

Association between *GSTM1* and *GSTT1* Allelic Variants and Head and Neck Squamous Cell Carcinoma

Yang Zhang¹, Yuanyuan Ni¹, Hao Zhang, Yongchu Pan, Junqing Ma*, Lin Wang*

Institute of Stomatology, Nanjing Medical University, Nanjing, Jiangsu, China

Abstract

Backgrounds: *GSTM1* and *GSTT1* are involved in the detoxification of carcinogens such as smoking by-products, and polymorphisms in these two genes with a result of loss of enzyme activity may increase risk of carcinogenesis. Although many epidemiological studies have investigated the association between *GSTM1* or *GSTT1* null genotype and head and neck squamous cell carcinoma (HNSCC), the results remain conflicting. To elucidate the overall association of *GSTM1*, *GSTT1* and HNSCC, we included all available studies and performed this meta-analysis.

Methodology/Principal Findings: A dataset including 42 articles for *GSTM1*, 32 articles for *GSTT1*, and 15 articles for *GSTM1* and *GSTT1* in combination were identified by a search in PubMed. Associations between HNSCC and polymorphisms of *GSTM1* and *GSTT1* alone and in combination were analysed by software RevMan 5.1. Stratification analysis on ethnicity and smoking status, sensitivity analysis, heterogeneity among studies and their publication bias were also tested. Association was found in overall analysis between HNSCC and *GSTM1* and *GSTT1* null genotype. Stratified by ethnicity, we found increased risks of HNSCC in carriers with *GSTM1* null genotype in Asian, *GSTT1* null genotype in South American, and dual null genotype in European and Asian. When stratified by smoking, a more significant association of *GSTM1* null genotype with HNSCC risk was observed in smokers.

Conclusions/Significance: This meta-analysis presented additional evidence of the association between *GSTM1* and *GSTT1* polymorphisms and HNSCC risk.

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* E-mail: junqingma2000@hotmail.com (JM); lw603@njmu.edu.cn (LW)

† These authors contributed equally to this work.

Introduction

Head and neck neoplasms are the sixth leading cause of death by cancer [1]. The most common histological type is the squamous cell carcinoma, accounting for about 90% of all cases [2,3]. Being a multifactorial disease, the etiology of head and neck squamous cell carcinoma (HNSCC) is still a much debated question. Smoking of cigarettes, consumption of alcohol and genetic causes are some of the foci of former etiological studies.

Enzymes of the glutathione S-transferase (GST) family are present in eukaryotes and in prokaryotes, which are composed of many cytosolic, mitochondrial, and microsomal proteins. They catalyze various reactions and participate in the phase II biotransformation of xenobiotics. GSTs contribute to the detoxification of by-products of smoking and alcohol and other exogenous chemical carcinogens which may induce HNSCC, so they have been considered as potential candidates for HNSCC susceptibility. Classes α and μ of the GST superfamily have been paid lots of attention, which are encoded by *GSTT1* and *GSTM1* genes. The *GSTM1* and *GSTT1* gene have been localized to chromosome 1p13.3 and 22q11.2. Both of the genes are polymorphic and frequent homozygous deletions of the genes

presenting null genotype are associated with loss of the corresponding enzyme activity. Therefore, carriers with null genotype will increase the risk of the development of HNSCC due to the decreased ability to detoxify carcinogens theoretically.

In 2003, a meta-analysis conducted by Hashibe *et al.* indicated modest associations of *GSTM1* and *GSTT1* genotypes with head and neck cancer risk [4]. However, more than twenty independent studies from various populations have further examined the relationships between these two genes and HNSCC risk, and still reported conflicting results. Some studies in HNSCC have indicated that the null genotype of *GSTM1* or *GSTT1* is a risk factor of HNSCC development [5–7]. However, such an association was not observed in some other groups [8–10]. Therefore, it is necessary to reevaluate the association of *GSTM1* or *GSTT1* null genotype with the risk of HNSCC by pooling the new published studies using meta-analysis. The present study included all eligible published case-control studies to establish a relatively comprehensive picture of the relationship between these two genes and HNSCC.

Materials and Methods

Selection criteria and identification of eligible studies

Candidate studies were identified through computer-aided literature searches in PubMed for relevant articles in English and Chinese (1995 to May 2012). To identify all articles that studied the association of *GSTT1* and *GSTM1* polymorphisms with HNSCC, we conducted the search using the following keywords and subject terms: 'GSTT1' or 'GSTM1', and 'squamous'. We also searched the references cited in the articles and included published works. Abstracts, case-only articles, editorials, review articles and repeated literatures were excluded. Of the articles with the overlapping data, we only included the publication with the most extensive information. The inclusion criteria in the current meta-analysis were as follows: (a) they are unrelated studies; (b) identification of squamous cell carcinoma was histologically confirmed; and (c) they have original data of genotype frequency and provided sufficient information to calculate the odds ratio (OR) or P-value.

Data extraction

Two reviewers (Zhang Y and Ni Y) independently examined the studies for inclusion in the meta-analysis and collected data on the genotype of *GSTT1* and *GSTM1*. We extracted the following

information from each study: first author, year of publication, country, ethnicity, numbers of case and control, smoking status and genotyping information. Disagreements between two reviewers were discussed and resolved with consensus. When essential information was not found in articles, we made effort to get the data from the authors (Figure 1).

Statistical analysis

The meta-analysis for *GSTM1* or *GSTT1* null genotype or dual null genotype compared HNSCC vs. controls. Odds ratio (OR) and its 95% confidence interval (CI) were assessed for each study. The Cochran's *Q*-statistic was used to test heterogeneity, and the heterogeneity was considered statistically significant when $P < 0.1$ [11]. The Mantel-Haenszel method was used to calculate the OR for the included data in a fixed effects model in the absence of between-study heterogeneity, while random effects model was used for those with heterogeneity. P -value < 0.05 was considered statistically significant, and $0.05 \leq P$ -value < 0.10 was indicated suggestive. In addition, we also performed stratification analyses on ethnicity, smoking and combined analyses of *GSTM1* and *GSTT1* on HNSCC risk. The sensitivity analysis was carried out to test the stability of the pooled effect after excluding individual studies. Begg's funnel plot was used to evaluate publication bias.

