



Higher Wheal Sizes of Dermatophagoides farinae Sensitization Exhibit Worse Nasal Symptoms in Allergic Rhinitis Patients

Siti Muhamad Nur Husna¹, Norasnieda Md Shukri^{2,3}, Hern-Tze Tina Tan¹, Noor Suryani Mohd Ashari^{1,3} and Kah Keng Wong^{1,3*}

OPEN ACCESS

Edited by:

Salvatore Leonardi, University of Catania, Italy

Reviewed by:

Sara Manti, University of Catania, Italy Giuseppe Fabio Parisi, University of Catania, Italy

*Correspondence:

Kah Keng Wong kahkeng@usm.my orcid.org/0000-0001-7359-6202

Specialty section:

This article was submitted to Pulmonary Medicine, a section of the journal Frontiers in Medicine

Received: 25 December 2021 Accepted: 07 February 2022 Published: 28 February 2022

Citation:

Nur Husna SM, Md Shukri N, Tan H-TT, Mohd Ashari NS and Wong KK (2022) Higher Wheal Sizes of Dermatophagoides farinae Sensitization Exhibit Worse Nasal Symptoms in Allergic Rhinitis Patients. Front. Med. 9:843432. doi: 10.3389/fmed.2022.843432 ¹ Department of Immunology, School of Medical Sciences, Universiti Sains Malaysia, Kubang Kerian, Malaysia, ² Department of Otorhinolaryngology, Head and Neck Surgery, School of Medical Sciences, Universiti Sains Malaysia, Kubang Kerian, Malaysia, ³ Hospital Universiti Sains Malaysia, Kubang Kerian, Malaysia

Allergic rhinitis (AR) is a global health burden and it manifests in both nasal and nonnasal symptoms. Skin prick test (SPT) is a routine procedure to diagnose AR sensitized to common allergens including house dust mites (HDMs). The degree of sensitivity of a patient toward allergens is determined by the size of the wheal formed by SPT procedure. SPT wheal sizes are influenced by recent anti-histamine usage, however it remains unclear if SPT wheal sizes are also influenced by other factors. In this study, we set out to investigate the association between SPT wheal sizes with the demographical, clinical and environmental characteristics, as well as nasal and non-nasal symptoms severity scores, of AR patients (n = 30) sensitized to common HDMs (i.e., Dermatophagoides pteronyssinus, Dermatophagoides farinae, and Blomia tropicalis). We showed that SPT wheal sizes of HDM allergens were not associated with clinical, demographical and environmental characteristics examined. Nonetheless, significant correlations were observed between SPT wheal sizes of D. farinae sensitization with worse severity scores of all five nasal symptoms examined (i.e., sneezing, runny nose, itchy nose, congestion and postnasal drip) and four of the six non-nasal symptoms examined (i.e., throat symptoms, ear symptoms, headache and mental function). Such relationships were not observed in SPT wheal sizes of D. pteronyssinus and B. tropicalis sensitization. We suggest that increased SPT wheal sizes for D. farinae sensitization may predict the likelihood of more severe nasal and, to a lesser extent, non-nasal manifestations in AR patients.

Keywords: allergic rhinitis, wheal size, Dermatophagoides farinae, nasal symptoms, house dust mite

1

INTRODUCTION

Allergic rhinitis (AR) is a global health burden that affects \sim 400 million people globally (1, 2). The disease significantly derails the quality of life and requires persistent treatment, leading to social-economic consequences. Moreover, AR is frequently accompanied with comorbidities such as asthma, sinusitis and conjunctivitis, complicating treatment and management of AR patients (3, 4).

Both skin prick test (SPT) and specific immunoglobulin E (sIgE) immunoassay have good correlation in terms of specificity and sensitivity in the diagnosis of sensitization to common allergens (5–7). The main house dust mite (HDM) allergens AR patients sensitized to include *Dermatophagoides pteronyssinus* (*D. pteronyssinus*), *Dermatophagoides farinae* (*D. farinae*) and *Blomia tropicalis* (*B. tropicalis*), and ~80% of AR patients are sensitized to these three HDM species (7–11).

The general principles of SPT include the introduction of relevant allergens into the skin, causing sIgE to bind surface receptors on mast cells, inducing mast cells to degranulate and release of inflammatory mediators including histamine (12). Histamine is responsible to promote capillary dilation, increases vascular permeability, induces pain nociceptors, and stimulates eosinophil chemotaxis to the inflamed tissue. The exudation then enters the skin and causes swelling accompanied by itching (12, 13). As a result, this produces a wheal and flare response whose diameter can be measured.

