



Past and Future of Neurotrophic Growth Factors Therapies in ALS: From Single Neurotrophic Growth Factor to Stem Cells and Human Platelet Lysates

Flore Gouel^{1†}, Anne-Sophie Rolland^{1†}, Jean-Christophe Devedjian¹, Thierry Burnouf^{2,3,4} and David Devos^{1,5*}

¹ Department of Medical Pharmacology, Lille University, INSERM UMRS_1171, University Hospital Center, LICEND COEN Center, Lille, France, ² Graduate Institute of Biomedical Materials and Tissue Engineering, College of Biomedical Engineering, Taipei Medical University, Taipei, Taiwan, ³ International PhD Program in Biomedical Engineering, College of Biomedical Engineering, Taipei Medical University, Taipei, Taiwan, ⁴ International PhD Program in Cell Therapy and Regeneration Medicine, College of Medicine, Taipei Medical University, Taipei, Taiwan, ⁵ Department of Neurology, Lille University, INSERM UMRS_1171, University Hospital Center, LICEND COEN Center, Lille, France

OPEN ACCESS

Edited by:

Pierre-francois Pradat, Hôpitaux Universitaires Pitié Salpêtrière, France

Reviewed by:

Javier Villadiego, University of Seville, Spain Fabrice Cognasse, Groupe Sur L'immunité Des Muqueuses Et Agents Pathogènes (GIMAP), France

> *Correspondence: David Devos david.devos@chru-lille.fr

[†]These authors have contributed equally to this work

Specialty section:

This article was submitted to Neurodegeneration, a section of the journal Frontiers in Neurology

Received: 22 February 2019 Accepted: 19 July 2019 Published: 02 August 2019

Citation:

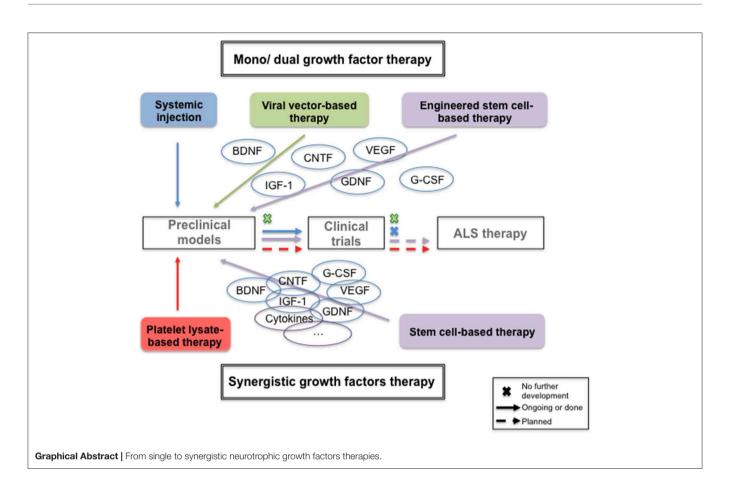
Gouel F, Rolland A-S, Devedjian J-C, Burnouf T and Devos D (2019) Past and Future of Neurotrophic Growth Factors Therapies in ALS: From Single Neurotrophic Growth Factor to Stem Cells and Human Platelet Lysates. Front. Neurol. 10:835. doi: 10.3389/fneur.2019.00835 Amyotrophic lateral sclerosis (ALS) is a fatal neurodegenerative disease that typically results in death within 3–5 years after diagnosis. To date, there is no curative treatment and therefore an urgent unmet need of neuroprotective and/or neurorestorative treatments. Due to their spectrum of capacities in the central nervous system—e.g., development, plasticity, maintenance, neurogenesis—neurotrophic growth factors (NTF) have been exploited for therapeutic strategies in ALS for decades. In this review we present the initial strategy of using single NTF by different routes of administration to the use of stem cells transplantation to express a multiple NTFs-rich secretome to finally focus on a new biotherapy based on the human platelet lysates, the natural healing system containing a mix of pleitropic NTF and having immunomodulatory function. This review highlights that this latter treatment may be crucial to power the neuroprotection and/or neurorestoration therapy requested in this devastating disease.

Keywords: Amyotrophic lateral sclerosis, growth factors, therapeutic, stem cell, human platelet lysate

INTRODUCTION

Amyotrophic lateral sclerosis (ALS) is a fatal neurodegenerative disease affecting the upper and lower motor neurons in the cerebral cortex, brainstem and spinal cord that lead to a progressive, irreversible muscle paralysis, and swallowing and respiratory dysfunctions. Death eventually occurs 3–5 years after diagnosis (1). The majority of ALS cases (90%) are sporadic with unknown cause (2). To date, there is no curative treatment in ALS. Therefore, the development of new and effective treatment is highly urgent. Among the different approaches, the delivery of neurotrophic factors (NTFs) is explored since the 90's because NTFs are necessary to regulate several physiological processes such as neuronal differentiation and survival, axonal outgrowth and synapses maintenance (3–5), proliferation and differentiation of stem cells in the nervous system (6–9). Therefore, these trophic factors represent a promising therapeutic strategy to treat neurodegenerative diseases (10) such as ALS.

1



PRECLINICAL EVIDENCE OF NEUROTROPHICS GROWTH FACTORS ABILITIES TO TREAT AMYOTROPHIC LATERAL SCLEROSIS (TABLE 1)

Recombinant NTFs Delivery by Injection

Some trophic factors have been demonstrated to promote cell survival and be protective in both *in vitro* and *in vivo* models of neuronal degeneration: Ciliary Neurotrophic Factor (CNTF), Brain-derived Neurotrophic Factor (BDNF), Glial-Derived Neurotrophic Factor (GDNF), Insulin-like Growth Factor 1 (IGF-1), Vascular Endothelial Growth Factor (VEGF), and Granulocyte-Colony Stimulating Factor (G-CSF). *In vivo* experiments performed in ALS models using single recombinant growth factors are described in this section.

CNTF, one of the first NTF studied in ALS models, injected intraperitonally in *pmn/pmn* mice, mouse model for human spinal motor neuron disease (11) or subcutaneously in wobbler mice (12) improved motor function and survival, and decreased neuronal degeneration and muscle atrophy (13). In addition, Mitsumoto et al. demonstrated a synergic effect of CNTF and BDNF, respectively, to arrest disease progression for 1 month (14).

The fusion protein BDNF with the c fragment of the tetanus toxin (BDNF-TTC) exhibited enhanced neuroprotective effect

in SOD1^{G93A} ALS mice model, but no synergic effect was observed compared to TTC alone (55). Recently, motor function improvement and less neuronal loss were observed in SOD1^{G93A} mice treated with the flavonoid 7,8-dihydroxyflavone, a small-molecule mimicking the effect of BDNF (56). Two receptors binding the BDNF, p75^{NTR} and TrkB.T1, were highlighted in SOD1^{G93A}: a decreased of p75^{NTR} expression correlated with a delay of mortality and motor impairment (57); a deletion of the TrkB.T1 increased survival and delayed motor deficit (58).

Treatment with encapsuled GDNF-secreting cells in *pmn/pmn* mice did not impact motor neuron degeneration and lifespan (15). The authors suggest a combined treatment for GDNF with others NTFs. Recently, astrocytic GDNF triggered by the tumor necrosis factor α (TNF α) was highlighted in the SOD1^{G93A} mice, and found to limit motor neuron degeneration and disease progression (59).

Intraperitoneal (16) or intracerebroventricular (17) injection of VEGF at doses of 1 g/kg/d and 0.2 μ g/kg/d in SOD1^{G93A} mice and rats, respectively, increased lifespan and improved motor performance. Similar data were observed in a sporadic model of ALS rats induced by excitotoxic administration of AMPA (60, 61).

Finally, protective properties of G-CSF were observed in SOD1^{G93A} mice when delivered continuously at dose of 30 μ g/kg/d (18). Indeed, disease progression was reduced and

TABLE 1 | Different routes of NTFs delivery and therapies in pre-clinical models.