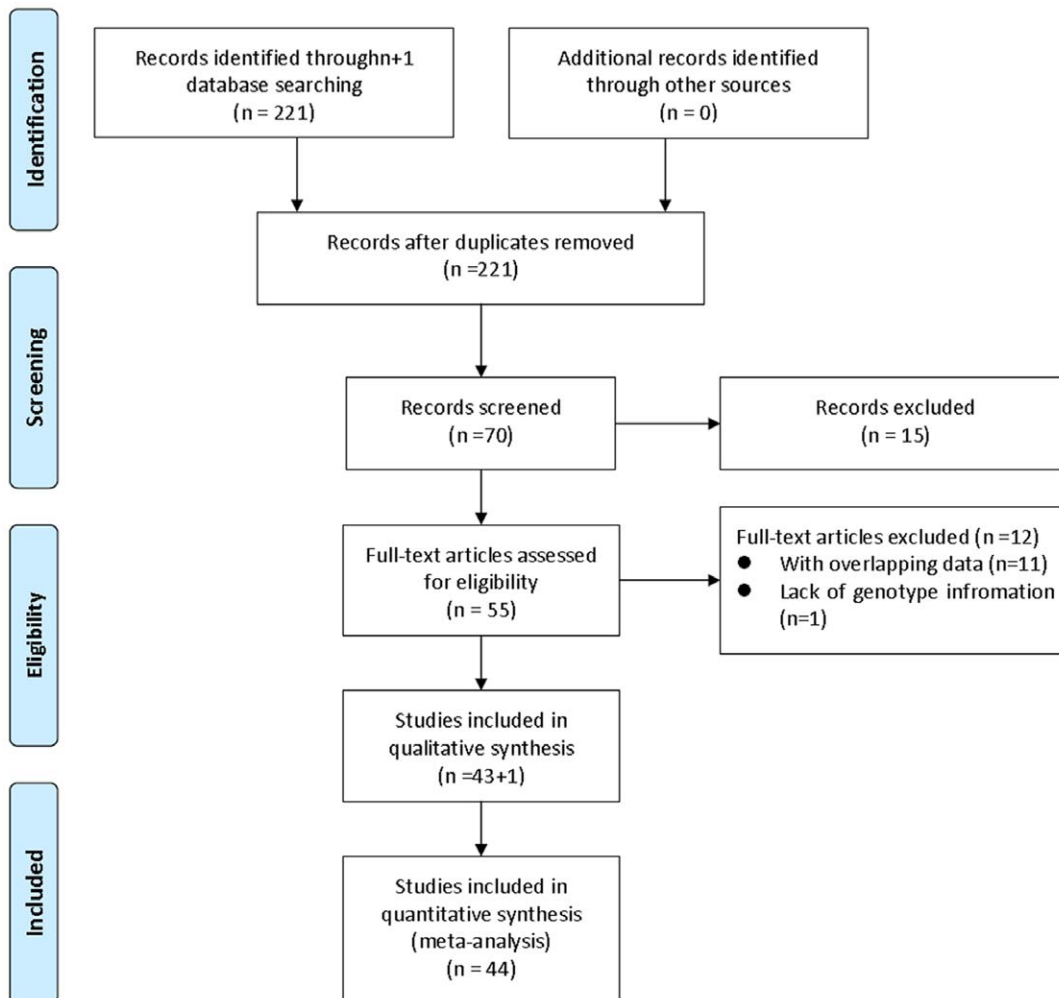


Figure 1. Flow diagram of study identification.

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Table 1. Characteristics of studies included in meta-analysis.

Author (Ref)	Year	Country	Ethnicity	Case	Control	Whether has genotype distribution information			
						<i>GSTM1</i>	<i>GSTT1</i>	Dual genes	Tobacco consumption
Jahnke et al. (23)	1996	Germany	European	269	216	Yes	Yes	No	No
Park et al. (24)	1997	USA	European	133	133	Yes	No	No	No
González et al. (25)	1998	Spain	European	75	200	Yes	No	No	No
Oude Ophuis et al. (13)	1998	Netherlands	European	185	207	Discarded	Discarded	Yes	No
Cheng et al. (26)	1999	USA	European	162	315	Yes	Yes	Yes	No
Kato et al. (27)	1999	Japan	Asian	92	147	Yes	Yes	No	No
Morita et al. (28)	1999	Japan	Asian	145	164	Yes	No	No	No
Nazar-Stewart et al. (29)	1999	USA	European	48	144	Yes	No	No	Yes
Sato et al. (30)	1999	Japan	Asian	142	142	Yes	No	No	No
Tanimoto et al. (31)	1999	Japan	Asian	100	100	Yes	No	No	No
Hamel et al. (32)	2000	Canada	European	90	90	Yes	Yes	No	No
Olshan et al. (33)	2000	USA	European	182	202	Yes	Yes	No	Yes
Hahn et al. (34)	2002	Germany	European	94	92	Yes	No	No	No
To-Figueras et al. (35)	2002	Spain	European	204	203	Yes	Yes	No	No
Gronau et al. (36)	2003	Germany	European	187	139	Yes	Yes	Yes	No
Drummond et al. (37)	2004	Brazil	South American	70	82	Yes	No	No	No
Evans et al. (38)	2004	USA	European	283	208	Yes	Yes	No	Yes
Li et al. (39)	2004	China	Asian	89	164	Yes	No	No	No
Drummond et al. (40)	2005	Brazil	South American	87	81	No	Yes	No	No
Gajecka et al. (41)	2005	Poland	European	292	321	Yes	Yes	No	No
Acar et al. (42)	2006	Turkey	Asian	110	197	Yes	Yes	No	No
Biselli et al. (10)	2006	Brazil	South American	60	60	Yes	Yes	Yes	No
Gatta's et al. (43)	2006	Brazil	South American	103	102	Yes	Yes	Yes	No
Oude Ophuis et al. (44)	2006	Netherlands	European	185	285	Yes	Yes	No	No
Peters et al. (45)	2006	USA	European	692	753	Yes	Yes	No	Yes
Sharma et al. (46)	2006	India	Asian	40	87	Yes	Yes	No	Yes
Sugimura et al. (47)	2006	Japan	Asian	122	241	Yes	Yes	No	Yes
Anatharaman et al. (48)	2007	India	Asian	451	727	Yes	Yes	No	Yes
Cha et al. (49)	2007	Korea	Asian	72	209	Yes	No	No	Yes
Suzen et al. (8)	2007	Turkey	Asian	98	120	Yes	Yes	Yes	Yes
Boccia et al. (9)	2008	Italy	European	210	245	Yes	Yes	No	No
Buch et al. (50)	2008	USA	European	196	414	Yes	Yes	No	No
Harth et al. (51)	2008	Germany	European	312	300	Yes	Yes	No	No
Hatagima et al. (52)	2008	Brazil	South American	231	212	Yes	Yes	No	No
Losi-Guembarovski et al. (53)	2008	Brazil	South American	91	81	Yes	Yes	No	No
Amtha et al. (54)	2009	Indonesia	Asian	81	162	Yes	Yes	No	No
Li et al. (55)	2009	China	Asian	76	76	Yes	Yes	No	Yes
Chatzimichalis et al. (56)	2010	Greek	European	88	102	Yes	Yes	No	No
Leme et al. (57)	2010	Brazil	South American	100	100	Yes	Yes	Yes	No
Sam et al. (58)	2010	India	Asian	408	220	Yes	Yes	Yes	No
Soucek et al. (59)	2010	Czech	European	122	179	Yes	Yes	No	No
Lourenço et al. (6)	2011	Brazil	South American	142	142	Yes	Yes	No	No
Ruwali et al. (5)	2011	India	Asian	500	500	Yes	Yes	Yes	Yes
Shukla et al. (60)	2012	India	Asian	150	141	Yes	No	No	No

