

Chapter 63

Genomics of Acute Lung Injury and Vascular Barrier Dysfunction

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Abstract Acute lung injury (ALI) is a devastating syndrome of diffuse alveolar damage that develops via a variety of local and systemic insults such as sepsis, trauma, pneumonia, and aspiration. It is interestingly to note that only a subset of individuals exposed to potential ALI-inciting insults develop the disorder and the severity of the disease varies from complete resolution to death. In addition, ALI susceptibility and severity are also affected by ethnicity as evidenced by the higher mortality rates observed in African-American ALI patients compared with other ethnic groups in the USA. Moreover, marked differences in strain-specific ALI responses to inflammatory and injurious agents are observed in preclinical animal models. Together, these observations strongly indicate genetic components to be involved in the pathogenesis of ALI. The identification of genes contributing to ALI would potentially provide a better understanding of ALI pathobiology, yield novel biomarkers, identify individuals or populations at risk, and prove useful for the development of novel and individualized therapies. Genome-wide searches in animal models have identified a number of quantitative trait loci that associate with ALI susceptibility. In this chapter, we utilize a systems biology approach combining cellular signaling pathway analysis with population-based association studies to review established and suspected candidate genes that contribute to dysfunction of endothelial cell barrier integrity and ALI susceptibility.

Keywords Alveolar capillary permeability • Lung injury • Genomics • Genetics • Bioinformatics • Polymorphism

1 Introduction

Acute lung injury (ALI) is a devastating syndrome of diffuse alveolar damage that develops via a variety of local and systemic insults such as sepsis, trauma, pneumonia, and

aspiration [1]. Deranged alveolar capillary permeability, profound inflammation, and extravasation of edematous fluids into the alveolar spaces are critical elements of ALI, reflecting the substantial surface area of the pulmonary vasculature needed for alveolar gas exchange. ALI, together with its severest form, acute respiratory distress syndrome (ARDS), afflicts approximately 190,000 patients per year in the USA and has a mortality rate of 35–50% [2, 3]. It is interestingly to note, however, that only a subset of individuals exposed to potential ALI-inciting insults develop the disorder and the severity of the disease varies from complete resolution to death. In addition, ALI susceptibility and severity are also affected by ethnicity as evidenced by the higher mortality rates observed in African-American ALI patients compared with other ethnic groups in the USA [4]. Moreover, marked differences in strain-specific ALI responses to inflammatory and injurious agents are observed in preclinical animal models [5]. Together, these observations strongly indicate genetic components to be involved in the pathogenesis of ALI.

The role that genetics plays in determining ALI risk or the subsequent severity of the outcome is one of the many unanswered questions regarding ALI pathogenesis and epidemiology. The identification of genes contributing to ALI would potentially provide a better understanding of the pathogenic mechanisms of ALI, yield novel biomarkers, identify individuals or populations at risk, and prove useful for the development of novel and individualized therapies. However, a traditional genetic approach to studies using family linkage mapping is not feasible given the sporadic nature of ALI and the necessity of an extreme environmental insult. Further, genetic studies of ALI are challenging owing to the substantial phenotypic variance in critically ill patients, diversity in the lung injury evoking stimuli, presence of varied comorbid illnesses common in the critically ill patient, complex gene–environment interactions, and potentially incomplete gene penetrance [6, 7]. Despite these inherent challenges, the unrivaled progress made in the post-human genome era combined with the utilization of sophisticated bioinformatics and high-throughput methods have allowed significant advances to be made. For example, these tools are now linked to escalating knowledge of the

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molecular mechanisms of lung endothelial permeability, a hallmark of ALI and an attractive target for the design of novel therapies, to identify candidate genes whose variants are potentially involved in ALI susceptibility. Genome-wide searches in animal models have identified a number of quantitative trait loci that associate with ALI susceptibility [8].

In this chapter, we utilize a systems biology approach combining cellular signaling pathway analysis with population-based association studies to review established and suspected candidate genes that contribute to dysfunction of endothelial cell barrier integrity and ALI susceptibility. The integration of high-throughput gene expression profiling in preclinical models of ALI with bioinformatics has led to the identification of differentially expressed genes in response to ALI whose variants are potentially involved in ALI susceptibility and severity. This approach confirmed long-suspected ALI-associated candidate genes, but more importantly, identified novel genes not previously implicated in ALI. Increasing knowledge of the molecular mechanisms of endothelial-barrier-regulatory pathways has also enhanced the ability to find novel ALI candidate genes. The analysis of the molecular pathways involving the cytoskeletal scaffolding and the dynamic cytoskeletal changes driving cell shape alterations, a key feature of vascular permeability, has identified additional genes contributing to the development and severity of ALI, thereby providing novel therapeutic targets in this devastating illness. Genes encoding proinflammatory cytokines, growth factors and mediators, receptors for barrier-regulatory agonists, and mechanical-stress-sensitive genes expressed in endothelium which regulate inflammatory responses also serve as attractive ALI candidate genes and are representative of the diverse but fertile areas of exploration for candidate SNPs affecting ALI susceptibility and severity.

2 Candidate Genes in Acute Lung Injury: Vascular Barrier Regulatory Cytokines, Growth Factors, and Mediators

2.1 Angiotensin-Converting Enzyme

Angiotensin-converting enzyme (ACE) is a member of the rennin-angiotensin system (RAS), balancing the levels of angiotensin I and angiotensin II, with significant expression in lung vascular endothelium as compared with other vascular beds [9]. The RAS is considered to be an important regulator of inflammation that contributes to ALI by altering vascular permeability, vascular tone, fibroblast activation, and endothelial-epithelial cell survival [10–12]. For example, angiotensin II activates inflammatory processes by upregulating proinflammatory cytokines and chemokines

via type I and type II angiotensin II receptors that subsequently activate the nuclear factor κ B (NF- κ B) pathway [13, 14]. The RAS is also involved in the fibrotic response to ALI via induction of transforming growth factor expression [15]. The most compelling evidence for RAS involvement in ALI has come from the effective attenuation of ALI pathobiology by ACE inhibitors or angiotensin receptor blocking drugs [16, 17] and ACE knockout mice in preclinical models of ALI [18].

An intronic insertion (I) or deletion (D) of a 287-bp Alu repeat sequence in the human ACE gene, located on chromosome 17q35, has been associated with ACE levels and activity in serum [19, 20]. The D allele possesses a higher enzyme activity which parallels the higher gene expression in individuals with DD genotype [21]. The initial association of the DD genotype in the ACE gene with increased ALI mortality provided the impetus for subsequent studies to more firmly establish a genetic basis of ALI and to identify ALI candidate genes [22]. Caucasian patients with ARDS show significantly higher frequencies of the DD genotype and the D allele as compared with ventilated intensive care unit (ICU) patients without ARDS, patients after coronary artery bypass surgery, or healthy controls. Moreover, ARDS patients with DD genotype show markedly higher mortality (54%) in comparison with the II genotype (11%) or strike “4” ID genotype (28%) [22]. The higher mortality rate in ARDS patients with DD or ID genotype as compared with II genotype was subsequently confirmed in Han Chinese patients in Taiwan, although the frequency of the D allele is significantly lower in the Chinese population as compared with Western populations [23]. Compared with Caucasians, a higher frequency of D allele has been reported among Africans (Nigerian and African-American populations) [24, 25], potentially contributing to the observed disparity in ALI-associated higher mortality rates in African-Americans [4]. However, to date, no association study of ACE polymorphisms and lung injury has been performed in African-Americans. In contrast, Mexican and Amerindian populations have slightly lower allelic frequencies of the D allele [25]. Thus, ACE represents a highly viable endothelial candidate gene and attractive target in acute inflammatory lung disease.

2.2 Tumor Necrosis Factor

Tumor necrosis factor (TNF) α , an early mediator of ALI development, is a potent proinflammatory cytokine which dramatically increases endothelial cell permeability, cytokine production, and a variety of cytotoxic and proinflammatory compounds which lead to subsequent vascular leakage and disturbed lung water balance. Both TNF α and TNF β subtypes appear in the circulation, in bronchoalveolar lavage (BAL) fluid and in pulmonary edema fluid during the onset of

lung injury. As such, the elevated levels of TNF and its soluble receptors are commonly used as markers of inflammation and are associated with morbidity and mortality in ALI patients [26]. Both the TNF α and TNF β genes lie in close proximity within the major histocompatibility complex, with several polymorphisms described in this region. The -308G/A promoter polymorphism in the TNF α gene and the NcoI restriction fragment length polymorphism in the TNF β gene appear to influence the expression of TNF α . The carriers of the -308A allele and homozygotes for the TNF β_2 allele exhibit increased TNF α expression and have increased susceptibility and mortality to sepsis [27, 28]. In patients with ARDS, the -308A allele is also associated with increased 60-day mortality, with the strongest association found among younger individuals [29]. However, in ARDS patients with direct or indirect pulmonary injury, these SNPs are associated with alterations in ALI susceptibility (TNF α -308 G/A SNP only in the direct pulmonary injury group, and TNF β NcoI only in the indirect pulmonary injury group). Owing to the extent of linkage disequilibrium in the region, it remains unclear as to whether these are regulatory SNPs or if the TNF protein level is modulated by a third locus or a haplotype [30]. Promoter SNPs within the TNF α gene (-238 G/A, -857 C/T) have been associated with inflammatory bowel disease along with the -308 G/A SNP [31]. Thus, the role of TNF variants in inflammatory disorders is apparent and indicates a need for further study of other TNF variants in association with ALI.

2.3 Interleukin-6

Interleukin-6 (IL-6) is an acute-phase response cytokine that plays a key role in the activation of B and T cells. Inflammatory cytokines, including IL-6, are essential for the immune system homeostasis; however, when IL-6 production is exaggerated as observed in inflammatory lung disorders including ALI [22, 32], clearly detrimental outcomes are observed. ALI-related increased levels of IL-6 have been established in the BAL fluid of critically ill patients with ARDS, sepsis, and trauma [33, 34] in association with ALI adverse outcome [35] and development of multisystem organ failure [36]. In prior reports, we observed significantly higher expression of IL-6 and the IL-6 receptor genes across multiple-species ALI models and in human lung endothelium exposed to ventilator-induced mechanical stress as well as in differential region-specific expression in lungs of the canine ALI model [37–39]. On the basis of these data, the IL-6 gene constitutes an excellent candidate gene to understand the genetic basis underlying ALI. A functional polymorphism in the IL-6 gene promoter region at the -174 position (G-174C) has been associated with alterations in both gene expression and IL-6 levels and lower circulating IL-6 concentrations and lower mortality rates in

patients with acute respiratory failure admitted to the ICU [22]. The contrasting correlation between G-174C alleles and circulating IL-6 levels has also been reported [32]. The haplotype involving -174 G/C, 1753C/G, and 2954G/C is associated with higher mortality (and other secondary clinical outcomes) in a cohort of septic patients of European descent [40].