NTF	Delivery route	Model	Outcomes	References
RECOMBINANT NEUROTROPHIC GROWTH FA	CTORS			
CNTF	I.P	<i>pmn/pmn</i> mice (20–21 d)	MP+, S+	(11)
	S.C	Wobbler mice	MP+, MC+	(12-14)
BDNF	S.C	Wobbler mice	MP+	(14)
GDNF	S.C	<i>pmn/pmn</i> mice (15–18 d)	No effect	(15)
/EGF	I.P	SOD1 ^{G93A} mice (74 d)	MP+, DDO+, S+11 d	(16)
	I.C.V	SOD1 ^{G93A/LSd} rats (60 d)	MP+, DDO+, S+10 d	(17)
	I.S.P	Excitotoxic model in rats	MP+, DDO+, S+10.5 d, +5 d	(18, 19)
/iral vector based gene therapy				
AV-NTF				
GF-1	I.M	SOD1 ^{G93A} mice (90 d)	MP+, S+22 d	(20)
	I.S.P	SOD1 ^{G93A} mice (60 d)	MP+, DDO+, S+12.3 d ♂	(21)
	In D.C.N	SOD1 ^{G93A} mice (88–90 d)	MP+, S+14 d	(22)
	I.M	SOD1 ^{G93A} mice (60 and 90 d)	MP+, DDO+, S+29 d and +15 d ♂, +24 d and +14 d ♀	(23)
	I.V	SOD1 ^{G93A} mice (90 d)	MP+, S+10 d	(24)
	I.C.V	SOD1 ^{G93A} mice (80–90 d)	DDO+, S+12 d	(25)
/EGF	I.C.V	SOD1 ^{G93A} mice (80–90 d)	DDO+, S+9 d ♂, +20 d ♀	(25)
	I.T	SOD1 ^{G93A} mice (90 d)	DDO+, S+12 d	(26)
DNF	I.M	SOD1 ^{G93A} mice (90 d)	MP+, DDO+, S+16.6 d	(27)
	I.V	SOD1 ^{G93A} rats (25 d)	MP +/-, S-	(28)
-CSF	I.S.P	SOD1 ^{G93A} mice (70 d)	MP+, DDO+, S+	(29)
tem cell based therapy				
AV-NTF				
SC-NSC	I.S.P	SOD1 ^{G93A} rats (56–62 d)	MP+, DDO+, S+11 d	(30, 31)
m hNSC line (VEGF)	I.T	SOD1 ^{G93A} mice (70 d)	DDO+, S+12 d	(32)
SC-NPC	I.S.P	SOD1 ^{G93A} mice (40 d)	MP+, S+5 d	(33)
m hNPC (GDNF)	I.S.P	SOD1 ^{G93A} rats (~80 d) rats (~80 d)	MP-, S-	(34, 35)
	Cortex	SOD1 ^{G93A} rats (~80 d) macaques	DDO+, S+14 d	(36)
BM-MSC	I.S.P	SOD1 ^{(G93A)dl} mice (28 w)	MP+	(37)
		SOD1 ^{G93A} mice	MP+	(38)
nBM-MSC	I.V	SOD1 ^{G93A} mice (90 d)	MP+, S+17.3 d	(39)
m hBM-MSC (GDNF, VEGF, GNDF/IGF-1, BDNF)	I.M	SOD1 ^{G93A} rats (80 d)	MP+, S+28 d and +18 d for GDNF, + 13 d for VEGF, +28 d for GDNF/VEGF	(40, 41)
nBM	I.S.P and I.M	mdf/ocd mice (6 weeks)	MP+	(42, 43)
nASC	I.V	SOD1 ^{G93A} mice (76–77 d)	MP+, S-	(44)
ASC	I.V and I.C.V	SOD1 ^{G93A} mice (70 d)	MP+, DDO+, S+	(45)
UCBC	I.V	SOD1 ^{G93A} mice (56 d, 66 d)	DDO+, S+21 d, +38.5 d, +23.8 d	(46–48)
		SOD1 ^{G93A} mice (60 and 90 d)	MP+, S+10 d	(49)
	I.T	SOD1 ^{G93A} mice	No effect	(50)
	I.S.P	SOD1 ^{G93A} mice (40 and 90 d)	MP+, S+6 d for 40 d mice	(51)
	I.C.V	SOD1 ^{G93A} (70 d) Wobbler mice (28 d)	MP+, S+18 d MP+	(52)
gm hUCBC (VEGF, GDNF, and/or NCAM)	I.V	SOD1 ^{(G93A)dl} mice	MP+, S+	(53, 54)

I.P, intraperitoneal; I.M, intramuscular; I.V, intravenous; I.C.V, intracerebrovascular; I.S.P, intraspinal; I.T, intrathecal; S.C, subcutaneous; DCN, deep cerebellar nuclei; gm, genetically modified for expression of NTFs in brackets; hSC-NSC: human spinal cord-neural stem cell; m/hBM-MSC, murine/human bone marrow-mesenchymal stem cell; m/hASC, murine/human adipose derived MSC; hUCBC, human umbilical cord blood cells. Main results are summarized as follow: MP, motor performance; DDO, delay of disease onset; S, survival. The age of the model at the treatment is noted in brackets (d, days old; w, weeks). +, improvement; –, deterioration. J, male; 9, female.

survival increased by rescuing motoneurons. Similar results were obtained with subcutaneous injection of pegfilgrastim, a more stable analog of G-CSF (19).

As protein infusion has known drawbacks (invasive method of delivery, protein stability over time, short half-life) others strategies, such as viral vector-based gene therapy and stem cellbased therapy have been developed to express NTFs of interest and avoid chronic injection.

NTFs Delivery by Viral Vector-Based Gene Therapy

Many studies focused on IGF-1. The intramuscular injection of adeno-associated viral (AAV)-IGF-1 in SOD1G93A mice before or at the time of disease symptoms delayed disease onset and increased lifespan (20). Intraparenchymal spinal cord delivery was also tested, showing higher expression of IGF-1 but partial rescue (21), whereas a stereotaxic injection into the deep cerebellar nuclei significantly extended mice lifespan (22). Recently the injection of self-complementary adeno-associated viral vector 9 (scAAV9), a more efficient transducing agent for IGF-1, extended survival, and motor performance of SOD1^{G93A} mice when injected either intramuscularly (23) or intravenously (24). Also, the intracerebroventricular injection of AAV4-VEGF was studied and gave similar results than AAV4-IGF-1 by slowing disease progression. No combined effect of these 2 constructions was observed in SOD1^{G93A} mice (25). Similarly the intrathecal injection of scAAV9-VEGF showed positive impact on lifespan and motor performance in mice (26). The AAV-GDNF, injected intramuscularly in SOD1^{G93A} allowed expression of the protein at the sites of injection, a retrograde transport in anterior horn neurons, and was associated with a delay in the onset and the progression of the disease (27). However, the systemic injection of AAV9-GDNF in SOD1^{G93A} rats showed limited functional improvement and no survival extension (28). Finally the efficacy of intraspinal delivery was showed for AAV-G-CSF in SOD1^{G93A} mice with minimal systemic effects (29).

NTFs Delivery by Stem Cell-Based Therapy

Different types of stem cells exist—based on their source, clonogenic capacity, differentiation potential and availability— and exert a paracrine effect, suitable for therapy in neurodegenerative disease such the ALS (62–65). We mainly focus here on stem cells with potential clinical application, engineered or used as such, e.g., a mix of NTFs.

Neuroprotection With Neural Stem Cells (NSC) and Neural Progenitor Cells (NPC)

Human NSC graft into lumbar protuberance of SOD1^{G93A} rats was shown to delay the onset and the progression of the disease, with their integration into the spinal cord (30, 31). Similarly, the intraspinal administration of human NPC delayed the progression of the disease in SOD1^{G93A} mice (33).

NSC were also engineered to secrete specific one. Intrathecal transplantation of human NSC overexpressing VEGF in SOD1^{G93A} mice delayed the onset of the disease and increased survival with an integration and differentiation of NSC-VEGF into the spinal cord (32). Human neural progenitor cells NPC

(hNPC) were also genetically modified to secrete GDNF. The transplantation of such engineered cells in SOD1 rats were integrated into the spinal cord, limited motoneuron degeneration but failed to improve motor function (34, 35). However, the transplantation of hPNC-GDNF into the cortex extended the survival of SOD1^{G93A} rats and was safe for primates (36).

