



Data Article

Generic mitigating and promoting effect of zeolite on anaerobic digestion: Physicochemical and metataxonomic data

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ABSTRACT

This article provides comprehensive data on degradation performance and microbial dynamics derived from a set of 24 lab-scale batch anaerobic digesters involving various types of inhibitors and the addition of zeolite as a support material. In the first series of 12 digesters, three inhibitors were investigated at the following concentrations: 20 g/L of sodium chloride, 400 mg/L of erythromycin, and 5 mg/L of S-metolachlor. Each inhibitor was tested in triplicate, along with a control condition without inhibition. A parallel series was set up identically, except that 15 g/L of zeolite was introduced into each digester to mitigate the inhibition and promote the degradation process. The provided data comprises information regarding the experimental setup, monitoring measurements that assess the degradation performance (production, composition, and apparent isotopic factor of biogas, pH, dissolved inorganic and organic carbon and volatile fatty acids concentrations), microbial samples information, and 16S rRNA gene sequencing data that decipher changes in microbial structure. This datapaper is associated with research article [1] and presents both the sequencing data and the associated physicochemical data in a structured table format. The sequencing data were generated using the Ion Torrent PGM sequencer and have been deposited in the

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European Nucleotide Archive (ENA) database at EMBL-EBI under accession number PRJEB65129 (<https://www.ebi.ac.uk/ena/browser/view/PRJEB65129>), with sample accession numbers ranging from ERS16257742 to ERS16257691 [2]. The data serves as a valuable resource for comparisons with data from other studies on lab-scale batch anaerobic digesters, particularly those utilizing zeolite as a support material or involving inhibition caused by similar types of inhibitors (salts, antibiotics, or pesticides).

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Specifications Table

Subject	Environmental Genomics and Metagenomics.
Specific subject area	Microbial ecology of anaerobic digestion.
Type of data	Table, Figure, Raw sequencing data
Data collection	Gas pressure in the digesters was measured using a differential manometer (Digitron 2082P, Margam, UK) to calculate gas production. Gas composition was analyzed by micro gas chromatography (CP4900, Varian, Palo Alto, USA). Isotopic fractionation of methane and carbon dioxide ($\delta^{13}\text{CH}_4$ and $\delta^{13}\text{CO}_2$) was measured using a Trace Gas Chromatograph Ultra coupled to an isotope ratio mass spectrometer Delta V Plus through a combustion machine GC III (Thermo Scientific) to calculate the apparent isotopic factor (α_{app}) following the description in [3]. Dissolved inorganic and organic carbon (DIC and DOC) were measured using a TOC-L CPN analyzer (Shimadzu) following the French standard NF EN 1484. Volatile fatty acids (VFAs) were measured using ionic chromatography (ICS 5000+, Thermo Fisher Scientific) equipped with an IonPac ICE-AS1 column (9 mm 250 mm) as described in [3]. pH of the liquid samples was measured with a pH meter (HANNA). DNA sequencing was performed on Ion Torrent Personal Genome Machine according to the manufacturer's instructions and following the procedure described in [4].
Data source location	INRAE, Antony, France.
Data accessibility	Data are available with the article. The sequencing data have been deposited in ENA at EMBL-EBI under accession number PRJEB65129 (https://www.ebi.ac.uk/ena/browser/view/PRJEB65129) with sample accession numbers ranging from ERS16257742 to ERS16257691. Repository name: ENA Data identification number: PRJEB65129 Direct URL to data: https://www.ebi.ac.uk/ena/browser/view/PRJEB65129
Related research article	Wang, X., Dürr, V., Guenne, A., Mazéas, L., Chapleur, O. 2024. Generic role of zeolite in enhancing anaerobic digestion and mitigating diverse inhibitions: Insights from degradation performance and microbial characteristics. <i>Journal of Environmental Management</i> , 356, 120,676. 10.1016/j.jenvman.2024.120676

Value of the Data

- The presented data provide connections between anaerobic digester performance (biogas production, dissolved inorganic/organic carbon and volatile fatty acids accumulation, methanogenesis pathway), inhibition types (salts, pesticides, antibiotics), zeolite addition as a support material, and microbial community structure at different time points. All data can serve as a valuable resource for comparative analysis with other studies on lab-scale batch anaerobic digesters utilizing zeolite or investigating similar inhibitors. The 16S rRNA sequencing data are particularly useful for identifying microbial characteristics influenced by zeolite

and different types of inhibition. Access to the sequencing data and associated metadata allows researchers to conduct new studies. The data can also be helpful for exploring relationships between zeolite and specific microbial functions.

- We present comprehensive information on anaerobic digestion performance and microbial structure characteristics in the presence of erythromycin and S-metolachlor (a typical antibiotic and pesticide, respectively), the influence of which has been limited studied but is of increasing interest in anaerobic digestion. The use of zeolite to mitigate their inhibition is also a novel research focus.
- All 8 tested conditions were designed in triplicate in the same experimental system, with identical inoculum and substrate, at a constant mesophilic temperature. The selected samples for sequencing were highly representative to reflect microbial structure changes in the different conditions

This data can be related to previous data from our group on AD inhibition by different inhibitors and mitigation using zeolite or other support materials, conducted under similar experimental conditions and measuring similar parameters [5–7]. Our previous studies investigated the influence of different concentrations of ammonia and phenol, the impact of various support media on the inhibition, and the effects of different co-inhibitors on AD. This paper provides additional knowledge by specifically analyzing the effect of zeolite on a wide range of inhibitors.

1. Background

Zeolite is an aluminosilicate mineral with favorable physicochemical properties. It has been demonstrated to mitigate the inhibition in anaerobic digestion caused by several inhibitors, such as long-chain fatty acids, ammonia, and phenolic compounds [8–10]. We verified the broad applicability of zeolite's mitigating effect on other types of inhibitors found in anaerobic digestion, including sodium chloride, erythromycin, and S-metolachlor, from aspects of anaerobic digestion performance and microbial structure changes. The data article adds value to the related research article by providing detailed raw degradation performance data and sequencing data, which enhances data transparency, reproducibility, and accessibility while allowing other researchers to treat the data as a reference.

