



A Multiscale View of the Mechanisms Underlying Ketamine's Antidepressant Effects: An Update on Neuronal Calcium Signaling

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Kawatake-Kuno A, Murai T and Uchida S (2021) A Multiscale View of the Mechanisms Underlying Ketamine's Antidepressant Effects: An Update on Neuronal Calcium Signaling. Front. Behav. Neurosci. 15:749180. doi: 10.3389/fnbeh.2021.749180 Major depressive disorder (MDD) is a debilitating disease characterized by depressed mood, loss of interest or pleasure, suicidal ideation, and reduced motivation or hopelessness. Despite considerable research, mechanisms underlying MDD remain poorly understood, and current advances in treatment are far from satisfactory. The antidepressant effect of ketamine is among the most important discoveries in psychiatric research over the last half-century. Neurobiological insights into the ketamine's effects have shed light on the mechanisms underlying antidepressant efficacy. However, mechanisms underlying the rapid and sustained antidepressant effects of ketamine remain controversial. Elucidating such mechanisms is key to identifying new therapeutic targets and developing therapeutic strategies. Accumulating evidence demonstrates the contribution of the glutamatergic pathway, the major excitatory neurotransmitter system in the central nervous system, in MDD pathophysiology and antidepressant effects. The hypothesis of a connection among the calcium signaling cascade stimulated by the glutamatergic system, neural plasticity, and epigenetic regulation of gene transcription is further supported by its associations with ketamine's antidepressant effects. This review briefly summarizes the potential mechanisms of ketamine's effects with a specific focus on glutamatergic signaling from a multiscale perspective, including behavioral, cellular, molecular, and epigenetic aspects, to provide a valuable overview of ketamine's antidepressant effects.

Keywords: ketamine, antidepressant action, neuroplasticity, epigenetics, gene expression, stress, glutamate receptor, calcium signaling

INTRODUCTION

Major depressive disorder (MDD) is the leading cause of disability worldwide. Despite considerable research, biological mechanisms underlying MDD pathophysiology remain unclear, with significant unmet needs for treatment. Typical antidepressants, including selective serotonin reuptake inhibitors (SSRIs) and serotonin and noradrenaline reuptake inhibitors, increase

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monoamine concentration in the synaptic cleft, resulting in antidepressant effects (Berton and Nestler, 2006). However, although increased monoamine concentration in the synapse occurs relatively quickly as an acute pharmacological action, recovery from depression takes several weeks to months in clinical practice (Krishnan and Nestler, 2008). Electroconvulsive therapy (ECT) is also an effective treatment for drug-resistant depression, although achieving clinically meaningful or sustained remission with ECT required at least 1 month (Yamasaki et al., 2020). Such substantial time lags are a major concern since patients with depression are at high risk for suicide. Thus, there is an urgent need to develop antidepressants with rapid onset and sustained effectiveness.

Ketamine, a non-competitive glutamate N-methyl-Daspartate receptor (NMDAR) antagonist, has gained considerable interest in the neuropsychiatric field. A single administration of ketamine elicits rapid and sustained antidepressant effects for 1-2 weeks in both humans and animals (Berman et al., 2000; Zarate et al., 2006; Li et al., 2010; Autry et al., 2011). This discovery offered new insight into the investigation of a whole new class of agents beyond the monoamine system to treat depression (Chaki, 2017). Esketamine, an enantiomer of (R,S)-ketamine, has been approved by the U.S. Food and Drug Administration (USFDA) for treating patients with treatmentresistant depression. Thus, research on pathophysiology and drug discovery for MDD has transitioned from the monoaminergic to the glutamatergic system. Recently, the importance of multiscale neuroscience to study cross-scale interactions at genetic, molecular, cellular, and macroscale levels of brain circuitry, connectivity, and behavior has been emphasized to establish a comprehensive understanding of neuropsychiatric disease (Van Den Heuvel et al., 2019). This mini-review aims to update the current knowledge regarding ketamine effect on the brain, focusing on the glutamatergic signaling pathway from a multiscale perspective at the behavioral, cellular, molecular, and epigenetic levels.

THE GLUTAMATERGIC SYSTEM IN NEUROPLASTICITY, INTRACELLULAR SIGNALING, AND GENE EXPRESSION

Glutamate is the major excitatory neurotransmitter in the brain, and increasing evidence indicates that dysfunction in glutamatergic signaling contributes to MDD pathophysiology (Popoli et al., 2011; Duman and Aghajanian, 2012; Thompson et al., 2015; Duman et al., 2019; Xia et al., 2021). The glutamatergic system is modulated by both ionotropic [NMDARs, α -amino-3-hydroxy-5-methyl-4-isoxazolepropionic acid receptors (AMPARs), and kainate receptors] and metabotropic glutamate receptors (mGluRs). NMDARs are found throughout the central nervous system and contribute to synaptic calcium (Ca²⁺) influx, which is required for activity-dependent synaptic plasticity (Koester and Sakmann, 1998; Reid et al., 2001; Ngo-Anh et al., 2005; Bloodgood and Sabatini, 2007; Carter et al., 2007). NMDAR function is tightly linked to AMPAR, which gates sodium and mediates fast excitatory

transmission. Increased AMPAR density in the postsynaptic membrane causes NMDAR-dependent long-term potentiation (LTP) (Huganir and Nicoll, 2013). AMPARs can also have several direct effects on synaptic transmission (i.e., LTP) and intracellular signals without the proper functioning of NMDARs. This NMDAR-independent and AMPAR-dependent intracellular signaling pathway is also hypothesized to underlie ketamine's antidepressant actions (Zanos et al., 2016; Duman et al., 2019; Wei et al., 2021).

Ca²⁺ influx into the postsynaptic neuron stimulates a signaling-cascade, such as calcium/calmodulin-dependent kinases [CAMKs; e.g., calcium/calmodulin-dependent kinase II (CaMKIIs), eukaryotic elongation factor 2 (eEF2) kinase]. Brain-derived neurotrophic factor (BDNF) and its receptor, neurotrophic receptor tyrosine kinase 2 (TrkB), also plays a key role in synaptic plasticity (Minichiello, 2009). TrkB activation stimulates phospholipase Cy1 (PLCy1), which results in CaMK activation (Minichiello, 2009). Calcium-signaling activation further sends its signal toward downstream epigenetic and transcription modulators, such as MEF2, MeCP2, and HDAC5. These pathways modulate gene expression that affects dendritic growth, synaptic development, and neuronal plasticity (Greer and Greenberg, 2008; Graff and Tsai, 2013; Takemoto-Kimura et al., 2017; Uchida and Shumyatsky, 2018a,b; Figure 1). Taken together, calcium-signaling stimulation through NMDARs and/or AMPARs activates multiple downstream nucleocytoplasmic pathways; it induces activity-dependent epigenetic genetic expression, contributing to depression and antidepressant action.

Chronic stress initiates and exacerbates several psychiatric illnesses. Indeed, adverse stressful environments are associated with the pathophysiology of major psychiatric disorders, including mood and anxiety disorders (Mcewen, 2007; Krishnan and Nestler, 2008; Duman and Aghajanian, 2012). There are several evidences demonstrating alterations in the expression and/or function of glutamatergic signaling and its downstream molecules (e.g., NMDARs, AMPARs, CaMKIIs, MEF2, MeCP2, and HDAC5), which is associated with plasticity and behaviors induced by chronic stress, traditional antidepressant drugs, and/or ketamine (Table 1). Moreover, molecular dysregulation associated with glutamatergic system is visible in postmortem brain tissues of patients with MDD (Table 1). Thus, such clinical and preclinical evidences suggest that calcium-signaling is a downstream target of the glutamatergic system in MDD pathophysiology and antidepressant effects.