We further evaluated 14 IL-6 gene tagging SNPs covering the entire gene for potential association in sepsis and ALI patients of European descent [32]. No single SNP was identified as significantly associated with ALI; however, a common haplotype (comprising -1363G/-572G/-174G/1208A/1305A/4835C) with a frequency of 63% in cases and 49% in controls showed a significant association with ALI susceptibility. In addition, homozygote carriers of the risk haplotype are twice as frequent in ALI cases (44.8%) than in controls (22.9%), yielding a highly significantly increased odds ratio for developing ALI (odds ratio 2.73; 95% confidence interval, 1.39–5.37; $p=0.003$). This haplotype spans the entire IL-6 gene including the G allele at position -174, i.e. the risk allele for susceptibility to ALI noted above. These data support the association of the IL-6 gene with ALI susceptibility and illustrate the value of haplotype analysis as a robust approach in association studies.

2.4 Vascular Endothelial Growth Factor

Vascular endothelial growth factor (VEGF) is an endothelial-cell-specific mitogen that regulates angiogenesis, migration, and cell permeability [41]. VEGF plays an important role in several organs by directly regulating vascular permeability to water and proteins. Lung overexpression of VEGF induces increased pulmonary vascular permeability, resulting in marked pulmonary edema [42], and plasma VEGF levels are significantly elevated in ALI patients [43]. Several studies have reported the association of low levels of VEGF with the severity of ARDS and elevated levels with the recovery from ARDS, indicating a role for VEGF in the repair process of lung injury [44]. Several polymorphisms have been described in the VEGF gene, primarily in association with cancer susceptibility and severity. The C/T SNP at position 936 of the 3' untranslated region (UTR) of the gene has been associated with higher VEGF plasma levels in healthy subjects [45]. Recently, the C936T SNP in the VEGF gene has been associated with ARDS susceptibility and severity (increased mortality) in subjects of European descent [46, 47]. The haplotype TCT at position C-460 T, C+405G, and C+936 T was significantly associated with a higher rate of mortality in ARDS patients and higher plasma levels of VEGF [47]. These studies highlight the VEGF gene as an attractive barrier-regulatory ALI candidate gene and molecular target in ALI therapeutic strategies.

Table 1 Genes with significant differential expression in multispecies models of acute lung injury and number of PubMatrix citations

| Genes | Gene symbol | Fold change ($p < 0.05$) | PubMatrix (tool for multiplex literature mining, http://pubmatrix.grc.nia.nih.gov), April 2009 | | | | | |
|---|-----------------|----------------------------|--|-------------|--------------------------|--------|--------------|---------------|
| | | | Acute lung injury | Endothelium | Endothelial permeability | Sepsis | Inflammation | Lung diseases |
| Interleukin-1 β | IL-1b | 1.53 | 168 | 829 | 66 | 523 | 4,219 | 649 |
| Interleukin-6 | IL-6 | 1.84 | 459 | 1,571 | 168 | 1,906 | 14,178 | 2,446 |
| Tissue factor/thromboplastin | F3 | 1.52 | 39 | 971 | 34 | 452 | 792 | 530 |
| Plasminogen activator inhibitor type I | PAI-1 | 1.47 | 54 | 1270 | 23 | 189 | 888 | 303 |
| Cyclooxygenase II | COX2 | 1.79 | 2 | 278 | 11 | 60 | 717 | 107 |
| Interleukin-13 | IL-13 | 1.3 | 16 | 85 | 12 | 52 | 1,467 | 1,070 |
| Aquaporin 1 | AQP-1 | 1.3 | 11 | 74 | 52 | 1 | 18 | 19 |
| Plasminogen activator urokinase receptor | PLAUR | 1.47 | 10 | 285 | 8 | 21 | 165 | 122 |
| Interleukin-1 receptor antagonist | IL-1RA | 2 | 24 | 83 | 4 | 208 | 1,200 | 166 |
| Pre-B cell colony enhancing factor | PBEF | 2.82 | 16 | 16 | 5 | 10 | 59 | 17 |
| Chemokine receptor 4 | CXCR4 | 1.62 | 24 | 316 | 8 | 90 | 1,265 | 394 |
| Growth arrest DNA damage inducible α | GADD45 α | 1.71 | 0 | 1 | 1 | 0 | 11 | 7 |

2.5 Chemokine Receptor 4

Chemokine receptor 4 (CXCR4) is an α -chemokine receptor specific for stromal-derived factor 1 (SDF-1; also known as CXCL12) that plays an important role in cell migration, inflammation, B lymphocyte development, angiogenesis, and human immunodeficiency virus (HIV) infection (HIV coreceptor) [48–50]. Chemokine receptors are G-protein-coupled receptors, which trigger diverse signaling cascades including activation of G proteins and the phosphatidylinositol 3-kinase, Janus kinase/signal transducer and activator of transcription, Rho-p160 Rho kinase, and mitogen-activated protein kinase signaling pathways [51]. The activation of these signaling pathways is often accompanied by the internalization of chemokine receptors and their trafficking back to the plasma membrane. This intracellular turnover determines the leukocyte responsiveness to chemokines [52]. Nonmuscle myosin II A is a molecular motor that binds with the cytoplasmic tail of CXCR4 and CCR552 and participates in the SDF-1-dependent endocytosis of CXCR4 via dynamic interaction with α -arrestin, a key component of the CXCR4 internalization pathway [50]. The CXCR4 gene was identified as a novel candidate gene in ALI as it survived two filtering strategies dedicated to identifying ALI-susceptibility genes associated with elevated levels of mechanical stress as observed in mechanical ventilator-associated lung injury (VALI). Our orthologous gene approach determined ALL-specific gene ontologies – coagulation, inflammation, chemotaxis/cell motility, and immune response [38] – involving recognized genes likely to participate in ALI pathogenesis [IL-6, aquaporin 1 (AQP-1), plasminogen activator inhibitor type I (PAI-1)], as well as novel genes not previously known to be mechanistically involved in ALI, including CXCR4 [38] (Table 1). We subsequently utilized a consomic rodent approach with introgression of rat chromosomes 2, 13, 16, and 17, which contained the highest density of VALI-responsive genes [39]. Introgression of the VALI-sensitive Brown Norway

(BN) rat chromosome 13, containing several genes, including CXCR-4, into the VALI-resistant Dahl salt-sensitive (SS) rat resulted in conversion of the SS consomic rats to a VALI-sensitive phenotype [39]. Surface expression of CXCR4 is downregulated by interleukin-4, interleukin-13, and granulocyte-macrophage colony-stimulating factor and upregulated by interleukin-10 and transforming growth factor- β (TGF β) [53], suggesting that CXCR4 may also play a role in the fibrotic response to ALI via TGF β signaling.

Polymorphisms in the CXCR4 gene have not yet been reported; however, a SNP in the 3' UTR of the SDF-1 gene (G801A), is associated with susceptibility to AIDS and type 1 diabetes [54, 55]. We are currently exploring CXCR4 as a potential ALI-associated candidate gene as suggested by the density of PubMatrix citations relating CXCR4 to inflammation (1,151 published papers), endothelium (297 published papers), ALI (28 published papers) and endothelial permeability (eleven published papers). PubMatrix is a Web-based tool that allows simple text-based mining of the NCBI literature search service PubMed using any two lists of keywords terms, resulting in a frequency matrix of term co-occurrence.

3 Strategies to Identify New Genes and Biomarkers in Acute Lung Injury

The advent of high-throughput gene sequencing and expression technologies, and complete genome sequencing of model organisms, now provides the tools to perform large-scale analyses of the genome in complex disorders such as ALI. Whole genome scans, in silico approaches, utilization of consomic rats, and a candidate gene approach involving expression profiling and pathway analysis are proving exceptionally useful in identifying novel candidate genes and genetic variations (Fig. 1).