2. Data Description

The experimental design of this study is depicted in Fig. 1. A total of 24 batch anaerobic digesters were set up and divided into two series. Within each series, the 3 inhibitors were examined in triplicate alongside a control condition without inhibitors. One of the series included the addition of zeolite to evaluate its potential in mitigating the inhibition and promoting the anaerobic digestion process. The nomenclature of the digesters, the type of inhibitor added, the presence of zeolite, the number of replicates, and the selected dates for 16S rRNA sequencing and associated sample names are presented in Table 1. Tables 2–4 present the cumulative total biogas, methane, and carbon dioxide production data over time for each digester, while Fig. 2 provides a graphical representation of the production curves using the mean values of the triplicates for each condition. Table 5 displays the apparent isotope factor (α_{app}) of the biogas over time calculated from the isotopic fractionation of methane and carbon dioxide ($\delta^{13}CH_4$ and $\delta^{13}CO_2$), serving as an indicator of the methanogenic pathways. This data is also illustrated in Fig. 3 using the mean values of the triplicates for each condition. As noted by the horizontal lines in the figure, α_{app} greater than 1.065 implies the hydrogenotrophic pathway as the dominant methanogenesis pathway, while α_{app} less than 1.055 indicates acetoclastic methanogenesis as the prominent pathway [11]. Tables 6 and 7 depict the concentration of dissolved inorganic and organic carbon (DIC and DOC) over time in each digester, with Fig. 4 presenting these datasets using the mean values of the triplicates. The pH values are recorded in Table 8 and illustrated

Table 1
Experimental setup information and dates and sample names used for 16S rRNA gene sequencing.

Name of digester	Type of inhibitor	Zeolite addition	Replicate	Dates used for 16S rRNA sequencing and associated sample names						
				Day 0	Day 6	Day 13	Day 20	Day 34	Day 41	Day 55
O_a	No inhibitor	No	a				O_a_D20	O_a_D34		
O_b	No inhibitor	No	b				O_b_D20	O_b_D34		
O_c	No inhibitor	No	c				O_c_D20	O_c_D34		
Na_a	Sodium chloride	No	a				Na_a_D20			Na_a_D55
Na_b	Sodium chloride	No	b	Inoculum_3			Na_b_D20			Na_b_D55
Na_c	Sodium chloride	No	c				Na_c_D20			Na_c_D55
ERY_a	Erythromycin	No	a				E_a_D20			E_a_D55
ERY_b	Erythromycin	No	b				E_b_D20			E_b_D55
ERY_c	Erythromycin	No	c				E_c_D20	E_c_D34		
MET_a	S-metolachlor	No	a	Inoculum_4			M_a_D20	M_a_D34		
MET_b	S-metolachlor	No	b				M_b_D20	M_b_D34		
MET_c	S-metolachlor	No	c				M_c_D20		M_c_D41	
Oz_a	No inhibitor	Yes	a		Oz_a_D6		Oz_a_D20			
Oz_b	No inhibitor	Yes	b		Oz_b_D6		Oz_b_D20			
Oz_c	No inhibitor	Yes	c	Inoculum_1	Oz_c_D6		Oz_c_D20			
Naz_a	Sodium chloride	Yes	a			Naz_a_D13	Naz_a_D20			
Naz_b	Sodium chloride	Yes	b	Inoculum_2		Naz_b_D13	Naz_b_D20			
Naz_c	Sodium chloride	Yes	c			Naz_c_D13	Naz_c_D20			
ERYz_a	Erythromycin	Yes	a		Ez_a_D6		Ez_a_D20			
ERYz_b	Erythromycin	Yes	b		Ez_b_D6		Ez_b_D20			
ERYz_c	Erythromycin	Yes	c		Ez_c_D6		Ez_c_D20			
METz_a	S-metolachlor	Yes	a		Mz_a_D6		Mz_a_D20			
METz_b	S-metolachlor	Yes	b		Mz_b_D6		Mz_b_D20			
METz_c	S-metolachlor	Yes	c		Mz_c_D6		Mz_c_D20			

Table 3 Cumulative methane production in each digester (mg of C).

Table with 36 rows (Day 0 to Day 36) and 36 columns (0_a to 0_z). Each cell contains a numerical value representing cumulative methane production in mg of C.

Table 4

Cumulative carbon dioxide production in each digester (mg of C).