MECHANISMS OF KETAMINE'S ANTIDEPRESSANT EFFECTS: A MULTISCALE VIEW

Less than one-third of patients with MDD achieve remission using traditional antidepressant pharmacotherapy (Trivedi et al., 2006). Treatment resistance occurs in up to 30% of patients with MDD (Fava, 2003). However, a single subanesthetic dose of ketamine produces a therapeutic response within a few hours that lasts for several days in patients with depression



FIGURE 1 | Proposed mechanisms of ketamine's antidepressant action. The binding of ketamine to N-methyl-D-aspartate receptors (NMDARs) on GABAergic interneurons disinhibits glutamatergic neurons, which results in increased synaptic glutamate release. AMPAR activation by glutamate increases brain-derived neurotrophic factor (BDNF) levels. Although the exact source of BDNF is yet to be determined, local release of BDNF is thought to stimulate TrkB receptors. This activation activates intracellular signaling, such as the Ca²⁺ pathway. Another mechanism is the direct inhibition of NMDAR by ketamine. Inhibiting postsynaptic NMDARs reduces eEF2 via the inactivation of CaMK (eEF2 kinase), which leads to enhanced local protein synthesis of BDNF. Increased intracellular Ca²⁺ stimulates CaMKs and their downstream targets, including MeCP2, MEF2, and HDAC5. MeCP2, a transcriptional regulator, binds to methylated CpG sites on the genomic region and interacts with other transcription repressors, including HDACs, CaMKII phosphorylates MeCP2, promotes its nuclear export, and increases activity-dependent transcription. MEF2 recruits HDAC5 and removes activating acetyl groups from histones, which results in a silenced or repressed state of transcription. CaMKII phosphorylates HDAC5, which promotes nuclear export and increases activity-dependent transcription. Ketamine is known to increase the phosphorylation of CaMKII, MeCP2, and HDAC5 (see detail in the main text). Thus, ketamine-mediated enhancement of intracellular Ca²⁺ signaling is linked to epigenetic regulation of transcription, which leads to long-term synaptic plasticity and, consequently, prolonged antidepressant-like effects.

(Berman et al., 2000; Zarate et al., 2006). Intravenous infusion of ketamine results in clinical response and remission in 70 and 30% of treatment-resistant patients with MDD, respectively (Zarate et al., 2006). Additionally, Ketamine reduces suicidal ideation (Krystal et al., 2013). In 2020, *esketamine* was approved by the USFDA for treating depressive symptoms in adults with MDD having acute suicidal ideation or behavior.

Ketamine elicits robust unwanted side effects, including prepulse-inhibition deficits, cognitive deficits, and schizophrenia-like psychotic symptoms in humans (Lahti et al., 1995; Chan et al., 2013; Giorgetti et al., 2015). Recent preclinical data indicate that ketamine's enantiomer (*R*)ketamine (Hashimoto, 2019; Wei et al., 2021) and its metabolites (*2R*, *6R*)-hydroxynorketamine (HNK) (Zanos et al., 2016) exert antidepressant effects with fewer adverse effects than do ketamine or (*S*)-ketamine. Since potential mechanisms underlying the rapid antidepressant actions of ketamine and its metabolites have been reviewed elsewhere (Fukumoto et al., 2017; Yang C. et al., 2018; Duman et al., 2019; Krystal et al., 2019; Sial et al., 2020; Highland et al., 2021; Shinohara et al., 2021; Wei et al., 2021; Xia et al., 2021), we review the recent progress in deciphering mechanisms underlying ketamine's sustained antidepressant effects, with a particular focus on the role of calcium signaling from a multiscale perspective.

Behavioral Effects of Ketamine

Several animal studies have demonstrated antidepressant-like responses to ketamine. A single intraperitoneal injection of ketamine or its metabolites produces rapid (30 min–1 h) and long-lasting (24 h–7 days) antidepressant effects (Autry et al., 2011; Koike et al., 2011; Zhou et al., 2014; Sun et al., 2016; Zanos et al., 2016; Yang C. et al., 2018; Kim et al., 2021). Moreover, such ketamine antidepressant effects have been observed in not only naïve, non-stressed animals but also in animals subjected to adverse stressful life events. Animals exposed to chronic stress show despair-like behavior, anhedonia, anxiety, and/or social avoidance, whereas a single injection of ketamine or its metabolites rapidly reverses these deleterious effects and exerts long-term effects (Li et al., 2011; Zanos et al., 2016; Duman et al., 2019; Wei et al., 2021).

Neurobiological Effects of Ketamine

Neuroimaging studies have shown structural and functional alterations in the hippocampus and dorsomedial prefrontal cortex (dmPFC) of patients with MDD (Price and Drevets, 2010; Macqueen and Frodl, 2011). Human functional magnetic resonance imaging (MRI) studies have demonstrated that a single dose of ketamine ameliorates reductions in functional connectivity in the prefrontal cortex (PFC), which is associated with the alleviation of depressive symptoms (Abdallah et al., 2017). Interestingly, a recent MRI study in animals demonstrated short- and long-term effects of ketamine on distinct brain circuitry. Gass et al. (2019) found in an animal model of depression that ketamine causes a rapid response in the amygdala, anterodorsal hippocampus, and ventral pallidum, which are related to cognitive, sensory, emotional, and reward functions. However, 48 h after administration, ketamine showed a long-term normalization of the habenula, midline thalamus, and hippocampal connectivity. They mediate cognitive flexibility for processing contextual information, distinguish contextual cues in safe versus threatening situations, and modulate fear and emotional responses in non-threatening environments (Gass et al., 2019).

There is increasing evidence suggesting altered neuronal and structural plasticity in animal models of depression as well as in patients with MDD (Duman and Aghajanian, 2012; Kang et al., 2012; Abe-Higuchi et al., 2016; Higuchi et al., 2016; Nie et al., 2018; Uchida et al., 2018; Sakai et al., 2021). Ketamine rapidly increases the number and function of spine synapses. Furthermore, Li et al. found that ketamine increases the number and function of spine synapses in the medial PFC (mPFC) TABLE 1 | Example evidence indicates alterations in behavior, glutamatergic signaling, and its downstream pathways regarding depression, chronic stress, and antidepressants: translational and multiscale views.

Benaviors		
	Findings	References
Ketamine's effects on a stress-induced animal model of depression	CUMS-induced increase of immobility in TST were reversed 0.5 and 72 h after ketamine treatment in rats	Sun et al., 2016
	CUS-induced reduction in sucrose preference in SPT was reversed by ketamine 24 h after injection in rats CSDS-induced reduction of social interaction was reversed 24 h after (<i>2R</i> , 6 <i>R</i>)-HNK treatment in mice	Li et al., 2011 Zanos et al., 2016 Yang C. et al. 2018
Ketamine's effects on pharmacological model of depression	CSDS-induced depression-like behaviors were reversed 24 in alter (A)-ketamine treatment in mice Chronic CORT effects on immobility in TST, open-arm exploration in an elevated plus maze and sucrose preference were reversed 24h after ketamine treatment in mice	Moda-Sava et al., 2019
	Chronic CORT-induced anhedonia in a sucrose preference test was recovered by (2S, 6S)-HNK treatment LPS-induced increase of immobility in FST was reversed by (<i>R</i>)- Ketamine, but not (<i>R</i>)- HNK, in mice Chronic CORT-induced anhedonia and increased immobility time in FST were improved by (2S, 6S)-HNK, but not (<i>2R</i> , 6 <i>R</i>)-HNK	Zanos et al., 2016 Yamaguchi et al., 2018 Yokoyama et al., 2020
Neuroplasticity		
	Findings	References
MDD patients	Postmortem brain of MDD patients showed a lower number of synapses in dIPFC Meta-analysis of structural imaging studies demonstrated that MDD patients have smaller hippocampus	Kang et al., 2012 Macqueen and Frodl, 2011
	Volumes	Drian and Dravata, 0010
	Meta-analysis of imaging showed the structural and functional decline in dmPFC of MDD patients	Price and Drevets, 2010
model of depression	Reduced spine density in the hippocampus and mPFC of mice susceptible to CUMS and CSDS	Abe-Higuchi et al., 2016; Higuchi et al., 2016; Nie et al., 2018; Sakai et al., 2021
	Repeated stress impairs glutamatergic transmission in PFC pyramidal neurons	Yuen et al., 2012
Ketamine's effect	(S)-ketamine normalized habenula, midline thalamus, and hippocampal connectivity at 48 h in fMRI imaging of stressed rats	Gass et al., 2019
	Ketamine blocks NMDAR spontaneous activity	Autry et al., 2011
	Ketamine treatment restores lost spines by chronic CORT exposure and promote generating functional synapses in mice	Moda-Sava et al., 2019
	Ketamine treatment increases the number and function of spine synapse in rat mPFC	Li et al., 2010
	(2R,6R)-HNK increased fEPSC slope in SC-CA1 of rats (2S,6S)-HNK caused no changes in sEPSC frequency or amplitudes in rat CA1 interneurons (but has	Zanos et al., 2016 Chen et al., 2012
Molecular pathway/Int		
	Findings	References
NMDARs	MDD and stress model	
	A postmortem prefrontal cortex showed increased levels of NR1 in MDD	Rodriquez-Munoz et al., 2017
	Reduced GluN2A in prefrontal cortex of MDD	Beneyto and Meador-Woodruft 2008
	MK801, a NMDAR antagonist, injection reduced immobility in FST	Autry et al., 2011
	CUS-induced reduction in sucrose preference in SPT was reversed by a selective NR2B antagonist, Ro 25-6981, 24 h after injection in rats	Li et al., 2011
	Ketamine treatment increases NR1 expression levels in mouse PFC Ketamine and a high dose of (2R, 6R)-HNK influences NMDAR-mediated eEF2 phosphorylation	Liu et al., 2011 Autry et al., 2011; Suzuki et al. 2017
	(2R. 6R)-HNK do not block NMDAR function	Lumsden et al., 2019
AMPARs	MDD and stress model	
	Postmortem cortical tissue from MDD patients showed decreased GluA1 levels	Beneyto et al., 2007
	Reduced GluA1 level in the hippocampus of stress-susceptible mice AMPAR potentiator drives stress resilience, whereas GluA1 inhibition leads to stress susceptibility Kataming	Sakai et al., 2021
	Ketamine increased the level of GluA1 subunit in the mouse biopocamous	Reurel et al. 2016
	(2R. 6R)-HNK increased synaptic GluA1 and GluA2 protein expression in the mouse hippocampus	Zanos et al., 2016
BDNF/TrkB	MDD and stress model Restmostern brein tissues from the biopocompute and profrontal contex in a violate subjects about a stress	Dwivedi et al. 2002
	expression of BDNF and TrkB	Dwiveur et al., 2003
	שטוער ieveis were lower in the anterior cingulate of postmortem brains of subjects with early life adversity and/or died by suicide	YOUSSET ET AL., 2018
		(Continued