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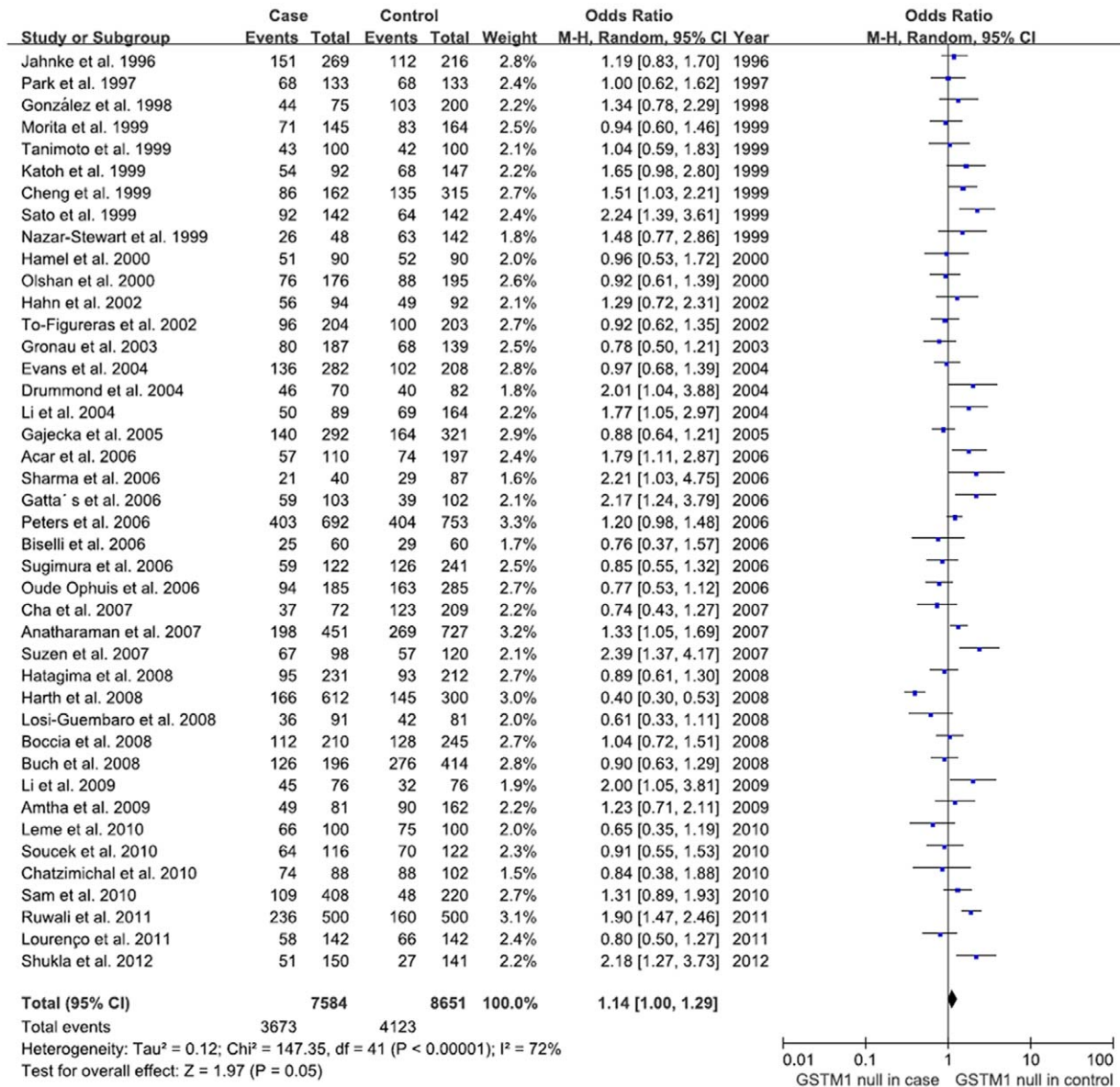


Figure 2. Forest plot of GSTM1 associated with HNSCC under random-effects model. Each study is shown by point estimate of OR and 95% CI by a horizontal line. The diamond shows the overall risk and the line represent the 95% CI for each meta-analysis. Events: null genotype. doi:10.1371/journal.pone.0047579.g002

Table 2 Genotype distribution of GSTM1 and GSTT1 in different Ethnicities.

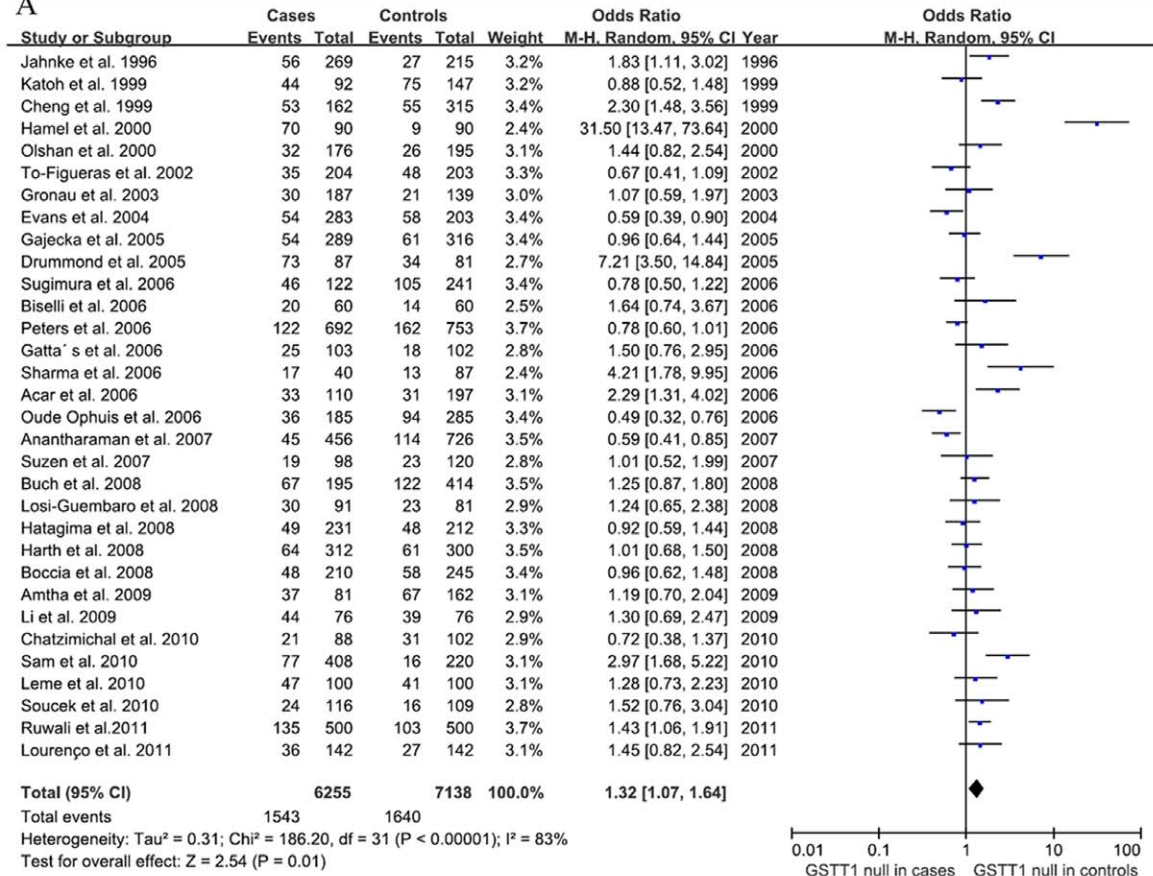
	GSTM1			GSTT1			GSTM1+GSTT1		
	European	Asian	South Am	European	Asian	South Am	European	Asian	South Am
Cases (n/N ^a)	2049/4111	1239/2676	385/797	766/3458	497/1983	280/814	83/534	200/1769	48/354
Controls (n/N)	2378/4475	1361/3397	384/779	849/3884	586/2476	205/778	57/661	185/2088	48/343
OR ^b	0.96	1.48	1.05	1.21	1.32	1.63	2.01	1.56	0.96
95% CI ^c	0.82–1.13	1.24–1.75	0.71–1.57	0.87–1.69	0.93–1.88	1.03–2.58	1.15–3.53	1.05–2.33	0.62–1.48
P ^d	0.64	<0.00001**	0.80	0.26	0.12	0.04*	0.01*	0.03*	0.85