Day	O_a	O_b	O_c	Na_a	Na_b	Na_c	Ery_a	Ery_b	Ery_c	Met_a	Met_b	Met_c	Oz_a	Oz_b	Oz_c	Naz_a	Naz_b	Naz_c	Eryz_a	Eryz_b	Eryz_c	Metz_a	Metz_b	Metz_c
0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
1	8.188	29.54	2.703	14.058	10.963	11.029	14.17	11.49	0.977	8.534	9.501	35.321	13.25	42.144	4.951	19.176	17.216	22.86	15.462	29.975	NA	13.024	12.802	12.003
4	12.403	105.968	8.54	41.054	29.972	79.49	52.938	100.113	9.471	43.779	39.608	49.268	66.587	127.839	13.195	62.866	52.409	44.103	66.534	98.905	16.016	64.861	55.445	61.638
5	22.931	118.197	NA	50.058	35.993	NA	NA	118.6 NA	47.053	40.028 NA	NA	NA	NA	154.667 NA	69.822	59.766	45.497	72.986	125.987	NA	81.733	64.526	65.246	
6	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	181.014	9.127 NA	NA	NA	NA	88.974	149.713	8.753 NA	NA	NA	68.37
7	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	69.89	217.867 NA	86.994	64.003	47.433	126.333	163.16	87.53	110.5	86.922	88.659	
8	71.411	120.951	66.177	72.362	92.946	103.251	95.054	132.849	79.573	99.12	99.382	104.559	164.974	240.575	98.723	138.265	129.803	121.919	146.038	179.36	111.395	153.879	138.889	149.002
11	103.017	148.27	99.795	99.829	101.61	114.067	99.942	143.255	85.465	125.41	109.785	108.291	183.157	261.094	140.365	173.074	186.041	152.254	174.53	219.055	152.388	189.632	168.327	183.618
12	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	222.673	294.277	175.657	194.943	191.323	174.403	209.575	244.257	184.223	220.762	197.972	219.943
13	123.352	165.106	114.426	NA	NA	NA	NA	NA	115.973	139.627 NA	NA	NA	251.154	321.06	206.015	212.757	211.746	194.996	235.437	277.894	212.86	244.485	221.052	244.725
14	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	278.883	347.856	231.961	230.661	227.51	211.436	260.669	306.198	241.129	267.683	244.401	271.403
15	133.102	181.357	128.178	125.696	124.024 NA	124.66	161.203	151.771	149.232	140.288	137.809	299.469	372.744	255.898	235.974	244.733	228.741	284.376	330.161	267.977	290.008	268.414	292.965	
18	161.02	206.208	151.222	137.537	133.701 NA	136.859	171.695	174.919	176.817	160.063	150.344	316.309	397.129	281.479	276.786	279.145	261.756	317.398	357.661	301.648	313.945	300.503	312.589	
19	176.037	219.418	162.133 NA	NA	NA	NA	NA	188.571	193.608	170.466 NA	NA	NA	323.59	411.146	298.697	305.974	309.03	291.972	338.208	375.698	323.303	329.087	321.636	325.599
20	191.757	238.701	177.825 NA	NA	NA	NA	NA	206.458	211.746	185.342 NA	NA	NA	327.641	417.579	305.996	318.898	326.648	308.76	347.398	384.002	334.241	337.105	330.526	332.349
21	209.041	256.119	193.086 NA	NA	NA	NA	155.487	192.891	224.386	229.581	200.666	168.014 NA	NA	NA	329.589	340.12	323.533	354.515	390.77	342.987	342.729	340.296	336.873	
22	225.35	271.973	208.07	151.372	145.262	137.212	156.659 NA	241.004	247.044	217.703	174.711 NA	NA	424.558	314.64	336.274	348.909	333.558	362.327	396.326	349.165	348.457	346.645 NA	NA	
25	259.047	312.425	239.039	166.066	159.126	150.724	168.396	208.167	269.168	274.249	245.928	197.733	334.837	429.808	321.756	345.608	359.429	344.104	371.282	404.213	359.395	352.864	356.948	344.739
26	286.582	333.726	265.18 NA	NA	NA	NA	NA	291.52	297.073	267.975	209.365 NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
27	303.591	350.831	282.378	174	161.888	160.046	185.324	214.756	307.133	309.911	283.096	219.146 NA	NA	NA	NA	350.885	367.002	352.915	380.234	409.965	367.792 NA	NA	NA	NA
28	317.75	365.392	297.64 NA	NA	NA	NA	NA	319.978	321.761	299.919	233.878 NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
29	329.278	377.442	311.326	180.024	173.916	167.41	198.091	224.09	327.569	329.985	312.511	245.345	341.482	435.574	327.166	355.992	372.227	357.792	383.467	413.257	372.985	360.537	364.428	352.018
32	343.676	383.925	325.919	192.078	185.873	179.965	215.263	238.17	342.308	340.668	329.049	266.675 NA	NA	NA	NA	362.473	378.395	364.179	388.96	418.587	378.18 NA	NA	NA	NA
33	354.234	395.417	344.369	197.841	190.164	183.364	222.581	245.333	355.112	344.28	343.578	280.761	346.52	440.614	332.352 NA	NA	NA	NA	NA	NA	NA	366.475	371.142	358.052
34	358.499	400.231	350.863 NA	NA	NA	NA	233.719	249.383	361.033 NA	NA	349.835	297.929 NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
35	NA	NA	NA	NA	NA	NA	NA	NA	NA	353.491	352.932	312.953 NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
36	364.636	406.336	358.144	217.363	202.797	203.675	249.089	266.099	370.002 NA	NA	NA	324.125 NA	NA	NA	NA	370.624	384.909	372.188	NA	424.714 NA	NA	NA	NA	NA
39	372.436	413.58	364.939	234.239	217.069	219.833	270.764	284.551	376.633	362.691	362.722	343.347 NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
41	374.335	417.596	371.312	251.406	228.232	235.376	292.534	302.186	383.451 NA	NA	367.373	362.339 NA	NA	NA	NA	NA	NA	NA	399.155 NA	NA	NA	NA	NA	NA
42	NA	NA	NA	261.869	235.895	244.58	309.952	316.052	385.545	367.869 NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
43	NA	NA	375.278	276.374	247.197	258.576	322.827	332.163 NA	374.095	372.604	381.097 NA	NA	356.174	452.258	343.552	375.032	389.849	378.19	402.9	430.726	382.36	379.772	381.817	369.454
46	NA	NA	NA	303.362	265.646	282.617	336.671	351.538	393.864	374.095	372.604	381.097 NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
47	NA	NA	NA	317.022	280.291	299.141	349.199	368.989 NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
48	382.58	427.063	381.185	328.182	293.332	313.628	356.38	380.506	395.755 NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
49	NA	NA	NA	334.859	303.411	322.442	364.737	391.272 NA	377.167	378.106	386.885 NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
53	NA	NA	NA	349.348	326.608	336.971	375.329	405.072	405.47 NA	NA	395.858 NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	393.072 NA	NA	NA	NA
54	391.644	434.316	392.064	354.249	341.831	346.898	381.301	414.294 NA	385.66	385.86 NA	364.209	461.669	352.361 NA	NA	NA	NA	NA	NA	413.771 NA	NA	388.124	390.392	378.095	
55	NA	NA	NA	360.613	349.959	350.838	387.548	421.88	408.617 NA	NA	399.712 NA	NA	NA	NA	NA	383.208	397.06	386.473 NA	NA	NA	NA	NA	NA	NA
56	NA	NA	NA	361.005	355.268	355.469	391.567	429.596 NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
57	NA	NA	NA	364.955	359.082	357.616 NA	NA	NA	393.534	385.867 NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	415.439	439.885	397.743 NA	NA	NA
60	NA	NA	NA	368.94	367.41	363.537	400.515	438.272	417.412 NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
61	NA	NA	NA	370.531 NA	402.532	442.519 NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
62	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
63	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
64	NA	NA	NA	394.758	373.165	378.525	368.469	410.115	452.358	423.783 NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
67	402.029	444.579	403.773	379.252	378.666	371.781	413.8	459.609	428.064	400.074	398.968	413.326	373.608	473.307	361.945 NA	NA	NA	NA	NA	NA	404.304	400.241	387.686	
68	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
69	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
71	NA	NA	NA	NA	NA	NA	419.54	466.147	432.631 NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	424.444	449.24	408.584 NA	NA	NA	NA
74	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
75	NA	NA	NA	385.829	386.649	393.566 NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	389.665	403.126	391.901 NA	NA	NA	NA	NA	NA	NA
78	NA	NA	NA	391.341 NA	427.038	473.51	436.867 NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA

Table 5
Apparent isotopic factor (α_{app}) in each digester.