TABLE 1 | (Continued)

Ketamine	
CUMS-induced reduction of the expression of BDNF was reversed 0.5 and 72 h after ketamine treatment in rats	Sun et al., 2016
The deletion of BDNF or TrkB in broad forebrain regions of mice blocks ketamine's antidepressant effects	Nosyreva et al., 2013, 2014
Neutralizing a BDNF antibody into the mPFC blocks the behavioral effects of ketamine in FST	Lepack et al., 2014
(2R,6R)-HNK increased BDNF protein levels 24 h after injection in mouse hippocampus	Zanos et al., 2016
(2S,6S)-HNK increased extracellular BDNF levels in the mouse prefrontal cortex	Anderzhanova et al., 2020
MDD and stress model	
A postmortem study showed decreased levels of CAMK2B in the anterior cingulate cortex of MDD	Seney et al., 2018
A postmortem prefrontal cortex study showed decreased levels of CAMK2A in MDD	Fuchsova et al., 2015
A postmortem prefrontal cortex study showed increased levels of CAMK2A in MDD	Tochigi et al., 2008
CaMKIIβ levels in the ventral HPC were lower in mice following CUMS. CaMKIIβ activation reversed depression-like behaviors	Sakai et al., 2021
Ketamine	
CaMKIIB activity is increased at 3 days after ketamine injection	Kim et al., 2021
MDD and stress model	
p-MeCP2 levels decreased in the hippocampus and prefrontal cortex of suicide victims	Misztak et al., 2020
MeCP2 complexes determine stress susceptibility and resilience in mice	Uchida et al., 2011
Ketamine	
p-MeCP2 is required for ketamine-induced metaplasticity and antidepressant effects	Kim et al., 2021
MDD and stress model	
MEF2C is one of the candidate risk genes for MDD	Hyde et al., 2016
Ketamine	
Ketamine enhances the transcriptional activity of MEF2 in mice hippocampus	Choi et al., 2015
MDD and stress model	
Increased HDAC5 level in MDD	lga et al., 2007; Hobara et al., 2010
HDAC5 overexpression in the hippocampus disrupts antidepressant-like effect of traditional antidepressant	Tsankova et al., 2006
HDAC 4/5 inhibitor induces antidepressant-like behavioral effects in mice	Higuchi et al., 2016
Ketamine	
Ketamine induces the phosphorylation of HDAC5 at 30 min and 24 h after administration in mice hippocampus	Choi et al., 2015
	 Ketamine CUMS-induced reduction of the expression of BDNF was reversed 0.5 and 72 h after ketamine treatment in rats The deletion of BDNF or TrkB in broad forebrain regions of mice blocks ketamine's antidepressant effects Neutralizing a BDNF antibody into the mPFC blocks the behavioral effects of ketamine in FST (<i>2R, 6P</i>)-HNK increased BDNF protein levels 24 h after injection in mouse hippocampus (<i>2S, SS</i>)-HNK increased extracellular BDNF levels in the mouse prefrontal cortex MDD and stress model A postmortem study showed decreased levels of <i>CAMK2B</i> in the anterior cingulate cortex of MDD A postmortem prefrontal cortex study showed decreased levels of <i>CAMK2A</i> in MDD CaMKliß levels in the ventral HPC were lower in mice following CUMS. CaMKliß activation reversed depression-like behaviors Ketamine CaMKliß activity is increased at 3 days after ketamine injection MDD and stress model p-MeCP2 levels decreased in the hippocampus and prefrontal cortex of suicide victims MeCP2 complexes determine stress susceptibility and resilience in mice Ketamine p-MeCP2 lis required for ketamine-induced metaplasticity and antidepressant effects MDD and stress model MEEP2C is one of the candidate risk genes for MDD Ketamine Ketamine enhances the transcriptional activity of MEF2 in mice hippocampus MDD and stress model Increased <i>HDACS</i> level in MDD HDACS overexpression in the hippocampus disrupts antidepressant-like effect of traditional antidepressant HDAC 4/5 Inhibitor induces antidepressant-like behavioral effects in mice Ketamine Ketamine ina Ketamine ina Ketamine ina Ketamine Ketamine Ketamine Ketamine Ketamine Ketamine enhances the transcriptional activity of MEF2 in mice hippocampus MDD and stress model Increased

CUMS, chronic unpredictable mild stress; CUS, chronic unpredictable stress; CSDS, chronic social defeat stress; CORT, corticosterone; LPS, lipopolysaccharide; HNK, hydroxynorketamine; MDD, major depressive disorder; SSRI, selective serotonin reuptake inhibitor; FST, forced-swimmed test; SPT, sucrose preference test; TST, tail suspension Test; sEPSC, spontaneous excitatory postsynaptic current; fEPSC, field excitatory postsynaptic current; dIPFC, dorsolateral prefrontal cortex; mPFC, medial prefrontal cortex; dmPFC, dorsomedial prefrontal cortex.

and rapidly reverses synaptic abnormalities caused by chronic stress exposure (Li et al., 2010). Although this evidence suggests an association between ketamine-induced spinogenesis and antidepressant-like behavior, the causal relationship is unclear. However, a recent report by Moda-Sava et al. has addressed this issue. They used a photoactivable proof to selectively reverse ketamine effects on spine formation in the PFC. They found that newly formed spines are necessary for and play a specific role in the sustained antidepressant-like behavior induced by ketamine treatment (Moda-Sava et al., 2019).

Ketamine-Induced Synaptic Plasticity

Brain-derived neurotrophic factor and its receptor TrkB play key roles in synaptic plasticity, stress, and depression (Duman and Monteggia, 2006; Minichiello, 2009; Castren and Monteggia, 2021). A recent report discovered that several antidepressants, including fluoxetine, imipramine, and ketamine, directly bind to TrkB, facilitating BDNF action and plasticity (Casarotto et al., 2021). In addition, increased BDNF-TrkB signaling in rodent frontocortical/hippocampal circuits has been observed following acute treatment with ketamine (Li et al., 2010; Autry et al., 2011).

Clinical evidence suggests that repeated ketamine administration allows cumulative and sustained antidepressant effects and that it is more effective than a single injection in patients with MDD (Aan Het Rot et al., 2010; Murrough et al., 2013; Phillips et al., 2019). The threshold and sensitivity of the persistent increase and decrease of synaptic strength are subject to activity-dependent regulation. This type of plasticity, called "metaplasticity," is important for stabilizing synaptic strength and preventing LTP saturation and long-term depression, leading to homeostatic alternations of synaptic activation (Bienenstock et al., 1982; Turrigiano et al., 1998; Kavalali and Monteggia, 2020). Notably, a preclinical study suggested that ketamine administration elicits metaplastic effects on LTP modulation and potentially other processes for long term. Kim et al. (2021) reported that, by using slice recordings of the Schaffer collateral-CA1 pathway in the hippocampus, ketamine induces AMPAR-mediated synaptic potentiation. Interestingly, this effect was more than two-fold higher in brain slices of mice that had received ketamine 7 days earlier, suggesting a priming effect of ketamine treatment such that subsequent ketamine augments synaptic potentiation. Further experiments to understand the mechanisms of this metaplasticity will provide critical insight into mechanisms underlying ketamine's potent and prolonged antidepressant effects.

