

## Novel Tools and Methods

# Validation of a New Coil Array Tailored for Dog Functional Magnetic Resonance Imaging Studies

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## Abstract

Comparative neuroimaging allows for the identification of similarities and differences between species. It provides an important and promising avenue, to answer questions about the evolutionary origins of the brain's organization, in terms of both structure and function. Dog functional magnetic resonance imaging (fMRI) has recently become one particularly promising and increasingly used approach to study brain function and coevolution. In dog neuroimaging, image acquisition has so far been mostly performed with coils originally developed for use in human MRI. Since such coils have been tailored to human anatomy, their sensitivity and data quality is likely not optimal for dog MRI. Therefore, we developed a multichannel receive coil (K9 coil, read "canine") tailored for high-resolution functional imaging in canines, optimized for dog cranial anatomy. In this paper we report structural ( $n=9$ ) as well as functional imaging data (resting-state,  $n=6$ ; simple visual paradigm,  $n=9$ ) collected with the K9 coil in comparison to reference data collected with a human knee coil. Our results show that the K9 coil significantly outperforms the human knee coil, improving the signal-to-noise ratio (SNR) across the imaging modalities. We noted increases of roughly 45% signal-to-noise in the structural and functional domain. In terms of translation to fMRI data collected in a visual flickering checkerboard paradigm, group-level analyses show that the K9 coil performs better than the knee coil as well. These findings demonstrate how hardware improvements may be instrumental in driving data quality, and thus, quality of imaging results, for dog-human comparative neuroimaging.

**Key words:** canine neuroimaging; comparative neuroimaging; dog fMRI; hardware; K9 coil

## Significance Statement

Comparative neuroimaging is a powerful avenue to discover evolutionary mechanisms at the brain level. However, data quality is a major constraint in nonhuman functional magnetic resonance imaging (fMRI). We describe a novel canine head coil for magnetic resonance imaging, designed specifically for dog cranial anatomy. Data quality performance and improvements over previously used human knee coils are described quantitatively. In brief, the canine coil improved signal quality substantially across both structural and functional imaging domains, with strongest improvements noted on the cortical surface.

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## Introduction

Comparative neuroimaging aims to find the commonalities and differences in brains and brain function of different species. The focus of comparative neuroimaging often lies on great apes and other nonhuman primates (Rilling, 2014; de Schotten et al., 2019), but by focusing on comparisons between primates, insights on convergent evolution are limited. Convergent evolution describes the advent of a trait, such as a neural mechanism, in phylogenetically distant species, where both species developed the trait independently (e.g., wings in bats and birds). Neuroscience research and neuroimaging in birds (Behroozi et al., 2018b; Güntürkün and Bugnyar, 2018; Behroozi et al., 2020) and reptiles (Behroozi et al., 2018a) have shown that cognition is not reliant on the presence of a neocortex. Therefore, looking at sophisticated behaviors in more distant species outside the primate lineage should not be neglected and indeed nonprimate neuroscience has seen a rise of interest in the past decades (De Groof et al., 2013; Mars et al., 2016; Bunford et al., 2017; Xu et al., 2020).

With regard to convergent evolution, dogs, *Canis lupus familiaris*, are a study species of the highest interest: they excel in social cognition, often outperforming great apes in their understanding of social cues from humans (Kirchhofer et al., 2012; Kaminski and Nitzschner, 2013; Huber, 2016). This places the dog at a prime position for investigating the convergent evolution of social cognition and other cognitive skills, mirrored in an increase of neuroimaging studies of dogs in recent years (for review, see Berns, 2013; Thompkins et al., 2016; Bunford et al., 2017; Huber and Lamm, 2017).

Dogs have the added advantage of being highly trainable, which makes it possible to perform awake, unrestrained and unsedated neuroimaging in dogs (Berns et al., 2012; Strassberg et al., 2019; Karl et al., 2020a), opening the possibility for classical functional magnetic resonance imaging (fMRI) studies in this species, something that is not easily possible in rodents (Keilholz et al., 2004), birds, or monkeys without fixating, restraining, or sedating the animals.

However, many challenges for canine neuroimaging remain to be met. Training dogs to lie still and voluntarily stay in the scanner environment while being attentive to the presented stimuli is very time consuming (Berns et al., 2012; Strassberg et al., 2019; Karl et al., 2020a). Canine neuroimaging runs also need to be shorter than those typically used in humans, and usually amount to a maximum

length of 5 min, as even highly trained dogs cannot maintain attention and stillness for longer. Moreover, dogs rarely manage to perform more than two such runs in one scanner session. These three constraints limit the amount of data that can be collected within a reasonable time frame. This increases the demands on the data, stressing the importance of data quality. In this report, we describe a hardware approach to circumvent data quantity limitations by increasing data quality.

One avenue to improve data quality is to focus on the “software” side of data analysis, e.g., optimizing data preprocessing, by taking into account the different physiology of dog skulls and brains. Increased data quality was obtained with an inhouse preprocessing pipeline based on SPM, as well as with determining a dog-tailored hemodynamic response function (HRF) for fMRI analysis (Boch et al., 2021). Another path to improve data quality and analysis sensitivity is the improvement of hardware, through specific dog-tailored hardware components, an avenue that has received less attention thus far.

Dog fMRI usually relies on human scanner systems, which cannot be easily replaced or exchanged to better fit the canine anatomy. Hence, we reasoned that data quality improvements through hardware can be achieved most straightforwardly and cost-effectively through a dog-tailored head coil.