Abbreviations: ^a, number of carriers with null genotype/ total number; ^b, odds ratio; ^c, confidence interval; ^d, value for heterogeneity; OR, odds ratio; 95% CI, 95% confidence interval.

**P < 0.01; * 0.01 ≤ P < 0.05

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A



B

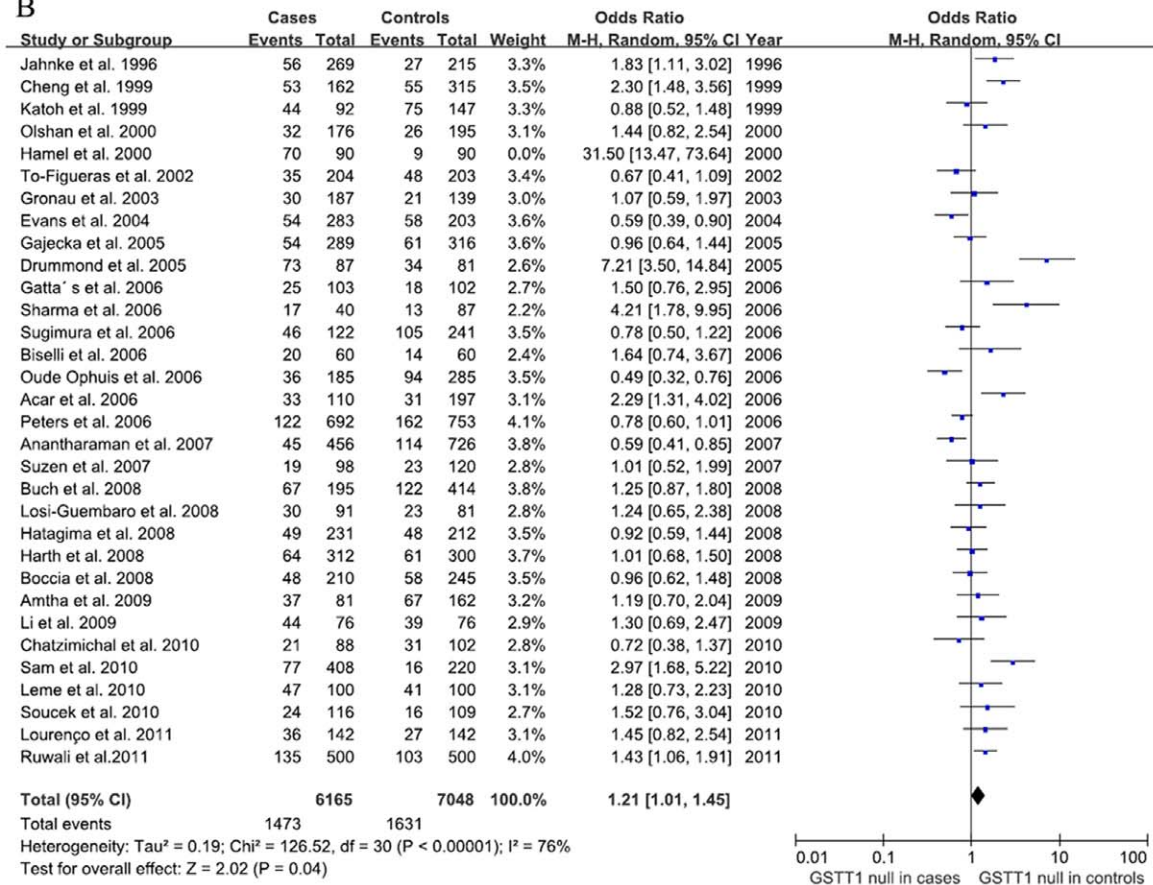


Figure 3. Forest plot of GSTT1 associated with HNSCC under random-effects model. A: Overall analysis. B: Sensitivity analysis with exclusion of the study by Hamel et al. 2000. The diamond shows the overall risk and the line represent the 95% CI for each meta-analysis. Events: null genotype. doi:10.1371/journal.pone.0047579.g003

All above statistical analysis was carried out using the software packages Review Manager (RevMan) 5.1.

Results

Eligible studies and meta-analysis databases

We identified 221 articles through the initial computerized search of published work. After reading titles, abstracts, 55 articles were retained. For the analysis of *GSTM1* or *GSTT1*, after discarding 11 articles [7,12–21] due to the overlapping data and 1 article [22] due to lack of essential genotype information, 44 case-control studies [5,6,8–10,23–60] finally met our criteria for inclusion. Among them, 42 studies described the association between *GSTM1* null genotype and HNSCC, and 32 between *GSTT1* null genotype and HNSCC. For the association between dual null genotype and HNSCC, 1 discarded article [13] containing the distribution information of dual null genotype was reincorporated, and 15 studies were included (Table 1). For the analyses stratified by smoking, eight studies [5,8,29,33,45,47,48,55] for *GSTM1*, and seven studies [5,33,38,45–48] for *GSTT1* were included.

Heterogeneity result

Cochran’s *Q* tests indicated heterogeneity exist in different studies in the analysis except studies of dual genes in South American ($P=0.51$, $I^2=0\%$) and *GSTM1* in non-smokers ($P=0.65$, $I^2=0\%$). The random or fixed effect model was selected for comparisons with or without heterogeneity, respectively.

Meta-analysis results

A total of 7584 HNSCC cases and 8576 controls for *GSTM1*, 6255 cases and 7138 controls for *GSTT1*, 2657 cases and 3092 controls for dual genes were investigated.

For *GSTM1* polymorphism, the overall meta-analysis showed a suggestively increased risk in null genotype as compared to wild

genotype (OR = 1.145, 95% CI: 1.00–1.29, $P=0.05$) (Figure 2). In sensitivity analysis by temporarily excluding individual studies, no single study substantially affected the pooled OR, indicating that the results of these meta-analyses are stable. Analysis after stratification by ethnicity indicated *GSTM1* null genotype tended to be associated with HNSCC in Asian (OR = 1.48, 95% CI: 1.24–1.75, $P<0.01$), while no significant association was found in European or South American (Table 2).

