Effect of simvastatin on expression of VEGF and TGF-β1 in atherosclerotic animal model of type 2 diabetes mellitus

HAIYAN WANG¹, JIAN LI², XUDONG FU¹, YU LIU², QINGHUA XU¹ and LILI SANG¹

Departments of ¹Cadre Health Protection and ²Endocrinology, Liaocheng People's Hospital, Liaocheng, Shandong 252000, P.R. China

Received April 17, 2018; Accepted July 25, 2018

DOI: 10.3892/etm.2018.6583

Abstract. Expression of vascular endothelial growth factor (VEGF) and transforming growth factor-β1 (TGF-β1) in atherosclerosis animal model of type 2 diabetes mellitus treated with simvastatin was investigated. Clean grade mature Sprague Dawley (SD) rats were divided into three groups: Normal control (n=10), model (n=13) and treatment group (n=13); lowdose simvastatin was administered. The changes of VEGF and TGF-β1 levels were analyzed by tail vein blood sampling. The relationship between levels of VEGF, TGF-β1 and treatment time was analyzed. The expression level of VEGF in the treatment group after 4 and 8 weeks of intervention was lower compared with the model group (P<0.05). The expression level of TGF-β1 in the treatment group after 8 weeks of intervention was higher than that in the model group (P<0.05). The expression level of VEGF in the treatment group after 8 weeks of intervention was lower than that after 1 week of intervention (P<0.05). The expression level of TGF-β1 was increased in the model group after 8 weeks of intervention compared with 1 week before and after the intervention (P<0.05). The expression level of TGF-β1 in the treatment group at 2, 4 and 8 weeks after intervention were significantly higher than that before intervention (P<0.05). The expression of TGF-β1 increased after 4 and 8 weeks after intervention compared with 1 week after intervention (P<0.05). The expression of VEGF was negatively correlated with TGF-β1 expression in the treatment group; negative correlation was found between VEGF and treatment time. There was a positive correlation between TGF-β1 and treatment time. VEGF and TGF-β1 may be involved in the development of type 2 diabetes (T2MD) atherosclerosis (AS). Simvastatin may play a therapeutic role in T2MD AS by downregulating VEGF and upregulating the expression of TGF-β1.

Key words: simvastatin, T2MD, atherosclerosis, VEGF, TGF-β1

Introduction

The incidence of type 2 diabetes (T2MD) is increasing as people's living standards increase, population ageing increases, and life structure changes. It is expected that T2MD will become the seventh largest cause of death in the world by 2030 (1,2). The major cause of death in patients with T2MD is major vascular complications (3). According to reports from the National Institutes of Health, 80% of patients with T2MD died of macrovascular complications directly (4). Therefore, how to treat T2MD macrovascular complications has become a global public health problem.

Atherosclerosis (AS) is the basis of T2MD macrovascular complications. It is prone to occur early in patients with T2MD, and the progression of AS is rapid and the prognosis of patients is poor. At present, the pathogenesis of AS in T2MD patients is not yet fully determined (5,6). Many researchers have found that the damage and dysfunction of the vascular endothelium, long-term chronic inflammatory response mediated by inflammatory factors and other reasons are important mechanisms for the occurrence and development of T2MD AS. Among them, vascular endothelial growth factor (VEGF) and transforming growth factor-β1 (TGF-β1) are the hot topics discussed in recent years. VEGF and TGF-β1 are abnormally expressed in patients with T2MD AS (5,7). Simvastatin is a competitive inhibitor of HMG-CoA reductase, and can block cholesterol synthesis (8). Studies have reported that simvastatin, in addition to regulating blood lipids, also has the effects of inhibiting proliferation of vascular smooth muscle cells, stabilizing AS plaques, resisting oxidative stress, preventing platelet aggregation, and improving endothelial function (9,10). However, its specific mechanism of action has not yet been studied clearly.

In this study, a rat model of T2MD was established to observe the effect of simvastatin on the expression of VEGF and TGF-β1 in rats and to investigate the mechanism of simvastatin.