Allergen sensitization is a key risk factor for the development of allergic diseases. Therefore, it is essential to determine the individual's allergen sensitization for the diagnosis and treatment of allergies (14). Characteristics such as environmental factors (e.g., geographical area) and demographic factors (e.g., age and sex) influence the pattern of inhalant allergen (including HDM) sensitization (15–19). The degree of sensitivity of a patient toward allergens is ascertained by the size of the wheal formed during SPT (20).

Nevertheless, it remains unclear if SPT wheal sizes could predict symptoms severity of AR patients (21, 22) or if they are influenced by factors other than recent anti-histamine usage or treatments. These factors include demographical, clinical and environmental characteristics, as well as nasal and non-nasal symptoms severity scores, of AR patients sensitized to HDMs. These intrinsic and extrinsic characteristics may influence or reflect SPT wheal sizes. Thus, this study attempted to examine these characteristics that might be associated with increased degree of sensitivity reflected by SPT wheal sizes in HDMsensitized AR patients.

MATERIALS AND METHODS

Study Population

The study population was part of our previously published cross-sectional gene expression study of AR patients (n = 30) conducted from February 2019 until January 2020 at the Otorhinolaryngology, Head and Neck Surgery (ORL-HNS) clinic in Hospital Universiti Sains Malaysia (HUSM), and research laboratory of Department of Immunology, Universiti Sains

Malaysia (USM) (23). Diagnosis of AR was conducted using SPT against the allergens from three HDM species (i.e., *D. pteronyssinus*, *D. farinae* and *B. tropicalis*). The mean age of the AR cohort was 28.7 years old consisting of 10 males and 20 females, and the complete clinico-demographical and environmental characteristics have previously been published (23). All subjects were recruited with signed informed consent and the study protocols were approved by the Human Research Ethics Committee of Universiti Sains Malaysia (JEPeM) (approved code: USM/JEPeM/18060273).

Skin Prick Test

Our exclusion criteria mandated our study subjects to stop using anti-allergy agents including steroids, anti-histamine and leukotriene receptor antagonists in the recent two weeks before the sample collection procedures were conducted. The study participants were screened using SPT to examine their sensitization toward HDM allergens (D. farinae, D. pteronyssinus or B. tropicalis). The forearm of study's subject was examined for any non-specific rash (i.e., without allergic/atopic characteristics) and chronic urticaria in the absence of allergic features on history. The study's subject had no food intolerance without allergic features (e.g., irritable bowel syndrome), chronic fatigue without allergic features, migraine headaches, behavioral disorders and reactions to respiratory irritants (e.g., smoke, fumes, perfumes). The study's subject was also asked about their medication status as it may interfere with SPT responses and the drugs may be contraindicated in skin testing.

Firstly, the test was initiated by sterilizing the prick area beforehand using an alcohol swab (i.e., not essential in the case of extreme dry skin and eczema). The forearm of the participant was pricked using intradermal sterile lancet in five separate pricks. The forearm was labeled with five different sections, positive control (+), negative control (-), *D. farinae* (f), *D. pteronyssinus* (p) and *B. tropicalis* (b) by using a skin marker pen. One drop of each allergen tested, histamine as positive control and saline as negative control was placed at the pricked area. The pricked area was observed for about 15–30 min to detect the sensitization toward the allergens. The size of wheals was measured using a ruler and was recorded. The patients with wheal size of 4 mm and more were considered positive for sensitization. A total of 30 AR patients met these criteria and were recruited in this study.

Nasal and Non-nasal Symptoms Severity Scores Assessment

The patients were assessed with AR nasal and non-nasal symptoms severity scores. The nasal symptoms assessed were sneezing, runny nose, itchy nose, congestion (stuffiness) and postnasal drip. Non-nasal symptoms assessed were throat symptoms, chronic cough, eye symptoms, ear symptoms, headache and mental function. The 7-point visual analog scale (VAS) was used in these assessments (24) (**Supplementary Table 1**).

Statistical Analysis

The numerical data for the clinico-demographical and environmental characteristics of AR patients and healthy

controls were described using median and interquartile rage (IQR). Correlation analysis between two continuous variables was conducted using the Spearman correlation coefficient. For comparison of each characteristic within the three groups (i.e., wheal sizes according to the sensitization to each of the three HDMs), Dunn's multiple comparisons *post hoc* test was performed following Kruskal-Wallis test. All statistical analysis was conducted using GraphPad Prism (v6; GraphPad Software Inc., USA). For all tests, two-tailed p < 0.05 was considered to be statistically significant.