Day	O_a	O_b	O_c	Na_a	Na_b	Na_c	Ery_a	Ery_b	Ery_c	Met_a	Met_b	Met_c	Oz_a	Oz_b	Oz_c	Naz_a	Naz_b	Naz_c	Eryz_a	Eryz_b	Eryz_c	Metz_a	Metz_b	Metz_c
14	1.075	1.072	1.072	1.063	1.068	1.068	1.072	1.068	1.075	1.076	1.077	1.075	1.038	1.042	1.041	1.041	1.045	1.046	1.052	1.048	1.051	1.044	1.05	1.041
21	1.075	1.068	1.072	1.066	1.073	1.075	1.081	1.078	1.065	1.066	1.079	1.083	1.038	1.033	1.029	1.031	NA	1.034	1.032	1.029	1.033	1.033	1.03	1.034
28	1.048	1.045	1.054	1.071	1.079	1.079	1.087	1.084	1.047	1.036	1.055	1.08	1.04	1.038	1.037	1.029	1.024	1.024	1.027	1.027	1.028	1.038	1.034	1.039
35	1.022	1.023	1.017	1.066	1.077	1.071	1.073	1.085	1.03	1.026	1.018	1.061	1.043	1.042	1.041	1.036	1.031	1.032	1.029	1.033	1.03	1.041	1.038	1.042
42	1.035	1.035	1.034	1.055	1.066	1.059	1.053	1.066	1.028	1.037	1.032	1.019	1.043	1.044	1.041	1.039	1.035	1.035	1.037	1.039	1.04	1.041	1.038	1.042
49	1.037	1.04	1.038	1.043	1.054	1.045	1.04	1.046	1.027	1.039	1.038	1.023	1.046	1.042	1.043	1.039	1.038	1.036	1.039	1.04	1.041	1.044	1.04	1.045

Table 6
Dissolved inorganic carbon (DIC) concentration in each digester (mgC/L).

Day	0_a	0_b	0_c	Na_a	Na_b	Na_c	Ery_a	Ery_b	Ery_c	Met_a	Met_b	Met_c	Oz_a	Oz_b	Oz_c	Naz_a	Naz_b	Naz_c	Eryz_a	Eryz_b	Eryz_c	Metz_a	Metz_b	Metz_c
0	1659.2	1602.4	1513.6	1663.2	1690.8	1702.8	1700	1687.2	1702.4	1691.2	1680	1804	1674.4	1652.4	1686.4	1664.8	1678.4	1657.6	1674.4	1673.6	1682.4	1670.8	1724	1668.4
6	1306.8	1308.8	1312.8	1331.2	1329.6	1356.8	1388.4	1371.6	1353.2	1316.4	1314.4	1294.4	1362	1323.2	1320.8	1286.8	1306	1278.8	1324.4	1253.6	1360.8	1309.2	1294	1291.2
13	1294.8	1294.4	1301.6	1368.4	1380.4	1419.2	1383.2	1400.8	1495.6	1360.8	1337.2	1358.4	1624	1567.6	1568.4	1543.6	1503.6	1454.8	1447.2	1446.8	1384	1540	1358.8	1520.4
20	1223.6	1250	1242	1429.6	1407.6	1400.8	1368.8	1370.8	1352.8	1286.8	1255.2	1310.4	1787.2	1948.4	1784	1744.8	1746.4	1710	1691.2	1713.2	1568.4	1788.8	1726.4	1754.8
27	1255.2	1320.8	1220	1198.8	1190	1193.6	1122	1134.4	1360.8	1484.8	1152.4	1055.2	1714.8	1732	1754.4	2115.2	2374	1694.8	1972.8	1699.2	1717.6	1771.6	1828.4	1820.8
34	2055.6	1320	1220.4	1255.6	1233.2	1277.2	1187.6	1127.2	1710	1804.4	1670.8	1203.6	1812	1804	1906.8	1758.4	1725.2	1765.6	1662.4	1871.6	1955.6	1741.6	1754.8	1711.2
41	1686.4	1273.6	1289.6	1090	898.4	996.8	989.2	700.8	1513.6	1834.4	2014.8	1197.6	1340.8	1299.2	1496.4	1720.4	1660.4	1631.6	1495.6	1403.2	1257.6	1104	1398.4	1386.8
48	1448.8	1578.8	1863.2	1252	1085.2	1289.2	1242.4	1044.8	1585.2	1474	1237.2	1563.2	1268.4	1449.2	1600.8	1521.2	1424.4	1422.4	1241.6	1502	1101.6	1172.4	1143.6	1546.4

Table 7
Dissolved organic carbon (DOC) concentration in each digester (mgC/L).

Day	0_a	0_b	0_c	Na_a	Na_b	Na_c	Ery_a	Ery_b	Ery_c	Met_a	Met_b	Met_c	Oz_a	Oz_b	Oz_c	Naz_a	Naz_b	Naz_c	Eryz_a	Eryz_b	Eryz_c	Metz_a	Metz_b	Metz_c
0	989.2	994.8	947.2	1004.4	1035.6	1016	1266.4	1359.2	1006	1282.8	1030	1091.6	1010.4	1063.2	1022.4	1005.6	1022	1026.8	1273.6	1229.6	1244	1004.4	915.6	1007.6
6	1235.2	1224.4	1256.8	1150.4	1180.8	1159.2	1393.6	1450.8	1364.8	1223.2	1220.8	1250.8	1043.6	1088.8	1122.8	1144.4	1142.4	1157.6	1376.8	1313.6	1340.4	1142.8	1165.6	1030
13	1343.6	1302.8	1357.6	1078	1122.8	1116.8	1402	1468	1688.8	1280.4	1259.6	1294	518.8	606.4	678.8	750.4	809.2	861.6	1059.2	998.4	1059.6	702	818	526
20	1402.8	1304	1337.2	1044.4	1087.2	1074.4	1389.2	1442.4	1377.2	1348.4	1406.4	1262.4	71.2	148.8	153.6	346.4	412.8	430.4	621.2	490	578.8	155.2	274.4	95.2
27	1205.2	1055.6	1275.6	1224	1303.6	1326	1652	1620.8	1186	768.4	1365.6	1566.4	56.4	55.2	60	189.6	1527.2	222	446.8	327.2	399.6	57.6	56.4	56
34	236.4	238	1567.6	1187.6	1284	1272	1697.6	1726.4	706.8	175.2	221.6	1407.2	33.2	38	43.6	93.6	107.6	112	335.2	279.6	359.2	41.2	36.8	28.4
41	69.2	58.8	94	1134.4	1156	1310.4	1443.6	1263.2	446	81.6	93.6	318.4	26.8	20.4	25.2	68.8	64	67.6	218	175.6	198.4	19.6	28.8	22
48	57.2	59.6	82.8	878.4	1276.4	1097.2	918.4	1180.4	362	48	45.6	205.2	20	21.6	25.6	56.4	45.6	55.6	147.6	158.8	142	18.4	17.2	22.4

Table 8
pH values in each digester.