Ketamine-Induced Ca²⁺ Signaling Cascades

N-methyl-D-aspartate receptors activate eEF2 via CaMKs (eEF2 kinases) and depress BDNF levels (Scheetz et al., 2000). Ketamine-induced suppression of postsynaptic NMDARs deactivates eEF2 kinase, leading to reduced eEF2 phosphorylation and increased translation of BDNF in the hippocampus (Autry et al., 2011; Suzuki and Monteggia, 2020). This signaling pathway then potentiates synaptic AMPAR responses through the insertion of GluA1/2 subunits (Autry et al., 2011). In contrast, ketamine's metabolite (*2R*, *6R*)-HNK has NMDAR inhibition-independent antidepressant actions (Zanos et al., 2016; Lumsden et al., 2019), whereas other reports have shown that NMDAR inhibition at a high dose of (*2R*, *6R*)-HNK triggers intracellular signaling via eEF2 (Suzuki et al., 2017).

A transient burst of glutamate via NMDAR blockade on GABAergic interneurons by ketamine activates postsynaptic AMPARs in excitatory neurons. This activation induces depolarization and activation of NMDARs that trigger Ca2+ influx, releasing BDNF (Krystal et al., 2019). Local release of BDNF is thought to activate TrkB on the postsynaptic membrane, stimulating the ERK and PI3K-Akt signaling pathways and mammalian target of rapamycin complex 1 (mTORC1) phosphorylation to promote synapse formation by stimulating synaptic proteins, such as GluA1 and PSD-95, which are required for synaptic plasticity (Cavalleri et al., 2018). Recently, mTORC1 effectors 4E-BP2 and 4-EB2 in excitatory or inhibitory neurons underlie behavioral and neurobiological responses to ketamine (Aguilar-Valles et al., 2021). Ketamine-induced activation of TrkB increases GSK-3β phosphorylation via the ERK signaling pathway, decreasing PSD-95 phosphorylation and internalizing the AMPA GluA1 subunit, which upregulates signaling through the GluA1 to promote synapse formation (Liu et al., 2013; Beurel et al., 2016). Ketamine-dependent changes in dendritic arborization and soma size are abolished by AMPAR antagonists or mTOR complex/signaling inhibitors (Cavalleri et al., 2018). Intracellular molecular signaling cascades stimulated by the glutamatergic pathway may be associated with ketamine-induced structural and synaptic plasticity and its antidepressant effects.

As mentioned earlier, CaMKIIs are major downstream target for the glutamatergic pathway and might be involved in

stress and depression. TrkB activation stimulates phospholipase Cy1 (PLCy1) and also results in the activation of CaMKs (Minichiello, 2009). Activated CaMKIIs further stimulate MeCP2 phosphorylation (Zhou et al., 2006), allowing the transcription of downstream target genes. A recent study showed that MeCP2 phosphorylation at S421 (p-MeCP2) is essential for the expression of metaplasticity and the sustained, but not acute, antidepressant effects of ketamine (Kim et al., 2021). Hippocampal BDNF protein levels were shown to increase rapidly 30 min after ketamine administration but returned to baseline 3 days after injection. In contrast, hippocampal p-MeCP2 levels increased 3 and 7 days, but not 30 min, after ketamine injection. CaMKIIß were elevated at 3 days after ketamine injection but returned to baseline at 7 days. These findings indicate that CaMKIIB plays a role in the intermediary process between BDNF activation and MeCP2 phosphorylation required for the sustained antidepressant effects of ketamine. This hypothesis is also supported, at least in part, by a recent finding that hippocampal CaMKIIB is downregulated in chronic stress-susceptible mice and that shortterm (within 4 days) CaMKIIß activation ameliorates depressionlike behaviors (Sakai et al., 2021).

Epigenetic Regulation of Gene Transcription by Ketamine

The interplay between genetic and environmental factors underlies depression pathophysiology, and epigenetic mechanisms might contribute to these interactions (Nestler et al., 2016; Uchida et al., 2018; Kawatake-Kuno et al., 2021). Although accumulating evidence demonstrated altered epigenetic functioning in animal models of depression and postpartum MDD-patient brains, few studies have used ketamine-induced transcriptome and epigenome analyses to characterize ketamine's antidepressant effects. Genome-wide transcriptome and epigenome mapping offer a template for several strategies to identify novel drug targets in unbiased ways to develop more effective treatments for MDD (Bagot et al., 2017). Here we summarize how ketamine-induced activation of Ca²⁺ signal influences epigenetic regulation of gene transcription.

MeCP2, MEF2, and HDAC5 functions are regulated by Ca²⁺ signaling and are associated with stress and depression (Table 1). As mentioned above, p-MeCP2 is necessary for sustained antidepressant response to ketamine (Kim et al., 2021). MeCP2 is a methylated cytosine reader that impacts chromatin organization with any change in DNA methylation. A previous report showed that chronic stress differentially modulates MeCP2 activity in stress-resilient and -susceptible mice and subsequent epigenetic gene transcription (Uchida et al., 2011). Thus, ketamine-induced enhancement of p-MeCP2 may be associated with the formation of chromatin-remodeling complexes on target genes and, thus, transcription regulation. HDAC5 is a histone deacetylase, and its phosphorylation by CaMKs is associated with transcription repression (Mckinsey et al., 2000). Hippocampal HDAC5 is associated with behavioral response to chronic stress and traditional antidepressants (e.g., imipramine and SSRIs) (Tsankova et al., 2006; Higuchi et al., 2016). A recent study suggested that ketamine rapidly induces HDAC5 phosphorylation and nuclear export through CaMKII-dependent pathways, which leads to enhanced MEF2 transcription that regulates neuronal structural and functional plasticity (Choi et al., 2015). Correspondingly, HDAC5 knockdown occludes the actions of ketamine. Moreover, MeCP2 is considered as a master regulator of metaplasticity (Chen et al., 2012). Ca^{2+} -signal-mediated modulation of MeCP2, HDAC5, and MEF2 functions may be involved in the sustained antidepressant response of ketamine through epigenetic transcription.

CONCLUSION

This mini-review highlights that the glutamatergic pathway is associated with behavioral, neuroplastic, neurobiological, molecular, and epigenetic effects of ketamine, focusing on Ca^{2+} signaling wherein its dysfunction is involved in depression pathophysiology according to both clinical and animal studies. Such (reverse) translational implications for bridging the research gap between human depression and animal models will provide a better understanding of how ketamine affects and modulates depression pathophysiology and ultimately contribute to the clinical application of ketamine or the development of related compounds for wide range of psychiatric disorders. Glutamatergic transmission and monoaminergic systems induce rapid biological changes that induce fast antidepressant effects. In contrast, ketamine's sustained antidepressant actions are likely mediated by intracellular Ca²⁺ signaling cascades that affect neurobiological processes, including dendritic spine formation, epigenetic modifications, and long-term synaptic plasticity, and consequently, maintain physiological functioning.

In this mini-review, we particularly focused on the hippocampus and prefrontal cortex, key brain regions associated with MDD pathophysiology and ketamine's antidepressant effect. However, other brain regions were suggested to also be involved in these processes, such as the lateral habenula. Emerging evidence from preclinical and clinical studies identified an important role of the lateral habenula in depression and ketamine's antidepressant effect through a glutamatergic pathway

REFERENCES

- Aan Het Rot, M., Collins, K. A., Murrough, J. W., Perez, A. M., Reich, D. L., Charney, D. S., et al. (2010). Safety and efficacy of repeated-dose intravenous ketamine for treatment-resistant depression. *Biol. Psychiatry* 67, 139–145. doi: 10.1016/j.biopsych.2009. 08.038
- Abdallah, C. G., Averill, L. A., Collins, K. A., Geha, P., Schwartz, J., Averill, C., et al. (2017). Ketamine Treatment and Global Brain Connectivity in Major Depression. *Neuropsychopharmacology* 42, 1210–1219. doi: 10.1038/npp.201 6.186
- Abe-Higuchi, N., Uchida, S., Yamagata, H., Higuchi, F., Hobara, T., Hara, K., et al. (2016). Hippocampal Sirtuin 1 Signaling Mediates Depressionlike Behavior. *Biol. Psychiatry* 80, 815–826. doi: 10.1016/j.biopsych.2016. 01.009

(Li et al., 2013; Cui et al., 2018a,b, 2019; Yang et al., 2018; Hu, 2019, Hu et al., 2020). In addition, dynamic molecular changes were observed in the nucleus accumbens of animal models of depression and ketamine-treated animals (Bagot et al., 2017). Thus, future studies are warranted to clarify how ketamine impacts neuronal circuit activity and identify underlying molecular and epigenetic mechanisms.