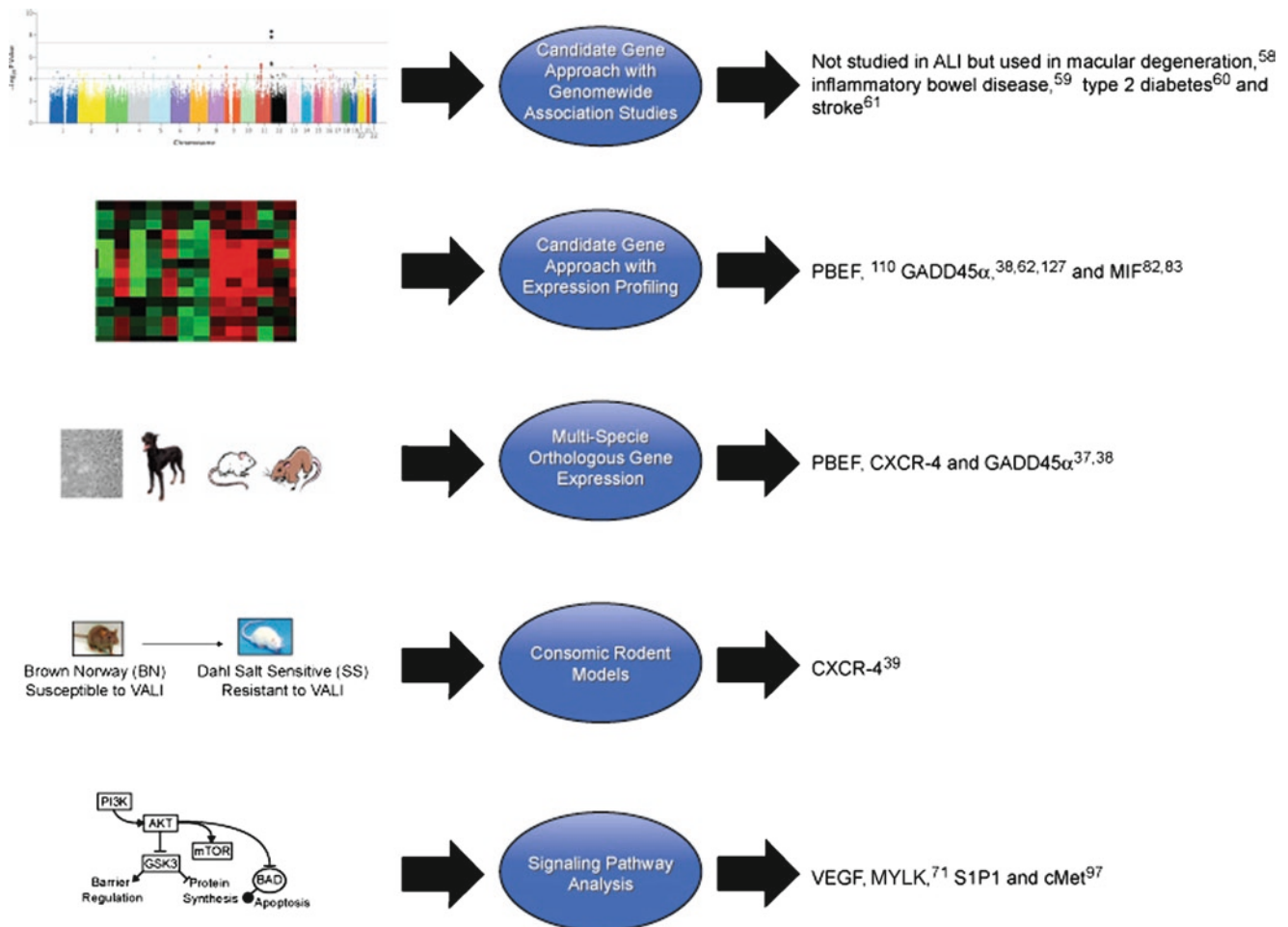


Fig. 1 Novel approaches to identify acute lung injury (ALI) genes. High-throughput gene sequencing and expression technologies, and complete genome sequencing of model organisms, now provide the tools to perform large-scale analyses of the genome in complex disorders such as ALI. Genome-wide association study (GWAS) platforms are effective and have been successfully used in diverse disorders, but although this approach has yet to be employed in either sepsis or ALI, the application of GWAS to the disease is clearly imminent. The differential gene expression between lung apex/base regions as well as between gravitationally dependent/nondependent regions of the lung base in a canine model of ventilator-associated lung injury (VALI) identified ALI-implicated lung genes in response to local mechanical stress within the lung. This approach identified the already established ALI gene macrophage migration inhibitory factor and novel genes such as growth

arrest DNA damage inducible (*GADD45*) and pre-B cell colony enhancing factor (*PBEF*). Our multispecies orthologous gene approach in human (endothelial cells), rat, mouse, and canine models of VALI exhibits expression of common ALI-implicated evolutionarily conserved genes (orthologues) across the species. The genes with a unidirectional 1.3-fold change ($p > 0.05$) are found to reside in high density on rat chromosomes 13 and 16, the chromosomal loci used to develop the consomic rodent model. Together, these approaches identified novel ALI genes such as *PBEF*, chemokine receptor (*CXCR-4*) *GADD45*. Interrogating the prospective pathways involved in endothelial permeability and correlation with these differentially expressed genes in VALI models identified the most putative ALI genes such as myosin light chain kinase (*MYLK*), sphingosine 1-phosphate receptor 1, cMet, and vascular endothelial growth factor (*VEGF*)

High-throughput whole genome scanning technology has recently emerged as a powerful tool, particularly in detecting disease-susceptibility genes with modest effects. The Haplotype Mapping Project [56], which identified blocks of SNPs associated with each other, has allowed selection of the most informative SNPs for further disease association studies [57]. Currently, the most commonly used high-throughput SNP platforms involve assessment of over one million SNPs spanning the genome, i.e. genome-wide association studies (GWAS). GWAS platforms are effective and have been successfully used in diverse disorders such as age-related macular degeneration [58], inflammatory bowel

disease [59], type 2 diabetes [60], and stroke [61]. Although this approach has yet to be employed in either sepsis or ALI, the application of GWAS to the disease is clearly imminent.

Another method to identify ALI candidate genes is an orthologue gene in silico approach. The basis of this approach is the hypothesis that patients with ALI and preclinical animal models of ALI would exhibit commonality in expression of evolutionarily conserved genes across species. For example, profiling results from more than 50 Affymetrix microarray chips obtained from ventilator-associated ALI models (human, rat, mouse, canine) identified 3,077 genes whose expression was altered across all four species in response to ventilator-associated

mechanical stress [37, 38]. Filtering these results for a unidirectional change in gene expression with greater than 1.3-fold change in expression refined the list to 69 genes, reflecting specific ALI-associated gene module/ontology categories: coagulation, inflammation, chemotaxis/cell motility, and immune response. This approach identified multiple genes already recognized as ALI genes (such as IL-6, AQP-1, and PAI-1), but also identified several novel genes that were not previously known to be mechanistically involved in ALI [38]. Complementing the *in silico* approach described above, a consomic rat approach can also be utilized to identify novel ALI gene candidates. In an experimental study, two strains of inbred rodents were determined to have differing susceptibility to VALI (20 mL/kg, 4 h): VALI-sensitive BN rats and the VALI-resistant Dahl SS rats. Using microarray analysis and a bioinformatic-intense candidate gene approach, we identified 245 differentially expressed potential VALI genes with ontologies such as transcription, chemotaxis, and inflammation. Because chromosomes 2, 13, 16, and 17 were found to contain the highest number of VALI-response genes, consomic SS rats containing substituted BN chromosome 13 were exposed to VALI mechanical stress, resulting in conversion of the resistant SS rat to VALI sensitivity [39].

Extensive expression profiling across preclinical ALI models can extend the identification of ALI gene candidates to determination of allelic frequencies of gene polymorphisms (SNPs) that may confer ALI risk or severity. This “candidate gene approach” has identified several candidates with hypothesized significant mechanistic roles in lung injury, inflammation, or repair in the setting of ALI and VALI [62]. Further, given the availability of sophisticated bioinformatic methods and increasing knowledge of the molecular and cellular mechanisms of lung injury, candidate genes can also be identified via analysis of cellular pathways involved in ALI pathogenesis [63, 67].

4 Novel Acute Lung Injury Candidate Genes and Biomarkers

The application of the novel techniques described in the previous section is proving to be exceptionally useful in identifying novel candidate genes and genetic variations in the study of the pathobiology of ALI. These novel gene and biomarkers are discussed in this section.

4.1 Myosin Light Chain Kinase

Myosin light chain kinase (MLCK) is an enzyme that phosphorylates regulatory myosin light chains, which allows myosin cross-bridging interactions with F-actin. In endothelial

cells, the contraction of the actomyosin complex generates a stronger centripetal force that overcomes the force keeping the adjacent endothelial cell tethered, leading to endothelial retraction, decreased intercellular adhesion, and increased vascular permeability [68, 69]. This phenomenon is physiologically relevant as evidenced by nonmuscle MLCK (nmMLCK) isoform knockout mice [which retain the smooth muscle MLCK (smMLCK) isoform] that are less susceptible to lipopolysaccharide (LPS)- and ventilator-induced ALI [70, 71]. Further, treatment with a MLCK inhibitor prior to LPS exposure in the wild-type mice attenuates endothelial cell barrier dysfunction and inflammation [70]. Thus, the myosin light chain kinase gene (*MYLK*), which encodes for MLCK, is an excellent ALI candidate gene.

Since initial cloning of the highly expressed nmMLCK in endothelium in our laboratory [72], we have identified substantial roles of nmMLCK in cytoskeleton rearrangement of endothelial cells regulating vascular barrier function [64, 68], angiogenesis, and leukocyte diapedesis [73], consistent with a potential mechanistic role for MLCK in the genesis of ALI. The human *MYLK* gene is located on chromosome 3q21 and encodes three proteins, including nmMLCK, smMLCK, and telokin. We sequenced exons, exon–intron boundaries, and 2 kb of the 5′ UTR of *MYLK* in healthy individuals, patients with sepsis alone, and patients with sepsis-associated ALI, all of European and African-American descent [66], and identified 51 SNPs (ten exonic, 31 intronic, nine in the 5′ UTR, and one in the noncoding exon 1), of which 28 were chosen for further linkage disequilibrium studies. Five of the ten coding *MYLK* SNPs confer an amino acid change (Pro21His, Pro147Ser, Val261Ala, Ser1341Pro, and Arg1450Gln) in MLCK. Subsequently, association analysis of both single SNPs and haplotypes demonstrated very strong associations in both ethnic groups [66]. In European Americans, the rs3845915A/*MYLK*_037C haplotype was associated with more than a fivefold increase in the risk of developing ALI and sepsis. In contrast, the haplotype *MYLK*_021G/*MYLK*_022G/*MYLK*_011T conferred specific risk for ALI but not sepsis [66]. The 5′ haplotype of the *MYLK* gene also conferred ALI-specific risk in both European- and African-descent subjects; however, the 3′ region haplotype was associated with ALI only in African-descent subjects. In African-Americans, the haplotype hcv1602689C/*MYLK*_037A/rs11707609G is substantially more prevalent in ALI (11%) as compared with sepsis (1%). This CAG haplotype is not found in European Americans, suggesting a potential genetic contribution to the observed ethnicity-specific differences in ALI/ARDS prevalence and susceptibility [4]. We noted similar findings in association studies involving a cohort with trauma-induced ALI [74]. We have also evaluated the association of 17 *MYLK* genetic variants with severe asthma in both European American and African-American populations and identified a SNP highly associated with severe asthma in

African-Americans [75] consistent with data linking this chromosomal locus (*MYLK*, 3q21.1) to asthma and asthma-related phenotypes [75]. Taken together, these data strongly implicate *MYLK* genetic variants as risk variants in inflammatory lung disorders, such as ALI and asthma.