Mesenchymal Stromal Cells (MSC)

Bone marrow (BM) MSC (BM-MSC), when injected intraspinally (37, 38) or intravenously (39) in SOD1^{G93A} mice, allowed decreased motoneurons degeneration, improved survival and motor function, prevented pro-inflammatory factors. Indeed, MSC display immunomodulatory properties by secreting anti-inflammatory cytokines such as TGF-B or IL-10 (66) Since neuroinflammatory markers were detected in neural tissues of ALS patients (67) promising results can be expected with MSC based therapy. Moreover, intramuscular transplantation of human BM-MSC genetically modified to secrete GDNF in SOD1G93A rats, showed a decrease in motoneuron loss and an overall increased lifespan (40). In addition they demonstrated a synergic effect of the combined intramuscular delivery of hMSC-GDNF and hMSC-VEGF with an increased survival, protection of neuromuscular junction and motoneuron degeneration, greater than either growth factor delivered individually (41). Even though human BM-MSC injections have positive effects on the disease progression, it should be noted that the whole BM intraspinally transplanted showed a greater improvement of motor functions than BM-MSC in *mdf/ocd* mice (42) and increased motoneurons survival when intramuscularly transplanted (43).

Others reported positive results with adipose derived MSC when administrated by systemic (44), or intracerebroventricular administration (45).

Human Umbilical Cord Blood (hUCB)

The first study performed on SOD1^{G93A} mice irradiated and transplanted intravenously with hUBC mononuclear cells (MNC), showed a delay in the onset of symptoms and increased the survival (46, 47). Transplanted cells integrated regions of motoneuron degeneration and expressed neural markers (48). Recently, the efficiency of chronic intravenous injections of UCB MNC in symptomatic SOD1^{G93A} mice was demonstrated, with increased lifespan and reduced inflammatory effectors (49). Similarly, the intraspinal as the intracerebroventricular injection of hUCB in pre-symptomatic SOD1^{G93A} or wobbler mice increased survival and motor performance (51, 52). However, intrathecal administration of hUCB did not affect the lifespan of motor function of ALS mice (50).

Some authors engineered hUCB MNC to secrete some NTFs or to enhance homing at the site of degeneration (68, 69). Recently, transplanted hUCB transduced with AAV encoding VEGF, GDNF and/or neural cell adhesion molecule (NCAM), led to a high rate of SOD1^{G93A} mice survival and improved motor function. Moreover, transplanted cells were detected 1 month after grafting into the lumbar spinal cord (53, 54).

CLINICAL TRIALS WITH GROWTH FACTORS: EVIDENCE AND HYPOTHESIS FOR THE FAILURE

Regarding the promising effects obtained in ALS animal models, clinical trials were conducted to examine the neuroprotective effects of these growth factors therapies in ALS patients (**Table 2**).

Trials Involving NTFs Protein Systemic Injections

In 90's the ALS CNTF Treatment study group published results obtained in phase I (70) and phase II/III (72) clinical trials where enrolled patients received subcutaneous administration of recombinant human CNTF (rHCNTF) at different doses, 15 or 30 μ g/kg, three times a week for 9 months. The phase II/III randomized, placebo-controlled evaluated the safety, tolerability, and efficacy. No statistically difference between rHCNTF-treated patients and placebo-treated patients were observed and side effects were sufficiently severe to limit dosing in many patients. A second trial, same year, did not show any positive effect either (71).

One year later, Penn et al. published results of a phase I clinical trial with intrathecal pump delivery (73). The disease progression was not modified either but no systemic side effects were observed. Thus, intrathecal administration may be the preferred route of administration. To our knowledge, no further clinical study are under investigation.

BDNF

Due to a promising phase I/II clinical trial showing the safety and efficacy of subcutaneous administration of BDNF in 1995, a phase III was designed (74). Results failed to demonstrate an effect on survival but *post-hoc* analyses showed that those ALS patients with early respiratory impairment showed benefit (75). One year later a phase I trial showed the feasibility of intrathecal method of delivery (76) but two other trials conducted in 2003 and 2005 felt to detect any efficacy (77, 78).

IGF-1

In the late 90's, two clinical trials used IGF-1 at a dose of 0.1 mg/kg/d by subcutaneous delivery and found contradictory and opposite results (79, 80). In 2008, a phase III showed no benefit of this route of delivery in 2 years of trials (82). In a pilot study conducted in 2005, intrathecal administration had beneficial effect using high doses of IGF-1 (3 μ g/kg every 2 weeks) but it was not placebo-controlled (81).

G-CSF

Ten years ago, two pilot clinical trials with subcutaneous G-CSF administration at a dose of 5 μ g/kg/d reported a trend for slowing down the disease progression (84) and a delay in motor decline (83). A Phase II clinical trial is under investigation but results are not yet available.

VEGF

Three clinical trials assessed the safety, tolerability, and the possible motor function improvement as well as survival time of the intracerebroventricular administration of 4 μ g/d VEGF. To our knowledge, no results are published.

6- Failure Hypothesis

Most of the clinical trials based on direct protein administration gave disappointing outcomes in view of the promising preclinical results. Different hypotheses can be raised to explain those failures (70–84):

- The route of administration: subcutaneous injection seems less efficient than the intrathecal one
- The minimal ability of these growth factors to cross the blood brain barrier
- The dose: highest safe dose in humans can be lower than those determined in animals, as the clinical trial with CNTF demonstrated
- The treatment start time: in animals, treatment start before the onset of the disease whereas in humans the diagnosis is performed at later stage
- The need of synergic association of numerous neurotrophic factors

Trials Involving Adeno-Associated Viral Gene Therapy

To our knowledge, there is no reported clinical trial using adeno-associated viral gene therapy despite promising results obtained with SOD1^{G93A} mice. AAV2 and AAV9 are vectors having the greatest potential, one specific for neuron tissue, one passing the blood brain barrier and exhibiting neuronal tropisms, respectively. One of the drawbacks of genes therapies for ALS can be the safety. Indeed to stop delivery will not be possible if serious adverse events occur during the treatment.

Trials Involving Stem Cell Therapy

Twenty-two trials involving stem cells-based therapy are registered on ClinicalTrials.gov. Most of them use MSC from different origins and few have results available. This section is an overview of all the known clinical trials.

Neural Stem Cells

In 2012, two trials sponsored by Neuralstem used NSC by intraspinal injection. The phase I did not show any adverse events (85, 86), but the phase II has an unknown status on the ClinicalTrials.gov website.

Recently, published results of a phase I trial, proposing transplantation of human NSCs into the lumbar spinal cord, demonstrated the safety and reproducibility of this cell therapy. Moreover, because the brain tissue used was from natural miscarriages, ethical concerns may be eliminated (87). An ongoing clinical trial concern neuronal progenitor cells engineered to produce GDNF. This is a phase I/IIA trial, active but not recruiting. No results are available for now.

TABLE 2 | Clinical trials with growth factors.

NCT number	NTF	Delivery method	Phase and status of the trial	Cohort size	Outcomes	References	Year
PROTEIN INFU	SION						
Not provided	CNTF	SC	Phase I, terminated	57	No adverse neurologic effects, safe, and tolerated	(70)	1995
Not provided		SC	Phase I, terminated	570	No beneficial effect, adverse events dose related, increased number of death at the highest dose, no beneficial effect on ALS progression	(71)	1996
Not provided		SC	Phase II/III	730	Disease progression not modified, minor adverse side effects	(72)	1996
Not provided		IT	Phase I	4	Pain syndromes dose-related, no systemic side effect, no improvement, or worsen of motor function	(73)	1997
Not provided	BDNF	SC	Phase I/II, terminated	283	Tolerated, Trend of improved survival, less deterioration of predicted FVC	(74)	1995
Not provided	BDNF	SC	Phase III	1 135	Disease progression not modified, Patients with early respiratory impairment and with altered bowel function showed benefit	(75)	1999
Not provided	BDNF	IT	Phase I/II, terminated	25	Well tolerated, feasible	(76)	2000
Not provided	BDNF	IT	Phase III, terminated	17	No adverse events, no effect	(77)	2003
Not provided	BDNF	IT	Phase II/III, terminated	13	No effect	(78)	2005
Not provided	IGF-1	SC	Not specify	266	Slowed the progression of functional impairment, slow the decline in health-related quality of life	(79)	1997
Not provided		SC	Not specify	183	Safe and well-tolerated, no effect	(80)	1998
Not provided		IT	Not specify	9	No serious adverse effect, modest beneficial effect	(81)	2005
NCT00035815		SC	Phase III, completed	330	No benefit	(82)	2008
Not provided	G-CSF	SC	Phase I, terminated	13	Safe, less decline of ALSFRS score	(83)	2009
Not provided		SC	Phase I, terminated	39	Safe, no significative effect on ALSFRS score	(84)	2010
NCT00397423		Not specify	Phase II, completed	40	Not available		
NCT01999803	VEGF	ICV	Phase I, terminated	15	Not available		
NCT02269436		ICV	Phase I, terminated	11	Not available		
NCT01384162		ICV	Phase I/II, terminated	15	Not available		
STEM CELLS							
NCT number	Type of stem cells	Delivery method	Phase and status of the trial	Cohort size	Results	References	Year
NCT01348451	NSC	ISP	Phase I	12	No major adverse events	(85, 86)	2012
NCT01730716	NSC	ISP	Phase II, unknown status	18	Not available		
NCT02943850	NPC	ISP	Phase I/IIa, active, not recruiting	18	Not available		
NCT01640067	NSC	ISP	Phase I, completed	6	Safe approach, no increase of disease progression	(87)	2015
NCT00781872	MSC	IT, IV	Phase I/II, terminated	19	Safe and feasible, ALS-FRS score stable the first 6 months	(88)	2010
NCT03085706	PBMC	ISP	Phase NA, completed	14	Not available		
NCT01933321	HSC	IT	Phase II/III, completed	14	Not available		
NCT01609283	MSC	IT	Phase I, active, not recruiting	27	Not available		
NCT01142856	MSC	IT	Phase I, completed	1	Not available		