RESULTS

Association of SPT Wheal Sizes of HDM Allergens With Demographical, Clinical and Environmental Characteristics of AR Patients

The associations of wheal sizes of HDM allergens (*D. pteronyssinus*, *D. farinae* and *B. tropicalis*) with demographical (age, sex), clinical (family history of allergic diseases, classification of AR, comorbidities) and environmental (exposure to secondhand smoke, home location, having pets, frequency of housekeeping, frequency of changing bedsheet) characteristics of AR patients were investigated. No significant difference was observed in all associations examined (**Table 1**).

Association of Nasal and Non-nasal Symptoms Severity Scores With the Number of HDM Allergens Sensitization of AR Patients

Any one of the three HDM allergens sensitized (monosensitized), two HDM allergens sensitized (double-sensitized), and three HDM allergens sensitized (triple-sensitized) represent the number of HDM allergens sensitization in these patients. No significant difference was observed for all associations investigated of nasal and non-nasal symptoms severity scores with the number of HDM allergens sensitization in AR patients. The association of nasal symptoms severity scores with the number of HDM allergens sensitization of AR patients are shown in **Supplementary Figure 1**. The association of non-nasal symptoms severity scores with the number of HDM allergens sensitization of AR patients are shown in **Supplementary Figure 2**.

Correlation of Nasal and Non-nasal Symptom Severity Scores With SPT Wheal Sizes of HDM Allergens of AR Patients

The severity scores of all five nasal symptoms examined (i.e., sneezing, runny nose, itchy nose, congestion and postnasal drip) were positively and significantly correlated with the wheal sizes of AR patients sensitized to *D. farinae* (Figure 1A). Likewise, significant positive relationships were also observed between four of the six non-nasal symptoms examined (i.e., throat symptoms, ear symptoms, headache and mental function) and wheal sizes of AR patients sensitized to *D. farinae* (Figure 1B), except eye symptoms and chronic cough (data not shown for

these insignificant correlations). Such significant relationships were not observed in the correlation of all the nasal and nonnasal symptoms' severity scores with wheal sizes of AR patients sensitized to *D. pteronyssinus* (Figures 1C,D) and *B. tropicalis* (Figures 1E,F).

DISCUSSION

In this study, no significant associations were observed between demographical, clinical and environmental characteristics with SPT wheal size in AR patients. We hypothesized that factors other than these characteristics such as nasal and non-nasal symptoms severity may be associated with SPT wheal sizes in each (or in combination) of the three HDMs sensitized. However, AR patients mono-sensitized, double-sensitized and triple-sensitized to HDM allergens did not show any significant difference in all nasal and non-nasal symptoms severity scores.

Polysensitization (i.e., sensitization to two or more allergens as confirmed by SPT) is associated with severe symptoms and poor QOL due to different immunologic phenotypes in monosensitized and polysensitized patients (25). In this study, symptoms severity scores were not associated with increased number of HDM sensitization. Polysensitized patients do not necessarily have polyallergy (i.e., documented, causal relationship between exposure to two or more specific, sensitizing allergens and the subsequent occurrence of relevant clinical symptoms of allergy), whereas all polyallergic patients are polysensitized (26). Our AR patients might not be polyallergic which may explain that the increased number of HDMs sensitization was not associated with increased symptoms severity scores.

Interestingly, almost all correlation of SPT wheal sizes of AR patients sensitized to D. farinae with nasal and non-nasal symptom severity scores showed significant positive correlations. These significant correlations were not observed in the rest of the HDM species examined (i.e., D. pteronyssinus and B. tropicalis). These results are comparable with previous report of asthma severity was significantly correlated with skin index of reactivity to D. farinae sensitization (27). Our observations of non-significance between D. pteronyssinus with any nasal symptoms also tally with an independent study demonstrating that SPT wheal sizes of AR patients sensitized to D. pteronyssinus did not correlate with the severity of nasal symptoms (28). Nonetheless, not all non-nasal symptoms (i.e., eye symptoms and chronic cough) were correlated with SPT wheal sizes for D. farinae. Allergic reactions produce signs and symptoms that vary according to the site of the reaction, and sensitization may not be accompanied by the presence of any allergic symptoms (29). In addition, as HDMs are inhaled through the nasal leading to AR onset, it is anticipated that the severity of non-nasal symptoms are less likely to be influenced by HDMs sensitization.