Materials and methods

Research objects. A total of 40 clean grade mature Sprague Dawley (SD) rats were purchased from Animal Center of Xinjiang Medical University [production license SCXK (new) 2003-0001] and were fed with normal dry feed (Beijing Zhecheng Technology Co., Ltd., Beijing, China). SD rats were

Correspondence to: Dr Lili Sang, Department of Cadre Health Protection, Liaocheng People's Hospital, 67 Dongchang Xi Road, Liaocheng, Shandong 252000, P.R. China E-mail: aseqau@163.com

 \sim 3 weeks old with an average age of 21.3 \pm 1.5 days, weight 150-160 g, average body weight 154.1±12.2 g, rearing temperature was 20-27˚C, relative humidity 70±10%, separate feeding in the terrarium, changing the litter every morning and evening, environmental noise below 85 dB, ammonia concentration not exceeding 20 ppm, ventilation 8-12 times per hour, fluorescent lamp light every 12 h, free food intake, free drinking water, feeding box was replaced 1-2 times a week. The humidity in the feeding box did not exceed 10%. Τhe water bottle was changed 1-2 times a week. Ten random-number table methods were used to select 10 rats as normal control groups, and the remaining 30 were used to establish T2MDAS rat models. After successful modeling, they were randomly divided into model and treatment group. The study was approved by the Ethics Committee of Liaocheng People's Hospital (Liaocheng, China).

Rat model establishment. Thirty rats were used to establish T2MD AS rat model: High-fat diet (Keaoxieli Co., Ltd., Beijing, China) for 8 weeks, immediately after 8 weeks of feeding, fasted for 12 h, and given a single intraperitoneal injection of Streptozocin solution (Shanghai Baoman Biotechnology Co., Ltd., Shanghai, China) 45 mg/kg, after 72 h measured tail vein blood glucose, fasting blood glucose ≥7.0 mmol/l, and postprandial blood glucose ≥11.1 mmol/l was set as the evaluation criteria for successful T2MD model. Establishment of AS rat model: T2MD successfully modeled rats were intragastrically administered with vitamin D3 injection (SFDA approval no. 31021259; Wuhan Dongkangyuan Technology Co., Ltd., Wuhan, China), and were administered at a total dose of 500,000 U/kg for 3 days. The normal control group was fed with normal dry feed.

Treatment method. After simvastatin tablets (Shenzhen Roark Standards Technology Co., Ltd., SFDA Approval no. H20123114) was ground into a powder, 0.9% physiological saline was formulated into a 1 mg/ml suspension, and the treatment group was given a 10 mg/kg/day simvastatin suspension intragastrically for drug intervention. The normal control and model group were given 0.9% physiological saline 5 mg/kg/day gavage as a control. Treatment was performed for 8 weeks. The normal control group was fed with normal dry feed, and the model and treatment group were still fed with high-fat diet.

ELISA. Serum VEGF and TGF-β1 were detected by ELISA. The procedure was with reference to the instructions of the kit. VEGF and TGF-β1 kits were purchased from Shanghai Jing Kang Biological Engineering Co., Ltd. (Shanghai, China).

Observation indicators. The changes of VEGF and TGF-β1 levels before and after modeling (before intervention), 1, 2, 4, and 8 weeks after intervention were analyzed by tail vein blood sampling. The relationship between VEGF, TGF-β1 levels and treatment time was analyzed.

Statistical analysis. SPSS 19.0 (SPSS, Inc., Chicago, IL, USA) was adopted. Enumeration data are expressed as rate, and rates were compared by using the χ^2 test. The measurement data are expressed as mean \pm SD. Analysis of variance was used for comparison among multiple groups, LSD tests were used for pairwise comparison, and repeated-variance measurement experiments or paired t-tests were used as comparison at different times within the group. The correlation between VEGF and TGF-β1 levels was analyzed by using Pearson's correlation. P<0.05 was statistically significant.

Results

Modeling results. T2MD rat models were first constructed in 30 rats. A total of 28 rats were successfully modeled. The success rate of modeling was 93.33%. Then AS rat model was established. Finally, 26 rats were successfully modeled. The success rate of modeling was 92.86%. There was no difference in body weight, fasting blood glucose, VEGF and TGF-β1 between the three groups before modeling (P>0.05). After successful modeling/before intervention, fasting blood glucose, VEGF, and TGF-β1 in the normal group were not different from those before modeling. Body weight, fasting blood glucose, VEGF, and TGF-β1 were increased in the model group and the treatment group at different degrees (P<0.05). The weight of rats before intervention in the normal group was higher than that before modeling (P<0.05). The vital signs before and after modeling in the three groups of rats are shown in Table I.