(Continued)

TABLE 2 | Continued

NCT number	NTF	Delivery method	Phase and status of the trial	Cohort size	Outcomes	References	Year
NCT00855400	MSC	ISP	Phase I/II completed	11	No severe adverse event, no acceleration in the rate of decline, possible neurotrophic activity	(89)	2012
NCT02286011	MC	IM	Phase I, active, not recruiting	20	Not available		
NCT00855400	MC	ISP	Phase I, completed	11	Safe approach, no worsening of the disease	(90)	2016
NCT03268603	MSC	IT	Phase II, recruiting	60	Not available		
NCT01254539	MSC	ISP, IT	Phase I/II, completed	63	Infusion of MSC produces spinal changes unrelated with clinical events and disease worsening	(91)	2013
NCT01363401	MSC	IT	Phase I/II, completed	64	Possible benefit lasting at least 6 months with safety	(92)	2018
NCT02917681	MSC	IT	Phase I/II, recruiting	28	Not available		
NCT02987413	MSC	IT	Phase I, completed	3	Not available		
NCT02290886	MSC	IV	Phase I/II, active, not recruiting	52	Not available		
NCT01051882	MSC	IM or IT	Phase I/II, completed	12	Safe and tolerated, no serious adverse event, possible benefits on ALS-FRS score, and percentage of FVC	(93)	2016
NCT01777646	MSC	IM + IT	Phase IIa, completed	14			
NCT03280056	MSC	IT	Phase III, Recruiting	200	Not available		
NCT02017912	MSC	IM, IT	Phase II, completed	48	Not available		
NCT01759797	MSC	IV	Phase I/II, completed	6	No adverse events, ALS-FRS score reduced, FVC percentage reduced	(94)	2019
NCT01771640	MSC	IT	Phase I, completed	8			

FVC, force vital capacity; HSC, hematopoietic stem cells; I, intramuscular; ISP, intraspinal; IT, intrathecal; IV, intravenous; MC, mononuclear cell; MSC, mesenchymal stem cells; NPC, neuronal progenitor cells; NSC, neural stem cells; NTF, neurotrophic factor, PBMC, peripheral blood mononuclear cell; SC, subcutaneous.

Blood Cells

Two clinical trials, one using autologous peripheral blood mononuclear cell for intraspinal transplantation and one in phase II/III using hematopoietic stem cells for intrathecal injection were conducted and completed but no results were reported to our knowledge. One trial using autologous bone marrow mononuclear cells (90) for intraspinal injection showed the safety of the procedure.

Mesenchymal Stromal Cells

Among 14 clinical trials using MSCs from diverse origin such as bone marrow, adipose tissue or engineered to secrete particular NTFs, through diverse types of delivery (intrathecal, intraspinal, intramuscular, intravenous, or intraventricular), 5 have no published results, 4 are ongoing, and 5 are completed with published results. All of them are listed in the **Table 2** and the last 5 are detailed below and involved the use of the bone marrow derived MSCs.

In 2012, a phase I/II, using autologous bone marrow MSCs administered by intraspinal delivery, was conducted. No severe adverse event were observed, no acceleration of the disease progression noticed and an increase of the motoneurons in the treated segments compared with the untreated segments for

patients who died for unrelated reasons to the procedure. Thus, this trial demonstrates the safety of intraspinal infusion of MSCs and suggests their neurotrophic activity (89). In 2013, a phase I/II confirmed the safety of BM-MSC infusion (91).

In 2016, two clinical trials in small groups of patients, phase I/II, used bone marrow MSCs engineered to secrete NTFs. Intramuscular transplantation for early ALS patients and intrathecal transplantation for progressive ALS patients were evaluated. They concluded that both route of administration are safe and provide indications of possible clinical benefits that need to be confirmed on a bigger cohort (93).

In 2018, a phase I/II trial was initiated to evaluate the safety and efficacy of these cells through intrathecal delivery. A possible benefit seems to last at least 6 months with apparent safety (92). A phase II is required to evaluate long-term efficacy and safety.

Finally, recent phase I/II trials showed safety and feasibility of intravenous and intrathecal transplantation of autologous bone marrow MSCs (94). Indeed, no adverse events were reported and the ALS-FRS score and the force vital capacity percentage were significantly reduced. Additional trials with bigger cohort are needed.

To conclude, stem cells-based therapy as a future therapy to treat ALS patients is premature due to the lack of results. As for the protein infusion, some questions need to be considered:

- The delivery method
- The timing of intervention
- The number of cells to transplant to obtain a therapeutic efficacy
- The capacity of transplanted cells to migrate to the area of interest and to mature in the hostile environment
- The evaluation of the long-term efficacy

Nevertheless, trophic factors remain essential for neuronal maintenance and survival and remain a promising candidate to treat ALS patients. Another source of those factors can be the natural healing system, namely the platelet lysate, and a continuous infusion into the brain by intracerebroventricular (ICV) injection can be a route of administration, avoiding the potential problem with the blood brain barrier crossing.

HOW TO IMPROVE GROWTH FACTORS THERAPEUTICS IN ALS: A NEW THERAPEUTIC APPROACH BASED ON THE HUMAN PLATELET LYSATE

The lack of clinical efficacy of single NTF infusion, despite a good diffusion, required increasing the dose to a point where they finally induced poor tolerance (i.e., μ g). A single NTF was therefore unable to induce the complex set of signaling pathways required to promote efficient neuroprotection. Platelets constitute abundant, natural sources of physiological balanced mixtures of many growth factors [e.g., Platelet Derived Growth Factor (PDGF), VEGF, IGF-1, EGF, or TGF β) (95) and are used to enhance wound healing and tissue repair (96). In addition, they express adhesion molecules, secret chemokines (97) giving thus neuroinflammatory property to the platelate lysate that could be of an additional interest in ALS therapy. Interestingly, it was demonstrated that ICV injection of human platelet lysates significantly reduced infarct volumes in rats with permanent