In summary, ketamine has great potential in the development of groundbreaking neuropsychiatric therapies. Our current understanding of depression pathophysiology and ketamine's action suggests that diverse drug actions converge around Ca²⁺-signaling-mediated neural plasticity. However, ketamine plays diverse roles in the glutamatergic pathway and other neurotransmitter systems, neurogenesis, inflammation, and even body-brain crosstalk. Furthermore, several studies have suggested the distinct roles of ketamine enantiomers ([S]ketamine and [R]-ketamine) and their metabolites ([2R,6R]-HNK and [2S,6S]-HNK) in plasticity and behavior (Zanos et al., 2016; Yamaguchi et al., 2018; Hashimoto, 2019; Lumsden et al., 2019; Yokovama et al., 2020; Highland et al., 2021; Wei et al., 2021). Thus, mechanisms underlying ketamine's actions remain controversial. Moreover, ketamine effects at the mesoscale of neural architecture and macroscale of neural connectivity, cognition, and behavior are poorly understood. Further investigations at both the multiscale and multisystem levels are necessary to comprehensively understand mechanisms underlying ketamine's antidepressant effects and develop novel drugs for treating MDD.

AUTHOR CONTRIBUTIONS

All authors listed have made a substantial, direct and intellectual contribution to the work, and approved it for publication.

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- Aguilar-Valles, A., De Gregorio, D., Matta-Camacho, E., Eslamizade, M. J., Khlaifia, A., Skaleka, A., et al. (2021). Antidepressant actions of ketamine engage cellspecific translation via eIF4E. *Nature* 590, 315–319. doi: 10.1038/s41586-020-03047-0
- Anderzhanova, E., Hafner, K., Genewsky, A. J., Soliman, A., Pohlmann, M. L., Schmidt, M. V., et al. (2020). The stress susceptibility factor FKBP51 controls S-ketamine-evoked release of mBDNF in the prefrontal cortex of mice. *Neurobiol. Stress* 13:100239. doi: 10.1016/j.ynstr.2020.100239
- Autry, A. E., Adachi, M., Nosyreva, E., Na, E. S., Los, M. F., Cheng, P. F., et al. (2011). NMDA receptor blockade at rest triggers rapid behavioural antidepressant responses. *Nature* 475, 91–95. doi: 10.1038/nature10130
- Bagot, R. C., Cates, H. M., Purushothaman, I., Vialou, V., Heller, E. A., Yieh, L., et al. (2017). Ketamine and Imipramine Reverse Transcriptional Signatures of Susceptibility and Induce Resilience-Specific Gene Expression Profiles. *Biol. Psychiatry* 81, 285–295. doi: 10.1016/j.biopsych.2016.06.012

- Beneyto, M., Kristiansen, L. V., Oni-Orisan, A., Mccullumsmith, R. E., and Meador-Woodruff, J. H. (2007). Abnormal glutamate receptor expression in the medial temporal lobe in schizophrenia and mood disorders. *Neuropsychopharmacology* 32, 1888–1902. doi: 10.1038/sj.npp.1301312
- Beneyto, M., and Meador-Woodruff, J. H. (2008). Lamina-specific abnormalities of NMDA receptor-associated postsynaptic protein transcripts in the prefrontal cortex in schizophrenia and bipolar disorder. *Neuropsychopharmacology* 33, 2175–2186. doi: 10.1038/sj.npp.1301604
- Berman, R. M., Cappiello, A., Anand, A., Oren, D. A., Heninger, G. R., Charney, D. S., et al. (2000). Antidepressant effects of ketamine in depressed patients. *Biol. Psychiatry* 47, 351–354. doi: 10.1016/s0006-3223(99)00230-9
- Berton, O., and Nestler, E. J. (2006). New approaches to antidepressant drug discovery: beyond monoamines. *Nat. Rev. Neurosci.* 7, 137–151. doi: 10.1038/ nrn1846
- Beurel, E., Grieco, S. F., Amadei, C., Downey, K., and Jope, R. S. (2016). Ketamine-induced inhibition of glycogen synthase kinase-3 contributes to the augmentation of alpha-amino-3-hydroxy-5-methylisoxazole-4-propionic acid (AMPA) receptor signaling. *Bipolar. Disord.* 18, 473–480. doi: 10.1111/bdi. 12436
- Bienenstock, E. L., Cooper, L. N., and Munro, P. W. (1982). Theory for the development of neuron selectivity: orientation specificity and binocular interaction in visual cortex. *J. Neurosci.* 2, 32–48. doi: 10.1523/JNEUROSCI.02-01-00032.1982
- Bloodgood, B. L., and Sabatini, B. L. (2007). Nonlinear regulation of unitary synaptic signals by CaV(2.3) voltage-sensitive calcium channels located in dendritic spines. *Neuron* 53, 249–260. doi: 10.1016/j.neuron.2006.12.017
- Carter, A. G., Soler-Llavina, G. J., and Sabatini, B. L. (2007). Timing and location of synaptic inputs determine modes of subthreshold integration in striatal medium spiny neurons. J. Neurosci. 27, 8967–8977. doi: 10.1523/JNEUROSCI. 2798-07.2007
- Casarotto, P. C., Girych, M., Fred, S. M., Kovaleva, V., Moliner, R., Enkavi, G., et al. (2021). Antidepressant drugs act by directly binding to TRKB neurotrophin receptors. *Cell* 184, 1299–1313e1219. doi: 10.1016/j.cell.2021.01.034
- Castren, E., and Monteggia, L. M. (2021). Brain-Derived Neurotrophic Factor Signaling in Depression and Antidepressant Action. *Biol. Psychiatry* 90, 128– 136. doi: 10.1016/j.biopsych.2021.05.008
- Cavalleri, L., Merlo Pich, E., Millan, M. J., Chiamulera, C., Kunath, T., Spano, P. F., et al. (2018). Ketamine enhances structural plasticity in mouse mesencephalic and human iPSC-derived dopaminergic neurons via AMPAR-driven BDNF and mTOR signaling. *Mol. Psychiatry* 23, 812–823. doi: 10.1038/mp.2017.241
- Chaki, S. (2017). Beyond Ketamine: New Approaches to the Development of Safer Antidepressants. Curr. Neuropharmacol. 15, 963–976. doi: 10.2174/ 1570159X15666170221101054
- Chan, K. W., Lee, T. M., Siu, A. M., Wong, D. P., Kam, C. M., Tsang, S. K., et al. (2013). Effects of chronic ketamine use on frontal and medial temporal cognition. *Addict. Behav.* 38, 2128–2132. doi: 10.1016/j.addbeh.2013.0 1.014
- Chen, S. X., Cherry, A., Tari, P. K., Podgorski, K., Kwong, Y. K., and Haas, K. (2012). The transcription factor MEF2 directs developmental visually driven functional and structural metaplasticity. *Cell* 151, 41–55. doi: 10.1016/j.cell. 2012.08.028
- Choi, M., Lee, S. H., Wang, S. E., Ko, S. Y., Song, M., Choi, J. S., et al. (2015). Ketamine produces antidepressant-like effects through phosphorylationdependent nuclear export of histone deacetylase 5 (HDAC5) in rats. *Proc. Natl. Acad. Sci. U S A* 112, 15755–15760. doi: 10.1073/pnas.151391 3112
- Cui, Y., Hu, S., and Hu, H. (2019). Lateral Habenular Burst Firing as a Target of the Rapid Antidepressant Effects of Ketamine. *Trends Neurosci.* 42, 179–191. doi: 10.1016/j.tins.2018.12.002
- Cui, Y., Yang, Y., Dong, Y., and Hu, H. (2018a). Decoding Depression: Insights from Glial and Ketamine Regulation of Neuronal Burst Firing in Lateral Habenula. *Cold Spr. Harb. Symp. Quant. Biol.* 83, 141–150. doi: 10.1101/sqb. 2018.83.036871
- Cui, Y., Yang, Y., Ni, Z., Dong, Y., Cai, G., Foncelle, A., et al. (2018b). Astroglial Kir4.1 in the lateral habenula drives neuronal bursts in depression. *Nature* 554, 323–327. doi: 10.1038/nature25752
- Duman, R. S., and Aghajanian, G. K. (2012). Synaptic dysfunction in depression: potential therapeutic targets. *Science* 338, 68–72. doi: 10.1126/science.1222939