For *GSTT1* polymorphism, null genotype was associated with an increased risk of HNSCC (OR = 1.32, 95% CI: 1.07–1.64, $P=0.01$) (Figure 3A). Sensitivity analysis showed that the association still exist even with exclusion of the study of Hamel et al. which was obviously deviating from others (OR = 1.21, 95% CI: 1.01–1.45, $P=0.04$) [32] (Figure 3B). Analysis stratified by ethnicity indicated that *GSTT1* null genotype increased the HNSCC risk in South American (OR = 1.63, 95% CI: 1.03–2.58, $P=0.04$) (Table 2).

Combined analysis of *GSTM1* and *GSTT1* on HNSCC risk showed that OR of individuals with dual null genotype was elevated (OR = 1.48, 95% CI: 1.12–1.96, $P=0.006$) compared to *GSTM1* or *GSTT1* individual null genotype (Figure 4). After stratification for ethnicity, we observed a significant association for HNSCC in European (OR = 2.01, 95% CI: 1.15–3.53, $P=0.01$) and Asian (OR = 1.56, 95% CI: 1.05–2.33, $P=0.03$) populations among *GSTM1* and *GSTT1* dual null individuals (Table 2). The exclusion of individual studies did not change these results qualitatively.

We further performed stratification analysis by smoking status. As shown in Table 3, significant association of *GSTM1* deletion with risk of HNSCC was observed in smoking group (OR = 1.51, 95% CI: 1.05–2.17, $P=0.03$) but not in non-smoking group (OR = 1.14, 95% CI: 0.90–1.43, $P=0.28$). However, we did not found any significant associations for *GSTT1* in either smokers (OR = 1.01, 95% CI: 0.64–1.60, $P=0.96$) or non-smokers

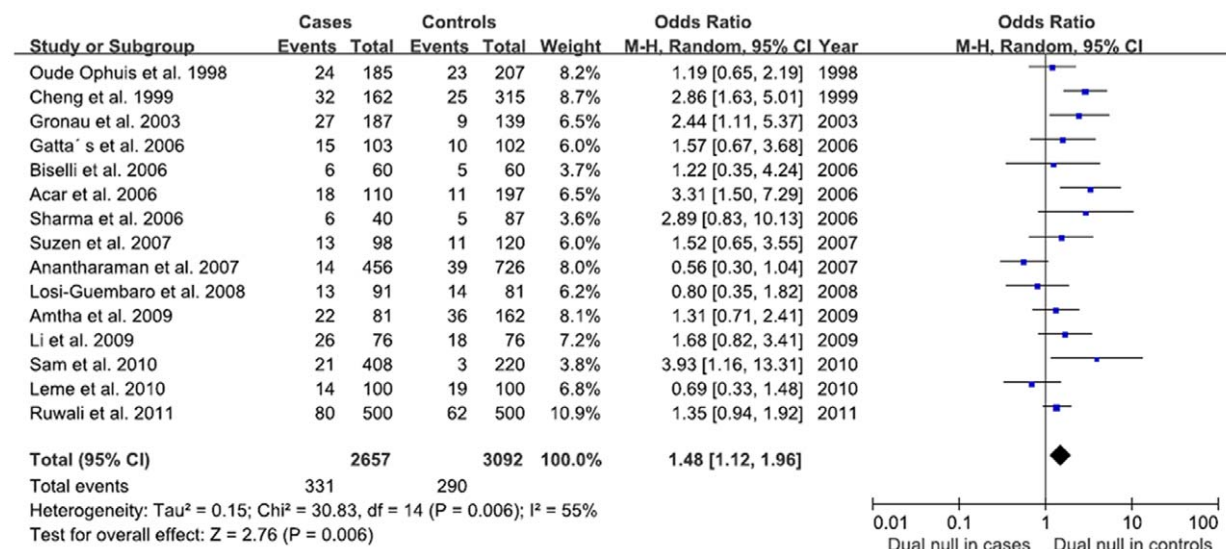


Figure 4. Forest plot of GSTM1 and GSTT1 associated with HNSCC under random-effects models. The diamond shows the overall risk and the line represent the 95% CI for each meta-analysis. Events: null genotype. doi:10.1371/journal.pone.0047579.g004

Table 3. Genotype distribution of *GSTM1* and *GSTT1* in different smoking status.

	<i>GSTM1</i>		<i>GSTT1</i>	
	Non-smoker	Smoker	Non-smoker	Smoker
Cases (n/N ^a)	255/455	862/1638	94/462	347/1671
Controls (n/N)	473/1031	658/1519	222/981	325/1527
OR ^b	1.14	1.51	1.13	1.01
95% CI ^c	0.9–1.43	1.05–2.17	0.68–1.86	0.64–1.6
<i>P</i> ^d	0.28	0.03*	0.64	0.96

Abbreviations: ^a, number of carriers with null genotype/total number; ^b, odds ratio; ^c, confidence interval; ^d, value for heterogeneity; OR, odds ratio; 95% CI, 95% confidence interval

*0.01 ≤ *P* < 0.05

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(OR = 1.13, 95% CI: 0.68–1.86, *P* = 0.64) (Table 3), which may be due to the limited number of study with smoking information.

Publication bias

Funnel plots were performed to assess the publication bias, and these shapes did not suggest any obvious evidence of asymmetry in the analyses of *GSTM1*, gene-gene interaction, and *GSTT1* analysis stratified by smoking status. When one study [32] for *GSTT1* analysis and two studies [46,49] for *GSTM1* analysis stratified by smoking status were omitted, funnel plots illustrated symmetric shape.

Discussion

Genetic factors play an important role in the etiology of tumors. For HNSCC, genes encoding xenobiotic-metabolizing enzymes (XMEs) are some of the most likely candidates that could affect individual's susceptibility to the disease, due to their involvement of the metabolic activation and detoxification of the environmental carcinogens [61]. Conjugation is one of the most common pathways of xenobiotic metabolism and is considered phase II metabolism which is catalyzed by multiple enzyme superfamilies including Glutathione S-transferases (GSTs). GSTs mediate the reactions of glutathione with electrophiles, resulting in the elimination of potentially carcinogenic chemicals [62]. *GSTM1* and *GSTT1* genes belonging to GSTs have been studied extensively due to their important detoxification function and high-frequency polymorphisms. *GSTM1* and *GSTT1* homozygous deletions (null genotype) may lead to deficient enzyme activity [63]. In the present study, the overall frequency of *GSTM1* and *GSTT1* null genotype in controls were 47.65% and 23.77% respectively in accordance with other studies [64–67]. After stratification for ethnicity, the frequency of *GSTM1* and *GSTT1* null genotype in controls in European, Asian and South American were 53.14%, 40.06%, 49.29% and 21.86%, 23.67%, 26.34% respectively, which indicated ethnic differences.

The importance of *GSTM1* and *GSTT1* polymorphisms effects on HNSCC has been a concern in recent years, but the data of existing studies are contradictory. An increase in the risk of HNSCC was observed in cases with null genotypes of *GSTM1* or *GSTT1* in some studies [5–8]. However the risk was not found in other studies. For example, Boccia [9] and Biselli [10] did not find the association between *GSTM1* or *GSTT1* and HNSCC. Although the confused effect of these polymorphisms may be a result of various reasons such as demographic features of subjects

and different life styles, comparatively small sample size in individual study might lead to lower statistical power and bias. The present meta-analyses of 42 studies including 7584 cases and 8651 controls for analysis of *GSTM1*, 32 studies including 6255 cases and 7138 controls for analysis of *GSTT1*, and 15 studies including 2657 cases and 3092 controls provide more comprehensive information on the relationships between two genes and HNSCC.