It has been demonstrated that protease activation cascade (30) possibly functions differently in *D. farinae* and *D. pteronyssinus* due to the presence of different protease isoforms in the feces of these two species (31). The major contributor of allergenic potential of mites is found in their feces (32, 33). Different protease isoforms could induce different degree of reactions of the protein with protease-activated receptors (PARs) which

TABLE 1 Association of clinico-demographical and environmental characteristics with SPT wheal sizes of HDM allergens in AR patients (n = 30).

Characteristics		Wheal size (D. pteronyssinus)		Wheal size (D. farinae)		Wheal size (B. tropicalis)	
		<i>r</i> -value or median (IQR)	p-value	<i>r</i> -value or median (IQR)	p-value	<i>r</i> -value or median (IQR)	p-value
Age (median: 27.5 years; IQR: 22.0–35.0)		r = 0.174	0.532	r = 0.340	0.215	r = 0.027	0.923
BMI (median: 26.5; IQR:23.3–28.6)		r = -0.488	0.065	r = -0.127	0.652	r = 0.308	0.264
Gender	Male	5.0 (2.0-11.0)	0.054	6.0 (3.0–10.3)	0.721	5.0 (2.3-11.0)	0.671
	Female	4.0 (2.3–5.0)		5.0 (4.0-9.0)		5.0 (3.0-6.8)	
Family history of allergic disease	Yes	4.0 (3.0–5.0)	0.481	5.0 (4.0–10.0)	0.523	5.0 (3.0–7.5)	0.548
	No	3.0 (0.0–9.0)		3.0 (1.5–10.0)		4.0 (1.5-8.0)	
Classification of AR	Persistent	4.0 (2.0-6.0)	>0.999	7.0 (3.5–10.0)	0.609	5.0 (4.0-9.0)	0.058
	Intermittent	4.0 (3.5–5.0)		5.0 (4.0-8.5)		3.0 (0.0–6.5)	
Comorbidity (pharyngitis)	Yes	4.0 (2.3–5.0)	0.597	7.0 (4.0–10.0)	0.627	5.0 (3.0–7.8)	0.831
	No	4.5 (2.8–6.5)		5.0 (3.8–10.0)		5.0 (2.3–7.3)	
Comorbidity (asthma)	Yes	5.0 (2.0-8.0)	0.214	7.0 (4.0–10.0)	0.316	5.0 (4.0-11.0)	0.214
	No	4.0 (3.0–5.0)		5.0 (3.0–10.0)		5.0 (3.0–7.0)	
Comorbidity (sinusitis)	Yes	4.0 (2.23-5.8)	0.871	5.0 (4.0-10.0)	0.641	5.0 (3.0-7.8)	0.366
	No	4.5 (4.0-5.0)		7.5 (4.0–11.0)		2.5 (0.0-5.0)	
Comorbidity (conjunctivitis)	Yes	4.0 (1.5–5.0)	0.062	5.0 (3.8–10.0)	0.457	5.0 (3.0–7.3)	0.567
	No	5.0 (4.3–7.5)		6.0 (4.0–10.8)		5.5 (1.0–12.3)	
Comorbidity (otitis media)	Yes	4.0 (1.0-4.5)	0.277	4.0 (0.0-10.0)	0.374	5.0 (3.5–6.0)	0.634
	No	5.0 (3.0-6.0)		5.0 (4.0–10.0)		5.0 (3.0–9.0)	
Exposure to secondhand smoke	Yes	4.0 (2.5–5.0)	0.662	5.0 (4.0–10.0)	0.681	5.0 (3.0–6.5)	0.125
	No	5.0 (1.5–7.0)		5.0 (2.0–9.0)		6.0 (4.0-12.0)	
Home location	Urban	4.0 (3.0–5.8)	0.893	6.0 (4.0–10.8)	0.295	5.0 (3.3–10.0)	0.582
	Rural	4.5 (2.0–5.3)		5.0 (3.8–8.0)		4.5 (3.0–7.3)	
Having pets	Yes	5.0 (1.5–6.0)	0.908	7.0 (4.5–10.5)	0.106	5.0 (3.0–9.5)	0.487
	No	4.0 (2.5–5.0)		4.0 (3.5–7.5)		4.0 (3.0-6.0)	
Frequency of housekeeping*	Daily	4.0 (0.0-5.0)	0.413	5.0 (4.0-8.0)	0.589	5.0 (3.0-8.0)	0.683
	Alternate daily	5.0 (2.0-5.0)		7.0 (3.0–10.0)		5.0 (4.0-5.0)	
	Weekly	5.0 (3.0–7.5)		6.0 (4.3–10.8)		6.0 (1.0 -11.0)	
Frequency of changing bedsheet*	Weekly	3.5 (0.5–4.8)	0.051	5.0 (3.3–6.5)	0.092	4.5 (3.0–9.0)	0.770
	Monthly	5.0 (3.8–6.8)		10.0 (4.0–10.3)		5.5 (3.0–7.3)	
	2-monthly	3.5 (0.8-4.8)		6.0 (1.0-11.0)		5.0 (4.3–9.5)	