4.2 Macrophage Migration Inhibitory Factor

Macrophage migration inhibitory factor (MIF) is an ALI candidate gene and recognized biomarker, initially discovered as a soluble product of activated T cells and named for its role in inhibiting random macrophage migration [76]. MIF is a proinflammatory cytokine which binds to CD44 and CD74 and is produced by many cell types, including monocytes/macrophages, pituitary cells, vascular endothelium, and respiratory epithelium [77, 78]. MIF may serve as a delicate regulator of the cytokine balance between immunity and inflammation as MIF counterregulates the immunosuppressive effects of glucocorticoids [79]. The role of MIF as an endogenous pro-survival factor has been demonstrated in vitro. LPS-mediated induction of Flice-like inhibitory protein (FLIP) by MIF confers resistance to LPS-mediated endothelial cell death [80]. Suppression of MIF by RNA interference induces cell death and sensitivity to apoptotic stimuli [80]. In addition, MIF interacts with the multidimensional nmMLCK [81] isoform which regulates TNF-mediated apoptosis in addition to its potent effects on endothelial cell barrier dysfunction as discussed earlier [68, 69]. Together, these findings implicate the role of MIF in regulation of nonmuscle cytoskeletal dynamics and vascular pathophysiology, which is evident from the enhanced MIF levels in the serum, BAL fluid, and alveolar endothelium of patients with ARDS as compared with other critically ill patients [76, 78, 82]. We found significant increases in MIF transcript and protein levels in murine and canine models of ventilator-induced lung injury (VILI) (using high mechanical ventilation and endotoxin exposure, respectively) [82] and in human lung endothelium cells exposed to 48 h of cyclic stretch [83]. MIF deficiency or immunoneutralization appears to protect mice or rats from fatal endotoxic shock or other inflammatory diseases [84] although these results are not without controversy [85] and our own studies in 8–12-week-old mice failed to demonstrate a VILI/ALI-related phenotype which was different from controls (data not shown). MIF also upregulates the expression of AQP-1, the water channels expressed in alveolar endothelial and epithelial cells, and a candidate gene we identified in models of VILI-associated mechanical stress [38]. MIF may serve to modulate fluid movement into alveolar spaces, a cardinal feature of ALI [86].

To extend the likelihood that MIF serves as a putative candidate gene in ALI and sepsis, we studied the association of eight MIF polymorphisms, including the most studied MIF

promoter G/C SNP at position -173, in a sepsis-induced ALI cohort ($n=506$) of African- and European-descent cases [82]. No individual SNP showed a significant association with either ALI or sepsis; however, the carriers of the CC genotype (rs755622) and the carriers of the TT genotype (rs2070767) showed more than twofold increased risk of developing sepsis and ALI, respectively. This association was lost, however, after age and gender adjustment in a logistic regression model. In contrast, MIF haplotypes at the 3' region of the gene display strong association with ALI and sepsis, conferring both protection as well as susceptibility to ALI, in European and African populations [82]. Furthermore, the haplotype at the 5' promoter region of the gene involving a short tandem repeat at position -794 (CATT)₅ and the -173 G allele show significant association with both ALI and trauma [82]; however, no association was found between promoter region haplotypes and MIF levels. Rheumatoid arthritis patients with the -173C allele have higher levels of MIF in the serum and synovial fluid than the carriers of the G allele and have a higher probability of developing idiopathic arthritis [87]. Thus, given these diverse MIF functions, MIF remains an attractive target in inflammatory diseases including the lung.

4.3 Sphingosine 1-Phosphate Receptor 1

The bioactive sphingolipid metabolite sphingosine 1-phosphate (S1P) is an important lipid mediator that enhances endothelial cell barrier function in vivo and in vitro by ligating S1P receptor 1 (S1P1), which is encoded by an endothelial differentiation gene (*EDG1* or *S1P1*) [88, 89]. S1P1 is a pertussis-toxin-sensitive, G_i-coupled receptor which induces Rac GTPase-dependent substantial increases in cortical actin polymerization critical to endothelial cell barrier enhancement [88, 90]. S1P1 activation enhances the organization and redistribution of vascular endothelial cadherin and β -catenin in junctional complexes in endothelium by phosphorylation of cadherin as well as p120-catenin and inducing the formation of cadherin/catenin/actin complexes [91]. Understanding the role of S1P in enhancing endothelial cell barrier function underscores its importance as a therapeutic target in reversing loss of endothelial cell barrier integrity. In vivo administration of selective S1P1 competitive antagonists induces a dose-dependent disruption of barrier integrity in pulmonary endothelium [92, 93], whereas S1P1 agonists, SEW2871 and FTY720, promote vascular endothelial barrier function [94–96]. A compelling argument for S1P1 as an attractive ALI candidate gene is not only its ability to transduce signals which restore barrier integrity but also that S1P1 is the target for transactivation by receptors for other potent barrier-protective agonists. These include EPCR (receptor for activated protein C) [65], c-Met [receptor for hepatocyte growth factor (HGF)] [97], CD44 (receptor for high molecular

weight hyaluronan) [67], and the ATP receptor [98]. We recently resequenced the SIP1 gene (14 African-Americans and 13 European Americans) to search for common variations in the *EDG1* gene and identified 39 SNPs in the *EDG1* gene, with several promoter SNPs associated with asthma, another inflammatory lung syndrome [99].

4.4 *c-Met* (Hepatocyte Growth Factor Receptor)

The role of HGF and its tyrosine kinase receptor *c-Met* has been investigated in lung development, inflammation, and repair [100] as well as in neoplastic processes such as cellular transformation, neoplastic invasion, and metastasis [101, 102]. SNPs causing underexpression of *c-Met* have been associated with autism and *c-Met* SNPs/mutations appear to be linked to lung cancer disparities in different ethnic groups. These include an N375S mutation in the HGF-binding domain of *c-Met*, an R988C SNP/mutation in the juxtamembrane domain, and an activating M1268T mutation in the tyrosine kinase domain (exon 19), all linked to development of solid tumors such as lung cancer, renal cancer, gastric cancer, and hepatocellular carcinoma [102]. HGF influences morphogenesis in epithelial cells from a variety of organs, including lungs, where HGF antisense oligonucleotides block alveolar and branching morphogenesis [103]. HGF expression and activity increase after 3–6 h of lung injury with intratracheally administered hydrochloric acid, suggesting that HGF plays a role in reparative responses to lung injury [104]. *c-Met* expression on type II pneumocytes is likely involved in increased type II pneumocyte proliferation and restoration of an intact alveolar epithelium [105]. *c-Met* is composed of a 50-kDa extracellular α subunit and a 145-kDa transmembrane β subunit [106] which contains tyrosine kinase domains, tyrosine phosphorylation sites, and tyrosine docking sites [107]. We demonstrated that HGF-mediated *c-Met* phosphorylation and *c-Met* recruitment to caveolin-enriched microdomains (CEMs) protects against the LPS-induced pulmonary vascular hyperpermeability that is regulated by high molecular weight hyaluronan (CD44 ligand) [108]. Our novel findings indicate that HGF/*c-Met*-mediated, CD44-regulated CEM signaling promotes Tiam1 (a Rac1 exchange factor)/dynamin 2 dependent Rac1 activation, and peripheral recruitment of cortactin (an actin cytoskeletal regulator), processes essential for endothelial cell barrier integrity. Understanding the mechanism(s) by which HGF/*c-Met* promotes increased endothelial cell barrier function may lead to novel treatments for diseases involving vascular barrier disruption, including inflammation, tumor angiogenesis, atherosclerosis, and ALI. However, on the contrary, the higher mortality rate in ALI patients with increased levels of HGF in BAL

fluids [109] and in pulmonary edematous fluids [110] indicates severer injury and inflammation in response to increased HGF levels. It has now become increasingly clear that HGF plays an important role in normal and injured lung and may have a therapeutic potential in lung diseases.

4.5 *Pre-B Cell Colony Enhancing Factor*

Pre-B cell colony enhancing factor (PBEF), was first identified by Samal and colleagues in 1994 as a protein secreted by activated lymphocytes in bone marrow stromal cells that stimulate early stage B cell formation in conjugation with stem cell factor and interleukin-7. A large body of work has now highlighted the power of a systems biology approach in the search for novel disease-susceptibility genes and potentially novel biomarkers, with PBEF serving as an excellent example of this approach (Fig. 2). We first identified marked upregulation of PBEF via microarray analyses of murine and canine models of VILI/ALI with increased gene/protein expression in BAL fluid and serum samples from critically ill ICU patients with ALI and sepsis [111]. With only a total of eight papers in PubMed at that time, we next directly sequenced the PBEF gene in 36 subjects with ALI, sepsis, and healthy controls and conducted a PBEF SNP-based association study in ALI subjects of European and African-American descent [111]. We identified 11 SNPs in the *PBEF* gene with two promoter SNPs, T-1001G and C-1543 T, associated with ALI and sepsis. Genotyping of PBEF C-1543 T and T-1001G SNPs revealed significant associations of sepsis and ALI, with the strongest association found with the -1543C/-1001G haplotype. Univariate analysis found carriers of the G allele (T1001G) to have 2.75-fold higher risk of developing ALI as compared with controls ($p=0.002$) [111]. These results were subsequently confirmed in a comparable but distinct replicate ALI population [112]. Interestingly, the -1543G/-1001C haplotype was also associated with increased ICU patient mortality, whereas the -1543 T/-1001 T haplotype was associated with fewer ventilator days and decreased ICU patient mortality [112].

A key challenge in genomic explorations is the ability to confirm the contribution of a SNP to a dysfunctional-gene-involved disease process. Additional reports have highlighted the capacity for the PBEF gene to have an influence far beyond any B-cell regulatory function, with a key role in regulating vascular permeability [113] as well as inhibiting neutrophil apoptosis [114]. To further explore mechanistic participation of PBEF in ALI and VILI, we focused on the contribution of PBEF to endothelial function. Our prior immunohistochemical staining of canine-injured lung tissues localized PBEF expression to vascular endothelial cells, in addition to infiltrating neutrophils and type 2 alveolar epithelial cells [111]. Our in vitro studies showed that expression