We validated a novel inhouse developed 16-channel receive coil (K9 coil), tailored to the dog’s cranial anatomy (Fig. 1). In collaboration with the other co-authors, this coil was developed by CW and EL at the Medical University Vienna. Our intention was to overcome the limitation of commonly used coils (human knee coils, Jia et al., 2016; Thompkins et al., 2016; Karl et al., 2020b, 2021; as well as FlexCoils, Cuaya et al., 2016; Szabó et al., 2019), which are not tailored to the anatomy of the dog’s skull and thus may result in suboptimal signal-to-noise ratios (SNRs) and data quality overall.

In the present paper, we apply the K9 coil and compare its images and image quality to a commonly used human knee coil (15 channel receive coil; Siemens Healthineers) we previously used to scan the same animals (Boch et al., 2021; Karl et al., 2021). To this end, we collected data from nine dogs in three different imaging modalities (structural, functional: task based, functional: resting-state), with the two different coils, using otherwise identical MR scanning parameters.

## Materials and Methods

### Sample

Dogs were recruited through the Clever Dog Lab at the Messerli Research Institute at the University of Veterinary Medicine Vienna. Only dogs who completed scanning with both coils were included in this comparison. In total, nine dogs were scanned for T1 imaging and in a functional flickering checkerboard condition with both coils. For resting-state measurements, six of the nine dogs were scanned with both coils and included in our analysis of these resting-state data (Table 1). On average, dogs were 8.1 years old (T1 and functional, 8.3 years in resting-state; note that

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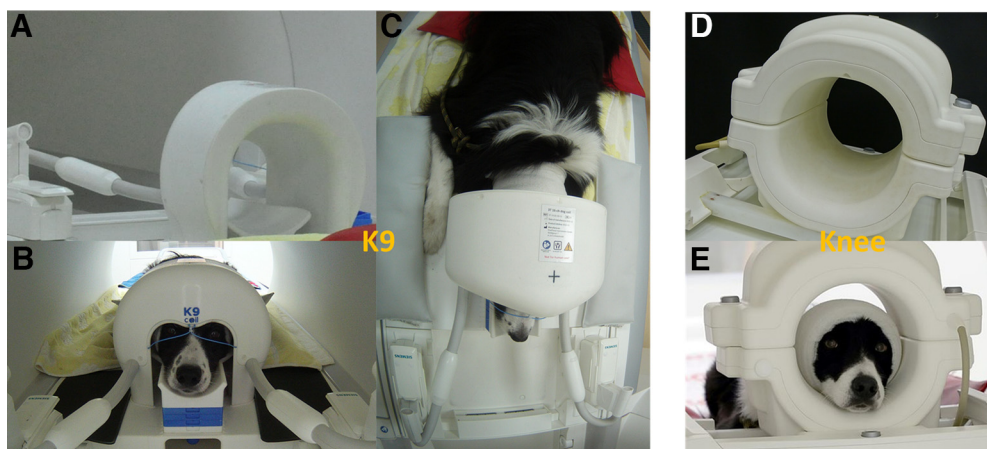
\*L.H. and C.L. shared senior authorship.

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**Figure 1.** **A**, Rear view of the K9 coil on the scanner bed. **B**, Front view with subject. Note the chin rest of adaptable height and the paws left and right of the coil. **C**, Bird's eye view of dog lying in K9 coil on the scanner bed. **D**, Rear view of the knee coil. **E**, Front view of the knee coil with participant. Note the sizable distance between the top of the head and the coil, which is likely reducing sensitivity of measurements.

part of the functional data with the human knee coil and with a different analysis focus was reported already by Boch et al., 2021). Most scanned dogs belonged to herding dog breeds (see Table 1). All dogs had been examined for potential problems with eyesight and general health condition. Dog owners did not receive any monetary compensation for their dogs' participation and gave written informed consent before data collection. All participants in this sample underwent extensive scanner training, based on reward-based positive reinforcement and operant conditioning (Karl et al., 2020a), which enabled them to lie unrestrained and still in the MRI scanner. If uncomfortable, dogs are able and allowed to interrupt the run and leave the coil and move on the scanner bed at any time during the examinations, on which the trainer will give the dog a short break, if so needed, or stop scanning for that day. The studies from which data for this comparative coil overview is taken were approved by the institutional ethics and animal welfare commission in accordance with Good Scientific Practice (GSP) guidelines and national legislation at the University of Veterinary Medicine Vienna (ETK-06/06/2017), based on a pilot study conducted at the University of Vienna. The current study complies with the ARRIVE Guidelines (Kilkenny et al., 2010).