This meta-analysis showed that both *GSTM1* and *GSTT1* null genotype confers susceptibility to HNSCC in the overall analysis. *GSTM1* can deal with large hydrophobic electrophiles including polycyclic aromatic hydrocarbons derived epoxides (PAH) [68,69], while *GSTT1* targets a more restricted kind of compounds, like monohalomethane and ethylene oxide [70]. Different GST isoforms exhibit overlapping substrate specificity, combinations of *GSTM1* and *GSTT1* null genotype may theoretically confer a higher risk to HNSCC. Comparing to homozygous deletion of *GSTM1* and *GSTT1* alone, deletion of two genes in combination significantly increases the risk of HNSCC as showed in our combined analysis, indicating a synergistic role of *GSTT1* and *GSTM1* in carcinogenesis.

Analyses after stratification by ethnicity revealed ethnicity-specific associations between two genes and HNSCC. Our findings indicate that *GSTM1* may be an important factor in Asians in the development of HNSCC, which is similar to the results reported by Hashibe et al. [4]. However, *GSTT1* but not *GSTM1* may be important in South Americans, while *GSTM1* and *GSTT1* in combination play a vital role in Europeans and Asians. This result may be attributed to the different habits of smoking, alcohol consumption, intake of food and different genetic backgrounds in different ethnic groups.

Both *GSTT1* and *GSTM1* can prevent the accumulation of tobacco smoke carcinogens, and compared with non-smokers, mutations of these two genes theoretically increase the risk of HNSCC in smokers. To investigate potential gene-environment interaction, we stratified the data by smoking status. A significant association was observed in smokers with *GSTM1*, whereas no difference was observed between smokers and non-smokers for *GSTT1*. Previous studies showed that *GSTT1* and *GSTM1* are involved in the detoxification of carcinogens such as smoking by-products, and polymorphisms in these two genes with a result of loss of enzyme activity may increase risk of carcinogenesis and have different role in detoxification. [68–70]. Although we found higher risk of *GSTM1* null genotype in smokers (OR = 1.51) than non-smokers (OR = 1.14), further individual large study are required to evaluate the interaction of *GSTM1* and smoking on HNSCC risk.

Although our result of this meta-analysis is constructive, its limitations and some potential bias should be addressed. First, despite that a well-designed search strategy was used to identify eligible studies, it was possible that some relevant studies were not included. This study only focused on full-text papers published in English and Chinese in PubMed, so some eligible studies in other languages or in other databases might be missed. Second, adjustments over age, gender and other environmental factors such as alcohol drinking might help better detect the association between *GSTM1*, *GSTT1* and HNSCC. If available detailed individual data are enough for an adjusted estimate in the future, a more precise analysis should be conducted. Third, ethnicity was determined roughly by subject's country due to inadequate available data, and this classification can help us have a regional concept of these genes functions. Fourth, the controls in the included studies were recruited in different ways and not uniformly defined, which may have distorted the meta-analysis. Finally,

because all the studies were designed with retrospective studies, we cannot clearly determine the causal relationship between the risk factor and HNSCC. Given the limitations and biases above, the conclusions or interpretations made from the results of this meta-analysis should be explained with caution.

Conclusions

The results of this meta-analysis suggest that *GSTM1* and *GSTT1* null genotypes may be associated with an increased risk of HNSCC. Further large well-designed studies are warranted to confirm these findings.

References

- Walker DM, Boey G, McDonald LA (2003) The pathology of oral cancer. *Pathology* 35: 376–383.
- Casiglia J, Woo SB (2001) A comprehensive review of oral cancer. *Gen Dent* 49: 72–82.
- Reichart PA (2001) Identification of risk groups for oral precancer and cancer preventive measures. *Clin Oral Investig* 5: 207–213.
- Hashibe M, Brennan P, Strange RC, Bhisey R, Cascorbi I, et al. (2003) Meta-analysis of Pooled Analyses of *GSTM1*, *GSTT1*, *GSTP1*, and *CYP1A1* Genotypes and Risk of Head and Neck Cancer. *Cancer Epidemiol Biomarkers Prev* 12:1509–1517.
- Ruwali M, Singh M, Pant MC, Parmar D (2011) Polymorphism in glutathione S-transferases: susceptibility and treatment outcome for head and neck cancer. *Xenobiotica* 41: 1122–1130.
- Lourenço GJ, Silva EF, Rinck-Junior JA, Chone CT, Lima CS (2011) CYP1A1, GSTM1 and GSTT1 polymorphisms, tobacco and alcohol status and risk of head and neck squamous cell carcinoma. *Tumor Biol* 32: 1209–1215.
- Singh M, Shah PP, Singh AP, Ruwali M, Mathur N, et al. (2008) Association of genetic polymorphisms in glutathione S-transferases and susceptibility to head and neck cancer. *Mutat Res* 638: 184–194.
- Suzen HS, Guvenc G, Turanli M, Comert E, Duydu Y, et al. (2007) The role of GSTM1 and GSTT1 polymorphisms in head and neck cancer risk. *Oncol Res* 16: 423–429.
- Boccia S, Cadoni G, Sayed-Tabatabaei FA, Volante M, Arzani D, et al. (2008) CYP1A1, CYP2E1, GSTM1, GSTT1, EPHX1 exons 3 and 4, and NAT2 polymorphisms, smoking, consumption of alcohol and fruit and vegetables and risk of head and neck cancer. *J Cancer Res Clin Oncol* 134: 93–100.
- Biselli JM, de Angelo Calsaverini Leal RC, Ruiz MT, Goloni-Bertollo EM, Maniglia JV, et al. (2006) GSTT1 and GSTM1 polymorphism in cigarette smokers with head and neck squamous cell carcinoma. *Braz J Otorhinolaryngol* 72: 654–658.
- Zintzaras E, Ioannidis JP (2005) HEGESMA: genome search meta-analysis and heterogeneity testing. *Bioinformatics* 21: 3672–3673.
- Trizna Z, Clayman GL, Spitz MR, Briggs KL, Goepfert H (1995) Glutathione S-transferase genotypes as risk factors for head and neck cancer. *Am J Surg* 170: 499–501.
- Oude Ophuis MB, van Lieshout EM, Roelofs HM, Peters WH, Manni JJ (1998) Glutathione S-transferase M1 and T1 and cytochrome P4501A1 polymorphisms in relation to the risk for benign and malignant head and neck lesions. *Cancer* 82: 936–943.
- McWilliams JE, Evans AJ, Beer TM, Andersen PE, Cohen JI, et al. (2000) Genetic polymorphisms in head and neck cancer risk. *Head Neck* 22: 609–617.
- Ko Y, Abel J, Harth V, Bröde P, Antony C, et al. (2001) Association of CYP1B1 codon 432 mutant allele in head and neck squamous cell cancer is reflected by somatic mutations of p53 in tumor tissue. *Cancer Res* 61:4398–4404.
- Gaudet MM, Olshan AF, Poole C, Weisler MC, Watson M, et al. (2004) Diet, GSTM1 and GSTT1 and head and neck cancer. *Carcinogenesis* 25:735–740.
- Capoluongo E, Almadori G, Concolino P, Bussu F, Santonocito C, et al. (2007) GSTT1 and GSTM1 allelic polymorphisms in head and neck cancer patients from Italian Lazio Region. *Clin Chim Acta* 376:174–178.
- Ruwali M, Khan AJ, Shah PP, Singh AP, Pant MC, et al. (2009) Cytochrome P450 2E1 and head and neck cancer: interaction with genetic and environmental risk factors. *Environ Mol Mutagen* 50: 473–482.
- Singh AP, Shah PP, Ruwali M, Mathur N, Pant MC, et al. (2009) Polymorphism in cytochrome P4501A1 is significantly associated with head and neck cancer risk. *Cancer Invest* 27: 869–876.
- Gronau S, Koenig-Greger D, Jerg M, Riechelmann H (2003) GSTM1 enzyme concentration and enzyme activity in correlation to the genotype of detoxification enzymes in squamous cell carcinoma of the oral cavity. *Oral Dis* 9: 62–67.
- Jahnke V, Strange R, Matthias C, Fryer A (1997) Glutathione S-transferase and cytochrome P450 genotypes as risk factors for laryngeal carcinoma. *Eur Arch Otorhinolaryngol* 254 Suppl 1:S147–149.
- Olivieri EH, da Silva SD, Mendonça FF, Urata YN, Vidal DO, et al. (2009) CYP1A2*1C, CYP2E1*5B, and GSTM1 polymorphisms are predictors of risk