- Duman, R. S., and Monteggia, L. M. (2006). A neurotrophic model for stressrelated mood disorders. *Biol. Psychiatry* 59, 1116–1127. doi: 10.1016/j.biopsych. 2006.02.013
- Duman, R. S., Shinohara, R., Fogaca, M. V., and Hare, B. (2019). Neurobiology of rapid-acting antidepressants: convergent effects on GluA1-synaptic function. *Mol. Psychiatry* 24, 1816–1832. doi: 10.1038/s41380-019-0400-x
- Dwivedi, Y., Rizavi, H. S., Conley, R. R., Roberts, R. C., Tamminga, C. A., and Pandey, G. N. (2003). Altered gene expression of brain-derived neurotrophic factor and receptor tyrosine kinase B in postmortem brain of suicide subjects. *Arch. Gen. Psychiatry* 60, 804–815. doi: 10.1001/archpsyc.60.8.804
- Fava, M. (2003). Diagnosis and definition of treatment-resistant depression. Biol. Psychiatry 53, 649–659. doi: 10.1016/s0006-3223(03)00231-2
- Fuchsova, B., Alvarez Julia, A., Rizavi, H. S., Frasch, A. C., and Pandey, G. N. (2015). Altered expression of neuroplasticity-related genes in the brain of depressed suicides. *Neuroscience* 299, 1–17. doi: 10.1016/j.neuroscience.2015.04.057
- Fukumoto, K., Toki, H., Iijima, M., Hashihayata, T., Yamaguchi, J. I., Hashimoto, K., et al. (2017). Antidepressant Potential of (R)-Ketamine in Rodent Models: Comparison with (S)-Ketamine. J. Pharmacol. Exp. Ther. 361, 9–16. doi: 10. 1124/jpet.116.239228
- Gass, N., Becker, R., Reinwald, J., Cosa-Linan, A., Sack, M., Weber-Fahr, W., et al. (2019). Differences between ketamine's short-term and long-term effects on brain circuitry in depression. *Transl. Psychiatry* 9:172. doi: 10.1038/s41398-019-0506-6
- Giorgetti, R., Marcotulli, D., Tagliabracci, A., and Schifano, F. (2015). Effects of ketamine on psychomotor, sensory and cognitive functions relevant for driving ability. *Forensic. Sci. Int.* 252, 127–142. doi: 10.1016/j.forsciint.2015.04.024
- Graff, J., and Tsai, L. H. (2013). Histone acetylation: molecular mnemonics on the chromatin. *Nat. Rev. Neurosci.* 14, 97–111. doi: 10.1038/nrn3427
- Greer, P. L., and Greenberg, M. E. (2008). From synapse to nucleus: calciumdependent gene transcription in the control of synapse development and function. *Neuron* 59, 846–860. doi: 10.1016/j.neuron.2008.09.002
- Hashimoto, K. (2019). Rapid-acting antidepressant ketamine, its metabolites and other candidates: A historical overview and future perspective. *Psychiatry Clin. Neurosci.* 73, 613–627. doi: 10.1111/pcn.12902
- Highland, J. N., Zanos, P., Riggs, L. M., Georgiou, P., Clark, S. M., Morris, P. J., et al. (2021). Hydroxynorketamines: Pharmacology and Potential Therapeutic Applications. *Pharmacol Rev* 73, 763–791. doi: 10.1124/pharmrev.120.000149
- Higuchi, F., Uchida, S., Yamagata, H., Abe-Higuchi, N., Hobara, T., Hara, K., et al. (2016). Hippocampal MicroRNA-124 Enhances Chronic Stress Resilience in Mice. J. Neurosci. 36, 7253–7267. doi: 10.1523/JNEUROSCI.0319-16.2016
- Hobara, T., Uchida, S., Otsuki, K., Matsubara, T., Funato, H., Matsuo, K., et al. (2010). Altered gene expression of histone deacetylases in mood disorder patients. J. Psychiatr. Res. 44, 263–270. doi: 10.1016/j.jpsychires.2009.08.015
- Hu, H. (2019). Advances in Molecular and Circuitry Mechanisms of Depressive Disorder-A Focus on Lateral Habenula. *Adv. Exp. Med. Biol.* 1180, 135–146. doi: 10.1007/978-981-32-9271-0_7
- Hu, H., Cui, Y., and Yang, Y. (2020). Circuits and functions of the lateral habenula in health and in disease. *Nat. Rev. Neurosci.* 21, 277–295. doi: 10.1038/s41583-020-0292-4
- Huganir, R. L., and Nicoll, R. A. (2013). AMPARs and synaptic plasticity: the last 25 years. *Neuron* 80, 704–717. doi: 10.1016/j.neuron.2013.10.025
- Hyde, C. L., Nagle, M. W., Tian, C., Chen, X., Paciga, S. A., Wendland, J. R., et al. (2016). Identification of 15 genetic loci associated with risk of major depression in individuals of European descent. *Nat. Genet.* 48, 1031–1036. doi: 10.1038/ng.3623
- Iga, J., Ueno, S., Yamauchi, K., Numata, S., Kinouchi, S., Tayoshi-Shibuya, S., et al. (2007). Altered HDAC5 and CREB mRNA expressions in the peripheral leukocytes of major depression. *Prog. Neuropsychopharmacol. Biol. Psychiatry* 31, 628–632. doi: 10.1016/j.pnpbp.2006.12.014
- Kang, H. J., Voleti, B., Hajszan, T., Rajkowska, G., Stockmeier, C. A., Licznerski, P., et al. (2012). Decreased expression of synapse-related genes and loss of synapses in major depressive disorder. *Nat. Med.* 18, 1413–1417. doi: 10.1038/nm.2886
- Kavalali, E. T., and Monteggia, L. M. (2020). Targeting Homeostatic Synaptic Plasticity for Treatment of Mood Disorders. *Neuron* 106, 715–726. doi: 10.1016/ j.neuron.2020.05.015
- Kawatake-Kuno, A., Murai, T., and Uchida, S. (2021). The Molecular Basis of Depression: Implications of Sex-Related Differences in Epigenetic Regulation. *Front. Mol. Neurosci.* 14:708004. doi: 10.3389/fnmol.2021.708004