### Coils

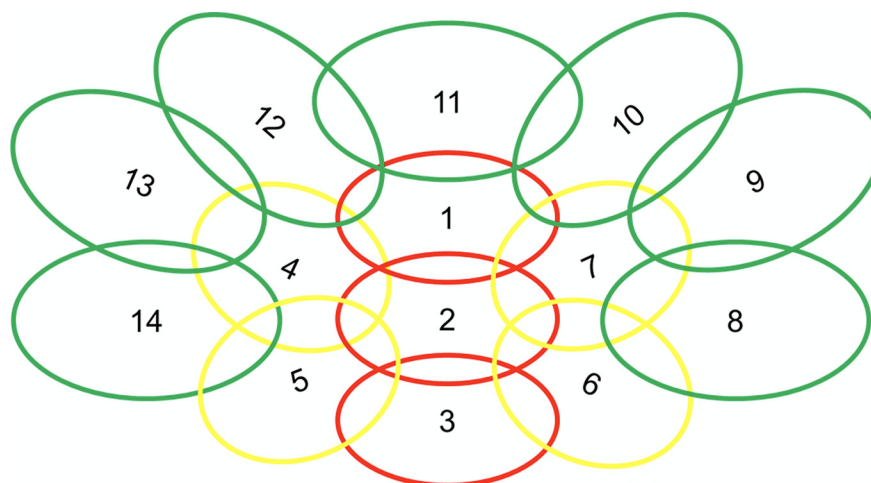
Data and images acquired with the Siemens (human) Tx/Rx 15-channel knee coil were compared with those acquired with the (dog) K9 coil. The K9 imaging coil was designed tailor-made, with special attention to dog head and brain anatomy. The coil is thus composed of 16 linearly polarized receive-only surface channels, 14 of which are mounted inside the coil housing, and two, the "eye-elements," are partly visible from the outside and consist of flexible cables. Each element was segmented in two halves, with tuning, matching and preamplifier (MPB-123R2-90, HiQA, Carleton Place) at one side, and active detuning, passive detuning, and protective fuse at the opposite side of the port. The largest coil elements of the array are elliptical with major axis lengths of 85 and 55 mm, resulting in a circumference of 222 mm. As a rule of thumb, segmentation of coils is recommended below a tenth of the wavelength, which is  $\sim 20$  cm. Therefore, one segmentation in two halves was considered sufficient. The layout of the coil elements (see Fig. 2), and the flexible rostral elements, were also particularly designed for the larger amount of muscle tissue in the dog's skull, i.e., by using seven smaller elements directly above the brain (elliptical with axes lengths  $\sim 4.5 \times 5.5$  cm) and seven larger

**Table 1. Demographic data of dogs included in the coil comparison**

| Name     | Sex | Age | Breed                                     | Weight (kg) | T1    | Functional | RS    |
|----------|-----|-----|-------------------------------------------|-------------|-------|------------|-------|
| Velvet   | F   | 5   | Labrador Retriever                        | 26          | x     | x          | x     |
| Maeva    | F   | 9   | Border Collie and Australian Shepherd Mix | 16          | x     | x          | x     |
| Amy      | F   | 10  | Border Collie                             | 23          | x     | x          | x     |
| Emily    | F   | 12  | Border Collie                             | 16          | x     | x          | x     |
| Linus    | M   | 6   | Australian Shepherd                       | 29          | x     | x          | x     |
| Aeden    | M   | 12  | Border Collie                             | 22.5        | x     | x          | x     |
| Cheyanna | F   | 6   | Australian Shepherd                       | 25          | x     | x          |       |
| Miley    | F   | 10  | Border Collie                             | 16          | x     | x          |       |
| Cameron  | M   | 7   | Border Collie                             | 18          | x     | x          |       |
| Total    |     | 8.1 |                                           |             | N = 9 | N = 9      | N = 6 |

Age indicates age at latest scan. The K9 coil only came into use in 2020, while scanning of the dogs using the knee coil began in 2018. RS = resting-state. All dogs had an 8-min RS run with each coil, except for Linus, who had a 6-min run. F = female, M = male.





**Figure 2.** Schematic of the K9 coil array.

lateral and caudal coil elements (elliptical with axes lengths  $\sim 5.5 \times 7$  cm). However, the coil dedicated to canine imaging not only incorporated a well-adjusted and tailored radio frequency part, but also a careful design of the housing and overall geometry. The design was tailored to optimize the achievable data quality and the comfort of the dog, so as to maximize the chances of a successful measurement. To achieve this, we incorporated an adjustable chin rest, that allowed for individual adjustment of each dog's head in a way that brought it as close to the inner surface of the coil as feasible, allowing for measuring dogs with heads of quite varying sizes (up to 45 cm head circumference), and improving data quality by increasing proximity of the skull to the coil. The coil dimensions were designed with the average size of dogs and dog breeds usually used for neuroimaging in mind, consisting largely of medium sized dogs (mean weight of roughly 20 kg), and a high proportion of Border Collies. This tailored chin rest also increases comfort for the dog, making the lying position adaptable to the individual needs of the subject. This was an improvement from the knee coil, where adjustments of head position had been incorporated using handmade foam chinrests, to increase comfort and minimize distance. Adding such cushion chinrests in the knee coil was also limited by the knee coil already having a higher bottom edge (see Fig. 1D,E), where the addition of further cushions could have resulted in discomfort in some of the dogs. Hence, we abstained from using them in those dogs. Note however that it implied that in some cases, their heads may have been naturally already very close to the coil elements. In the here reported knee coil measurements, four of the dogs had additional chinrests in the resting state scans, three for the visual flickering checkerboard and two in the T1 scans. More chinrests were used in the resting state scans since these were by far the longest, giving more opportunities for run failure and thus experimentation with the addition of further chin rests (this is true for both coils). The foam chinrests were between 1 and 2 cm in thickness, while the average of used chinrests in the K9 coil was 2.3 blocks of 1-cm thickness each. Additionally, the coil is smaller in width than the human knee coil,

allowing the dog to comfortably rest its paws on either side of the coil while its head is inside. An added benefit of higher comfort for the dogs is increased compliance to finish the runs, since dogs will be more reluctant to remain in an uncomfortable setting.

### Visual presentation during scanning

For structural imaging (3:12 min), dogs were either looking at the trainer sitting in front of the scanner or presented with a video engaging their continuous attention (e.g., showing small animals foraging, such as mice or rooks). The latter approach helped the dogs stay still while they could focus on the screen. During resting-state data acquisition, dogs were presented with a white cross on a black background (run durations between 6 and 8 min, see below). The functional task consisted of 10-s blocked presentation of a flickering black and white checkerboard (8 Hz) interspersed with 10-s cross (green on black background). In total, the run was 2:14 min long, including six blocks of visual stimulation and six blocks of baseline in a fixed order, starting with the visual baseline condition.