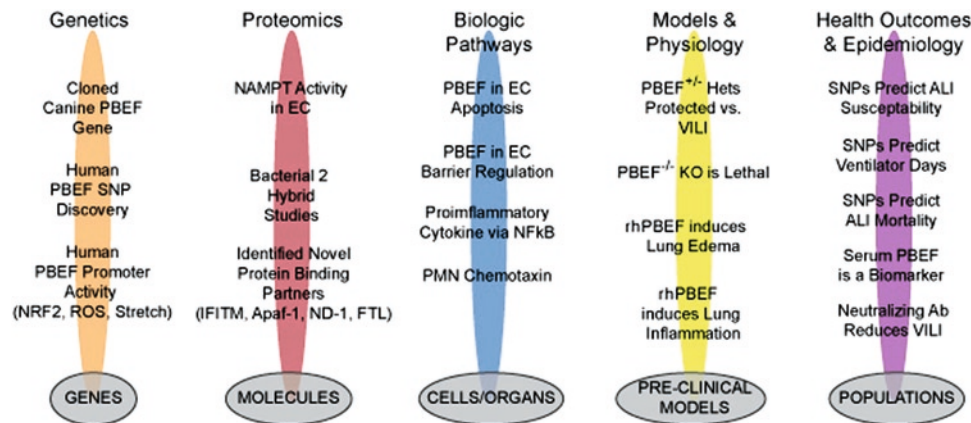


Fig. 2 Systems biology approach in defining the role of *PBEF* in ALI and ventilator-induced lung injury (*VILI*). Each systems biology compartment has been utilized to address *PBEF* involvement across the entire systems biology spectrum. This includes genes (microarrays, SNP discovery, small interfering RNAs, microRNAs, promoter luciferase assays), proteins (bacterial two hybrid studies, recombinant

human *PBEF* (*rhPBEF*), cell signaling, site-directed mutagenesis), organelles (nuclear events, mitochondrial function, cytoskeleton), cells (endothelial cells, neutrophils), organs (lung), preclinical animal models (genetically engineered *PBEF*^{+/-} mice, *VILI*, and sepsis models), and populations [bronchoalveolar lavage (*BAL*) studies, biomarker studies in intensive care unit (*ICU*) subjects, SNP association studies]

of *PBEF* in pulmonary artery endothelial cells increases thrombin-mediated vascular permeability [111], suggesting that enhanced *PBEF* expression may mediate the early increase in vascular permeability that is characteristic of ALI. Neutrophils harvested from the circulation of septic and ALI patients show marked inhibition of the apoptotic process in association with evidence of enhanced respiratory burst capacity [115, 116], with both activities largely restored with administration of *PBEF* antisense oligonucleotides. Our initial in vitro studies further demonstrated recombinant human *PBEF* (*rhPBEF*) as a direct rat neutrophil chemotactic factor, with in vivo studies demonstrating marked increases in *BAL* fluid leukocytes (polymorphonuclear leukocytes, PMNs) following intratracheal injection in C57BL/6 J mice [117]. These changes were accompanied by increased *BAL* fluid levels of PMN chemoattractants (KC and MIP2) and modest increases in lung vascular and alveolar permeability. We also noted synergism between *rhPBEF* challenge and a model of limited *VILI* and observed dramatic increases in *BAL* fluid PMNs, *BAL* protein, and cytokine levels (IL-6, TNF α , KC) compared with either challenge alone. Gene expression profiling identified induction of ALI- and *VILI*-associated gene modules (NF- κ B, leukocyte extravasation, apoptosis, toll-receptor pathways). Heterozygous *PBEF*^{+/-} mice were significantly protected (reduced *BAL* fluid protein levels, *BAL* fluid IL-6 levels, peak inspiratory pressures) when exposed to a model of severe *VILI* (4 h, 40 mL/kg tidal volume) and exhibited significantly reduced gene expression of *VILI*-associated modules. Finally, strategies to reduce *PBEF* availability (neutralizing antibody) resulted in significant protection from *VILI* [117]. *PBEF* is now recognized as associated with modestly increased risk of type 2 diabetes and elevated levels of acute-phase proteins [118] and a C-948G SNP has been associated with an increased diastolic blood pressure in obese

children [119]. These studies implicate *PBEF*, now associated with a number of inflammatory disorders such as inflammatory bowel disease, multiple sclerosis, cystic fibrosis, and asthma [120–122], as a key inflammatory mediator intimately involved in both the development and the severity of ventilator-induced ALI.

4.6 Growth Arrest DNA Damage Inducible α (*GADD45 α*)

Growth arrest DNA damage inducible α (*GADD45 α*), a member of an evolutionarily conserved gene family, is implicated as a stress sensor that modulates the response of mammalian cells to genotoxic or physiological stress [123, 124]. *GADD45 α* is a small 21-kDa predominantly nuclear protein that interacts with other proteins implicated in stress responses, including proliferating cell nuclear antigen, p21, Cdc2/cyclin B1, MEKK4, and p38 kinase [125, 126]. *GADD45* induces cell cycle arrest and apoptosis in most of cells as well as promoting DNA repair functions and survival [126]. Growth Arrest and DNA Damage gene (*GADD45*) also maintains genomic stability in a p53-responsive manner [127]. Despite the multiple known functions of *GADD45*, its role ALI, endothelial/epithelial barrier dysfunction, or repair of injured lung is unknown [38]. *GADD45* exhibited differential expression in orthologous global gene expression profiling, in multispecies ALI models [38], in region-specific lung tissue expression profiling [62], and was markedly upregulated in response to the *VILI* [128]. We explored the mechanistic involvement of *GADD45 α* in endotoxin (LPS)- and ventilator-induced inflammatory lung injury (*VILI*) by comparing multiple biochemical and genomic parameters of

inflammatory lung injury in wild-type C57Bl/6 and GADD45 α ^{-/-} knockout mice exposed to high tidal volume ventilation (VILI) or intratracheally administered LPS [129]. GADD45 α ^{-/-} mice were modestly susceptible to LPS-induced injury but were profoundly susceptible to VILI, demonstrating increased inflammation and increased microvascular permeability. VILI-exposed GADD45 α ^{-/-} mice manifested striking neutrophilic alveolitis with increased BAL fluid levels of protein, IgG, and inflammatory cytokines. Expression profiling of lung homogenates revealed strong dysregulation in the B cell receptor signaling pathway in GADD45 α ^{-/-} mice, suggesting the involvement of phosphatidylinositol 3-kinase/Akt signaling components. Western blots confirmed a threefold reduction in Akt protein and phosphorylated Akt levels observed in GADD45 α ^{-/-} lungs. Electrical resistance measurements across human lung endothelial cell monolayers transfected with small interfering RNAs to reduce GADD45 α or Akt expression revealed significant potentiation of LPS-induced endothelial barrier dysfunction which was attenuated by overexpression of a constitutively active Akt1 transgene. Whereas other lung injury studies failed to demonstrate a role for GADD45 in hyperoxic lung injury [130, 131], our studies validate GADD45 α as a novel inflammatory lung injury candidate gene and a significant participant in vascular barrier regulation via effects on Akt-mediated endothelial signaling [132]. Thus, both Akt and GADD45 are extremely attractive ALI candidate genes. The human GADD45 α gene contains 25 validated SNPs (National Center for Biotechnology Information SNP database) whose role in ALI pathogenesis is completely unknown [123]. We are currently pursuing further characterization of the role of GADD45 α and its association of genetic variants with sepsis and ALI.

5 Novel Therapeutics

The identification of novel pathways involved in the pathobiology of ALI also opens doors for the exploration of new therapeutic targets for the disease. As such, the use of agents that attenuate the endothelial barrier dysfunction and the inflammatory response characteristic of ALI have shown promise in preclinical studies which will hopefully lead to their use in trials of human ALI (Fig. 3).

5.1 Sphingosine 1-Phosphate

S1P, an important lipid mediator generated by the phosphorylation of sphingosine by sphingosine kinase, decreases endothelial permeability to both water and solute via cytoskeletal reorganization and adherens junction assembly

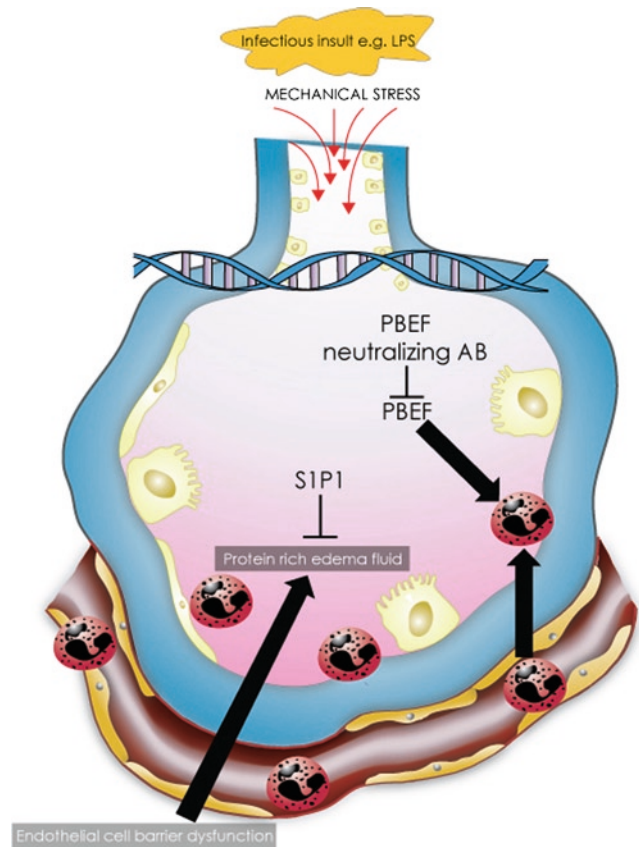


Fig. 3 Mechanism-based novel therapies for ALI. The identification of novel pathways involved in the pathobiology of ALI also facilitates the exploration of new therapeutic targets. Sphingosine 1-phosphate (S1P) attenuates the endothelial barrier dysfunction associated with ALI, whereas blocking of PBEF attenuates VALI

[88, 89]. S1P-induced barrier protective effects could serve to attenuate the increased pulmonary vascular permeability essential factor in the development of ALI. The S1P analogue FTY720 (0.1 mg/kg), when administered to C57Bl/6 mice with endotoxin-induced lung injury, decreases lung edema formation, solute transport across the alveolar capillary endothelium, and inflammatory cell infiltration into lung parenchyma [94]. Similarly, the prophylactic administration of S1P attenuates both alveolar and vascular barrier dysfunction while significantly reducing shunt formation associated with lung injury in rodent and canine models of ALI induced by combined intrabronchial endotoxin administration and high-tidal-volume mechanical ventilation [133]. In a recent study of a canine model of ALI, we demonstrated that when bacterial endotoxin was instilled intratracheally followed in 1 h by intravenous administration of S1P (85 μ g/kg) or vehicle and 8 h of high-tidal-volume mechanical ventilation [134], S1P treatment attenuated the severity of ALI-induced increases in shunt fraction and the presence of both protein and neutrophils in BAL fluid compared with vehicle controls. Interestingly, BAL fluid cytokine production was not altered

significantly by intravenous administration of S1P and S1P potentiated the endotoxin-induced systemic production of the inflammatory cytokines TNF α , C-X-C chemokine ligand-1, and IL-6, without resulting in end-organ dysfunction. These data suggest that S1P may represent a viable therapy for the prevention and treatment of ALI.