Supporting Information

Prisma Checklist S1 (DOC)

Author Contributions

Conceived and designed the experiments: JM LW. Analyzed the data: YZ YN HZ. Wrote the paper: JM YP.

- and poor outcome in head and neck squamous cell carcinoma patients. *Oral Oncol* 45: e73–79.
- Jahnke V, Matthias C, Fryer A, Strange R (1996) Glutathione S-transferase and cytochrome-P450 polymorphism as risk factors for squamous cell carcinoma of the larynx. *Am J Surg* 172:671–673.
- Park JY, Muscat JE, Ren Q, Schantz SP, Harwick RD, et al. (2009) CYP1A1 and GSTM1 polymorphisms and oral cancer risk. *Cancer Epidemiol Biomarkers Prev* 6: 791–797.
- González MV, Alvarez V, Pello MF, Menéndez MJ, Suárez C, et al. (1998) Genetic polymorphism of N-acetyltransferase-2, glutathione S-transferase-M1, and cytochromes P450H1E1 and P450H1D6 in the susceptibility to head and neck cancer. *J Clin Pathol* 51: 294–298.
- Cheng L, Sturgis EM, Eicher SA, Char D, Spitz MR, et al. (1999) Glutathione-S-transferase polymorphisms and risk of squamous-cell carcinoma of the head and neck. *Int J Cancer* 84: 220–224.
- Katoh T, Kaneko S, Kohshi K, Munaka M, Kitagawa K, et al. (1999) Genetic polymorphisms of tobacco- and alcohol-related metabolizing enzymes and oral cavity cancer. *Int J Cancer* 83: 606–609.
- Morita S, Yano M, Tsujinaka T, Akiyama Y, Taniguchi M, et al. (1999) Genetic polymorphisms of drug-metabolizing enzymes and susceptibility to head-and-neck squamous-cell carcinoma. *Int J Cancer* 80: 685–688.
- Nazar-Stewart V, Vaughan TL, Burt RD, Chen C, Berwick M, et al. (1998) Glutathione S-transferase M1 and susceptibility to nasopharyngeal carcinoma. *Cancer Epidemiol Biomarkers Prev* 8: 547–551.
- Sato M, Sato T, Izumo T, Amagasa T (1999) Genetic polymorphism of drug-metabolizing enzymes and susceptibility to oral cancer. *Carcinogenesis* 20:1927–1931.
- Tanimoto K, Hayashi S, Yoshiga K, Ichikawa T (1999) Polymorphisms of the CYP1A1 and GSTM1 gene involved in oral squamous cell carcinoma in association with a cigarette dose. *Oral Oncol* 35: 191–196.
- Hamel N, Karimi S, Hébert-Blouin MN, Brunet JS, Gilfix B, et al. (2000) Increased risk of head and neck cancer in association with GSTT1 nullizygosity for individuals with low exposure to tobacco. *Int J Cancer* 87: 452–454.
- Olshan AF, Weisler MC, Watson MA, Bell DA (2000) GSTM1, GSTT1, CYP1A1, and NAT1 polymorphisms, tobacco use, and the risk of head and neck cancer. *Cancer Epidemiol Biomarkers Prev* 9: 185–191.
- Hahn M, Hagedorn G, Kuhlisch E, Schackert HK, Eckelt U (2002) Genetic polymorphisms of drug-metabolizing enzymes and susceptibility to oral cavity cancer. *Oral Oncol* 38: 486–490.
- To-Figueroas J, Gené M, Gómez-Catalán J, Piqué E, Borrego N, et al. (2002) Microsomal epoxide hydrolase and glutathione S-transferase polymorphisms in relation to laryngeal carcinoma risk. *Cancer Lett* 187: 95–101.
- Gronau S, Koenig-Greger D, Jerg M, Riechelmann H (2003) Gene polymorphisms in detoxification enzymes as susceptibility factor for head and neck cancer? *Otolaryngol Head Neck Surg* 128: 674–680.
- Drummond SN, De Marco L, Noronha JC, Gomez RS (2004) GSTM1 polymorphism and oral squamous cell carcinoma. *Oral Oncol* 40: 52–55.
- Evans AJ, Henner WD, Eilers KM, Montalto MA, Wersinger EM, et al. (2004) Polymorphisms of GSTT1 and related genes in head and neck cancer risk. *Head Neck* 26: 63–70.
- Li L, Lin P, Deng YF, Zhu ZL, Lu HH (2004) Relationship between susceptibility and prognosis of laryngeal cancer and genetic polymorphisms in CYP1A1 and GSTM1. *Zhonghua Er Bi Yan Hou Ke Za Zhi* 39: 2–7.
- Drummond SN, Gomez RS, Motta Noronha JC, Pordeus IA, Barbosa AA, et al. (2005) Association between GSTT-1 gene deletion and the susceptibility to oral squamous cell carcinoma in cigarette-smoking subjects. *Oral Oncol* 41: 515–519.
- Gajdecka M, Ryzdzanicz M, Jaskula-Sztul R, Kujawski M, Szyfyer W, et al. (2005) CYP1A1, CYP2D6, CYP2E1, NAT2, GSTM1 and GSTT1 polymorphisms or their combinations are associated with the increased risk of the laryngeal squamous cell carcinoma. *Mutat Res* 574: 112–123.
- Acar H, Ozturk K, Muslumanoglu MH, Yildirim MS, Cora T, et al. (2006) Relation of glutathione S-transferase genotypes (GSTM1 and GSTT1) to laryngeal squamous cell carcinoma risk. *Cancer Genet Cytogenet*. 2006 169: 89–93.

