, A. N. Evans, and W. G. Anderson, unpublished data). Here, we report the genome sequence of the isolate designated Rhodococcus sp. UM008, determined by singlemolecule real-time (SMRT) sequencing.

The genome of the Rhodococcus isolate was assembled using Pacific Biosciences (PacBio) RS II long reads. Briefly, cultures were grown in tryptic soy broth (Difco), and genomic DNA was extracted using the UltraClean microbial DNA isolation kit (Mo Bio Laboratories), modifying the manufacturer's direction with the addition of incubation with mutanolysin (Sigma-Aldrich) to digest the cell wall (6), in accordance with the established protocol (7). A sequencing-ready PacBio RS II library with 15-kb inserts was prepared at the Génome Québec facility (Montreal, Québec, Canada) and sequenced using C2 chemistry on 3 single-molecule real-time (SMRT) cells, with 180-min movie collection. The combined data consisted of 205,395 reads, with an N_{50} size of 9.4 kb. The continuous long reads (CLR) were assembled de novo with the PacBio SMRT Analysis software (version 2.3) using the HGAP protocol (8), followed by polishing with Quiver. The NCBI Prokaryotic Genome Annotation Pipeline (9) was then used for functional genome annotation. The chromosome resolved to a single 6,570,200-bp contig with a G+C content of 62.35%. Functional genome annotation with the NCBI Prokaryotic Genome Annotation Pipeline (9) identified 6,050 genes, 15 rRNAs, 53 tRNAs, and 1 noncoding RNA. The chromosome is similar in size, content, and organization to *Rhodococcus erythropolis* PR4 (10). The genome also included three separate contigs representing distinct plasmids.

Genome sequence analysis has identified enzymes orthologous to those characterized in steroid-catabolizing *Actinobacteria* (11), indicating that intracellular *Rhodococcus* bacteria may be involved in the biosynthesis of corticosteroids. We suggest that *Rhodococcus* bacteria may act in a novel mutualistic relationship with their elasmobranch host, serving as biocatalysts in the steroidogenic pathway of 1α -OH-B.

Accession number(s). This genome project has been deposited in GenBank under accession numbers CP012749 (chromosome), CP015203 (plasmid 1), CP015204 (plasmid 2), and CP015205 (plasmid 3).

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REFERENCES

- 1. Idler DR, Truscott B. 1966. 1α-hydroxycorticosterone from cartilaginous fish: a new adrenal steroid in blood. J Fisheries Res Bd Can 23:615–619. http://dx.doi.org/10.1139/f66-053.
- Anderson WG. 2012. The endocrinology of 1α-hydroxycorticosterone in elasmobranch fish: a review. Comp Biochem Physiol 162:73–80. http:// dx.doi.org/10.1016/j.cbpa.2011.08.015.
- Schwarz V, Ulrich M, Syhora K. 1964. The isolation and the characterization of 1β-hydroxy-11-deoxycortisol as a by-product of a microbial

11-hydroxylation. Steroids 4:645–656. http://dx.doi.org/10.1016/0039 -128X(64)90064-9.

- 4. Dodson RM, Goldkamp AH, Muir RD. 1957. Microbiological hydroxylation of C₁₉-steroids at positions C-1 and C-2. J Am Chem Soc **79:3**921. http://dx.doi.org/10.1021/ja01571a081.
- 5. Ambrus G, Szarka E, Barta I, Horváth G. 1974. Microbiological 1α -hydroxylation of norethisterone. Steroids 25:99–106.
- 6. Assaf NA, Dick WA. 1993. Spheroplast formation and plasmid isolation from *Rhodococcus* spp. BioTechniques 15:1010–1012.
- 7. Lessard PA. Genomic DNA preparation from Corynebacterium and Rhodococcus, mini-protocol. Massachusetts Institute of Technology, Cambridge, MA. http://ocw.mit.edu/courses/biology/7-13-experimental -microbial-genetics-fall-2003/labs/Genomic_DNA_miniprep_v2.pdf.
- 8. Chin C-S, Alexander DH, Marks P, Klammer AA, Drake J, Heiner C, Clum A, Copeland A, Huddleston J, Eichler EE, Turner SW, Korlach J.

2013. Nonhybrid, finished microbial genome assemblies from long-read SMRT sequencing data. Nat Methods 10:563–569. http://dx.doi.org/ 10.1038/nmeth.2474.

- 9. Tatusova T, DiCuccio M, Badretdin A, Chetvernin V, Ciufo S, Li W. 2013. Prokaryotic genome annotation pipeline. The NCBI handbook, 2nd ed. National Center for Biotechnology, Bethesda, MD.
- Sekine M, Tanikawa S, Omata S, Saito M, Fujisawa T, Tsukatani N, Tajima T, Sekigawa T, Kosugi H, Matsuo Y, Nishiko R, Imamura K, Ito M, Narita H, Tago S, Fujita N, Harayama S. 2006. Sequence analysis of three plasmids harbored in *Rhodococcus erythropolis* strain PR4. Environ Microbiol 8:334–346. http://dx.doi.org/10.1111/j.1462-2920.2005.00899.x.
- Shtratnikova VY, Schelkunov MI, Fokina VV, Pekov YA, Ivashina T, Donova MV. 2016. Genome-wide bioinformatics analysis of steroid metabolism-associated genes in *Nocardioides simplex* VKM Ac-2033D. Curr Genet 62:643–656. http://dx.doi.org/10.1007/s00294-016-0568-4.