REFERENCES

- 1. Wijesekera LC, Leigh PN. Amyotrophic lateral sclerosis. Orphanet J Rare Dis. (2009) 4:3. doi: 10.1186/1750-1172-4-3
- Taylor JP, Brown RH, Cleveland DW. Decoding ALS: from genes to mechanism. *Nature*. (2016) 539:197–206. doi: 10.1038/nature20413
- Korsching S. The neurotrophic factor concept: a reexamination. J Neurosci. (1993) 13:2739–48. doi: 10.1523/JNEUROSCI.13-07-02739.1993
- Boonman Z, Isacson O. Apoptosis in neuronal development and transplantation: role of caspases and trophic factors. *Exp Neurol.* (1999) 156:1–15. doi: 10.1006/exnr.1999.7056
- Hou ST, Jiang SX, Smith RA. Permissive and repulsive cues and signalling pathways of axonal outgrowth and regeneration. *Int Rev Cell Mol Biol.* (2008) 267:125–81. doi: 10.1016/S1937-6448(08)00603-5
- Lamballe F, Smeyne RJ, Barbacid M. Developmental expression of trkC, the neurotrophin-3 receptor, in the mammalian nervous system. *J Neurosci*. (1994) 14:14–28. doi: 10.1523/JNEUROSCI.14-01-00014.1994
- Tuttle R, O'Leary DD. Neurotrophins rapidly modulate growth cone response to the axon guidance molecule, collapsin-1. *Mol Cell Neurosci*. (1998) 11:1–8. doi: 10.1006/mcne.1998.0671
- Lo DC. Instructive roles of neurotrophins in synaptic plasticity. *Prog Brain Res.* (1998) 117:65–70. doi: 10.1016/S0079-6123(08)64008-X

middle cerebral artery occlusion, improved motor function and promoted endogenous neural stem cells proliferation (98). Similar results were obtained with platelet rich plasma in ischemic rats (99). Moreover, intranasal (IN) administration of platelet lysates was demonstrated to be neuroprotective in Alzheimer and Parkinson's disease animal models (100, 101). To pursue with the neuroprotective potential of platelets lysate in neurodegenerative diseases, we developed a heated low protein human purified platelet lysate (HPPL) preparation, compatible with ICV and IN intermittent administration, to deplete fibrinogen, avoid thrombogenic, and proteolytic activities. We demonstrated its neuroprotective effect in in vitro and in vivo model of Parkinson's disease and its anti-inflammatory properties (102). To extend the concept to ALS, HPPL was tested on a motoneuron-like model and strongly protected from apoptosis and oxidative stress (103). Higher neuroprotection was obtained with HPPL compare to single growth factor or combination of 4 (PDGF, BDNF, BFGF, VEGF) and involved specific signaling pathway such as Akt and MEK (103). These results give a real hope for neuroprotective therapy and need to be confirmed in in vivo ALS model with ICV or IN administration of HPPL.

AUTHOR CONTRIBUTIONS

FG and A-SR wrote the manuscript. J-CD, TB, and DD critically revised the manuscript. All authors read and approved the submitted version.

FUNDING

The authors wish to acknowledge support from the ARSLA charity (*Association pour la Recherche sur la Sclérose Latérale Amyotrophique et autres maladies du motoneurone*), I-SITE-ULNE Foundation, Credit Agricole Foundation, and SATT NORD.

- Cattaneo E, McKay R. Proliferation and differentiation of neuronal stem cells regulated by nerve growth factor. *Nature*. (1990) 347:762– 5. doi: 10.1038/347762a0
- Kotzbauer PT, Holtzman DM. Expectations and challenges in the therapeutic use of neurotrophic factors. *Ann Neurol.* (2006) 59:444– 7. doi: 10.1002/ana.20794
- Sendtner M, Schmalbruch H, Stöckli KA, Carroll P, Kreutzberg GW, Thoenen H. Ciliary neurotrophic factor prevents degeneration of motor neurons in mouse mutant progressive motor neuronopathy. *Nature*. (1992) 358:502–4. doi: 10.1038/358502a0
- Mitsumoto H, Ikeda K, Holmlund T, Greene T, Cedarbaum JM, Wong V, et al. The effects of ciliary neurotrophic factor on motor dysfunction in wobbler mouse motor neuron disease. *Ann Neurol.* (1994) 36:142– 8. doi: 10.1002/ana.410360205
- Ikeda K, Wong V, Holmlund TH, Greene T, Cedarbaum JM, Lindsay RM, et al. Histometric effects of ciliary neurotrophic factor in wobbler mouse motor neuron disease. *Ann Neurol.* (1995) 37:47–54. doi: 10.1002/ana.410370110
- Mitsumoto H, Ikeda K, Klinkosz B, Cedarbaum JM, Wong V, Lindsay RM. Arrest of motor neuron disease in wobbler mice cotreated with CNTF and BDNF. *Science*. (1994) 265:1107–1110. doi: 10.1126/science. 8066451

- Sagot Y, Tan SA, Hammang JP, Aebischer P, Kato AC. GDNF slows loss of motoneurons but not axonal degeneration or premature death of pmn/pmn mice. J Neurosci. (1996) 16:2335–41. doi: 10.1523/JNEUROSCI.16-07-02335.1996
- Zheng C, Nennesmo I, Fadeel B, Henter J-I. Vascular endothelial growth factor prolongs survival in a transgenic mouse model of ALS. *Ann Neurol.* (2004) 56:564–7. doi: 10.1002/ana.20223
- Storkebaum E, Lambrechts D, Dewerchin M, Moreno-Murciano M-P, Appelmans S, Oh H, et al. Treatment of motoneuron degeneration by intracerebroventricular delivery of VEGF in a rat model of ALS. *Nat Neurosci.* (2005) 8:85–92. doi: 10.1038/nn1360
- Pitzer C, Krüger C, Plaas C, Kirsch F, Dittgen T, Müller R, et al. Granulocyte-colony stimulating factor improves outcome in a mouse model of amyotrophic lateral sclerosis. *Brain.* (2008) 131:3335–47. doi: 10.1093/brain/awn243
- Rando A, Gasco S, de la Torre M, García-Redondo A, Zaragoza P, Toivonen JM, et al. Granulocyte colony-stimulating factor ameliorates skeletal muscle dysfunction in amyotrophic lateral sclerosis mice and improves proliferation of SOD1-G93A myoblasts *in vitro*. *Neurodegener Dis*. (2017) 17:1–13. doi: 10.1159/000446113
- Kaspar BK, Lladó J, Sherkat N, Rothstein JD, Gage FH. Retrograde viral delivery of IGF-1 prolongs survival in a mouse ALS model. *Science*. (2003) 301:839–42. doi: 10.1126/science.1086137
- Lepore AC, Haenggeli C, Gasmi M, Bishop KM, Bartus RT, Maragakis NJ, et al. Intraparenchymal spinal cord delivery of adeno-associated virus IGF-1 is protective in the SOD1G93A model of ALS. *Brain Res.* (2007) 1185:256–65. doi: 10.1016/j.brainres.2007.09.034
- Dodge JC, Haidet AM, Yang W, Passini MA, Hester M, Clarke J, et al. Delivery of AAV-IGF-1 to the CNS extends survival in ALS mice through modification of aberrant glial cell activity. *Mol Ther.* (2008) 16:1056– 64. doi: 10.1038/mt.2008.60
- Lin H, Hu H, Duan W, Liu Y, Tan G, Li Z, et al. Intramuscular delivery of scAAV9-hIGF1 prolongs survival in the hSOD1G93A ALS mouse model via upregulation of D-amino acid oxidase. *Mol Neurobiol.* (2018) 55:682– 95. doi: 10.1007/s12035-016-0335-z
- 24. Wang W, Wen D, Duan W, Yin J, Cui C, Wang Y, et al. Systemic administration of scAAV9-IGF1 extends survival in SOD1G93A ALS mice via inhibiting p38 MAPK and the JNK-mediated apoptosis pathway. *Brain Res Bull.* (2018) 139:203–10. doi: 10.1016/j.brainresbull.2018.02.015
- Dodge JC, Treleaven CM, Fidler JA, Hester M, Haidet A, Handy C, et al. AAV4-mediated expression of IGF-1 and VEGF within cellular components of the ventricular system improves survival outcome in familial ALS mice. *Mol Ther.* (2010) 18:2075–84. doi: 10.1038/mt.2010.206
- 26. Wang Y, Duan W, Wang W, Di Wen, Liu Y, Liu Y, et al. scAAV9-VEGF prolongs the survival of transgenic ALS mice by promoting activation of M2 microglia and the PI3K/Akt pathway. *Brain Res.* (2016) 1648:1–10. doi: 10.1016/j.brainres.2016.06.043
- Wang LJ, Lu YY, Muramatsu S, Ikeguchi K, Fujimoto K, Okada T, et al. Neuroprotective effects of glial cell line-derived neurotrophic factor mediated by an adeno-associated virus vector in a transgenic animal model of amyotrophic lateral sclerosis. *J Neurosci.* (2002) 22:6920– 8. doi: 10.1523/JNEUROSCI.22-16-06920.2002
- Thomsen GM, Alkaslasi M, Vit JP, Lawless G, Godoy M, Gowing G, et al. Systemic injection of AAV9-GDNF provides modest functional improvements in the SOD1G93A ALS rat but has adverse side effects. *Gene Ther.* (2017) 24:245–52. doi: 10.1038/gt.2017.9
- Henriques A, Pitzer C, Dittgen T, Klugmann M, Dupuis L, Schneider A. CNS-targeted viral delivery of G-CSF in an animal model for ALS: improved efficacy and preservation of the neuromuscular unit. *Mol Ther.* (2011) 19:284–92. doi: 10.1038/mt.2010.271
- Xu L, Yan J, Chen D, Welsh AM, Hazel T, Johe K, et al. Human neural stem cell grafts ameliorate motor neuron disease in SOD-1 transgenic rats. *Transplantation*. (2006) 82:865–75. doi: 10.1097/01.tp.0000235532.00920.7a
- Xu L, Ryugo DK, Pongstaporn T, Johe K, Koliatsos VE. Human neural stem cell grafts in the spinal cord of SOD1 transgenic rats: differentiation and structural integration into the segmental motor circuitry. *J Comp Neurol.* (2009) 514:297–309. doi: 10.1002/cne.22022