*The p-values were conducted according to Kruskal-Wallis test and all comparison between groups were insignificant by Dunn's multiple comparisons post hoc test.

ultimately influence allergenicity (34) as well as the cleavage of TJ molecules of epithelial cells (35–37).

In addition, the allergen found in *B. tropicalis*, Blo t 5, is structurally monomeric (38), while the allergen of *D. farinae*, Der f 5, is present as dimers (i.e., structure with large hydrophobic cavity) (39). The hydrophobic cavities in allergens are known to bind hydrophobic ligands, which are thought to stimulate the innate immune system and have adjuvant-like effects on IgE-mediated inflammatory responses in allergic diseases (38, 40). This suggests that Der f 5 confers greater immunogenicity than Blo t 5, and this at least partially explains why SPT wheal sizes of *D. farinae* sensitization demonstrated positive association with nasal and non-nasal symptoms severity in AR patients. However, the structure of *D. pteronyssinus* specific allergen is monomeric and it remains unknown why *D*.

pteronyssinus sensitization, unlike *D. farinae* sensitization, is not associated with nasal and non-nasal symptoms severity scores.

Allergic inflammation results from not only cellular and humoral responses but also other mediators including neurosensory structures and growth factors such as neurotrophins (41, 42). Moreover, two biomarkers i.e., nasal nitric oxide and nasal cytology have been shown to predict treatment efficacy of sublingual immunotherapy in AR patients sensitized to HDMs (43). Future studies should also examine the potential contribution of these mediators and biomarkers to SPT wheal sizes and nasal or non-nasal manifestations in HDMs-sensitized AR patients.

We acknowledge the limitations of the study as follows: (1) Small sample size; (2) Sample size calculation had been





conducted previously according to past gene expression studies, but not studies pertaining to SPT wheal sizes in AR patients, as this study was part of our previously published cross-sectional gene expression study (23); (3) Lack of correlation studies with other symptom questionnaires such as the total nasal symptom score (TNSS) (44–46), or with the levels of total and specific IgE. Future investigations addressing these limitations are thus recommended.

In conclusion, we demonstrated the significant correlation between wheal sizes of *D. farinae* sensitization with all nasal and majority of non-nasal symptoms severity scores. Such relationships were not observed in *D. pteronyssinus* and *B. tropicalis.* We suggest that increased SPT wheal sizes for *D. farinae* sensitization may predict the likelihood of more severe nasal severity in AR patients, and that these patients should be routinely monitored or managed to avoid the potential onset of severe nasal manifestations.

DATA AVAILABILITY STATEMENT

The original contributions presented in the study are included in the article's/**Supplementary Material**, further inquiries can be directed to the corresponding author.

ETHICS STATEMENT

The studies involving human participants were reviewed and approved by Human Research Ethics Committee of Universiti Sains Malaysia (JEPeM) (approved code:

REFERENCES

- 1. Pawankar R. Allergic diseases and asthma: a global public health concern and a call to action. *World Allergy Organ J.* (2014) 7:12. doi: 10.1186/1939-4551-7-12
- Greiner AN, Hellings PW, Rotiroti G, Scadding GK. Allergic rhinitis. Lancet. (2011) 378:2112–22. doi: 10.1016/S0140-6736(11)60130-X
- Meltzer EO. Allergic rhinitis: burden of illness, quality of life, comorbidities, and control. *Immunol Allergy Clin N Am.* (2016) 36:235–48. doi: 10.1016/j.iac.2015.12.002
- Siti Sarah CO, Md Shukri N, Mohd Ashari NS, Wong KK. Zonula occludens and nasal epithelial barrier integrity in allergic rhinitis. *PeerJ.* (2020) 8:e9834. doi: 10.7717/peerj.9834
- Wongpiyabovorn J, Suratannon N, Boonmee S, Chatchatee P. Comparison of specific IgE detection by immunoblotting and fluorescence enzyme assay with in vivo skin prick test. *Asian Pac J Allergy Immunol.* (2018) 36:159–65. doi: 10.12932/AP-270217-0035
- Nam YH, Lee SK. Comparison between skin prick test and serum immunoglobulin E by CAP system to inhalant allergens. Ann Allergy Asthma Immunol. (2017) 118:608–13. doi: 10.1016/j.anai.2017.03.005
- Azid NA, Sani MM, Zamry AA, Ashari NSM, Tan TH-T, Wong KK, et al. Total IgE levels and their relevance in the diagnosis of allergy among Malaysian population in the North-East Region of Peninsular Malaysia. J Biomed Clin Sci. (2019) 4:1–7.
- Lim FL, Hashim Z, Than LT, Md Said S, Hisham Hashim J, Norback D. Asthma, airway symptoms and rhinitis in office workers in malaysia: associations with house dust mite (HDM) allergy, cat allergy and levels of house dust mite allergens in office dust. *PLoS ONE.* (2015) 10:e0124905. doi: 10.1371/journal.pone.0124905

USM/JEPeM/18060273). The patients/participants provided their written informed consent to participate in this study.

AUTHOR CONTRIBUTIONS

H-TT, NSMA, and KKW conceived the project and recruited research grants. SMNH, NSMA, and NMS recruited the participants and performed the investigations. SMNH prepared the tables and figures. KKW supervised data analysis. SMNH and KKW designed, wrote, and revised the manuscript.

FUNDING

This work was supported by Universiti Sains Malaysia (USM) grants comprising of the Research University Grant (1001/PPSP/8012349) awarded to KKW and Research University Grant (1001.PPSP.8012285) awarded to NSMA.

SUPPLEMENTARY MATERIAL

The Supplementary Material for this article can be found online at: https://www.frontiersin.org/articles/10.3389/fmed. 2022.843432/full#supplementary-material

Supplementary Figure 1 Association of nasal symptoms severity scores with the number of HDM allergens sensitization of AR patients.

Supplementary Figure 2 Association of non-nasal symptoms severity scores with number of HDM allergens sensitization of AR patients.

Supplementary Table 1 | The 7-point visual analog scale (VAS) indicator to assess AR nasal and non-nasal symptoms severity scores.

- Majdiah W, Khaiza, NY, Suzina, SAH, Maraina, CC, Suryani N. Association between specific IgE levels and severity of symptoms among patients with rhinitis in North East Malaysia. *Int Med J.* (2011) 18:348–9.
- Sani MM, Ashari NSM, Abdullah B, Wong KK, Musa KI, Mohamud R, et al. Reduced CD4⁺ terminally differentiated effector memory T cells in moderatesevere house dust mites sensitized allergic rhinitis patients. *Asian Pac J Allergy Immunol.* (2019) 37:138–46. doi: 10.12932/AP-191217-0220
- Ho TM, Murad S, Kesavapillai R, Singaram SP. Prevalence of allergy to some inhalants among rhinitis patients in Malaysia. *Asian Pac J Allergy Immunol.* (1995) 13:11–6.
- Bousquet J, Heinzerling L, Bachert C, Papadopoulos NG, Bousquet PJ, Burney PG, et al. Practical guide to skin prick tests in allergy to aeroallergens. *Allergy*. (2012) 67:18–24. doi: 10.1111/j.1398-9995.2011. 02728.x
- Bousquet J, Schünemann HJ, Samolinski B, Demoly P, Baena-Cagnani CE, Bachert C, et al. Allergic Rhinitis and its Impact on Asthma (ARIA): achievements in 10 years and future needs. J Allergy Clin Immunol. (2012) 130:1049–62. doi: 10.1016/j.jaci.2012.07.053
- Ichinose M, Sugiura H, Nagase H, Yamaguchi M, Inoue H, Sagara H, et al. Japanese guidelines for adult asthma 2017. *Allergol Int.* (2017) 66:163–89. doi: 10.1016/j.alit.2016.12.005
- Kato M, Yamada Y, Maruyama K, Hayashi Y. Age at onset of asthma and allergen sensitization early in life. *Allergol Int.* (2014) 63(Suppl. 1):23–8. doi: 10.2332/allergolint.13-OA-0631
- Newson RB, van Ree R, Forsberg B, Janson C, Lotvall J, Dahlen SE, et al. Geographical variation in the prevalence of sensitization to common aeroallergens in adults: the GA(2) LEN survey. *Allergy*. (2014) 69:643–51. doi: 10.1111/all.12397