Changes of serum VEGF levels in three groups before and after intervention. There was no difference in serum VEGF levels between the three groups before modeling (P>0.05). Αnalysis of variance showed there were differences in the expression of VEGF among three groups of rats before intervention, 1, 2, 4, and 8 weeks after intervention (P<0.05). The LSD test results showed that the VEGF expression levels in the model and the treatment group at the five time-points were higher than those in the normal control group (P<0.05). The expression level of VEGF in the treatment group was lower than the model group after 4 weeks and 8 weeks after the intervention (P<0.05), and there was no difference at other time-points (P>0.05). The results of repeated analysis of variance at different time-points within the group showed that the expression levels of VEGF in the normal control group and model group had no significant changes before and after the intervention (P>0.05). Τhe expression level of VEGF in the treatment group after 4 and 8 weeks of intervention was lower than that before the intervention (P<0.05). Τhe expression level of VEGF in the treatment group after 8 weeks of intervention was lower than that after 1 week (P<0.05), and there was no difference between any of the other time-points (P>0.05) (Table II and Fig. 1).

Changes of serum TGF-β1 levels in three groups before and after intervention. There was no difference in the levels of serum TGF-β1 between the three groups before modeling (P>0.05). The analysis of variance showed that the expression levels of TGF-β1 of the three groups of rats were different before the intervention, 1, 2, 4, and 8 weeks after the intervention (P<0.05). The results of LSD showed that the expression levels of TGF-β1 in the model and the treatment group at the five time-points were higher than those in the normal control group (P<0.05). The expression level of TGF-β1 in the treatment was higher than that in the model group after

Table I. Vital signs before and after modeling in three groups of rats.

a Comparison within group, P<0.05 compared with before modeling. VEGF, vascular endothelial growth factor.

Table II. Changes of serum VEGF levels in rats before and after intervention $(\mu g/I)$.

^aP<0.05 before intervention in the same group; ^bP<0.05 after 1 week of intervention in the same group; ^ccompared with the control group at the same time-point $(P<0.05)$; $\text{d}P<0.05$ compared with the model group at the same time-point. VEGF, vascular endothelial growth factor.

8 weeks of intervention (P<0.05), and there was no difference at other time-points (P>0.05). The results of repeated analysis of variance at different time-points within the group showed that the expression level of TGF-β1 in normal control rats did not change significantly before and after intervention (P>0.05). Τhe expression level of TGF-β1 in the model group 8 weeks after intervention was higher than that before intervention and 1 week after intervention (P<0.05), and there was no difference in the expression level of TGF-β1 at other time-points in the model group (P>0.05). The expression level of TGF-β1 in the treatment group after 2, 4 and 8 weeks after intervention was higher than before intervention (P<0.05). After 4 and 8 weeks of intervention, the expression level of TGF-β1 was higher than that after 1 week of intervention (P<0.05). There was no difference between the other time-points in the treatment group (P>0.05) (Table III and Fig. 2).

Correlation analysis of VEGF and TGF-β1. Pearson's correlation analysis showed that the expression of VEGF was negatively correlated with TGF-β1 expression in the treatment group (r=-0.712, P=0.015). Νegative correlation was found between VEGF and treatment time (r=-0.769, P=0.003). There was a positive correlation between TGF-β1 and treatment time (r=0.812, P=0.001). (Data not shown).

Discussion

Studies have reported that simvastatin can mediate CD40-CD40L signaling pathway in diabetic rats, reduce inflammation in arterial walls of diabetic rats, and improve vascular endothelium and its function in rats (11). However, there are many factors that cause T2MD AS, in addition to CD40-CD40L-related factors, whether VEGF and TGF-β1 play an important role in the treatment of diabetic rats AS with simvastatin is not yet clear and was the focus of this study.