5.2 Pre-B Cell Colony Enhancing Factor Blockage

As previously described in this chapter, PBEF appears to play a central role in the promotion of several pathogenetic aspects of ALI and VALI. Therefore, interventions aimed at attenuating the effects of PBEF could have a potential therapeutic effect in these disorders. To begin to address the potential for PBEF to serve as a therapeutic target in ameliorating VILI, we assessed the effect of PBEF neutralizing antibody on rhPBEF-stimulated lung inflammation [117]. Simultaneous instillation of rhPBEF and PBEF neutralizing antibody produced dramatic reductions in rhPBEF-induced neutrophil recruitment. Further, the intratracheal delivery of PBEF neutralizing antibody (30 min before high-tidal-volume mechanical ventilation) abolished VILI-induced increases in total BAL fluid cell counts and significantly decreased neutrophil influx into the alveolar space as well as VILI-mediated increases in the level of lung tissue albumin.

6 Summary

ALI is a major cause of morbidity and mortality in critically ill patients. Given the unacceptably high mortality rate observed in ALI and the paucity of novel therapies and biomarkers, it is essential to recognize molecular targets associated with ALI to identify individuals at risk and to develop novel therapeutic targets and biomarkers. It is clear that derangements in endothelial cell barrier regulation play a major role in the pathobiology of ALI and genetic variants regulate endothelial cell barrier function, thereby determining ALI risk or subsequent severity of outcome. High-throughput gene sequencing and expression technologies, and complete genome sequencing of model organisms, have allowed for the performance of large-scale analyses of the genome in ALI. In this chapter, we have highlighted how global gene expression profiling in multispecies ALI models served to broaden our net knowledge of ALI-implicated genes and provide a basis for hope that increased insights and therapies may be forthcoming. As genotyping becomes more rapid and easily accessed, combining advanced bioinformatics techniques with high-throughput methods will be the future practice of personalizing treatment strategies. Continued challenges will be the gene–gene and

gene–environment interactions, which add complexity to our understanding of the genome. These novel genetic approaches may prove exceptionally useful in ushering in the era of personalized medicine for critically ill individuals.

References

1. Ware LB, Matthay MA (2000) The acute respiratory distress syndrome. *N Engl J Med* 342:1334–1349
2. Flores C, Ma SF, Maresco K, Ahmed O, Garcia JGN (2006) Genomics of acute lung injury. *Semin Respir Crit Care Med* 27:389–395
3. Pepe PE, Potkin RT, Reus DH, Hudson LD, Carrico CJ (1982) Clinical predictors of the adult respiratory distress syndrome. *Am J Surg* 144:124–130
4. Moss M, Mannino DM (2002) Race and gender differences in acute respiratory distress syndrome deaths in the United States: an analysis of multiple-cause mortality data (1979–1996). *Crit Care Med* 30:1679–1685
5. Savov JD, Whitehead GS, Wang J et al (2004) Ozone-induced acute pulmonary injury in inbred mouse strains. *Am J Respir Cell Mol Biol* 31:69–77
6. Kamp R, Sun X, Garcia JG (2008) Making genomics functional: deciphering the genetics of acute lung injury. *Proc Am Thorac Soc* 5:348–353
7. Meyer NJ, Garcia JGN (2007) Wading into the genomic pool to unravel acute lung injury genetics. *Proc Am Thorac Soc* 4:69–76
8. Bauer AK, Malkinson AM, Kleeberger SR (2004) Susceptibility to neoplastic and non-neoplastic pulmonary diseases in mice: genetic similarities. *Am J Physiol Lung Cell Mol Physiol* 287:L685–L703
9. Ng KK, Vane JR (1968) Fate of angiotensin I in the circulation. *Nature* 218:144–150
10. Dimmeler S, Rippmann V, Weiland U, Haendeler J, Zeiher AM (1997) Angiotensin II induces apoptosis of human endothelial cells. Protective effect of nitric oxide. *Circ Res* 81:970–976
11. Kiely DG, Cargill RI, Wheeldon NM, Coutie WJ, Lipworth BJ (1997) Haemodynamic and endocrine effects of type I angiotensin II receptor blockade in patients with hypoxaemic cor pulmonale. *Cardiovasc Res* 33:201–208
12. Marshall RP, Webb S, Bellingan GJ et al (2002) Angiotensin converting enzyme insertion/deletion polymorphism is associated with susceptibility and outcome in acute respiratory distress syndrome. *Am J Respir Crit Care Med* 166:646–650
13. Esteban V, Lorenzo O, Rupérez M et al (2004) Angiotensin II, via AT1 and AT2 receptors and NF- κ B pathway, regulates the inflammatory response in unilateral ureteral obstruction. *J Am Soc Nephrol* 15:1514–1529
14. Suzuki Y, Ruiz-Ortega M, Lorenzo O, Ruperez M, Esteban V, Egido J (2003) Inflammation and angiotensin II. *Int J Biochem Cell Biol* 35:881–900
15. Marshall RP, Gohlke P, Chambers RC et al (2004) Angiotensin II and the fibroproliferative response to acute lung injury. *Am J Physiol Lung Cell Mol Physiol* 286:L156–L164
16. Wang R, Zagariya A, Ibarra-Sunga O et al (1999) Angiotensin II induces apoptosis in human and rat alveolar epithelial cells. *Am J Physiol* 276:L885–L889
17. Wang R, Ibarra-Sunga O, Verlinski L, Pick R, Uhal BD (2000) Abrogation of bleomycin-induced epithelial apoptosis and lung fibrosis by captopril or by a caspase inhibitor. *Am J Physiol Lung Cell Mol Physiol* 279:L143–L151
18. Imai Y, Kuba K, Rao S et al (2005) Angiotensin-converting enzyme 2 protects from severe acute lung failure. *Nature* 436:112–116

19. Rigat B, Hubert C, Alhenc-Gelas F, Cambien F, Corvol P, Soubrier F (1990) An insertion/deletion polymorphism in the angiotensin I-converting enzyme gene accounting for half the variance of serum enzyme levels. *J Clin Invest* 86:1343–1346
20. Tiret L, Rigat B, Visvikis S et al (1992) Evidence, from combined segregation and linkage analysis, that a variant of the angiotensin I-converting enzyme (ACE) gene controls plasma ACE levels. *Am J Hum Genet* 51:197–205
21. Costerousse O, Allegrini J, Lopez M, Alhenc-Gelas F (1993) Angiotensin I-converting enzyme in human circulating mononuclear cells: genetic polymorphism of expression in T-lymphocytes. *Biochem J* 290:33–40
22. Marshall RP, Webb S, Hill MR, Humphries SE, Laurent GJ (2002) Genetic polymorphisms associated with susceptibility and outcome in ARDS. *Chest* 121:68S–69S
23. Jerng JS, Yu CJ, Wang HC, Chen KY, Cheng SL, Yang PC (2006) Polymorphism of the angiotensin-converting enzyme gene affects the outcome of acute respiratory distress syndrome. *Crit Care Med* 34:1001–1006
24. Mathew J, Basheeruddin K, Prabhakar S (2001) Differences in frequency of the deletion polymorphism of the angiotensin-converting enzyme gene in different ethnic groups. *Angiology* 52:375–379
25. Vargas-Alaróon G, Hernández-Pacheco G, Rodríguez-Pérez JM et al (2003) Angiotensin-converting enzyme gene (ACE) insertion/deletion polymorphism in Mexican populations. *Hum Biol* 75:889–896
26. Parsons PE, Matthay MA, Ware LB, Eisner MD (2005) Elevated plasma levels of soluble TNF receptors are associated with morbidity and mortality in patients with acute lung injury. *Am J Physiol Lung Cell Mol Physiol* 288:L426–L431
27. Maier LA, Sawyer RT, Bauer RA et al (2001) High beryllium-stimulated TNF- α is associated with the -308 TNF- α promoter polymorphism and with clinical severity in chronic beryllium disease. *Am J Respir Crit Care Med* 164:1192–1199
28. Tang GJ, Huang SL, Yien HW et al (2000) Tumor necrosis factor gene polymorphism and septic shock in surgical infection. *Crit Care Med* 28:2733–2736
29. Gong MN, Zhou W, Williams PL et al (2005) *308GA* and *TNFB* polymorphisms in acute respiratory distress syndrome. *Eur Respir J* 26:382–389
30. Hajeer AH, Hutchinson IV (2001) Influence of TNF α gene polymorphisms on TNF α production and disease. *Hum Immunol* 62:1191–1199
31. Ferguson LR, Huebner C, Petermann I et al (2008) Single nucleotide polymorphism in the tumor necrosis factor-alpha gene affects inflammatory bowel diseases risk. *World J Gastroenterol* 14:4652–4661
32. Flores C, Ma SF, Maresso K, Wade MS, Villar J, Garcia JGN (2008) IL6 gene-wide haplotype is associated with susceptibility to acute lung injury. *Transl Res* 152:11–17
33. Park WY, Goodman RB, Steinberg KP et al (2001) Cytokine balance in the lungs of patients with acute respiratory distress syndrome. *Am J Respir Crit Care Med* 164:1896–1903
34. Takala A, Jousela I, Takkunen O et al (2002) A prospective study of inflammation markers in patients at risk of indirect acute lung injury. *Shock* 17:252–257
35. Meduri GU, Headley S, Kohler G et al (1995) Persistent elevation of inflammatory cytokines predicts a poor outcome in ARDS. Plasma IL-1 beta and IL-6 levels are consistent and efficient predictors of outcome over time. *Chest* 107:1062–1073
36. Ranieri VM, Zhang H, Mascia L et al (2000) Pressure-time curve predicts minimally injurious ventilatory strategy in an isolated rat lung model. *Anesthesiology* 93:1320–1328
37. Grigoryev DN, Finigan JH, Hassoun P, Garcia JGN (2004) Science review: searching for gene candidates in acute lung injury. *Crit Care* 8:440–447
38. Grigoryev DN, Ma SF, Irizarry RA, Ye SQ, Quackenbush J, Garcia JGN (2004) Orthologous gene-expression profiling in multi-species models: search for candidate genes. *Genome Biol* 5:R34
39. Nonas SA, Moreno-Vinasco L, Ma SF et al (2007) Use of consomic rats for genomic insights into ventilator-associated lung injury. *Am J Physiol Lung Cell Mol Physiol* 293:L292–L302
40. Sutherland AM, Walley KR, Manocha S, Russell JA (2005) The association of interleukin 6 haplotype clades with mortality in critically ill adults. *Arch Intern Med* 165:75–82
41. Mura M, dos Santos CC, Stewart D, Liu M (2004) Vascular endothelial growth factor and related molecules in acute lung injury. *J Appl Physiol* 97:1605–1617
42. Kaner RJ, Ladetto JV, Singh R, Fukuda N, Matthay MA, Crystal RG (2000) Lung overexpression of the vascular endothelial growth factor gene induces pulmonary edema. *Am J Respir Cell Mol Biol* 22:657–664
43. Thickett DR, Armstrong L, Christie SJ, Millar AB (2001) Vascular endothelial growth factor may contribute to increased vascular permeability in acute respiratory distress syndrome. *Am J Respir Crit Care Med* 164:1601–1605
44. Abadie Y, Bregeon F, Papazian L et al (2005) Decreased VEGF concentration in lung tissue and vascular injury during ARDS. *Eur Respir J* 25:139–146
45. Renner W, Kotschan S, Hoffmann C, Obermayer-Pietsch B, Pilger E (2000) A common 936 C/T mutation in the gene for vascular endothelial growth factor is associated with vascular endothelial growth factor plasma levels. *J Vasc Res* 37:443–448
46. Medford AR, Keen LJ, Bidwell JL, Millar AB (2005) Vascular endothelial growth factor gene polymorphism and acute respiratory distress syndrome. *Thorax* 60:244–248
47. Zhai R, Gong MN, Zhou W et al (2007) Genotypes and haplotypes of the VEGF gene are associated with higher mortality and lower VEGF plasma levels in patients with ARDS. *Thorax* 62:718–722
48. Berger EA, Murphy PM, Farber JM (1999) Chemokine receptors as HIV-1 coreceptors: roles in viral entry, tropism, and disease. *Annu Rev Immunol* 17:657–700
49. Muller A, Homey B, Soto H et al (2001) Involvement of chemokine receptors in breast cancer metastasis. *Nature* 410:50–56
50. Rey M, Valenzuela-Fernández A, Urzainqui A et al (2007) Myosin IIA is involved in the endocytosis of CXCR4 induced by SDF-1 α . *J Cell Sci* 120:1126–1133
51. Ganju RK, Brubaker SA, Meyer J et al (1998) The α -chemokine, stromal cell-derived factor-1 α , binds to the transmembrane G-protein-coupled CXCR-4 receptor and activates multiple signal transduction pathways. *J Biol Chem* 273:23169–23175
52. Fan GH, Lapierre LA, Goldenring JR, Sai J, Richmond A (2004) Rab11-family interacting protein 2 and myosin Vb are required for CXCR2 recycling and receptor-mediated chemotaxis. *Mol Biol Cell* 15:2456–2469
53. Wang J, Guan E, Roderiquez G, Calvert V, Alvarez R, Norcross MA (2001) Role of tyrosine phosphorylation in ligand-independent sequestration of CXCR4 in human primary monocytes-macrophages. *J Biol Chem* 276:49236–49243
54. Dubois-Laforgue D, Hendel H, Caillat-Zucman S et al (2001) A common stromal cell-derived factor-1 chemokine gene variant is associated with the early onset of type 1 diabetes. *Diabetes* 50:1211–1213
55. Winkler C, Modi W, Smith MW et al (1998) Genetic restriction of AIDS pathogenesis by an SDF-1 chemokine gene variant. ALIVE Study, Hemophilia Growth and Development Study (HGDS), Multicenter AIDS Cohort Study (MACS), Multicenter Hemophilia Cohort Study (MHCS), San Francisco City Cohort (SFCC). *Science* 279:389–393
56. International HapMAP Project. <http://www.hapmap.org/>
57. Daly MJ, Rioux JD, Schaffner SF, Hudson TJ, Lander ES (2001) High-resolution haplotype structure in the human genome. *Nat Genet* 29:229–232