### Data acquisition

The structural image was obtained using a voxel size of 0.7 mm isotropic [TR/TE=2100/3.13 ms, field of view (FoV)= $230 \times 230 \times 165$  mm<sup>3</sup>]. Functional imaging data for both the flickering checkerboard task and the resting-state data were obtained from 24 axial slices (interleaved acquisition in descending order, spanning the whole brain) using a twofold multiband-accelerated echo planar imaging (EPI) sequence with a voxel size of  $1.5 \times 1.5 \times 2$  mm<sup>3</sup> (TR/TE=1000/38 ms, FoV= $144 \times 144 \times 58$  mm<sup>3</sup>, flip angle=61°, 20% slice gap). The functional flickering checkerboard task consisted of 134 volumes, the resting-state scans were at least 6 min (360 volumes), and at most 8 min long (480 volumes), depending on the dog's capability to lie still for such a prolonged time, without visual input beyond a fixation cross. The structural image was obtained using a voxel size of 0.7 mm isotropic (TR/TE=2100/3.13 ms, FoV= $230 \times 230 \times 165$

mm<sup>3</sup>). Images in these three modalities were acquired in separate sessions. Note that imaging parameters were chosen to be identical for both coils, so that possible differences in image quality could not be attributed to differences in imaging parameters. We used a Siemens Magnetom Skyra with a field strength of 3 Tesla for all measurements.

### Preprocessing

Preprocessing was run in MATLAB version 2020a, using the SPM12 toolbox. Images were slice-time corrected to the middle slice (see Sladky et al., 2011), and realigned. Thereafter, we performed manual reorientation for the structural and EPI images, and proceeded to manually skull-strip the images with itk-SNAP (Yushkevich et al., 2006). This step is of particular importance in dog MRI, where the skull is bordered by massive musculature which can hinder successful coregistration, which was performed onto the mean image of each run. Structural segmentation of the brain was performed using the canine tissue probability maps provided by Nitzsche et al. (2019). Normalization of functional and structural data were performed using the “Old Normalization” module in SPM (originally implemented in SPM8), finally reslicing images to 1.5-mm isotropic voxel size, and smoothing of 3 mm (with a Gaussian FWHM kernel). Data were motion scrubbed by calculating framewise displacement, and excluding volumes with a displacement larger than 0.5 mm in comparison to the previous volume (Power et al., 2012, 2014). Roughly 16 volumes had to be excluded on average in the K9 coil, roughly five volumes in the knee coil (based on flickering checkerboard runs). Since dogs were older when the data were collected with the K9 coil compared with the knee coil, this likely contributed to the increase in motion, as older age is often linked to musculoskeletal issues in dogs. Likewise, the re-training from the known to an unknown coil may also have increased susceptibility for motion.

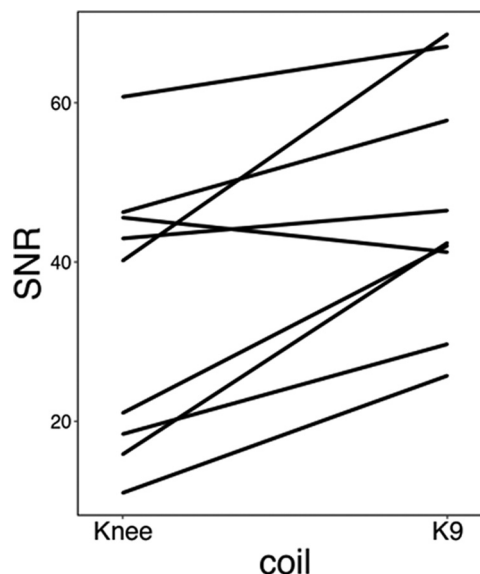
### Data analysis

#### Signal-to-noise ratio (SNR) for structural data

SNR is an important measure of data quality, as it describes the relative contribution of signal of interest versus noise (of no interest) to the overall recorded signal. One major aim of the K9 coil was to improve SNR by improving signal intensity, foremost by reducing distance between the dog’s brain and the coil elements. We calculated SNR for structural images and temporal SNR (tSNR) for functional images (visual flickering checkerboard and resting-state) using the “SPMUP” toolbox (Pernet, 2021). This toolbox defines SNR as the ratio between mean signal intensity in the tissue (gray and white matter) by the signal variance outside of the brain, expressed through the SD, or:

$$\frac{GM_{mean} + WM_{mean}/2}{SD_{nonbrain}}.$$

The tSNR (temporal SNR) was calculated as the voxel-wise mean over all time points divided by the SD over all time points (Murphy et al., 2007). For the calculation of SNR, we used unsmoothed and unwarped data. *t* tests



**Figure 3.** SNR values (AU) for each dog (indicated by different colors) in each coil for the structural data.

and percent differences between coils were calculated using R (version 4.1.0).

#### fMRI data (resting-state and visual stimulation)

Resting-state data were used to calculate subject-specific tSNR maps. Task data were used to estimate the subject-specific BOLD response to visual stimulation using SPM12’s default settings for a first-level single-subject *t* test (task > 0). However, instead of SPM12’s canonical hemodynamic response function (HRF), we used a tailored dog HRF in the analysis of the data (Boch et al., 2021) to account for the faster BOLD response in dogs. The resulting single-subject statistical parametric maps of *t* values were transformed into *z*-values to allow for second-level group analysis. On the group level, we compared tSNR and activation maps between the two coils statistically using paired *t* tests in SPM with a threshold of *p* < 0.05. We used a canine brain atlas (Nitzsche et al., 2019) for parcellation to investigate brain area specific differences.

