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T-cell immunodeficiency and reconstruction based on TCR rearrangement analysis in hematological malignancy: update from 2011 ASH annual meeting

Yangqiu Li

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Introduction

Poor cellular immune function may relate to carcinogenic processes and to worse prognosis in solid tumor patients as well as in leukemia. Moreover, the progression of tumor might further induce the cellular immune suppression. Therefore, a set of molecular immunological techniques to analyze and monitor the changes of host T-cell immune status is needed, which can fully characterize the feature of T-cell immunodeficiency in different malignancies, providing information and direction for immune reconstruction, in particular for enhancement the specific anti-tumor immune function.

The feature of T-cell immunodeficiency in hematological malignancies

In recent years, molecular analysis of the T cell receptor (TCR) utilization feature based on the principle of TCR α , β , γ and δ gene rearrangement and deletion rearrangement, has proven to be an effective technique for studying the distribution of T cell repertoire, the diversity of TCR subfamilies [1,2], the antigen specific expansion of T-cell clones and the recent thymic output function [3,4]. This in turn can help to characterize the feature of host T cell immune status, the identification of T-cell populations of interest in cancer, as well as the peripheral immune repertoire reconstitution after hematopoietic stem cell transplantation (HSCT).

T-cell immunodeficiency is a common feature in different hematological malignancies, including the absence

of TCR V α and V β subfamilies, decreased diversity of TCR repertoires, reduced thymic recent output function (naïve T cells) and lower frequencies of TCR subfamily naïve T cells. An impaired thymic export function and, as a consequence, altered ability to maintain T cell homeostasis may play an important pathogenic role in hematological malignancies. On the other hand, clonally expanded T cells could be identified in some TCR subfamilies in leukemia patients, which display specific anti-leukemia cytotoxicity like WT1 or BCR-ABL specific CTL, indicating that specific anti-leukemic T cells could be generated in vivo. This suggests that the host could have the ability of specific immune response to leukemia associated antigens, despite of T cell immunodeficiency.

T-cell immune reconstitution and establishment of specific anti-tumor and virus immunity

Prolonged period of immunodeficiency and poor immune reconstitution after stem cell transplantation place patients at high risk for viral infection and disease relapse, resulting in significant morbidity and mortality. Reversion of the cellular immunodeficiency is one of the crucial steps for improvement the outcome of tumor therapy in hematological malignancies. Moreover, the T-cell immune reconstitution is a key determinacy of long-term outcome in patients with hematological malignancies post chemotherapy or stem cell transplantation.

T-cell immune reconstitution requests not only the recovery of the comprehensive T-cell immunity, a broad TCR repertoire and recent thymic emigrants, more importantly, also the enhancement of the specific anti-tumor cellular immune function, which plays determinant role on elimination of minimal residual disease,

Correspondence: yangqiu@hotmail.com
Institute of Hematology, Medical College, and Key Laboratory for Regenerative Medicine of Ministry of Education, Jinan University, Guangzhou 510632, China

relapse prevention and improvement of prognosis in hematological malignancies. Antigen specific T-cell immune reconstitution could be carried out by active (cancer vaccine) or adoptive immunotherapy (T-cells transfusion) [5-8]. Cancer vaccines induce expansion and functional differentiation of tumor antigen-specific effectors and memory cells. The latter are particularly relevant for prevention of disease relapse. Adoptive antigen specific immunotherapy is one of the best approaches for tumor immunotherapy. The antigen specific CTL can directly kill tumor cells, ignoring the host immune status.

Antigen specific CTL could be amplified by cellular or gene engineering techniques. Peptide -specific stimulation in vitro can induce high-affinity CTL (auto- or allogenic) capable of recognizing tumor cells expressing the appropriate tumor antigen. For example, Epstein-Barr virus (EBV)-specific CTL were used to treat the post transplantation lymphoproliferative disease (PTLD) or EBV+ lymphoma, CMV-specific CTL were used to establish anti-CMV immunity in immunodeficiency patients post allogeneic stem cell transplantation [9,10]. Genetically-modified CTL were obtained by engineering antigen specific TCR gene, thus altering their original antigen specificity and arming them with new cytotoxicity for tumor cells. The approach provides a new strategy for adoptive specific immunotherapy in malignancies and so on. A lot of TCR-modified CTL against different leukemia and lymphoma were developed, like mHagHA-2, EBV, WT1, CML or DLBCL-specific TCR modified CTL [5-8,11,12], as well as the single-chain antibody-derived chimeric antigen receptors (CARs) modified T cells that specifically recognize surface molecules expressed on malignant B cells (CD19) or acute myeloid leukemia cells (CD33) independent from HLA [13,14].

In summary, dynamic detection of the alteration of host immune function in patients is important for the defense of host against neoplastic transformation and so on. The new techniques of tumor immunology, molecular biology and increased knowledge of the optimal methodology for generation of T-cell products and optimization of gene therapy approaches make it possible to enhance the function of adoptively transferred T cells. This enhances tumor- specific response and can reverse the host immunodeficiency status.