A T2MD AS rat model was established in 2 steps. Clean SD rats were used to establish a T2MD rat model by feeding of high-fat diet for 8 weeks and using Streptozocin solution

Table III. Changes of serum TGF-β1 levels in rats before and after intervention $(\mu g/I)$.

^aP<0.05 before intervention in the same group; ^bP<0.05 after 1 week of intervention in the same group; ^ccompared with the control group at the same time-point (P<0.05); ^dP<0.05 compared with the model group at the same time-point. TGF-β1, transforming growth factor-β1.

Figure 1. Changes of serum VEGF levels in three groups of rats before and after intervention. The expression of VEGF in the three groups before the intervention, 1, 2, 4 and 8 weeks after the intervention were all different (P<0.05). The expression level of VEGF in the non-intervention model and the treatment group at the five time‑points was higher than that in the normal control group (P<0.05). The expression level of VEGF was lower in the treatment group than in the non-intervention model group 4 and 8 weeks after intervention (P<0.05). The expression of VEGF in the treatment group 4 and 8 weeks after intervention was lower than that before the intervention (P<0.05). The expression level of VEGF in the treatment group 8 weeks after intervention was lower than that of 1 week after intervention $(P<0.05)$. P<0.05 compared with pre-intervention; ^bP<0.05 compared with 1 week after intervention; °P<0.05 compared with no intervention model group at the same time-point. VEGF, vascular endothelial growth factor.

to destroy rat islet B cells. High-dose vitamin D disrupted the integrity of rat artery wall and replicated AS pathological model. After successful modeling, body weight and fasting blood glucose were significantly increased in rats, and VEGF and TGF-β1 were also significantly elevated. Therefore, we initially speculated that VEGF and TGF-β1 is involved in the development of rat T2MD AS. Then we used low-dose simvastatin to treat Figure 2. Changes of serum TGF-β1 levels in the three groups before and after intervention. The expression of TGF-β1 in the three groups before the intervention, 1, 2, 4 and 8 weeks after the intervention were all different (P<0.05). The expression levels of TGF-β1 in the non-intervention model and the treatment group at the five time-points were higher than those in the normal control group (P<0.05). The expression level of TGF-β1 in the treatment group was higher 8 weeks after the intervention than in the non-intervention model group (P<0.05). The expression level of TGF-β1 in the non-intervention model group was higher than that before the intervention and 1 week after the intervention (P<0.05). There was no significant difference in the expression level of TGF-β1 at the other time-points in the non-intervention model group (P>0.05). The expression level of TGF-β1 in the treatment group was higher at 2, 4, and 8 weeks after intervention than that before the intervention (P<0.05), and the expression level of TGF- $β1$ after 4 and 8 weeks after intervention was higher than that at 1 week after intervention (P<0.05). ^aP<0.05 compared with pre-intervention; ^bindicates P<0.05 compared with 1 week after intervention; °P<0.05 compared with no intervention model group at the same time-point. VEGF, vascular endothelial growth factor; TGF-β1, transforming growth factor-β1.

13 T2MD AS rats in the treatment group. With the increase of treatment time, VEGF showed a decreasing trend. Until 4 weeks after treatment, the VEGF levels in the treatment group rats showed a significant difference compared with those before the intervention, and there was also a statistically significant difference in VEGF levels between the treatment and the model group, indicating that simvastatin can improve VEGF levels in T2MD rats. The occurrence of AS in humans or rats causes insufficient blood supply to the arteries, resulting in endothelial cells secreting higher VEGF (8,12). Some studies have reported (13) that VEGF can activate the VEGFR2/KDR/flk1 signaling pathway, increase vascular permeability, promote adhesion of inflammatory cells in the blood vessel wall, and infiltration of inflammatory cells, promote neovascularization, and aggravate AS conditions. VEGF plays an important role in the occurrence of T2MD AS. In the present study, the results of TGF-β1 detection found that with the increase of treatment time, TGF-β1 showed an upward trend. After 2 weeks of treatment, serum TGF-β1 levels were statistically different from those before the intervention, indicating that simvastatin can increase TGF-β1 levels. Correlation analysis showed that VEGF was negatively correlated with treatment time, and TGF-β1 was positively correlated with treatment time. The inflammatory response is an important cause of plaque instability in AS, so improving the excessive inflammatory response is an important process for the treatment of AS (14,15). TGF-β1 has a strong non-specific anti-inflammatory effect (16). It was reported (17) that TGF-β1 can control the local inflammatory response due to AS by downregulating E-selectin and inhibiting the activity of CD4+ T cells. The proliferation and migration of endothelial cells and the activation of the coagulation system due to endothelial cell damage are also important causes of AS (18,19). Some reports (20) indicated that TGF-β1 can activate the TGF-β1/ALK5/Smad3 signaling pathway and promote the expression of endothelin-1, thus hindering the proliferation and migration of endothelial cells. It has also been reported (21) that TGF-β1 can promote endothelial cells to express more prostacyclin, inhibit platelet aggregation, and prevent thrombosis. Therefore, TGF-β1 also plays an important role in the development of T2MD AS. Simvastatin can improve AS by improving the levels of VEGF and TGF-β1 in serum of T2MD rats. However, the trial of simvastatin in the treatment of diabetes generally lasted for more than 2 years. Since the animal model was used as the subject for analysis in this experiment, the analysis results only represented short-term therapeutic effects, and the long-term treatment effect could not be evaluated, and whether it has the same effect in human still needs to be studied.