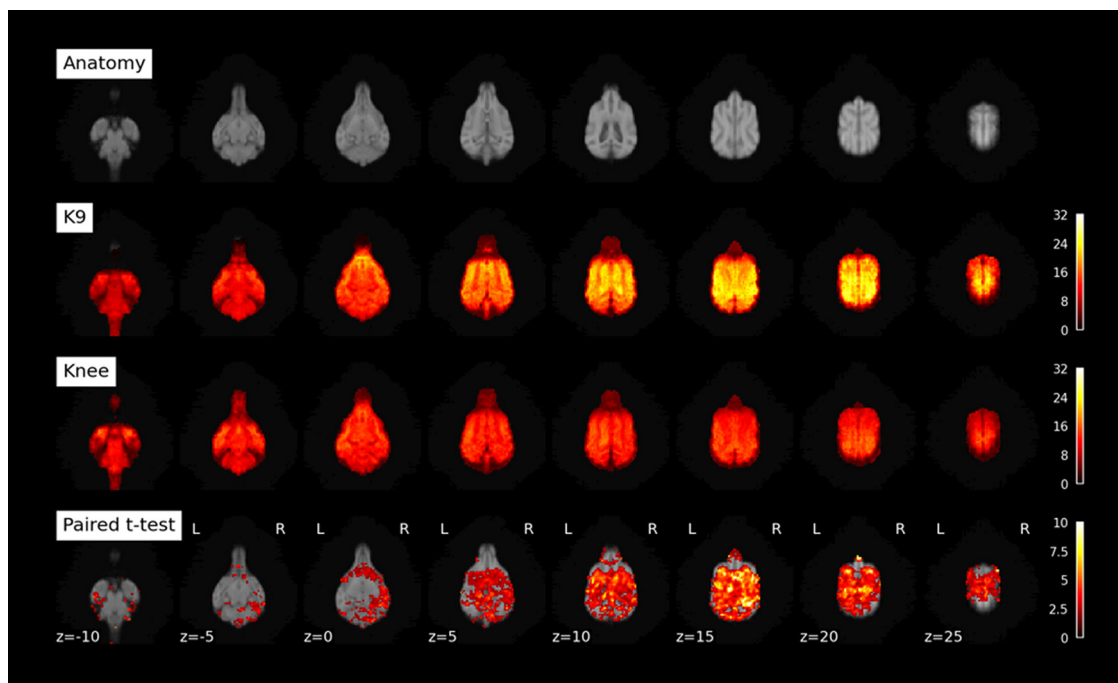
### Data and code availability statement

Data and code can be made available on written reasonable request to the corresponding authors. The SPMup (<https://github.com/CPernet/spmup>) and SPM12 toolboxes (<https://www.fil.ion.ucl.ac.uk/spm/software/spm12/>) are available to the community.

## Results

### T1 data quality/SNR

For structural images (*N* = 9), overall group-averaged SNR was 45.28 a.u. (13.78 SD) for the K9 coil, and 31.66 a.u. (15.22 SD) for the knee coil, corresponding to a 43.01% increase of SNR in the K9 coil compared with the knee coil (see Fig. 3). The difference was significant, with a large effect size (Cohen’s *d* = 0.94 and *t*<sub>(8)</sub> = 3.98,



**Figure 4.** Upper row, Anatomic scan from K9 coil T1 images of six dogs included in resting-state analysis. K9: tSNR maps for resting-state data collected with K9 coil. Knee: tSNR maps for resting-state data collected with knee coil (both unsmoothed data). Paired  $t$  test: contrasting K9 > Knee (smoothed data). Colorbar represents  $t$  values.

$p < 0.01$ ). We also analyzed SNR for gray and white matter separately. For gray matter, SNR increased (K9 > knee) by 47.03% ( $t_{(8)} = 4.3$ ,  $p < 0.005$ ), while it increased by 39.44% for white matter ( $t_{(8)} = 3.68$ ,  $p < 0.01$ ; Cohen's  $d = 1.02$  and 0.87, respectively).

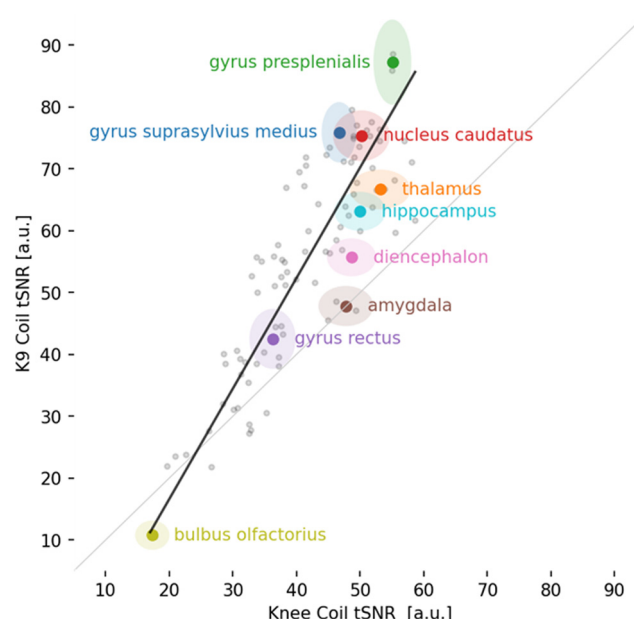
#### Functional neuroimaging: tSNR in resting-state data