58. Klein RJ, Zeiss C, Chew EY et al (2005) Complement factor H polymorphism in age-related macular degeneration. *Science* 308:385–389
59. Duerr RH, Taylor KD, Brant SR et al (2006) A genome-wide association study identifies IL23R as an inflammatory bowel disease gene. *Science* 314:1461–1463
60. Sladek R, Rocheleau G, Rung J et al (2007) A genome-wide association study identifies novel risk loci for type 2 diabetes. *Nature* 445:881–885
61. Ikram MA, Seshadri S, Bis JC et al (2009) Genomewide association studies of stroke. *N Engl J Med* 360:1718–1728
62. Simon BA, Easley RB, Grigoryev DN et al (2006) Microarray analysis of regional cellular responses to local mechanical stress in acute lung injury. *Am J Physiol Lung Cell Mol Physiol* 291:L851–L861
63. Dudek SM, Birukov KG, Zhan X, Garcia JGN (2002) Novel interaction of cortactin with endothelial cell myosin light chain kinase. *Biochem Biophys Res Commun* 298:511–519
64. Dudek SM, Garcia JGN (2001) Cytoskeletal regulation of pulmonary vascular permeability. *J Appl Physiol* 91:1487–1500
65. Finigan JH, Dudek SM, Singleton PA et al (2005) Activated protein C mediates novel lung endothelial barrier enhancement: role of sphingosine 1-phosphate receptor transactivation. *J Biol Chem* 280:17286–17293
66. Gao L, Grant A, Halder I et al (2006) Novel polymorphisms in the myosin light chain kinase gene confer risk for acute lung injury. *Am J Respir Cell Mol Biol* 34:487–495
67. Singleton PA, Dudek SM, Ma SF, Garcia JGN (2006) Transactivation of sphingosine 1-phosphate receptors is essential for vascular barrier regulation. Novel role for hyaluronan and CD44 receptor family. *J Biol Chem* 281:34381–34393
68. Garcia JGN, Davis HW, Patterson CE (1995) Regulation of endothelial cell gap formation and barrier dysfunction: role of myosin light chain phosphorylation. *J Cell Physiol* 163:510–522
69. Garcia JGN, Schaphorst KL (1995) Regulation of endothelial cell gap formation and paracellular permeability. *J Investig Med* 43:117–126
70. Wainwright MS, Rossi J, Schavocky J et al (2003) Protein kinase involved in lung injury susceptibility: evidence from enzyme isoform genetic knockout and in vivo inhibitor treatment. *Proc Natl Acad Sci U S A* 100:6233–6238
71. Mirzapioazova T, Moitra J, Sammani S et al (2009) Critical role of non-muscle MLCK in ventilator-induced lung injury. *Am J Respir Crit Care Med* 179:A3822
72. Garcia JGN, Lazar V, Gilbert-McClain LI, Gallagher PJ, Verin AD (1997) Myosin light chain kinase in endothelium: molecular cloning and regulation. *Am J Respir Cell Mol Biol* 16:489–494
73. Garcia JGN, Verin AD, Herenyiova M, English D (1998) Adherent neutrophils activate endothelial myosin light chain kinase: role in transendothelial migration. *J Appl Physiol* 84:1817–1821
74. Christie JD, Ma SF, Aplenc R et al (2008) Variation in the MYLK gene is associated with development of acute lung injury after major trauma. *Crit Care Med* 36:2794–2800
75. Flores C, Ma SF, Maresco K, Ober C, Garcia JG (2007) A variant of the myosin light chain kinase gene is associated with severe asthma in African Americans. *Genet Epidemiol* 31:296–305
76. Donnelly SC, Bucala R (1997) Macrophage migration inhibitory factor: a regulator of glucocorticoid activity with a critical role in inflammatory disease. *Mol Med Today* 3:502–507
77. Baugh JA, Bucala R (2002) Macrophage migration inhibitory factor. *Crit Care Med* 30:S27–S35
78. Lai KN, Leung JC, Metz CN, Lai FM, Bucala R, Lan HY (2003) Role for macrophage migration inhibitory factor in acute respiratory distress syndrome. *J Pathol* 199:496–508
79. Calandra T, Bernhagen J, Metz CN et al (1995) MIF as a glucocorticoid-induced modulator of cytokine production. *Nature* 377:68–71
80. Damico RL, Chesley A, Johnston L et al (2008) Macrophage migration inhibitory factor governs endothelial cell sensitivity to LPS-induced apoptosis. *Am J Respir Cell Mol Biol* 39:77–85
81. Wadgaonkar R, Dudek SM, Zaiman AL et al (2005) Intracellular interaction of myosin light chain kinase with macrophage migration inhibition factor (MIF) in endothelium. *J Cell Biochem* 95:849–858
82. Gao L, Flores C, Fan-Ma S et al (2007) Macrophage migration inhibitory factor in acute lung injury: expression, biomarker, and associations. *Transl Res* 150:18–29
83. Gao L, Ye SQ, Maloney JP, Garcia JG (2003) Role of macrophage migration inhibitory factor (MIF) in human and animal models of acute lung injury (ALI) and sepsis: association of a promoter polymorphism and increased gene expression. *Am J Respir Crit Care Med* 167:A162
84. Tohyama S, Onodera S, Tohyama H et al (2008) A novel DNA vaccine-targeting macrophage migration inhibitory factor improves the survival of mice with sepsis. *Gene Ther* 15:1513–1522
85. Korsgren M, Kallstrom L, Uller L et al (2000) Role of macrophage migration inhibitory factor (MIF) in allergic and endotoxin-induced airway inflammation in mice. *Mediators Inflamm* 9: 15–23
86. King KL, Barnes KC, Ashworth R (2003) Aquaporin-1: a candidate gene in sepsis and lung injury. *Am J Respir Crit Care Med* 167:A662
87. Donn RP, Shelley E, Ollier WE, Thomson W (2001) A novel 5'-flanking region polymorphism of macrophage migration inhibitory factor is associated with systemic-onset juvenile idiopathic arthritis. *Arthritis Rheum* 44:1782–1785
88. Garcia JGN, Liu F, Verin AD et al (2001) Sphingosine 1-phosphate promotes endothelial cell barrier integrity by Edg-dependent cytoskeletal rearrangement. *J Clin Invest* 108:689–701
89. Singleton PA, Dudek SM, Chiang ET, Garcia JGN (2005) Regulation of sphingosine 1-phosphate-induced endothelial cytoskeletal rearrangement and barrier enhancement by S1P1 receptor, PI3 kinase, Tiam1/Rac1, and alpha-actinin. *FASEB J* 19:1646–1656
90. Arce FT, Whitlock JL, Birukova AA et al (2008) Regulation of the micromechanical properties of pulmonary endothelium by S1P and thrombin: role of cortactin. *Biophys J* 95:886–894
91. Chiang ET, Camp SM, Dudek SM et al (2009) Protective effects of high-molecular weight polyethylene glycol (PEG) in human lung endothelial cell barrier regulation: role of actin cytoskeletal rearrangement. *Microvasc Res* 77:174–186
92. Foss FW Jr, Snyder AH, Davis MD et al (2007) Synthesis and biological evaluation of γ -aminophosphonates as potent, subtype-selective sphingosine 1-phosphate receptor agonists and antagonists. *Bioorg Med Chem* 15:663–677
93. Sanna MG, Wang SK, Gonzalez-Cabrera PJ et al (2006) Enhancement of capillary leakage and restoration of lymphocyte egress by a chiral S1P1 antagonist in vivo. *Nat Chem Biol* 2:434–441
94. Peng X, Hassoun PM, Sammani S et al (2004) Protective effects of sphingosine 1-phosphate in murine endotoxin-induced inflammatory lung injury. *Am J Respir Crit Care Med* 169:1245–1251
95. Rosen H, Sanna MG, Cahalan SM, Gonzalez-Cabrera PJ (2007) Tipping the gatekeeper: S1P regulation of endothelial barrier function. *Trends Immunol* 28:102–107
96. Sammani S, Mirzapioazova T, Moreno-Vinasco L, Garcia J (2008) Effect of S1P1 receptor agonists on murine lung airway function. *J Investig Med* 56:659
97. Liu F, Verin AD, Wang P et al (2001) Differential regulation of sphingosine-1-phosphate- and VEGF-induced endothelial cell chemotaxis. Involvement of G_{i2} -linked Rho kinase activity. *Am J Respir Cell Mol Biol* 24:711–719
98. Jacobson JR, Dudek SM, Singleton PA, Kolosova IA, Verin AD, Garcia JGN (2006) Endothelial cell barrier enhancement by ATP