- Park HJ, Kim EJ, Yoon D, Lee JK, Chang WS, Lim YM, et al. Prevalence of selfreported allergic diseases and IgE levels: A 2010 KNHANES analysis. *Allergy Asthma Immunol Res.* (2017) 9:329–39. doi: 10.4168/aair.2017.9.4.329
- Park SC, Hwang CS, Chung HJ, Purev M, Al Sharhan SS, Cho HJ, et al. Geographic and demographic variations of inhalant allergen sensitization in Koreans and non-Koreans. *Allergol Int.* (2019) 68:68–76. doi: 10.1016/j.alit.2018.07.005
- Nur Husna SM, Tan HT, Md Shukri N, Mohd Ashari NS, Wong KK. Nasal epithelial barrier integrity and tight junctions disruption in allergic rhinitis: overview and pathogenic insights. *Front Immunol.* (2021) 12:663626. doi: 10.3389/fimmu.2021.663626
- Tourlas K, Burman D. Allergy testing. Prim Care. (2016) 43:363–74. doi: 10.1016/j.pop.2016.04.001
- Tatar EC, Surenoglu UA, Saylam G, Isik E, Ozdek A, Korkmaz H. Is there any correlation between the results of skin-prick test and the severity of symptoms in allergic rhinitis? *Am J Rhinol Allergy.* (2012) 26:e37–9. doi: 10.2500/ajra.2012.26.3750
- Madani S, Zandieh F, Ahmadi M, Parvizi M, Rezaei N. Does the reaction size of skin prick test associated with the allergic rhinitis symptom severity? *Allergol Immunopathol.* (2021) 49:60–2. doi: 10.15586/aei.v49i6.211
- Nur Husna SM, Siti Sarah CO, Tan HT, Md Shukri N, Mohd Ashari NS, Wong KK. Reduced occludin and claudin-7 expression is associated with urban locations and exposure to second-hand smoke in allergic rhinitis patients. *Sci Rep.* (2021) 11:1245. doi: 10.1038/s41598-020-79208-y
- Spector SL, Nicklas RA, Chapman JA, Bernstein IL, Berger WE, Blessing-Moore J, et al. Symptom severity assessment of allergic rhinitis: part 1. Ann Allergy Asthma Immunol. (2003) 91:105–14. doi: 10.1016/S1081-1206(10)62160-6
- 25. Savi E, Peveri S, Capelli O. Allergic sensitization in rhinitis and asthma. *Prim Care Pract.* (2016) 143:143–63. doi: 10.5772/63009
- Migueres M, Dávila I, Frati F, Azpeitia A, Jeanpetit Y, Lhéritier-Barrand M, et al. Types of sensitization to aeroallergens: definitions, prevalences and impact on the diagnosis and treatment of allergic respiratory disease. *Clin Transl Allergy*. (2014) 4:16. doi: 10.1186/2045-7022-4-16
- Li J, Sun B, Huang Y, Lin X, Zhao D, Tan G, et al. A multicentre study assessing the prevalence of sensitizations in patients with asthma and/or rhinitis in China. *Allergy*. (2009) 64:1083–92. doi: 10.1111/j.1398-9995.2009.01967.x
- Wanjun W, Qiurong H, Yanqing X, Mo X, Nili W, Jing L. Responsiveness of nasal provocation testing-but not skin test and specific immunoglobulin E blood level-correlates with severity of allergic rhinitis in dermatophagoides species-sensitized patients. *Am J Rhinol Allergy.* (2018) 32:236–43. doi: 10.1177/1945892418779435
- Galli SJ, Tsai M, Piliponsky AM. The development of allergic inflammation. Nature. (2008) 454:445–54. doi: 10.1038/nature07204
- Dumez ME, Herman J, Campizi V, Galleni M, Jacquet A, Chevigne A. Orchestration of an uncommon maturation cascade of the house dust mite protease allergen quartet. *Front Immunol.* (2014) 5:138. doi: 10.3389/fimmu.2014.00138
- Erban T, Harant K, Hubert J. Detailed two-dimensional gel proteomic mapping of the feces of the house dust mite *Dermatophagoides pteronyssinus* and comparison with *D. farinae*: Reduced trypsin protease content in *D. pteronyssinus* and different isoforms. *J Proteomics*. (2017) 162:11–9. doi: 10.1016/j.jprot.2017.04.021
- Chevigné A, Jacquet A. Emerging roles of the protease allergen Der p 1 in house dust mite-induced airway inflammation. J Allergy Clin Immunol. (2018) 142:398. doi: 10.1016/j.jaci.2018.05.027
- Cao H, Liu Z. Clinical significance of dust mite allergens. *Mol Biol Rep.* (2020) 47:6239–46. doi: 10.1007/s11033-020-05613-1
- Heuberger DM, Schuepbach RA. Protease-activated receptors (PARs): mechanisms of action and potential therapeutic modulators in PAR-driven inflammatory diseases. *Thromb J.* (2019) 17:1–24. doi: 10.1186/s12959-019-0194-8