We calculated tSNR maps for the K9 and knee coil resting-state data collected in 6 dogs (Fig. 4). The K9 coil shows statistically significant tSNR increases in all dorsal brain regions and most ventral brain areas ( $p < 0.05$ ). No statistically significant tSNR decreases were found. Importantly, no voxels in the knee coil dataset had an increased tSNR when tested with a paired  $t$  test with a threshold of  $p < 0.05$ . To quantify the region-specific tSNR increases we performed a comparison based on mean values from a brain parcellation (Nitzsche et al., 2019). In line with the voxel-based analysis, the overwhelming majority of atlas areas showed a statistically significant increase, while no statistically significant tSNR decreases were found (paired  $t$  test,  $p < 0.05$  one-sided; Fig. 5; Table 2). Importantly, over the whole cortex (see encephalon; Table 2) there was a 46.5% increase in tSNR from knee to K9 coil. Some minor decreases were noted in the olfactory bulb, among a few other regions (see Table 2, negative  $t$  values and discussion). Figure 5 gives an overview of all brain areas and their change in tSNR from the Knee to the K9 coil in the resting-state data.

#### Functional neuroimaging: activation in the visual flickering checkerboard

For the visual flickering checkerboard, we had data from both coils from nine dogs. Contrasting activation to

baseline, we found activation in the visual cortex with both coils (see Fig. 6, top two rows). A paired  $t$  test (bottom row, K9 > Knee) shows areas in which the K9 coil outperformed the knee coil in the visual cortex.



**Figure 5.** Scatterplot of 90 brain areas averaged across all six dogs in the analysis. Almost all brain areas (visualized as gray dots) fall above the gray identity line, hence showing a tSNR increase for the K9 coil. Some important brain areas of interest are color-coded, labeled, and displayed with their 95% confidence intervals.

**Table 2: tSNR differences between K9 and knee coil based on mean regional tSNR for brain parcellations derived from the Nitzsche canine brain atlas (2019)**