Day	0_a	0_b	0_c	Na_a	Na_b	Na_c	Ery_a	Ery_b	Ery_c	Met_a	Met_b	Met_c	Oz_a	Oz_b	Oz_c	Naz_a	Naz_b	Naz_c	Eryz_a	Eryz_b	Eryz_c	Metz_a	Metz_b	Metz_c
0	8.2	8.2	8.2	8.2	8.3	8.2	8	8.1	8	8.3	8.3	8.3	7.9	8.2	7.5	8.2	8.2	8	8	8.1	8.1	8.3	8.2	8.2
6	7.4	7.3	7.3	7.2	7.2	7.2	7.2	7.3	7.3	7.2	7.2	7.2	7.1	7.1	7.1	7	7	7	7.2	7.2	7.2	7.1	7.2	7.2
13	7.2	7.2	7.2	7.1	7.1	7.2	7.2	7.2	7.2	7.2	7.2	7.1	7.5	7.4	7.4	7.2	7.1	7.1	7.4	7.3	7.3	7.4	7.2	7.4
20	7	7.1	7.1	6.9	6.9	6.9	7.1	7.1	7.1	7.1	7	7.1	7.5	7.5	7.5	7.3	7.2	7.3	7.5	7.5	7.5	7.5	7.5	7.5
27	7.3	7.4	7.3	7	7	7	7.2	7.2	7.4	7.5	7.3	7.1	7.6	7.6	7.7	7.4	7.4	7.4	7.7	7.7	7.7	7.7	7.7	7.6
34	7.8	7.8	7.8	7	7	7.1	7.3	7.2	7.8	7.8	7.8	7.3	7.7	7.8	7.7	7.4	7.5	7.5	7.8	7.8	7.8	7.7	7.7	7.7
41	7.7	7.8	7.8	7.2	7.1	7.1	7.4	7.2	7.7	7.8	7.9	7.7	7.7	7.7	7.8	7.5	7.5	7.5	7.8	7.8	7.8	7.7	7.7	7.7
48	7.7	7.9	7.9	7.5	7.3	7.3	7.7	7.5	7.9	7.8	7.9	7.8	7.8	7.7	7.7	7.7	7.7	7.6	7.8	7.9	7.8	7.7	7.7	7.7

in Fig. 5 using the mean values of the triplicates. Tables 9–11, and 12 provide data on the accumulation of acetic acid, propionic acid, butyric acid, and total volatile fatty acids (VFAs) over time in each digester, and Fig. 6 visually represents these datasets with the mean values of the triplicates.

The sequencing data has been deposited in the form of fastq.gz files in the European Nucleotide Archive. These files contain 16S rRNA gene sequencing data generated using the Ion Torrent PGM platform. The sequencing data captures information from two time points for each digester. The first time point corresponds to the peak of VFAs accumulation, focusing on the hydrolysis and acidogenesis processes (referred to as early degradation stage in the related research article). The second time point corresponds to the peak of biogas production, aiming at

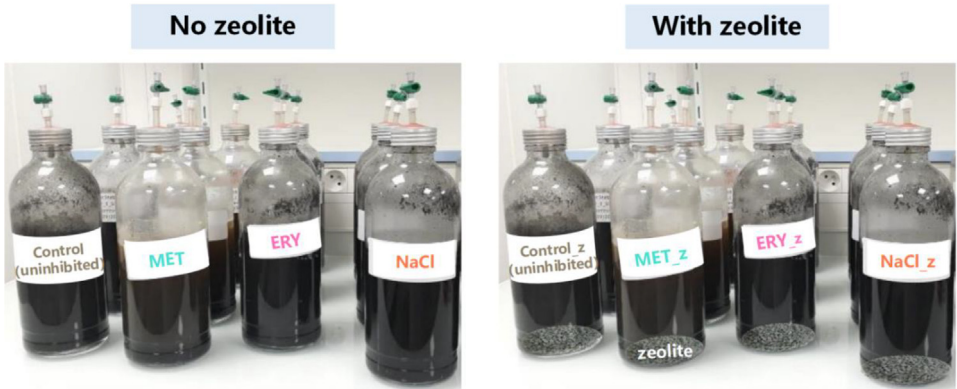


Fig. 1. Experimental design.

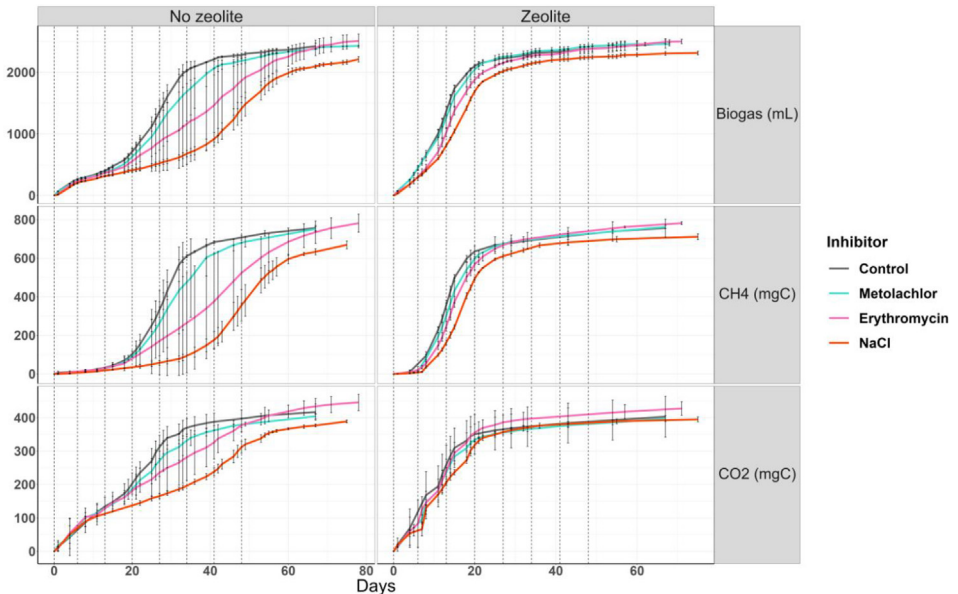


Fig. 2. Cumulative total biogas, CH₄, and CO₂ production (mg of C) over time (number of days) for different conditions. The data used are mean values of the triplicate digesters, and standard deviations are indicated with error bars. Vertical lines represent the sampling dates.