- 32. Hwang DH, Lee HJ, Park IH, Seok JI, Kim BG, Joo IS, et al. Intrathecal transplantation of human neural stem cells overexpressing VEGF provide behavioral improvement, disease onset delay and survival extension in transgenic ALS mice. *Gene Ther.* (2009) 16:1234–44. doi: 10.1038/gt.2009.80
- 33. Knippenberg S, Rath KJ, Böselt S, Thau-Habermann N, Schwarz SC, Dengler R, et al. Intraspinal administration of human spinal cordderived neural progenitor cells in the G93A-SOD1 mouse model of ALS delays symptom progression, prolongs survival and increases expression of endogenous neurotrophic factors. *J Tissue Eng Regen Med.* (2017) 11:751– 64. doi: 10.1002/term.1972
- Klein SM, Behrstock S, McHugh J, Hoffmann K, Wallace K, Suzuki M, et al. GDNF delivery using human neural progenitor cells in a rat model of ALS. *Hum Gene Ther*. (2005) 16:509–21. doi: 10.1089/hum.2005.16.509
- Suzuki M, McHugh J, Tork C, Shelley B, Klein SM, Aebischer P, et al. GDNF secreting human neural progenitor cells protect dying motor neurons, but not their projection to muscle, in a rat model of familial ALS. *PLoS ONE*. (2007) 2:e689. doi: 10.1371/journal.pone.0000689
- 36. Thomsen GM, Avalos P, Ma AA, Alkaslasi M, Cho N, Wyss L, et al. Transplantation of neural progenitor cells expressing glial cell line-derived neurotrophic factor into the motor cortex as a strategy to treat amyotrophic lateral sclerosis. *Stem Cells.* (2018) 36:1122–31. doi: 10.1002/stem.2825
- 37. Vercelli A, Mereuta OM, Garbossa D, Muraca G, Mareschi K, Rustichelli D, et al. Human mesenchymal stem cell transplantation extends survival, improves motor performance and decreases neuroinflammation in mouse model of amyotrophic lateral sclerosis. *Neurobiol Dis.* (2008) 31:395–405. doi: 10.1016/j.nbd.2008.05.016
- Boido M, Piras A, Valsecchi V, Spigolon G, Mareschi K, Ferrero I, et al. Human mesenchymal stromal cell transplantation modulates neuroinflammatory milieu in a mouse model of amyotrophic lateral sclerosis. *Cytotherapy*. (2014) 16:1059–72. doi: 10.1016/j.jcyt.2014.02.003
- Uccelli A, Milanese M, Principato MC, Morando S, Bonifacino T, Vergani L, et al. Intravenous mesenchymal stem cells improve survival and motor function in experimental amyotrophic lateral sclerosis. *Mol Med.* (2012) 18:794–804. doi: 10.2119/molmed.2011.00498
- Suzuki M, McHugh J, Tork C, Shelley B, Hayes A, Bellantuono I, et al. Direct muscle delivery of GDNF with human mesenchymal stem cells improves motor neuron survival and function in a rat model of familial ALS. *Mol Ther*. (2008) 16:2002–10. doi: 10.1038/mt.2008.197
- Krakora D, Mulcrone P, Meyer M, Lewis C, Bernau K, Gowing G, et al. Synergistic effects of GDNF and VEGF on lifespan and disease progression in a familial ALS rat model. *Mol Ther.* (2013) 21:1602– 10. doi: 10.1038/mt.2013.108
- 42. Pastor D, Viso-Leon MC, Jones J, Jaramillo-Merchan J, Toledo-Aral JJ, Moraleda JM, et al. Comparative effects between Bone Marrow and mesenchymal stem cell transplantation in GDNF expression and motor function recovery in a motorneuron degenerative mouse model. *Stem Cell Rev.* (2012) 8:445–58. doi: 10.1007/s12015-011-9295-x
- Pastor D, Viso-Leon MC, Botella-Lopez A, Jaramillo-Merchan J, Moraleda JM, Jones J, et al. Bone marrow transplantation in hindlimb muscles of motorneuron degenerative mice reduces neuronal death and improves motor function. *Stem Cells Dev.* (2013) 22:1633–44. doi: 10.1089/scd.2012.0487
- 44. Marconi S, Bonaconsa M, Scambi I, Squintani GM, Rui W, Turano E, et al. Systemic treatment with adipose-derived mesenchymal stem cells ameliorates clinical and pathological features in the amyotrophic lateral sclerosis murine model. *Neuroscience*. (2013) 248:333–43. doi: 10.1016/j.neuroscience.2013.05.034
- 45. Kim KS, Lee HJ, An J, Kim YB, Ra JC, Lim I, et al. Transplantation of human adipose tissue-derived stem cells delays clinical onset and prolongs life span in ALS mouse model. *Cell Transplant*. (2014) 23:1585– 97. doi: 10.3727/096368913X673450
- Ende N, Weinstein F, Chen R, Ende M. Human umbilical cord blood effect on sod mice (amyotrophic lateral sclerosis). *Life Sci.* (2000) 67:53– 9. doi: 10.1016/S0024-3205(00)00602-0
- 47. Chen R, Ende N. The potential for the use of mononuclear cells from human umbilical cord blood in the treatment of amyotrophic lateral sclerosis in SOD1 mice. *J Med.* (2000) 31:21–30.