| ID | Label                          | K9            | Knee          | Difference | Paired <i>t</i> test                |
|----|--------------------------------|---------------|---------------|------------|-------------------------------------|
| 1  | Encephalon                     | 71.12 ± 9.25  | 48.54 ± 8.37  | 46.50%     | <i>t</i> = 4.0, <i>p</i> = 0.005**  |
| 2  | Gyrus frontalis L              | 21.90 ± 4.51  | 19.69 ± 2.24  | 11.20%     | <i>t</i> = 1.1, n.s.                |
| 3  | Gyrus frontalis R              | 23.59 ± 4.20  | 20.92 ± 2.64  | 12.80%     | <i>t</i> = 1.5, n.s.                |
| 4  | Gyrus proreus L                | 39.34 ± 6.95  | 31.16 ± 5.50  | 26.30%     | <i>t</i> = 2.1, <i>p</i> = 0.043*   |
| 5  | Gyrus proreus R                | 40.70 ± 7.21  | 30.67 ± 6.06  | 32.70%     | <i>t</i> = 2.7, <i>p</i> = 0.020*   |
| 6  | Gyrus compositus rostralis L   | 54.91 ± 8.56  | 38.10 ± 8.90  | 44.10%     | <i>t</i> = 3.2, <i>p</i> = 0.012*   |
| 7  | Gyrus compositus rostralis R   | 50.06 ± 9.65  | 33.84 ± 9.44  | 48.00%     | <i>t</i> = 2.7, <i>p</i> = 0.022*   |
| 8  | Gyrus precruciatu L            | 55.78 ± 17.84 | 33.67 ± 5.60  | 65.70%     | <i>t</i> = 3.2, <i>p</i> = 0.012*   |
| 9  | Gyrus precruciatu R            | 55.03 ± 16.83 | 34.50 ± 6.00  | 59.50%     | <i>t</i> = 3.8, <i>p</i> = 0.006**  |
| 10 | Gyrus postcruciatu L           | 69.47 ± 19.81 | 40.46 ± 8.30  | 71.70%     | <i>t</i> = 4.1, <i>p</i> = 0.005**  |
| 11 | Gyrus postcruciatu R           | 67.02 ± 19.83 | 38.30 ± 8.60  | 75.00%     | <i>t</i> = 4.6, <i>p</i> = 0.003**  |
| 12 | Gyrus marginalis L             | 52.68 ± 12.11 | 32.99 ± 8.33  | 59.70%     | <i>t</i> = 3.3, <i>p</i> = 0.010*   |
| 13 | Gyrus marginalis R             | 57.79 ± 12.95 | 37.08 ± 9.02  | 55.80%     | <i>t</i> = 3.6, <i>p</i> = 0.007**  |
| 14 | Gyrus ectomarginalis L         | 55.83 ± 8.88  | 36.48 ± 5.84  | 53.10%     | <i>t</i> = 4.1, <i>p</i> = 0.004**  |
| 15 | Gyrus ectomarginalis R         | 55.33 ± 8.93  | 37.66 ± 7.40  | 46.90%     | <i>t</i> = 4.3, <i>p</i> = 0.004**  |
| 16 | Gyrus occipitalis L            | 38.46 ± 6.53  | 28.81 ± 5.29  | 33.50%     | <i>t</i> = 2.7, <i>p</i> = 0.022*   |
| 17 | Gyrus occipitalis R            | 40.15 ± 4.62  | 28.53 ± 4.35  | 40.70%     | <i>t</i> = 3.5, <i>p</i> = 0.009**  |
| 18 | Gyrus suprasylvius rostralis L | 73.48 ± 14.66 | 45.25 ± 8.05  | 62.40%     | <i>t</i> = 5.6, <i>p</i> = 0.001**  |
| 19 | Gyrus suprasylvius rostralis R | 70.57 ± 13.12 | 41.48 ± 8.02  | 70.10%     | <i>t</i> = 6.1, <i>p</i> < 0.001*** |
| 20 | Gyrus suprasylvius medius L    | 79.57 ± 12.77 | 48.72 ± 6.19  | 63.30%     | <i>t</i> = 5.6, <i>p</i> = 0.001**  |
| 21 | Gyrus suprasylvius medius R    | 72.25 ± 10.54 | 44.68 ± 6.96  | 61.70%     | <i>t</i> = 6.6, <i>p</i> < 0.001*** |
| 22 | Gyrus suprasylvius caudalis L  | 51.63 ± 12.10 | 42.88 ± 5.37  | 20.40%     | <i>t</i> = 1.8, n.s.                |
| 23 | Gyrus suprasylvius caudalis R  | 51.21 ± 5.75  | 38.21 ± 8.85  | 34.00%     | <i>t</i> = 3.6, <i>p</i> = 0.008**  |
| 24 | Gyrus ectosylvius rostralis L  | 76.31 ± 11.66 | 51.06 ± 6.70  | 49.50%     | <i>t</i> = 5.2, <i>p</i> = 0.002**  |
| 25 | Gyrus ectosylvius rostralis R  | 71.89 ± 9.96  | 41.40 ± 10.64 | 73.70%     | <i>t</i> = 6.7, <i>p</i> < 0.001*** |
| 26 | Gyrus ectosylvius medius L     | 74.88 ± 11.96 | 48.89 ± 6.35  | 53.20%     | <i>t</i> = 4.0, <i>p</i> = 0.005**  |
| 27 | Gyrus ectosylvius medius R     | 67.25 ± 9.21  | 41.17 ± 10.48 | 63.40%     | <i>t</i> = 6.3, <i>p</i> < 0.001*** |
| 28 | Gyrus ectosylvius caudalis L   | 58.50 ± 9.36  | 46.21 ± 5.33  | 26.60%     | <i>t</i> = 2.8, <i>p</i> = 0.020*   |
| 29 | Gyrus ectosylvius caudalis R   | 56.72 ± 7.55  | 41.36 ± 10.41 | 37.20%     | <i>t</i> = 3.9, <i>p</i> = 0.006**  |
| 30 | Gyrus sylvius rostralis L      | 63.97 ± 12.48 | 47.63 ± 8.96  | 34.30%     | <i>t</i> = 2.8, <i>p</i> = 0.019*   |
| 31 | Gyrus sylvius rostralis R      | 64.30 ± 9.62  | 43.36 ± 13.05 | 48.30%     | <i>t</i> = 2.9, <i>p</i> = 0.016*   |
| 32 | Gyrus sylvius caudalis L       | 59.92 ± 9.77  | 49.91 ± 7.59  | 20.10%     | <i>t</i> = 1.9, n.s.                |
| 33 | Gyrus sylvius caudalis R       | 60.01 ± 8.71  | 41.84 ± 11.10 | 43.40%     | <i>t</i> = 3.1, <i>p</i> = 0.013*   |
| 34 | Gyrus compositus caudalis L    | 38.16 ± 6.48  | 37.23 ± 6.71  | 2.50%      | <i>t</i> = 0.3, n.s.                |
| 35 | Gyrus compositus caudalis R    | 38.55 ± 4.61  | 33.68 ± 4.17  | 14.50%     | <i>t</i> = 2.1, <i>p</i> = 0.046*   |
| 36 | Gyrus rectus L                 | 44.63 ± 11.57 | 37.62 ± 10.46 | 18.60%     | <i>t</i> = 1.8, n.s.                |
| 37 | Gyrus rectus R                 | 40.40 ± 12.39 | 34.89 ± 7.38  | 15.80%     | <i>t</i> = 1.3, n.s.                |
| 38 | Gyrus genualis L               | 27.72 ± 5.41  | 26.22 ± 5.31  | 5.70%      | <i>t</i> = 0.6, n.s.                |
| 39 | Gyrus genualis R               | 32.03 ± 8.38  | 28.40 ± 6.68  | 12.80%     | <i>t</i> = 1.3, n.s.                |
| 40 | Area subcallosa L              | 73.57 ± 14.35 | 49.88 ± 10.36 | 47.50%     | <i>t</i> = 2.7, <i>p</i> = 0.022*   |
| 41 | Area subcallosa R              | 76.44 ± 13.69 | 53.11 ± 11.18 | 43.90%     | <i>t</i> = 2.9, <i>p</i> = 0.018*   |
| 42 | Gyrus cinguli L                | 71.84 ± 9.74  | 49.00 ± 5.78  | 46.60%     | <i>t</i> = 4.9, <i>p</i> = 0.002**  |
| 43 | Gyrus cinguli R                | 71.17 ± 10.03 | 47.37 ± 6.24  | 50.30%     | <i>t</i> = 5.7, <i>p</i> = 0.001**  |
| 44 | Gyrus presplenialis L          | 85.93 ± 16.18 | 54.99 ± 6.71  | 56.30%     | <i>t</i> = 4.4, <i>p</i> = 0.004**  |
| 45 | Gyrus presplenialis R          | 88.57 ± 18.25 | 55.16 ± 8.18  | 60.60%     | <i>t</i> = 5.3, <i>p</i> = 0.002**  |
| 46 | Gyrus splenialis L             | 67.80 ± 8.24  | 49.41 ± 4.64  | 37.20%     | <i>t</i> = 3.9, <i>p</i> = 0.005**  |
| 47 | Gyrus splenialis R             | 65.95 ± 6.90  | 48.93 ± 7.33  | 34.80%     | <i>t</i> = 4.1, <i>p</i> = 0.005**  |
| 48 | Gyrus parahippocampalis L      | 56.96 ± 7.63  | 47.19 ± 6.76  | 20.70%     | <i>t</i> = 2.4, <i>p</i> = 0.029*   |
| 49 | Gyrus parahippocampalis R      | 56.60 ± 6.48  | 44.56 ± 9.50  | 27.00%     | <i>t</i> = 2.7, <i>p</i> = 0.020*   |
| 50 | Hippocampus L                  | 63.81 ± 8.23  | 51.85 ± 7.54  | 23.10%     | <i>t</i> = 2.8, <i>p</i> = 0.020*   |
| 51 | Hippocampus R                  | 62.45 ± 7.59  | 48.13 ± 11.91 | 29.70%     | <i>t</i> = 2.7, <i>p</i> = 0.022*   |
| 52 | Lobus piriformis L             | 39.59 ± 5.47  | 37.13 ± 9.25  | 6.60%      | <i>t</i> = 0.6, n.s.                |
| 53 | Lobus piriformis R             | 43.34 ± 7.44  | 37.81 ± 7.50  | 14.60%     | <i>t</i> = 1.4, n.s.                |
| 54 | Tuberculum olfactorium L       | 53.39 ± 14.54 | 38.48 ± 12.15 | 38.80%     | <i>t</i> = 2.4, <i>p</i> = 0.032*   |
| 55 | Tuberculum olfactorium R       | 52.54 ± 14.80 | 37.57 ± 10.68 | 39.90%     | <i>t</i> = 2.3, <i>p</i> = 0.034*   |
| 56 | Gyrus diagonalis L             | 56.42 ± 11.43 | 45.23 ± 12.86 | 24.70%     | <i>t</i> = 1.5, n.s.                |
| 57 | Gyrus diagonalis R             | 60.58 ± 11.06 | 46.82 ± 12.74 | 29.40%     | <i>t</i> = 1.9, n.s.                |
| 58 | Gyrus paraterminalis L         | 74.58 ± 12.52 | 53.10 ± 12.88 | 40.40%     | <i>t</i> = 2.2, <i>p</i> = 0.041*   |
| 59 | Gyrus paraterminalis R         | 74.55 ± 12.98 | 56.88 ± 13.81 | 31.10%     | <i>t</i> = 1.9, n.s.                |
| 60 | Gyrus olfactorius lateralis L  | 52.17 ± 13.00 | 39.86 ± 12.57 | 30.90%     | <i>t</i> = 1.8, n.s.                |
| 61 | Gyrus olfactorius lateralis R  | 51.04 ± 13.72 | 36.55 ± 11.01 | 39.70%     | <i>t</i> = 2.4, <i>p</i> = 0.032*   |