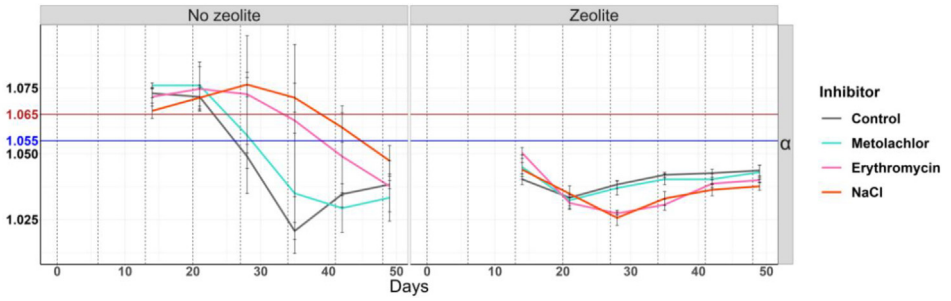


Fig. 3. Apparent isotope factor (α_{app}) of the biogas over time (number of days) for different conditions. α_{app} is an indicator of the methanogenic pathway. It is commonly assumed that α_{app} greater than 1.065 implies the hydrogenotrophic pathway as the dominant methanogenesis pathway, while α_{app} less than 1.055 indicates acetoclastic methanogenesis as the prominent pathway [6]. The horizontal red and blue lines denote the thresholds for hydrogenotrophic and acetoclastic methanogenesis, respectively. The data used are mean values of the triplicate digesters, and standard deviations are indicated with error bars. Vertical lines represent the sampling dates.

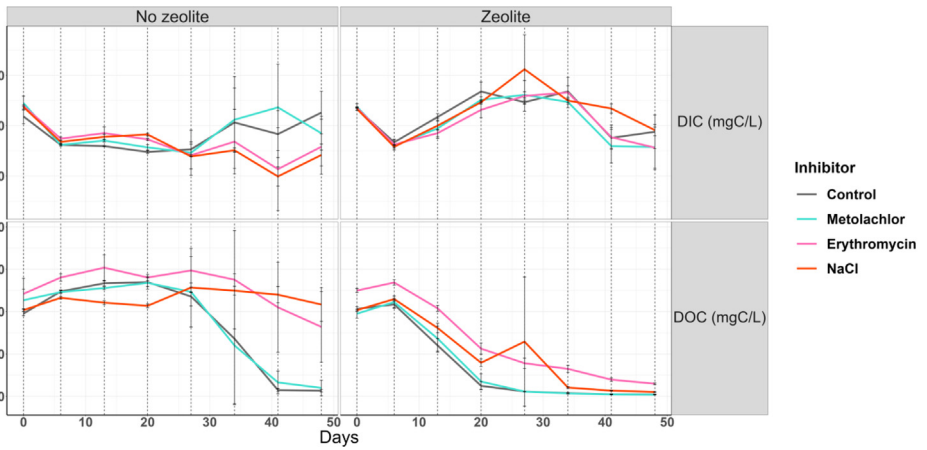


Fig. 4. Dissolved inorganic and organic carbon (DIC and DOC) concentrations (mg of C/L) over time (number of days) for different conditions. The data used are mean values of the triplicate digesters, and standard deviations are indicated with error bars. Vertical lines represent the sampling dates.

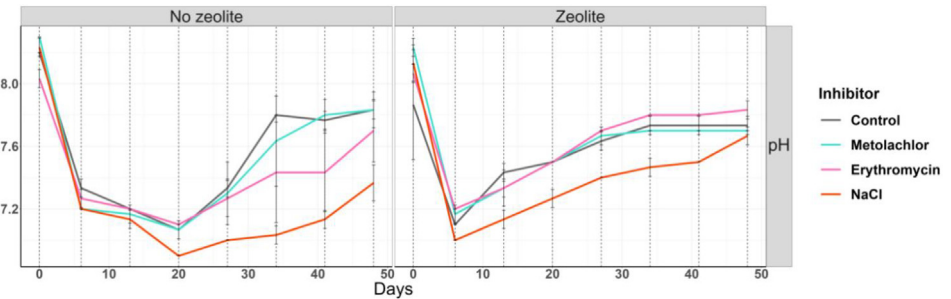


Fig. 5. pH values over time (number of days) for different conditions. The data used are mean values of the triplicate digesters, and standard deviations are indicated with error bars. Vertical lines represent the sampling dates.

Table 9
Acetic acid concentration in each digester (mgC/L).

Day	0_a	0_b	0_c	Na_a	Na_b	Na_c	Ery_a	Ery_b	Ery_c	Met_a	Met_b	Met_c	Oz_a	Oz_b	Oz_c	Naz_a	Naz_b	Naz_c	Eryz_a	Eryz_b	Eryz_c	Metz_a	Metz_b	Metz_c
0	27.2	27.3	28.4	88.5	53.1	44.1	26.1	28.4	26.4	26.6	55.1	57.1	48.6	41.5	43.7	31	67.6	36.6	28.6	27	30.8	49.4	49.7	23.4
6	414.8	446.5	446.5	4.6	36.9	20.7	421.5	335	369.5	444.7	417.3	396.4	402.9	377.3	406.1	46.4	81	10.1	342.5	410.3	352.2	422.2	486.9	508.6
13	701.7	717.2	742.6	548.1	563.2	540.7	504.8	499.2	554.2	781.7	737.7	701.9	257.1	337.7	386.8	424.3	502.3	523.3	569.2	538.1	576.5	416.1	541.6	288.4
20	1052.4	974.6	975.6	660.7	657.5	635.1	772.7	750.3	913.8	948.4	1058.7	888.7	0	0	0	109.2	193.6	211.8	218.9	99	220.7	9.7	80.9	0
27	886	738	966	868	914	940	1038	996	686	464	1068	1228	0	0	0	23.4	21.8	46.8	7.5	33.6	0	0	0	0
34	7	6.7	15.6	928	1076	966	1146	1126	200	0	6.7	1202	0	0	0	0	0	0	0	5.6	0	0	0	0
41	6	4.9	6.1	812.6	754.8	782.1	288.6	607.1	61.2	2.3	3.6	45.7	3.7	2.2	2.3	3.5	3.4	4.3	13.3	1.4	7.2	2.9	2.9	3.2
48	4.2	5.4	5	337.5	723.8	423.8	243.8	207.9	37	2	3.2	4.3	1.3	1.9	1.8	5	4.8	3.6	2.6	1.3	3	1.8	1.3	2

Table 10
Propionic acid concentration in each digester (mgC/L).