- Garbuzova-Davis S, Willing AE, Zigova T, Saporta S, Justen EB, Lane JC, et al. Intravenous administration of human umbilical cord blood cells in a mouse model of amyotrophic lateral sclerosis: distribution, migration, and differentiation. J Hematother Stem Cell Res. (2003) 12:255– 70. doi: 10.1089/152581603322022990
- Garbuzova-Davis S, Rodrigues MCO, Mirtyl S, Turner S, Mitha S, Sodhi J, et al. Multiple intravenous administrations of human umbilical cord blood cells benefit in a mouse model of ALS. *PLoS ONE*. (2012) 7:e31254. doi: 10.1371/journal.pone.0031254
- Habisch HJ, Janowski M, Binder D, Kuzma-Kozakiewicz M, Widmann A, Habich A, et al. Intrathecal application of neuroectodermally converted stem cells into a mouse model of ALS: limited intraparenchymal migration and survival narrows therapeutic effects. J Neural Transm. (2007) 114:1395– 406. doi: 10.1007/s00702-007-0748-y
- Knippenberg S, Thau N, Schwabe K, Dengler R, Schambach A, Hass R, et al. Intraspinal injection of human umbilical cord blood-derived cells is neuroprotective in a transgenic mouse model of amyotrophic lateral sclerosis. *Neurodegener Dis.* (2012) 9:107–20. doi: 10.1159/000331327
- 52. Bigini P, Veglianese P, Andriolo G, Cova L, Grignaschi G, Caron I, et al. Intracerebroventricular administration of human umbilical cord blood cells delays disease progression in two murine models of motor neuron degeneration. *Rejuvenation Res.* (2011) 14:623–39. doi: 10.1089/rej.2011.1197
- 53. Islamov RR, Rizvanov AA, Mukhamedyarov MA, Salafutdinov II, Garanina EE, Fedotova VY, et al. Symptomatic improvement, increased life-span and sustained cell homing in amyotrophic lateral sclerosis after transplantation of human umbilical cord blood cells genetically modified with adeno-viral vectors expressing a neuro-protective factor and a neural cell adhesion molecule. *Curr Gene Ther.* (2015) 15:266–76. doi: 10.2174/1566523215666150126122317
- Islamov RR, Rizvanov AA, Fedotova VY, Izmailov AA, Safiullov ZZ, Garanina EE, et al. Tandem delivery of multiple therapeutic genes using umbilical cord blood cells improves symptomatic outcomes in ALS. *Mol Neurobiol.* (2017) 54:4756–63. doi: 10.1007/s12035-016-0017-x
- 55. Calvo AC, Moreno-Igoa M, Mancuso R, Manzano R, Oliván S, Muñoz MJ, et al. Lack of a synergistic effect of a non-viral ALS gene therapy based on BDNF and a TTC fusion molecule. *Orphanet J Rare Dis.* (2011) 6:10. doi: 10.1186/1750-1172-6-10
- Korkmaz OT, Aytan N, Carreras I, Choi J-K, Kowall NW, Jenkins BG, et al. 7,8-Dihydroxyflavone improves motor performance and enhances lower motor neuronal survival in a mouse model of amyotrophic lateral sclerosis. *Neurosci Lett.* (2014) 566:286–91. doi: 10.1016/j.neulet.2014.02.058
- 57. Turner BJ, Cheah IK, Macfarlane KJ, Lopes EC, Petratos S, Langford SJ, et al. Antisense peptide nucleic acid-mediated knockdown of the p75 neurotrophin receptor delays motor neuron disease in mutant SOD1 transgenic mice. J Neurochem. (2003) 87:752–63. doi: 10.1046/j.1471-4159.2003.02053.x
- Yanpallewar SU, Barrick CA, Buckley H, Becker J, Tessarollo L. Deletion of the BDNF truncated receptor TrkB.T1 delays disease onset in a mouse model of amyotrophic lateral sclerosis. *PLoS ONE*. (2012) 7:e39946. doi: 10.1371/journal.pone.0039946
- Brambilla L, Guidotti G, Martorana F, Iyer AM, Aronica E, Valori CF, et al. Disruption of the astrocytic TNFR1-GDNF axis accelerates motor neuron degeneration and disease progression in amyotrophic lateral sclerosis. *Hum Mol Genet.* (2016) 25:3080–95. doi: 10.1093/hmg/ddw161
- Tovar-Y-Romo LB, Zepeda A, Tapia R. Vascular endothelial growth factor prevents paralysis and motoneuron death in a rat model of excitotoxic spinal cord neurodegeneration. J Neuropathol Exp Neurol. (2007) 66:913– 22. doi: 10.1097/nen.0b013e3181567c16
- 61. Tovar-y-Romo LB, Tapia R. Delayed administration of VEGF rescues spinal motor neurons from death with a short effective time frame in excitotoxic experimental models *in vivo*. *ASN Neuro*. (2012) 4: e0008. doi: 10.1042/AN20110057
- Czarzasta J, Habich A, Siwek T, Czaplinski A, Maksymowicz W, Wojtkiewicz J. Stem cells for ALS: an overview of possible therapeutic approaches. *Int J Dev Neurosci*. (2017) 57:46–55. doi: 10.1016/j.ijdevneu.2017.01.003
- 63. Mathieu P, Battista D, Depino A, Roca V, Graciarena M, Pitossi F. The more you have, the less you get: the functional role of

inflammation on neuronal differentiation of endogenous and transplanted neural stem cells in the adult brain. *J Neurochem.* (2010) 112:1368-85. doi: 10.1111/j.1471-4159.2009.06548.x

- 64. Reekmans K, Praet J, Daans J, Reumers V, Pauwels P, Van der Linden A, et al. Current challenges for the advancement of neural stem cell biology and transplantation research. *Stem Cell Rev.* (2012) 8:262–78. doi: 10.1007/s12015-011-9266-2
- Lewis CM, Suzuki M. Therapeutic applications of mesenchymal stem cells for amyotrophic lateral sclerosis. *Stem Cell Res Ther.* (2014) 5:32. doi: 10.1186/scrt421
- Gugliandono A, Bramanti P, Mazzon E. Mesenchymal stem cells: a potential approach for Amyotrophic Lateral Sclerosis? *Stem Cells Int.* (2019) 2019:3675627. doi: 10.1155/2019/3675627
- Beers DR, Appel SH. Immune dysregulation in amyotrophic lateral sclerosis: mechanisms and emerging therapies. *Lancet Neurol.* (2019) 18:211– 20. doi: 10.1016/S1474-4422(18)30394-6
- 68. Rizvanov AA, Kiyasov AP, Gaziziov IM, Yilmaz TS, Kaligin MS, Andreeva DI, et al. Human umbilical cord blood cells transfected with VEGF and L(1)CAM do not differentiate into neurons but transform into vascular endothelial cells and secrete neuro-trophic factors to support neuro-genesisa novel approach in stem cell therapy. *Neurochem Int.* (2008) 53:389–94. doi: 10.1016/j.neuint.2008.09.011
- Rizvanov AA, Guseva DS, Salafutdinov II, Kudryashova NV, Bashirov FV, Kiyasov AP, et al. Genetically modified human umbilical cord blood cells expressing vascular endothelial growth factor and fibroblast growth factor 2 differentiate into glial cells after transplantation into amyotrophic lateral sclerosis transgenic mice. *Exp Biol Med.* (2011) 236:91– 8. doi: 10.1258/ebm.2010.010172
- 70. A phase I study of recombinant human ciliary neurotrophic factor (rHCNTF) in patients with amyotrophic lateral sclerosis. The ALS CNTF Treatment Study (ACTS) Phase I-II Study Group. *Clin Neuropharmacol.* (1995) 18:515–32. doi: 10.1097/00002826-199512000-00004
- Miller RG, Petajan JH, Bryan WW, Armon C, Barohn RJ, Goodpasture JC, et al. A placebo-controlled trial of recombinant human ciliary neurotrophic (rhCNTF) factor in amyotrophic lateral sclerosis. rhCNTF ALS Study Group. *Ann Neurol.* (1996) 39:256–60. doi: 10.1002/ana.410390215
- A double-blind placebo-controlled clinical trial of subcutaneous recombinant human ciliary neurotrophic factor (rHCNTF) in amyotrophic lateral sclerosis. ALS CNTF Treatment Study Group. *Neurology*. (1996) 46:1244–9. doi: 10.1212/WNL.46.5.1244
- Penn RD, Kroin JS, York MM, Cedarbaum JM. Intrathecal ciliary neurotrophic factor delivery for treatment of amyotrophic lateral sclerosis (phase I trial). *Neurosurgery*. (1997) 40:94–9; discussion: 99–100. doi: 10.1227/00006123-199701000-00021
- 74. Works in progress abstracts. Ann Neurol. (1995) 38:971– 3. doi: 10.1002/ana.410380629
- A controlled trial of recombinant methionyl human BDNF in ALS: The BDNF Study Group (Phase III). *Neurology*. (1999) 52:1427–33. doi: 10.1212/WNL.52.7.1427
- 76. Ochs G, Penn RD, York M, Giess R, Beck M, Tonn J, et al. A phase I/II trial of recombinant methionyl human brain derived neurotrophic factor administered by intrathecal infusion to patients with amyotrophic lateral sclerosis. Amyotroph Lateral Scler Other Motor Neuron Disord. (2000) 1:201–6. doi: 10.1080/14660820050515197
- 77. Kalra S, Genge A, Arnold DL. A prospective, randomized, placebocontrolled evaluation of corticoneuronal response to intrathecal BDNF therapy in ALS using magnetic resonance spectroscopy: feasibility and results. *Amyotroph Lateral Scler Other Motor Neuron Disord*. (2003) 4:22– 6. doi: 10.1080/14660820310006689
- Beck M, Flachenecker P, Magnus T, Giess R, Reiners K, Toyka KV, et al. Autonomic dysfunction in ALS: a preliminary study on the effects of intrathecal BDNF. *Amyotroph Lateral Scler Other Motor Neuron Disord*. (2005) 6:100–3. doi: 10.1080/14660820510028412
- Lai EC, Felice KJ, Festoff BW, Gawel MJ, Gelinas DF, Kratz R, et al. Effect of recombinant human insulin-like growth factor-I on progression of ALS. A placebo-controlled study. The North America ALS/IGF-I Study Group. *Neurology*. (1997) 49:1621–30. doi: 10.1212/WNL.49. 6.1621