- 35. Steelant B, Farre R, Wawrzyniak P, Belmans J, Dekimpe E, Vanheel H, et al. Impaired barrier function in patients with house dust miteinduced allergic rhinitis is accompanied by decreased occludin and zonula occludens-1 expression. J Allergy Clin Immunol. (2016) 137:1043–53.e5. doi: 10.1016/j.jaci.2015.10.050
- Platts-Mills TA, Woodfolk JA. Allergens and their role in the allergic immune response. *Immunol Rev.* (2011) 242:51–68. doi: 10.1111/j.1600-065X.2011.01021.x
- London NR Jr, Ramanathan M Jr. The role of the sinonasal epithelium in allergic rhinitis. Otolaryngol Clin N Am. (2017) 50:1043–50. doi: 10.1016/j.otc.2017.08.002
- Mueller GA, Gosavi RA, Krahn JM, Edwards LL, Cuneo MJ, Glesner J, et al. Der p 5 crystal structure provides insight into the group 5 dust mite allergens. *J Biol Chem.* (2010) 285:25394–401. doi: 10.1074/jbc.M110.128306
- 39. Khemili S, Kwasigroch JM, Hamadouche T, Gilis D. Modelling and bioinformatics analysis of the dimeric structure of house dust mite allergens from families 5 and 21: Der f 5 could dimerize as Der p 5. J Biomol Struct Dyn. (2012) 29:663–75. doi: 10.1080/073911012010525018
- Asam C, Batista AL, Moraes AH, de Paula VS, Almeida FC, Aglas L, et al. Bet v 1–a Trojan horse for small ligands boosting allergic sensitization? *Clin Exp Aller*. (2014) 44:1083–93. doi: 10.1111/cea.12361
- Voisin T, Bouvier A, Chiu IM. Neuro-immune interactions in allergic diseases: novel targets for therapeutics. *Int Immunol.* (2017) 29:247–61. doi: 10.1093/intimm/dxx040
- Manti S, Brown P, Perez MK, Piedimonte G. The role of neurotrophins in inflammation and allergy. *Vitam Horm.* (2017) 104:313–41. doi: 10.1016/bs.vh.2016.10.010
- 43. Parisi GF, Manti S, Papale M, Amato M, Licari A, Marseglia GL, et al. Nasal nitric oxide and nasal cytology as predictive markers of short-term sublingual allergen-specific immunotherapy efficacy in children with allergic rhinitis. *Am J Rhinol Allergy.* (2021). doi: 10.1177/19458924211060592. [Epub ahead of print].
- 44. Downie SR, Andersson M, Rimmer J, Leuppi JD, Xuan W, Akerlund A, et al. Symptoms of persistent allergic rhinitis during a full calendar year in house dust mite-sensitive subjects. *Allergy*. (2004) 59:406–14. doi: 10.1111/j.1398-9995.2003.00420.x
- Restimulia L, Pawarti DR, Ekorini HM. The relationship between serum vitamin D levels with allergic rhinitis incidence and total nasal symptom score in allergic rhinitis patients. *Open Access Maced J Med Sci.* (2018) 6:1405–9. doi: 10.3889/oamjms.2018.247
- Tamasauskiene L, Gasiuniene E, Sitkauskiene B. Translation, adaption and validation of the total nasal symptom score (TNSS) for Lithuanian population. *Health Qual Life Outcomes*. (2021) 19:54. doi: 10.1186/s12955-020-01659-8

Conflict of Interest: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Publisher's Note: All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

Copyright © 2022 Nur Husna, Md Shukri, Tan, Mohd Ashari and Wong. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.