Day	O_a	O_b	O_c	Na_a	Na_b	Na_c	Ery_a	Ery_b	Ery_c	Met_a	Met_b	Met_c	Oz_a	Oz_b	Oz_c	Naz_a	Naz_b	Naz_c	Eryz_a	Eryz_b	Eryz_c	Metz_a	Metz_b	Metz_c
0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
6	53.6	79.9	75.3	0	6.3	0	85.7	52.1	54.6	63.1	50.4	51.4	81.1	67.5	74.5	0	0	0	79.1	66.6	70.3	70.5	87	100.3
13	101.6	104.1	104.9	76.2	82.5	91.5	112.8	120.9	94.4	95.8	99.6	94.9	107.1	111.8	118.1	72.4	75.2	89	105.7	80.6	120.7	103.9	96	110.9
20	110.3	110.1	97.7	82.7	83.5	65.5	121.8	126.7	112.1	73.9	99.3	70.3	0	10	45	84	89.6	82.8	106	82.8	112.8	22.6	79.2	0
27	133.1	133.3	132.6	91.5	98	109.2	183.9	193.4	121.9	127.9	130.1	123.8	0	0	0	76.1	98.8	98.5	120.4	100.2	111.4	0	0	0
34	102.2	101.4	100.2	89.3	153.5	83.7	195.1	121.1	90.2	64.2	102.6	120.2	0	0	0	0	0	0	36.2	0	102.4	0	0	0
41	0	0	20.3	143.8	117.8	142.9	64	112.4	94.8	0.5	0	71.5	0.6	0.6	0.7	0	0	0	21.6	0	16.7	0.6	0.6	0.7
48	0	0	0	98.3	133.7	115.8	98.5	58.9	117.1	0.5	0.6	82.8	0.6	0.6	0.7	0	0	0	0	0	0.7	0.6	0.6	0.6

Table 12
Total volatile fatty acids (VFAs) concentration in each digester (mgC/L).

Day	O_a	O_b	O_c	Na_a	Na_b	Na_c	Ery_a	Ery_b	Ery_c	Met_a	Met_b	Met_c	Oz_a	Oz_b	Oz_c	Naz_a	Naz_b	Naz_c	Eryz_a	Eryz_b	Eryz_c	Metz_a	Metz_b	Metz_c
0	125.4	103.8	110.8	148.1	75.5	67.1	123.7	135.1	72.9	127.9	190.4	209.5	230.5	180.7	214.7	154.2	196.7	166.3	148.7	127.8	142.6	186.3	201	225.3
6	483.4	707.7	643.6	262.5	51.2	30.8	935.8	846.6	823.3	520.2	488.5	499.4	645.7	501.3	599	46.4	81	10.1	862.7	864.7	802.6	652.1	573.9	878.2
13	1066.2	1100.7	1175.5	951.6	999	969.5	1061.7	1099	997.2	1102.2	1120.6	1154.4	371.9	456.4	512.8	632.6	724.1	762.9	750.7	709.6	774.5	520	637.6	399.4
20	1247.1	1192.6	1197.1	960.1	1013.5	899.2	1160.8	1184.8	1141.3	1110.7	1264	1047.7	0	10	45	193.1	290.4	302.1	324.9	181.8	333.5	32.3	160.1	0
27	1019.1	871.3	1119.6	1045	1102.5	1124.9	1282.4	1254	807.9	591.9	1223.1	1389.8	0	0	0	76.1	122.2	120.3	167.2	107.7	145	0	0	0
34	109.2	108.1	115.8	1017.3	1229.5	1049.7	1399.7	1312.6	290.2	64.2	109.4	1322.2	0	0	0	0	0	0	36.2	0	108	0	0	0
41	6	4.9	26.5	958.8	875	927.7	362.5	754.4	156.3	3.1	4	117.6	4.4	2.8	3	3.5	3.4	4.3	35.2	1.4	24.3	3.5	3.5	4.2
48	4.5	5.7	5.4	435.8	857.4	541	342.3	268.3	154.4	2.6	4.1	87.5	2.2	2.8	2.8	5	4.8	3.6	2.9	1.5	4	2.4	1.9	2.6

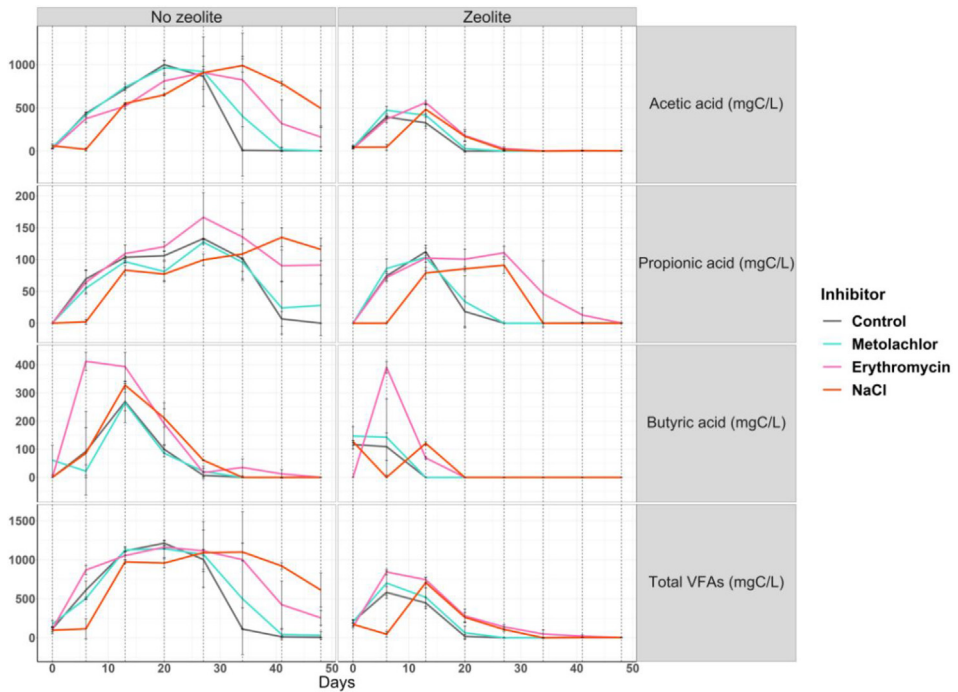


Fig. 6. Acetic acid, propionic acid, butyric acid, and total volatile fatty acids concentrations (mg of C/L) over time (number of days) for different conditions. The data used are mean values of the triplicate digesters, and standard deviations are indicated with error bars. Vertical lines represent the sampling dates.

the methanogenesis process (referred to as late degradation stage in the related research article). In addition, 4 samples taken on day 0 were analyzed to characterize the initial microbial community (inoculum). Details are provided in [Table 1](#).