- Borasio GD, Robberecht W, Leigh PN, Emile J, Guiloff RJ, Jerusalem F, et al. A placebo-controlled trial of insulin-like growth factor-I in amyotrophic lateral sclerosis. European ALS/IGF-I Study Group. *Neurology*. (1998) 51:583– 6. doi: 10.1212/WNL.51.2.583
- Nagano I, Shiote M, Murakami T, Kamada H, Hamakawa Y, Matsubara E, et al. Beneficial effects of intrathecal IGF-1 administration in patients with amyotrophic lateral sclerosis. *Neurol Res.* (2005) 27:768–2. doi: 10.1179/016164105X39860
- Sorenson EJ, Windbank AJ, Mandrekar JN, Bamlet WR, Appel SH, Armon C, et al. Subcutaneous IGF-1 is not beneficial in 2-year ALS trial. *Neurology*. (2008) 71:1770–5. doi: 10.1212/01.wnl.0000335970.78664.36
- Zhang Y, Wang L, Fu Y, Song H, Zhao H, Deng M, et al. Preliminary investigation of effect of granulocyte colony stimulating factor on amyotrophic lateral sclerosis. *Amyotroph Lateral Scler.* (2009) 10:430– 1. doi: 10.3109/17482960802588059
- Nefussy B, Artamonov I, Deutsch V, Naparstek E, Nagler A, Drory VE. Recombinant human granulocyte-colony stimulating factor administration for treating amyotrophic lateral sclerosis: A pilot study. *Amyotroph Lateral Scler.* (2010) 11:187–93. doi: 10.3109/17482960902933809
- Riley J, Federici T, Polak M, Kelly C, Glass J, Raore B, et al. Intraspinal stem cell transplantation in amyotrophic lateral sclerosis: a phase I safety trial, technical note, and lumbar safety outcomes. *Neurosurgery*. (2012) 71:405–16; discussion: 416. doi: 10.1227/NEU.0b013e31825ca05f
- Glass JD, Boulis NM, Johe K, Rutkove SB, Federici T, Polak M, et al. Lumbar intraspinal injection of neural stem cells in patients with amyotrophic lateral sclerosis: results of a phase I trial in 12 patients. *Stem Cells.* (2012) 30:1144–51. doi: 10.1002/stem.1079
- Mazzini L, Gelati M, Profico DC, Sgaravizzi G, Projetti Pensi M, Muzi G, et al. Human neural stem cell transplantation in ALS: initial results from a phase I trial. *J Transl Med.* (2015) 13:17. doi: 10.1186/s12967-014-0371-2
- Karussis D, Karageorgiou C, Vaknin-Dembinsky A, Gowda-Kurkalli B, Gomori JM, Kassis I, et al. Safety and immunological effects of mesenchymal stem cell transplantation in patients with multiple sclerosis and amyotrophic lateral sclerosis. *Arch Neurol.* (2010) 10:1187– 94. doi: 10.1001/archneurol.2010.248
- Blanquer M, Moraleda JM, Iniesta F, Gómez-Espuch J, Meca-Lallana J, Villaverde R, et al. Neurotrophic bone marrow cellular nests prevent spinal motoneuron degeneration in amyotrophic lateral sclerosis patients: a pilot safety study. *Stem Cells.* (2012) 30:1277–85. doi: 10.1002/ stem.1080
- Ruiz-López FJ, Guardiola J, undIzura Verline, Gómez-Espuch J, uIniesta Fnderline, Blanquer Munderline, et al. Breathing pattern in a phase I clinical trial of intraspinal injection of autologous bone marrow mononuclear cells in patients with amyotrophic lateral sclerosis. *Respir Physiol Neurobiol.* (2016) 221:54–8. doi: 10.1016/j.resp.2015.11.007
- 91. García Santos JM, Blanquer Munderline, Torres del Río S, uIniesta Fnderline, uEspuch JGnderline, Pérez-Espejo MÁ, et al. Acute and chronic MRI changes in the spine and spinal cord after surgical stem cell grafting in patients with definite amyotrophic lateral sclerosis: post-infusion injuries are unrelated with clinical impairment. *Magn Reson Imaging*. (2013) 8:1298– 308. doi: 10.1016/j.mri.2013.05.006
- Oh KW, Noh MY, Kwon MS, Kim HY, Oh SI, Park J, et al. Repeated intrathecal mesenchymal stem cells for amyotrophic lateral sclerosis. *Ann Neurol.* (2018) 84:361–73. doi: 10.1002/ana.25302

- 93. Petrou P, Gothelf Y, Argov Z, Gotkine M, Levy YS, Kassis I, et al. Safety and clinical effects of mesenchymal stem cells secreting neurotrophic factor transplantation in patients with amyotrophic lateral sclerosis: results of phase 1/2 and 2a clinical trials. *JAMA Neurol.* (2016) 73:337– 44. doi: 10.1001/jamaneurol.2015.4321
- Nabavi SM, Arab L, Jarooghi N, Bolurieh T, Abbasi F, Mardpour S, et al. Safety, feasibility of intravenous and intrathecal injection of autologous Bone marrow derived mesenchymal stromal cells in patients with amyotrophic lateral sclerosis: an open label phase I clinical trial. *Cell J.* (2019) 20:592– 8. doi: 10.22074/cellj.2019.5370
- 95. Nurden AT, Nurden P, Sanchez M, Andia I, Anitua E. Platelets and wound healing. *Front Biosci.* (2008) 13:3532–48. doi: 10.2741/2947
- Burnouf T, Goubran HA, Chen TM, Ou KL, El-Ekiaby M, Radosevic M. Blood-derived biomaterials and platelet growth factors in regenerative medicine. *Blood Rev.* (2013) 27:77–89. doi: 10.1016/j.blre.2013.02.001
- 97. Nurden AT. The biology of the platelet with special reference to inflammation, wound healing and immunity. *Front Biosci.* (2018) 23:726–51. doi: 10.2741/4613
- Hayon Y, Dashevsky O, Shai E, Varon D, Leker RR. Platelet lysates stimulate angiogenesis, neurogenesis and neuroprotection after stroke. *Thromb Haemost.* (2013) 110:323–30. doi: 10.1160/TH12-11-0875
- 99. Zhang Y, Ying G, Ren C, Jizhang Y, Brogan D, Liu Z, et al. Administration of human platelet-rich plasma reduces infarction volume and improves motor function in adult rats with focal ischemic stroke. *Brain Res.* (2015) 1594:267–73. doi: 10.1016/j.brainres.2014.10.035
- 100. Anitua E, Pascual C, Antequera D, Bolos M, Padilla S, Orive G, et al. Plasma rich in growth factors (PRGF-Endoret) reduces neuropathologic hallmarks and improves cognitive functions in an Alzheimer's disease mouse model. *Neurobiol Aging.* (2014) 35:1582–95. doi: 10.1016/j.neurobiolaging.2014.01.009
- 101. Anitua E, Pascual C, Perez-Gonzalez R, Orive G, Carro E. Intranasal PRGF-Endoret enhances neuronal survival and attenuates NF-kappaB-dependent inflammation process in a mouse model of Parkinson's disease. J Control Release. (2015) 203:170–80. doi: 10.1016/j.jconrel.2015.02.030
- 102. Chou ML, Wu JW, Gouel F, Jonneaux A, Timmerman K, Renn TY, et al. Tailor-made purified human platelet lysate concentrated in neurotrophins for treatment of Parkinson's disease. *Biomaterials.* (2017) 142:77–89. doi: 10.1016/j.biomaterials.2017.07.018
- 103. Gouel F, Do Van B, Chou ML, Jonneaux A, Moreau C, Bordet R, et al. The protective effect of human platelet lysate in models of neurodegenerative disease: involvement of the Akt and MEK pathways. J Tissue Eng Regen Med. (2017) 11:3236–40. doi: 10.1002/term.2222

Conflict of Interest Statement: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Copyright © 2019 Gouel, Rolland, Devedjian, Burnouf and Devos. This is an openaccess article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.