- Kim, J. W., Autry, A. E., Na, E. S., Adachi, M., Bjorkholm, C., Kavalali, E. T., et al. (2021). Sustained effects of rapidly acting antidepressants require BDNF-dependent MeCP2 phosphorylation. *Nat. Neurosci.* 2021:8. doi: 10.1038/ s41593-021-00868-8
- Koester, H. J., and Sakmann, B. (1998). Calcium dynamics in single spines during coincident pre- and postsynaptic activity depend on relative timing of back-propagating action potentials and subthreshold excitatory postsynaptic potentials. *Proc. Natl. Acad. Sci. U S A* 95, 9596–9601. doi: 10.1073/pnas.95.16. 9596
- Koike, H., Iijima, M., and Chaki, S. (2011). Involvement of AMPA receptor in both the rapid and sustained antidepressant-like effects of ketamine in animal models of depression. *Behav. Brain Res.* 224, 107–111. doi: 10.1016/j.bbr.2011.05.035
- Krishnan, V., and Nestler, E. J. (2008). The molecular neurobiology of depression. *Nature* 455, 894–902. doi: 10.1038/nature07455
- Krystal, J. H., Abdallah, C. G., Sanacora, G., Charney, D. S., and Duman, R. S. (2019). Ketamine: A Paradigm Shift for Depression Research and Treatment. *Neuron* 101, 774–778. doi: 10.1016/j.neuron.2019.02.005
- Krystal, J. H., Sanacora, G., and Duman, R. S. (2013). Rapid-acting glutamatergic antidepressants: the path to ketamine and beyond. *Biol. Psychiatry* 73, 1133– 1141. doi: 10.1016/j.biopsych.2013.03.026
- Lahti, A. C., Koffel, B., Laporte, D., and Tamminga, C. A. (1995). Subanesthetic doses of ketamine stimulate psychosis in schizophrenia. *Neuropsychopharmacology* 13, 9–19. doi: 10.1016/0893-133X(94)00131-I
- Lepack, A. E., Fuchikami, M., Dwyer, J. M., Banasr, M., and Duman, R. S. (2014). BDNF release is required for the behavioral actions of ketamine. *Int. J. Neuropsychopharmacol.* 18:33. doi: 10.1093/ijnp/pyu033
- Li, K., Zhou, T., Liao, L., Yang, Z., Wong, C., Henn, F., et al. (2013). betaCaMKII in lateral habenula mediates core symptoms of depression. *Science* 341, 1016– 1020. doi: 10.1126/science.1240729
- Li, N., Lee, B., Liu, R. J., Banasr, M., Dwyer, J. M., Iwata, M., et al. (2010). mTOR-dependent synapse formation underlies the rapid antidepressant effects of NMDA antagonists. *Science* 329, 959–964. doi: 10.1126/science.1190287
- Li, N., Liu, R. J., Dwyer, J. M., Banasr, M., Lee, B., Son, H., et al. (2011). Glutamate N-methyl-D-aspartate receptor antagonists rapidly reverse behavioral and synaptic deficits caused by chronic stress exposure. *Biol. Psychiatry* 69, 754–761. doi: 10.1016/j.biopsych.2010.12.015
- Liu, F., Paule, M. G., Ali, S., and Wang, C. (2011). Ketamine-induced neurotoxicity and changes in gene expression in the developing rat brain. *Curr. Neuropharmacol.* 9, 256–261. doi: 10.2174/157015911795017155
- Liu, R. J., Fuchikami, M., Dwyer, J. M., Lepack, A. E., Duman, R. S., and Aghajanian, G. K. (2013). GSK-3 inhibition potentiates the synaptogenic and antidepressant-like effects of subthreshold doses of ketamine. *Neuropsychopharmacology* 38, 2268–2277. doi: 10.1038/npp.2013.128
- Lumsden, E. W., Troppoli, T. A., Myers, S. J., Zanos, P., Aracava, Y., Kehr, J., et al. (2019). Antidepressant-relevant concentrations of the ketamine metabolite (2R,6R)-hydroxynorketamine do not block NMDA receptor function. *Proc. Natl. Acad. Sci. U S A* 116, 5160–5169. doi: 10.1073/pnas.181607 1116
- Macqueen, G., and Frodl, T. (2011). The hippocampus in major depression: evidence for the convergence of the bench and bedside in psychiatric research? *Mol. Psychiatry* 16, 252–264. doi: 10.1038/mp.2010.80
- Mcewen, B. S. (2007). Physiology and neurobiology of stress and adaptation: central role of the brain. *Physiol. Rev.* 87, 873–904. doi: 10.1152/physrev.00041.2006
- Mckinsey, T. A., Zhang, C. L., Lu, J., and Olson, E. N. (2000). Signal-dependent nuclear export of a histone deacetylase regulates muscle differentiation. *Nature* 408, 106–111. doi: 10.1038/35040593
- Minichiello, L. (2009). TrkB signalling pathways in LTP and learning. Nat. Rev. Neurosci. 10, 850–860. doi: 10.1038/nrn2738
- Misztak, P., Panczyszyn-Trzewik, P., Nowak, G., and Sowa-Kucma, M. (2020). Epigenetic marks and their relationship with BDNF in the brain of suicide victims. *PLoS One* 15:e0239335. doi: 10.1371/journal.pone.0239335
- Moda-Sava, R. N., Murdock, M. H., Parekh, P. K., Fetcho, R. N., Huang, B. S., Huynh, T. N., et al. (2019). Sustained rescue of prefrontal circuit dysfunction by antidepressant-induced spine formation. *Science* 364:8078. doi: 10.1126/ science.aat8078
- Murrough, J. W., Perez, A. M., Pillemer, S., Stern, J., Parides, M. K., Aan Het, et al. (2013). Rapid and longer-term antidepressant effects of repeated ketamine

infusions in treatment-resistant major depression. *Biol. Psychiatry* 74, 250–256. doi: 10.1016/j.biopsych.2012.06.022

- Nestler, E. J., Pena, C. J., Kundakovic, M., Mitchell, A., and Akbarian, S. (2016). Epigenetic Basis of Mental Illness. *Neuroscientist* 22, 447–463. doi: 10.1177/ 1073858415608147
- Ngo-Anh, T. J., Bloodgood, B. L., Lin, M., Sabatini, B. L., Maylie, J., and Adelman, J. P. (2005). SK channels and NMDA receptors form a Ca2+-mediated feedback loop in dendritic spines. *Nat. Neurosci.* 8, 642–649. doi: 10.1038/nn1449
- Nie, X., Kitaoka, S., Tanaka, K., Segi-Nishida, E., Imoto, Y., Ogawa, A., et al. (2018). The Innate Immune Receptors TLR2/4 Mediate Repeated Social Defeat Stress-Induced Social Avoidance through Prefrontal Microglial Activation. *Neuron* 99, 464–479e467. doi: 10.1016/j.neuron.2018.06.035
- Nosyreva, E., Autry, A. E., Kavalali, E. T., and Monteggia, L. M. (2014). Age dependence of the rapid antidepressant and synaptic effects of acute NMDA receptor blockade. *Front. Mol. Neurosci.* 7:94. doi: 10.3389/fnmol.2014.00094
- Nosyreva, E., Szabla, K., Autry, A. E., Ryazanov, A. G., Monteggia, L. M., and Kavalali, E. T. (2013). Acute suppression of spontaneous neurotransmission drives synaptic potentiation. *J. Neurosci.* 33, 6990–7002. doi: 10.1523/ JNEUROSCI.4998-12.2013
- Phillips, J. L., Norris, S., Talbot, J., Birmingham, M., Hatchard, T., Ortiz, A., et al. (2019). Single, Repeated, and Maintenance Ketamine Infusions for Treatment-Resistant Depression: A Randomized Controlled Trial. Am. J. Psychiatry 176, 401–409. doi: 10.1176/appi.ajp.2018.18070834
- Popoli, M., Yan, Z., Mcewen, B. S., and Sanacora, G. (2011). The stressed synapse: the impact of stress and glucocorticoids on glutamate transmission. *Nat. Rev. Neurosci.* 13, 22–37. doi: 10.1038/nrn3138
- Price, J. L., and Drevets, W. C. (2010). Neurocircuitry of mood disorders. Neuropsychopharmacology 35, 192–216. doi: 10.1038/npp.2009.104
- Reid, C. A., Fabian-Fine, R., and Fine, A. (2001). Postsynaptic calcium transients evoked by activation of individual hippocampal mossy fiber synapses. *J. Neurosci.* 21, 2206–2214. doi: 10.1523/JNEUROSCI.21-07-02206.2001
- Rodriguez-Munoz, M., Sanchez-Blazquez, P., Callado, L. F., Meana, J. J., and Garzon-Nino, J. (2017). Schizophrenia and depression, two poles of endocannabinoid system deregulation. *Transl. Psychiatry* 7:1291. doi: 10.1038/ s41398-017-0029-y
- Sakai, Y., Li, H., Inaba, H., Funayama, Y., Ishimori, E., Kawatake-Kuno, A., et al. (2021). Gene-environment interactions mediate stress susceptibility and resilience through the CaMKIIbeta/TARPgamma-8/AMPAR pathway. *iScience* 24:102504. doi: 10.1016/j.isci.2021.102504
- Scheetz, A. J., Nairn, A. C., and Constantine-Paton, M. (2000). NMDA receptormediated control of protein synthesis at developing synapses. *Nat. Neurosci.* 3, 211–216. doi: 10.1038/72915
- Seney, M. L., Huo, Z., Cahill, K., French, L., Puralewski, R., Zhang, J., et al. (2018). Opposite Molecular Signatures of Depression in Men and Women. *Biol. Psychiatry* 84, 18–27. doi: 10.1016/j.biopsych.2018.01.017
- Shinohara, R., Aghajanian, G. K., and Abdallah, C. G. (2021). Neurobiology of the Rapid-Acting Antidepressant Effects of Ketamine: Impact and Opportunities. *Biol. Psychiatry* 90, 85–95. doi: 10.1016/j.biopsych.2020.12.006
- Sial, O. K., Parise, E. M., Parise, L. F., Gnecco, T., and Bolanos-Guzman, C. A. (2020). Ketamine: The final frontier or another depressing end? *Behav. Brain Res.* 383:112508. doi: 10.1016/j.bbr.2020.112508
- Sun, H. L., Zhou, Z. Q., Zhang, G. F., Yang, C., Wang, X. M., Shen, J. C., et al. (2016). Role of hippocampal p11 in the sustained antidepressant effect of ketamine in the chronic unpredictable mild stress model. *Transl. Psychiatry* 6:e741. doi: 10.1038/tp.2016.21
- Suzuki, K., and Monteggia, L. M. (2020). The role of eEF2 kinase in the rapid antidepressant actions of ketamine. *Adv. Pharmacol.* 89, 79–99. doi: 10.1016/ bs.apha.2020.04.005
- Suzuki, K., Nosyreva, E., Hunt, K. W., Kavalali, E. T., and Monteggia, L. M. (2017). Effects of a ketamine metabolite on synaptic NMDAR function. *Nature* 546, E1–E3. doi: 10.1038/nature22084
- Takemoto-Kimura, S., Suzuki, K., Horigane, S. I., Kamijo, S., Inoue, M., Sakamoto, M., et al. (2017). Calmodulin kinases: essential regulators in health and disease. *J. Neurochem.* 141, 808–818. doi: 10.1111/jnc.14020
- Thompson, S. M., Kallarackal, A. J., Kvarta, M. D., Van Dyke, A. M., Legates, T. A., and Cai, X. (2015). An excitatory synapse hypothesis of depression. *Trends Neurosci.* 38, 279–294. doi: 10.1016/j.tins.2015.03.003