In conclusion, VEGF, TGF-β1 may be involved in the occurrence of T2MD AS, and simvastatin may play a therapeutic role in T2MD AS by downregulating VEGF and upregulating TGF-β1 expression.

Acknowledgements

Not applicable.

Funding

No funding was received.

Availability of data and materials

The datasets used and/or analyzed during the present study are available from the corresponding author on reasonable request.

Authors' contributions

HW wrote the manuscript. HW, JL and XF constructed T2MD rat models. YL and QX were responsible for ELISA. LS helped with statistical analysis. All authors read and approved the final manuscript.

Ethics approval and consent to participate

The study was approved by the Ethics Committee of Liaocheng People's Hospital (Liaocheng, China).

Patient consent for publication

Not applicable.

Competing interests

The authors declare that they have no competing interests.

References

- 1. American Diabetes Association: Standards of medical care in diabetes-2015 abridged for primary care providers. Clin Diabetes 33: 97-111, 2015.
- 2. Ferrannini E, Baldi S, Frascerra S, Astiarraga B, Heise T, Bizzotto R, Mari A, Pieber TR and Muscelli E: Shift to fatty substrate utilization in response to sodium-glucose cotransporter 2 inhibition in subjects without diabetes and patients with type 2 diabetes. Diabetes 65: 1190-1195, 2016.
- 3. Nolan CJ, Ruderman NB, Kahn SE, Pedersen O and Prentki M: Insulin resistance as a physiological defense against metabolic stress: Implications for the management of subsets of type 2 diabetes. Diabetes 64: 673-686, 2015.
- 4. American Diabetes Association: Standards of medical care in diabetes - 2014. Diabetes Care 37 (Suppl 1): S14-S80, 2014.
- 5. Catapano AL, Graham I, De Backer G, Wiklund O, Chapman MJ, Drexel H, Hoes AW, Jennings CS, Landmesser U, Pedersen TR, *et al*; Authors/Task Force Members: 2016 ESC/ EAS guidelines for the management of dyslipidaemias: The task force for the management of dyslipidaemias of the European Society of Cardiology (ESC) and European Atherosclerosis Society (EAS) Developed with the special contribution of the European Assocciation for Cardiovascular Prevention & Rehabilitation (EACPR). Atherosclerosis 253: 281-344, 2016.
- 6. Li H, Horke S and Förstermann U: Vascular oxidative stress, nitric oxide and atherosclerosis. Atherosclerosis 237: 208-219, 2014.
- 7. Alexopoulos N, Katritsis D and Raggi P: Visceral adipose tissue as a source of inflammation and promoter of atherosclerosis. Atherosclerosis 233: 104-112, 2014.
- 8. Bhakdi S: Pathogenesis of atherosclerosis: Infectious versus immune pathogenesis. A new concept. Herz 25: 84-86, 2000.
- 9. Murphy SA, Cannon CP, Blazing MA, Giugliano RP, White JA, Lokhnygina Y, Reist C, Im K, Bohula EA, Isaza D, et al: Reduction in total cardiovascular events with ezetimibe/simv-
astatin post-acute coronary syndrome: The IMPROVE-IT Trial. J Am Coll Cardiol 67: 353-361, 2016.