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References

1. Krell PFI, Weber S, Reuther S, Gombert M, Keller T, Schuster FR, Asang C, Stoye J, Borkhardt A, Fischer U: **Next Generation Sequencing Spectratyping (NGS-S) Comprehensively Monitors T Cell Receptor Diversity in Children with T Cell Abnormalities.** *ASH Annual Meeting Abstracts* 2011, **118**(21):#2173.
2. Xuan L, Xiuli Wu X, Liu Q, Zhang Y, Fan Z, Li Y, Ling Y: **G-CSF Affects the Distribution and Clonality of TRGV and TRDV Repertoire of T Cells and the Expression Pattern of CD3 Genes.** *ASH Annual Meeting Abstracts* 2011, **118**(21):#1946.
3. Wu X, Zhu K, Du X, Chen S, Yang L, Wu J, Liu Q, Li Y: **Frequency analysis of TRBV subfamily sJTRECs to characterize T-cell reconstitution in acute leukemia patients after allogeneic hematopoietic stem cell transplantation.** *J Hematol Oncol* 2011, 4:19.
4. Brown J, Kim HT, Cutler C, McDonough S, Alyea E, Ho V, Attar EC, Dey BR, McAfee SL, Spitzer T, et al: **Immune Reconstitution After Cord Blood Transplantation in Adults Depends on Activity of Thymic Epithelial Cells and Vascular Endothelial Elements.** *ASH Annual Meeting Abstracts* 2011, **118**(21):#4075.
5. Porter DL: **Allogeneic immunotherapy to optimize the graft-versus-tumor effect: concepts and controversies.** *ASH Education Program Book* 2011, 2011(1):292-298.
6. Zhang M, Sukhumalchandra P, Enyenihi AA, St. John LS, Hunsucker SA, Ruisaard K, Atrache Z, Ropp PA, Rodriguez-Cruz T, Mittendorf E, et al: **A Novel HLA-A2 Restricted Peptide Derived From Cathepsin G Is An Effective Immunotherapeutic Target for Myeloid Leukemia.** *ASH Annual Meeting Abstracts* 2011, **118**(21):#2986.
7. Zohren F, Imperato MR, Usanarat A, Spencer DM, Heslop HE, Brenner MK, Rooney CM, Leen AM, Vera JF, Gerdemann U: **Genetic Modification of Multi Leukemia Antigen-Specific Cytotoxic T Lymphocytes (CTL) to Enhance In Vivo Safety and Persistency.** *ASH Annual Meeting Abstracts* 2011, **118**(21):#644.
8. Weber G, Gerdemann U, Hensel NF, Leen AM, Bollard CM, Barrett AJ: **Generation of Multi-Antigen Specific T Cells for Adoptive Immunotherapy of Myeloid Leukemia and Identification of MHC Class I and II-Restricted Peptides for WT1, Proteinase 3 and Human Neutrophil Elastase.** *ASH Annual Meeting Abstracts* 2011, **118**(21):#2985.
9. Bao L, Cowan MJ, Dunham K, Horn BN, McGuirk J, Gilman A, Lucas K: **Adoptive Immunotherapy with CMV Specific Cytotoxic T Lymphocytes for Stem Cell Transplant Patients with Refractory CMV Infections.** *ASH Annual Meeting Abstracts* 2011, **118**(21):#4033.
10. Hanley PJ, Martinez C, Leung K, Savaldo B, Dotti G, Gee AP, Rooney CM, Heslop HE, Krance RA, Shpall EJ, et al: **Phase I Study to Improve Virus-Specific Immune Reconstitution After Cord Blood Transplantation Using Cord Blood-Derived Virus-Specific Cytotoxic T Lymphocytes.** *ASH Annual Meeting Abstracts* 2011, **118**(21):#155.
11. Yin Q, Zha X, Yang L, Chen S, Zhou Y, Wu X, Li Y: **Generation of diffuse large B cell lymphoma-associated antigen-specific Va6/β13+T cells by TCR gene transfer.** *J Hematol Oncol* 2011, 4:2.
12. Ochi T, Fujiwara H, Okamoto S, Asai H, Miyazaki Y, Shirakata T, Mineno J, Kuzushima K, Shiku H, Yasukawa M: **Redirected CD4+ T Cells Using WT1-Specific T-Cell Receptor Gene Transfer Can Supply Multifactorial Help to Enhance the Anti-Leukemia Reactivity Mediated by Similarly Redirected CD8+ T Cells Using the Identical Gene Transfer.** *ASH Annual Meeting Abstracts* 2011, **118**(21):#645.
13. Terakura S, Yamamoto TN, Gardner RA, Turtle CJ, Jensen MC, Riddell SR: **Generation and Signaling Function of CD19 Chimeric Antigen Receptor Modified CD8+ T Cells Derived From Virus-Specific Central Memory Cells for Adoptive Therapy After Allogeneic Hematopoietic Stem Cell Transplant.** *ASH Annual Meeting Abstracts* 2011, **118**(21):#2978.
14. Cartellieri M, Michalk I, von Bonin M, Krüger T, Stamova S, Koristka S, Arndt C, Feldmann A, Schmitz M, Wermke M, et al: **Chimeric Antigen Receptor-Engineered T Cells for Immunotherapy of Acute Myeloid Leukemia.** *ASH Annual Meeting Abstracts* 2011, **118**(21):#2618.

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