- is mediated by the small GTPase Rac and cortactin. *Am J Physiol Lung Cell Mol Physiol* 291:L289–L295
99. Sun X, Ma SF, Wade MS et al (2009) Sphingosine-1-phosphate receptor 1 variant increases promoter activity and decreases susceptibility to human asthma. *Am J Respir Crit Care Med* 175:A5420
 100. Brinkmann V, Foroutan H, Sachs M, Weidner KM, Birchmeier W (1995) Hepatocyte growth factor/scatter factor induces a variety of tissue-specific morphogenic programs in epithelial cells. *J Cell Biol* 131:1573–1586
 101. Lesko E, Majka M (2008) The biological role of HGF-MET axis in tumor growth and development of metastasis. *Front Biosci* 13:1271–1280
 102. Ma PC, Kijima T, Maulik G et al (2003) c-MET mutational analysis in small cell lung cancer: novel juxtamembrane domain mutations regulating cytoskeletal functions. *Cancer Res* 63:6272–6281
 103. Itakura A, Kurauchi O, Morikawa S, Okamura M, Furugori K, Mizutani S (1997) Involvement of hepatocyte growth factor in formation of bronchoalveolar structures in embryonic rat lung in primary culture. *Biochem Biophys Res Commun* 241:98–103
 104. Yanagita K, Matsumoto K, Sekiguchi K, Ishibashi H, Niho Y, Nakamura T (1993) Hepatocyte growth factor may act as a paracrine factor on lung regeneration after acute lung injury. *J Biol Chem* 268:21212–21217
 105. Panos RJ, Patel R, Bak PM (1996) Intratracheal administration of hepatocyte growth factor/scatter factor stimulates rat alveolar type II cell proliferation in vivo. *Am J Respir Cell Mol Biol* 15:574–581
 106. Hammond DE, Carter S, Clague MJ (2004) Met receptor dynamics and signalling. *Curr Top Microbiol Immunol* 286:21–44
 107. Kermorgant S, Parker PJ (2005) c-Met signalling: spatio-temporal decisions. *Cell Cycle* 4:352–355
 108. Singleton PA, Salgia R, Moreno-Vinasco L et al (2007) CD44 regulates hepatocyte growth factor-mediated vascular integrity. Role of c-Met, Tiam1/Rac1, dynamin 2, and cortactin. *J Biol Chem* 282:30643–30657
 109. Stern JB, Fierobe L, Paugam C et al (2000) Keratinocyte growth factor and hepatocyte growth factor in bronchoalveolar lavage fluid in acute respiratory distress syndrome patients. *Crit Care Med* 28:2326–2333
 110. Verghese GM, McCormick-Shannon K, Mason RJ, Matthay MA (1998) Hepatocyte growth factor and keratinocyte growth factor in the pulmonary edema fluid of patients with acute lung injury. Biologic and clinical significance. *Am J Respir Crit Care Med* 158:386–394
 111. Ye SQ, Simon BA, Maloney JP et al (2005) Pre-B-cell colony-enhancing factor as a potential novel biomarker in acute lung injury. *Am J Respir Crit Care Med* 171:361–370
 112. Bajwa EK, Yu CL, Gong MN, Thompson BT, Christiani DC (2007) Pre-B-cell colony-enhancing factor gene polymorphisms and risk of acute respiratory distress syndrome. *Crit Care Med* 35:1290–1295
 113. Ye SQ, Zhang LQ, Adyshev D et al (2005) Pre-B-cell-colony-enhancing factor is critically involved in thrombin-induced lung endothelial cell barrier dysregulation. *Microvasc Res* 70:142–151
 114. Jia SH, Li Y, Parodo J et al (2004) Pre-B cell colony-enhancing factor inhibits neutrophil apoptosis in experimental inflammation and clinical sepsis. *J Clin Invest* 113:1318–1327
 115. Jimenez MF, Watson RW, Parodo J et al (1997) Dysregulated expression of neutrophil apoptosis in the systemic inflammatory response syndrome. *Arch Surg* 132:12639, discussion 9–70
 116. Taneja R, Parodo J, Jia SH, Kapus A, Rotstein OD, Marshall JC (2004) Delayed neutrophil apoptosis in sepsis is associated with maintenance of mitochondrial transmembrane potential and reduced caspase-9 activity. *Crit Care Med* 32:1460–1469
 117. Hong SB, Huang Y, Moreno-Vinasco L et al (2008) Essential role of pre-B-cell colony enhancing factor in ventilator-induced lung injury. *Am J Respir Crit Care Med* 178:605–617
 118. Zhang YY, Gottardo L, Thompson R et al (2006) A visfatin promoter polymorphism is associated with low-grade inflammation and type 2 diabetes. *Obesity (Silver Spring)* 14:2119–2126
 119. Körner A, Böttcher Y, Enigk B, Kiess W, Stumvoll M, Kovacs P (2007) Effects of genetic variation in the visfatin gene (*PBEF1*) on obesity, glucose metabolism, and blood pressure in children. *Metabolism* 56:772–777
 120. Koppelman GH, Stine OC, Xu J et al (2002) Genome-wide search for atopy susceptibility genes in Dutch families with asthma. *J Allergy Clin Immunol* 109:498–506
 121. Satsangi J, Parkes M, Louis E et al (1996) Two stage genome-wide search in inflammatory bowel disease provides evidence for susceptibility loci on chromosomes 3, 7 and 12. *Nat Genet* 14:199–202
 122. Vandenbroeck K, Fiten P, Heggarty S et al (2002) Chromosome 7q21–22 and multiple sclerosis: evidence for a genetic susceptibility effect in vicinity to the protachykinin-1 gene. *J Neuroimmunol* 125:141–148
 123. Lal A, Gorospe M (2006) Egad, more forms of gene regulation: the *gadd45a* story. *Cell Cycle* 5:1422–1425
 124. Liebermann DA, Hoffman B (2007) Gadd45 in the response of hematopoietic cells to genotoxic stress. *Blood Cells Mol Dis* 39:329–335
 125. Fornace AJ Jr, Jackman J, Hollander MC, Hoffman-Liebermann B (1992) Liebermann DA Genotoxic-stress-response genes and growth-arrest genes *gadd*, *MyD*, and other genes induced by treatments eliciting growth arrest. *Ann N Y Acad Sci* 663:139–153
 126. Liebermann DA, Hoffman B (2002) Myeloid differentiation (*MyD*)/growth arrest DNA damage (*GADD*) genes in tumor suppression, immunity and inflammation. *Leukemia* 16:527–541
 127. Hollander MC, Philburn RT, Patterson AD, Wyatt MA, Fornace AJ Jr (2005) Genomic instability in *Gadd45a*^{-/-} cells is coupled with S-phase checkpoint defects. *Cell Cycle* 4:704–709
 128. Dolinay T, Szilasi M, Liu M, Choi AM (2004) Inhaled carbon monoxide confers antiinflammatory effects against ventilator-induced lung injury. *Am J Respir Crit Care Med* 170:613–620
 129. Meyer NJ, Huang Y, Singleton PA et al (2009) *GADD45a* is a novel candidate gene in inflammatory lung injury via influences on Akt signaling. *FASEB J* 23(5):1325–1337
 130. O'Reilly MA, Staversky RJ, Watkins RH, Maniscalco WM, Keng PC (2000) p53-independent induction of *GADD45* and *GADD153* in mouse lungs exposed to hyperoxia. *Am J Physiol Lung Cell Mol Physiol* 278:L552–L559
 131. Roper JM, Gehen SC, Staversky RJ, Hollander MC, Fornace AJ Jr, O'Reilly MA (2005) Loss of *Gadd45a* does not modify the pulmonary response to oxidative stress. *Am J Physiol Lung Cell Mol Physiol* 288:L663–L671
 132. Altemeier WA, Matute-Bello G, Gharib SA, Glenn RW, Martin TR, Liles WC (2005) Modulation of lipopolysaccharide-induced gene transcription and promotion of lung injury by mechanical ventilation. *J Immunol* 175:3369–3376
 133. McVerry BJ, Peng X, Hassoun PM, Sammani S, Simon BA, Garcia JGN (2004) Sphingosine 1-phosphate reduces vascular leak in murine and canine models of acute lung injury. *Am J Respir Crit Care Med* 170:987–993
 134. Szczepaniak WS, Zhang Y, Hagerty S et al (2008) Sphingosine 1-phosphate rescues canine LPS-induced acute lung injury and alters systemic inflammatory cytokine production in vivo. *Transl Res* 152:213–224