43. Gattás GJ, de Carvalho MB, Siraque MS, Curioni OA, Kohler P, et al. (2006) Genetic polymorphisms of CYP1A1, CYP2E1, GSTM1, and GSTT1 associated with head and neck cancer. *Head Neck* 28: 819–826.
44. Oude Ophuis MB, Manni JJ, Peters WH (2006) Glutathione S-transferase T1 null polymorphism and the risk for head and neck cancer. *Acta Otolaryngol* 126: 311–317.
45. Peters ES, McClean MD, Marsit CJ, Luckett B, Kelsey KT (2006) Glutathione S-transferase polymorphisms and the synergy of alcohol and tobacco in oral, pharyngeal, and laryngeal carcinoma. *Cancer Epidemiol Biomarkers Prev* 15: 2196–2202.
46. Sharma A, Mishra A, Das BC, Sardana S, Sharma JK (2006) Genetic polymorphism at GSTM1 and GSTT1 gene loci and susceptibility to oral cancer. *Neoplasma* 53: 309–315.
47. Sugimura T, Kumimoto H, Tohnoi I, Fukui T, Matsuo K, et al. (2006) Gene-environment interaction involved in oral carcinogenesis: molecular epidemiological study for metabolic and DNA repair gene polymorphisms. *J Oral Pathol Med* 35: 11–18.
48. Anantharaman D, Chaubal PM, Kannan S, Bhisey RA, Mahimkar MB (2007) Susceptibility to oral cancer by genetic polymorphisms at CYP1A1, GSTM1 and GSTT1 loci among Indians: tobacco exposure as a risk modulator. *Carcinogenesis* 28: 1455–1462.
49. Cha IH, Park JY, Chung WY, Choi MA, Kim HJ, et al. (2007) Polymorphisms of CYP1A1 and GSTM1 genes and susceptibility to oral cancer. *Yonsei Med J* 48: 233–239.
50. Buch SC, Nazar-Stewart V, Weissfeld JL, Romkes M (2008) Case-control study of oral and oropharyngeal cancer in whites and genetic variation in eight metabolic enzymes. *Head Neck* 30: 1139–1147.
51. Harth V, Schafer M, Abel J, Maintz L, Neuhaus T, et al. (2008) Head and neck squamous-cell cancer and its association with polymorphic enzymes of xenobiotic metabolism and repair. *J Toxicol Environ Health A* 71: 887–897.
52. Hatagima A, Costa EC, Marques CF, Koifman RJ, Boffetta P, et al. (2008) Glutathione S-transferase polymorphisms and oral cancer: a case-control study in Rio de Janeiro, Brazil. *Oral Oncol* 44:200–207.
53. Losi-Guembarovski R, Cólus IM, De Menezes RP, Polisei F, Chaves VN, et al. (2008) Lack of association among polymorphic xenobiotic-metabolizing enzyme genotypes and the occurrence and progression of oral carcinoma in a Brazilian population. *Anticancer Res* 28:1023–1028.
54. Amtha R, Ching CS, Zain R, Razak IA, Basuki B, et al. (2009) GSTM1, GSTT1 and CYP1A1 polymorphisms and risk of oral cancer: a case-control study in Jakarta, Indonesia. *Asian Pac J Cancer Prev* 10:21–26.
55. Li Q, Wang L, Chen Y, Du Y, Kong P, et al. (2009) Polymorphisms of GSTM1, GSTT1 and susceptibility of laryngeal and hypopharyngeal carcinomas. *Lin Chung Er Bi Yan Hou Tou Jing Wai Ke Za Zhi* 23:1105–7, 1111.
56. Chatzimichalis M, Xenellis J, Tzagaroulakis A, Sarof P, Banis K, et al. (2010) GSTT1, GSTM1, GSTM3 and NAT2 polymorphisms in laryngeal squamous cell carcinoma in a Greek population. *J Laryngol Otol* 124: 318–323.
57. Leme CV, Raposo LS, Ruiz MT, Biselli JM, Galbiatti AL, et al. (2010) GSTM1 and GSTT1 genes analysis in head and neck cancer patients. *Rev Assoc Med Bras* 56: 299–303.
58. Sam SS, Thomas V, Reddy KS, Surianarayanan G, Chandrasekaran A (2010) Gene-gene interactions of drug metabolizing enzymes and transporter protein in the risk of upper aerodigestive tract cancers among Indians. *Cancer Epidemiol* 34: 626–633.
59. Soucek P, Susova S, Mohelnikova-Duchonova B, Gromadzinska J, Moravicec-Sztandera A, et al. (2010) Polymorphisms in metabolizing enzymes and the risk of head and neck squamous cell carcinoma in the Slavic population of the central Europe. *Neoplasma* 57: 415–421.
60. Shukla D, Kale AD, Hallikerimath S, Vivekanandhan S, Venkatakantiah Y (2012) Genetic polymorphism of drug metabolizing enzymes (GSTM1 and CYP1A1) as risk factors for oral premalignant lesions and oral cancer. *Biomed Pap Med Fac Univ Palacky Olomouc Czech Repub* [Epub ahead of print].
61. Puga A, Nebert DW, McKinnon RA, Menon AG (1997) Genetic polymorphisms in human drug-metabolizing enzymes: potential uses of reverse genetics to identify genes of toxicological relevance. *Crit Rev Toxicol* 27: 1999–2222.
62. Keen JH, Jakoby WB (1978) Glutathione transferases. Catalysis of nucleophilic reactions of glutathione. *J Biol Chem* 253: 5654–5657.
63. Hayes JD, Strange RC (2000) Glutathione S-transferase polymorphisms and their biological consequences. *Pharmacology* 61: 154–166.
64. Seidegard J, Vorachek WR, Pero RW, Pearson WR (1988) Hereditary differences in the expression of the human glutathione transferase active on trans-stilbene oxide are due to a gene deletion. *Proc Natl Acad Sci USA*, 85: 7293–7297.
65. Strange RC, Jones PW, Fryer AA (2000) Glutathione S-transferase. Genetics and role in toxicology. *Toxicol Lett* 112–113: 357–363.
66. Schroder KR, Hallier E, Meyer DJ, Wiebel FA, Muller AM, et al. (1996) Purification and characterization of a new glutathione S-transferase, class t, from human erythrocytes. *Arch Toxicol* 70: 559–566.
67. Pemble S, Schroeder KR, Spencer SR, Meyer DJ, Hallier E, et al. (1994) Human glutathione S-transferase t (GSTT1): cDNA cloning and the characterization of a genetic polymorphism. *Biochem J* 300: 271–276.
68. Hayes JD, Pulford DJ (1995) The glutathione S-transferase supergene family: Regulation of GST and the contribution of the isoenzymes to cancer chemoprotection and drug resistance. *Crit Rev Biochem Mol Biol* 30: 445–600.
69. Hayes JD, Flanagan JU, Jowsey IR (2005) Glutathione transferases. *Annu Rev Pharmacol Toxicol* 45: 51–88.
70. Landi S (2000) Mammalian class theta GST and differential susceptibility to carcinogens: A review. *Mutat Res* 463: 247–283.