- 10. Criner GJ, Connett JE, Aaron SD, Albert RK, Bailey WC, Casaburi R, Cooper JA Jr, Curtis JL, Dransfield MT, Han MK, *et al*; COPD Clinical Research Network; Canadian Institutes of Health Research: Simvastatin for the prevention of exacerbations in moderate-to-severe COPD. N Engl J Med 370: 2201-2210, 2014.
- 11. Xue L, Zhu XH, Yang XF, Bao XC, Gao XQ, Qiu YH, Wu Z, Ji XP and Li HW: Effect of pioglitazone combined with simvastatin on the CD40-CD40 ligand system in rabbits with atherosclerosis. Eur Rev Med Pharmacol Sci 19: 322-327, 2015.
- 12. Al Rifai M, Silverman MG, Nasir K, Budoff MJ, Blankstein R, Szklo M, Katz R, Blumenthal RS and Blaha MJ: The association of nonalcoholic fatty liver disease, obesity, and metabolic sclerosis: The Multi-Ethnic Study of Atherosclerosis (MESA). Atherosclerosis 239: 629-633, 2015.
- 13. Kim SH, Pei QM, Jiang P, Yang M, Qian XJ and Liu JB: Effect of active vitamin D3 on VEGF-induced ADAM33 expression and proliferation in human airway smooth muscle cells: Implications for asthma treatment. Respir Res 18: 7, 2017.
- 14. Mangge H, Weghuber D, Prassl R, Haara A, Schnedl W, Postolache TT and Fuchs D: The role of vitamin D in atherosclerosis inflammation revisited: More a bystander than a player? Curr Vasc Pharmacol 13: 392-398, 2015.
- 15. Lutgens E, Lievens D, Beckers L, Wijnands E, Soehnlein O, Zernecke A, Seijkens T, Engel D, Cleutjens J, Keller AM, *et al*: Deficient CD40-TRAF6 signaling in leukocytes prevents atherosclerosis by skewing the immune response toward an antiinflammatory profile. J Exp Med 207: 391-404, 2010.
- 16. Kotlarz D, Marquardt B, Barøy T, Lee WS, Konnikova L, Hollizeck S, Magg T, Lehle AS, Walz C, Borggraefe I, *et al*: Human TGF-β1 deficiency causes severe inflammatory bowel disease and encephalopathy. Nat Genet 50: 344-348, 2018.
- 17. Edwards JP, Hand TW, Morais da Fonseca D, Glass DD, Belkaid Y and Shevach EM: The GARP/Latent TGF-β1 complex on Treg cells modulates the induction of peripherally derived Treg cells during oral tolerance. Eur J Immunol 46: 1480-1489, 2016.
- 18. Gerdes N, Seijkens T, Lievens D, Kuijpers MJ, Winkels H, Projahn D, Hartwig H, Beckers L, Megens RT, Boon L, *et al*: Platelet CD40 exacerbates atherosclerosis by transcellular activation of endothelial cells and leukocytes. Arterioscler Thromb Vasc Biol 36: 482-490, 2016.
- 19. Gimbrone MA Jr and García-Cardeña G: Endothelial cell dysfunction and the pathobiology of atherosclerosis. Circ Res 118: 620-636, 2016.
- 20. Li Y, Zou N, Wang J, Wang KW, Li FY, Chen FX, Sun BY and Sun DJ: TGF-β1/Smad3 signaling pathway mediates T-2 toxin-induced decrease of type II collagen in cultured rat chondrocytes. Toxins (Basel) 9: 9, 2017.
- 21. Hwangbo C, Tae N, Lee S, Kim O, Park OK, Kim J, Kwon SH and Lee JH: Syntenin regulates TGF-β1-induced Smad activation and the epithelial-to-mesenchymal transition by inhibiting caveolinmediated TGF-β type I receptor internalization. Oncogene 35: 389-401, 2016.

COOO This work is licensed under a Creative Commons Attribution-NonCommercial-NoDerivatives 4.0 International (CC BY-NC-ND 4.0) License.