3. Experimental Design, Materials and Methods

3.1. Experimental design and sampling

The batch anaerobic digesters used in the study were 1 L glass bottles with a working volume of 700 mL. Each bottle was inoculated with 5.7 g of methanogenic sludge and fed 12.7 g of food waste, reaching a substrate-to-inoculum ratio of 7.9 g COD/1.8 g COD. This ratio was chosen to prevent excessive gas production that might lead to the rupture of the bottles. The inoculum came from a 60 L laboratory anaerobic bioreactor regularly fed with food waste at 35 °C and was centrifuged at 10,000 g for 10 min before use. The food waste was obtained from the institute's restaurant containing 0.45 g COD/g. All food waste was finely ground using a grinder and stored at -20 °C before use. In the first series of 12 digesters, 20 g/L of sodium chloride (Acros Organics), 400 mg/L of erythromycin (Acros Organics), and 5 mg/L of S-metolachlor (Honeywell) were added in triplicate, with three control digesters without any inhibitors. A second series of another 12 digesters was set up precisely as the first, but 15 g/L of zeolite (Siliz 24®, Somez company, France) was introduced as a support material, directly in the batch digesters. All digesters were sealed with rubber septa and caps. A short, flexible tube was inserted into the hole at the center of the rubber septum, above which was controlled by a valve that switched to open and close to collect the produced biogas. All digesters were incubated in the dark at 35 °C without agitation.

8 mL of liquid samples were collected from each digester weekly using a syringe of 10 mL through the septa. The samples were distributed into 2 mL Eppendorf tubes and centrifuged at 10,000 g for 10 min at 4 °C. The supernatant and pellet samples were stored at -20 °C before analyses. 7 mL of gas samples were taken every week from the headspace using a glass syringe and stored in vacuum tubes (BD Vacutainer dry tubes) at room temperature before the isotopic composition analysis.

3.2. Biodegradation performance monitoring

Gas production and composition were measured using a differential manometer (Digitron 2082P, Margam, UK) and a micro gas chromatograph (CP4900, Varian, Palo Alto, USA) respectively, as described in [3]. Volatile fatty acid concentrations were quantified using ionic chromatography (ICS 5000+, Thermo Fisher Scientific) with an IonPAC ICE-AS1 column, as described in [3]. Dissolved organic and inorganic carbon (DOC and DIC) were measured according to the French standard NF EN 1484 using a TOC-L CPN analyzer (Shimadzu). Isotopic fractionation of methane and carbon dioxide ($\delta^{13}\text{C}_{\text{CH}_4}$ and $\delta^{13}\text{C}_{\text{CO}_2}$) was measured with a Trace Gas Chromatograph Ultra (Thermo Scientific) connected to a Delta V Plus isotope ratio mass spectrometer (Thermo Scientific) via a combustion interface GC III (Thermo Scientific), to calculate apparent isotopic fractionation as described in [3].

3.3. DNA extraction, amplification and sequencing

The total DNA was extracted from the pellet samples using the DNeasy PowerSoilPro Isolation Kit (QIAGEN) with the QIAcube Instrument following the manufacturer's instructions. The concentration of extracted DNA was quantified with Qubit 2.0 fluorometer using dsDNA kit (Invitrogen), and the purity was checked with Epoch 2 Microplate Spectrophotometer (Agilent BioTek).

The extracted DNA was used to amplify the V4-V5 hypervariable region of the 16S rRNA genes of bacterial and archaeal populations. Briefly, the IonAmplicon Library Preparation (FusionMethod) Protocol, Revision C, was employed [4]. Amplicons were prepared using primers 515F (5'-GTGYCAGCMGCCGCGTA-3') and 928R (5'-CCCCGYCAATTCMTTTRAGT-3'). The forward primer was modified by adding a PGM sequencing adaptor (adaptor A: 5'-CCATCTCATCCCTGCGTGTCTCCGACTCAG-3') and a barcode (5'-adaptor A-Barcode-515F-3'). The reverse primer was modified with the addition of a PGM sequencing adaptor (adaptor trP1: 5'-CCTCTCTATGGG CAGTCGGTGAT-3') (5'-adaptor trP1-928R-3'). The V4-V5 region was amplified following the Platinum Pfx protocol (Life Technologies). PCR products were cleaned using the Agencourt AMPure XP magnetic beads purification system (Beckman Coulter). Automated electrophoresis (2200 TapeStation with D1000 ScreenTape, Agilent Technologies) was incorporated to verify the quantity and size of the amplicons. All libraries were pre-diluted to 500 pM in molecular-grade water and pooled equimolarly. The pooled library was then diluted to 26 pM and processed on the Ion OneTouch 2 Instrument using the Ion PGM Hi-Q View OT2 Kit to prepare template-positive Ion Sphere Particles (ISPs) containing clonally amplified DNA by emulsion PCR. These templated ISPs were quantified and enriched on the Ion OneTouch ES according to the manufacturer's instructions.

Sequencing was performed on Ion Torrent PGM (Life Technologies) using Ion 316 V2 chips and the Ion PGM Hi-Q View Sequencing Kit following the manufacturer's instructions.

3.4. Sequence read processing

Upon completion of sequencing, the sequencing instrument generated DAT files containing the raw traces of electrical signals. These raw traces were converted into single numeric values

for each flow per well, resulting in 1.wells files. The information in the 1.wells files was then transformed into a sequence of bases using the BaseCaller, producing unaligned BAM (Binary Sequence Alignment/Map) files. The BAM files were subsequently converted to FASTQ format using the FileExporter plugin. Finally, the data were processed with Torrent Suite software to filter out low-quality and polyclonal sequence reads, ultimately yielding high-quality data in FASTQ format.

Limitations

The outcomes are influenced by the type of sludge used to inoculate the digesters and may vary if a different inoculum is employed. Likewise, the composition of the waste introduced into the digesters also affects the results.

Ethics Statement

The authors have read and follow the ethical requirements for publication in Data in Brief and confirm that the current work does not involve human subjects, animal experiments, or any data collected from social media platforms.

Data availability

[16S rRNA gene sequencing data \(Original data\)](#) (ENA).

CRedit Author Statement

Xiaoqing Wang: Conceptualization, Methodology, Investigation, Data curation, Formal analysis, Visualization, Writing – original draft; **Vincent Dürr:** Conceptualization, Methodology, Investigation, Data curation, Formal analysis, Validation; **Angéline Guenne:** Investigation, Data curation; **Nadine Derlet:** Investigation, Data curation; **Christelle Bureau:** Investigation, Data curation; **Elodie Gittard:** Investigation, Data curation; **Laurent Mazéas:** Supervision; **Olivier Chapleur:** Conceptualization, Methodology, Writing – review & editing, Supervision, Project administration, Funding acquisition.

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Declaration of Competing Interest

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

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