- Tochigi, M., Iwamoto, K., Bundo, M., Sasaki, T., Kato, N., and Kato, T. (2008). Gene expression profiling of major depression and suicide in the prefrontal cortex of postmortem brains. *Neurosci. Res.* 60, 184–191. doi: 10.1016/j.neures.2007.10. 010
- Trivedi, M. H., Rush, A. J., Wisniewski, S. R., Nierenberg, A. A., Warden, D., Ritz, L., et al. (2006). Evaluation of outcomes with citalopram for depression using measurement-based care in STAR*D: implications for clinical practice. Am. J. Psychiatry 163, 28–40. doi: 10.1176/appi.ajp.163.1.28
- Tsankova, N. M., Berton, O., Renthal, W., Kumar, A., Neve, R. L., and Nestler, E. J. (2006). Sustained hippocampal chromatin regulation in a mouse model of depression and antidepressant action. *Nat. Neurosci.* 9, 519–525. doi: 10.1038/ nn1659
- Turrigiano, G. G., Leslie, K. R., Desai, N. S., Rutherford, L. C., and Nelson, S. B. (1998). Activity-dependent scaling of quantal amplitude in neocortical neurons. *Nature* 391, 892–896. doi: 10.1038/36103
- Uchida, S., Hara, K., Kobayashi, A., Otsuki, K., Yamagata, H., Hobara, T., et al. (2011). Epigenetic status of Gdnf in the ventral striatum determines susceptibility and adaptation to daily stressful events. *Neuron* 69, 359–372. doi: 10.1016/j.neuron.2010.12.023
- Uchida, S., and Shumyatsky, G. P. (2018a). Epigenetic regulation of Fgf1 transcription by CRTC1 and memory enhancement. *Brain Res. Bull.* 141, 3–12. doi: 10.1016/j.brainresbull.2018.02.016
- Uchida, S., and Shumyatsky, G. P. (2018b). Synaptically Localized Transcriptional Regulators in Memory Formation. *Neuroscience* 370, 4–13. doi: 10.1016/j. neuroscience.2017.07.023
- Uchida, S., Yamagata, H., Seki, T., and Watanabe, Y. (2018). Epigenetic mechanisms of major depression: Targeting neuronal plasticity. *Psychiatry Clin. Neurosci.* 72, 212–227. doi: 10.1111/pcn.12621
- Van Den Heuvel, M. P., Scholtens, L. H., and Kahn, R. S. (2019). Multiscale Neuroscience of Psychiatric Disorders. *Biol. Psychiatry* 86, 512–522. doi: 10. 1016/j.biopsych.2019.05.015
- Wei, Y., Chang, L., and Hashimoto, K. (2021). Molecular mechanisms underlying the antidepressant actions of arketamine: beyond the NMDA receptor. *Mol. Psychiatry* 2021:1. doi: 10.1038/s41380-021-01121-1
- Xia, C. Y., He, J., Du, L. D., Yan, Y., Lian, W. W., Xu, J. K., et al. (2021). Targeting the dysfunction of glutamate receptors for the development of novel antidepressants. *Pharmacol. Ther.* 226:107875. doi: 10.1016/j.pharmthera.2021. 107875
- Yamaguchi, J. I., Toki, H., Qu, Y., Yang, C., Koike, H., Hashimoto, K., et al. (2018). (2R,6R)-Hydroxynorketamine is not essential for the antidepressant actions of (R)-ketamine in mice. *Neuropsychopharmacology* 43, 1900–1907. doi: 10.1038/ s41386-018-0084-y
- Yamasaki, S., Aso, T., Miyata, J., Sugihara, G., Hazama, M., Nemoto, K., et al. (2020). Early and late effects of electroconvulsive therapy associated with different temporal lobe structures. *Transl. Psychiatry* 10:344. doi: 10.1038/ s41398-020-01025-8
- Yang, C., Ren, Q., Qu, Y., Zhang, J. C., Ma, M., Dong, C., et al. (2018). Mechanistic Target of Rapamycin-Independent Antidepressant Effects of (R)-Ketamine in a Social Defeat Stress Model. *Biol. Psychiatry* 83, 18–28. doi: 10.1016/j.biopsych. 2017.05.016

- Yang, Y., Cui, Y., Sang, K., Dong, Y., Ni, Z., Ma, S., et al. (2018). Ketamine blocks bursting in the lateral habenula to rapidly relieve depression. *Nature* 554, 317–322. doi: 10.1038/nature25509
- Yokoyama, R., Higuchi, M., Tanabe, W., Tsukada, S., Naito, M., Yamaguchi, T., et al. (2020). (S)-norketamine and (2S,6S)-hydroxynorketamine exert potent antidepressant-like effects in a chronic corticosterone-induced mouse model of depression. *Pharmacol. Biochem. Behav.* 191:172876. doi: 10.1016/j.pbb.2020.17 2876
- Youssef, M. M., Underwood, M. D., Huang, Y. Y., Hsiung, S. C., Liu, Y., Simpson, N. R., et al. (2018). Association of BDNF Val66Met Polymorphism and Brain BDNF Levels with Major Depression and Suicide. *Int. J. Neuropsychopharmacol.* 21, 528–538. doi: 10.1093/ijnp/pyy008
- Yuen, E. Y., Wei, J., Liu, W., Zhong, P., Li, X., and Yan, Z. (2012). Repeated stress causes cognitive impairment by suppressing glutamate receptor expression and function in prefrontal cortex. *Neuron* 73, 962–977. doi: 10.1016/j.neuron.2011. 12.033
- Zanos, P., Moaddel, R., Morris, P. J., Georgiou, P., Fischell, J., Elmer, G. I., et al. (2016). NMDAR inhibition-independent antidepressant actions of ketamine metabolites. *Nature* 533, 481–486. doi: 10.1038/nature1 7998
- Zarate, C. A. Jr., Singh, J. B., Carlson, P. J., Brutsche, N. E., Ameli, R., Luckenbaugh, D. A., et al. (2006). A randomized trial of an N-methyl-D-aspartate antagonist in treatment-resistant major depression. *Arch. Gen. Psychiatry* 63, 856–864. doi: 10.1001/archpsyc.63.8.856
- Zhou, W., Wang, N., Yang, C., Li, X. M., Zhou, Z. Q., and Yang, J. J. (2014). Ketamine-induced antidepressant effects are associated with AMPA receptors-mediated upregulation of mTOR and BDNF in rat hippocampus and prefrontal cortex. *Eur. Psychiatry* 29, 419–423. doi: 10.1016/j.eurpsy.2013. 10.005
- Zhou, Z., Hong, E. J., Cohen, S., Zhao, W. N., Ho, H. Y., Schmidt, L., et al. (2006). Brain-specific phosphorylation of MeCP2 regulates activity-dependent Bdnf transcription, dendritic growth, and spine maturation. *Neuron* 52, 255–269. doi: 10.1016/j.neuron.2006.09.037

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