(Continued)



Table 2: Continued

| ID | Label                                              | K9            | Knee          | Difference | Paired <i>t</i> test               |
|----|----------------------------------------------------|---------------|---------------|------------|------------------------------------|
| 62 | Thalamus L                                         | 66.75 ± 8.05  | 53.81 ± 11.14 | 24.00%     | <i>t</i> = 1.8, n.s.               |
| 63 | Thalamus R                                         | 66.59 ± 8.08  | 52.51 ± 12.69 | 26.80%     | <i>t</i> = 1.9, n.s.               |
| 64 | Bulbus olfactorius L                               | 10.99 ± 5.86  | 17.01 ± 6.94  | −35.40%    | <i>t</i> = −1.7, n.s.              |
| 65 | Bulbus olfactorius R                               | 10.57 ± 6.05  | 17.66 ± 6.55  | −40.20%    | <i>t</i> = −2.1, n.s.              |
| 66 | Nucleus caudatus L                                 | 75.36 ± 10.79 | 51.47 ± 10.61 | 46.40%     | <i>t</i> = 3.5, <i>p</i> = 0.009** |
| 67 | Nucleus caudatus R                                 | 75.36 ± 9.77  | 48.97 ± 11.63 | 53.90%     | <i>t</i> = 3.6, <i>p</i> = 0.008** |
| 68 | Insular cortex L                                   | 68.19 ± 12.41 | 55.40 ± 9.17  | 23.10%     | <i>t</i> = 1.7, n.s.               |
| 69 | Insular cortex R                                   | 70.23 ± 12.95 | 51.89 ± 15.36 | 35.30%     | <i>t</i> = 2.1, <i>p</i> = 0.044*  |
| 70 | Hypophysis                                         | 23.82 ± 7.45  | 22.60 ± 6.68  | 5.40%      | <i>t</i> = 0.3, n.s.               |
| 71 | Vermis cerebelli                                   | 44.52 ± 7.29  | 36.69 ± 5.93  | 21.30%     | <i>t</i> = 2.3, <i>p</i> = 0.033*  |
| 72 | Pons                                               | 30.53 ± 2.94  | 35.31 ± 3.63  | −13.50%    | <i>t</i> = −2.4, n.s.              |
| 73 | Medulla oblongata                                  | 27.76 ± 4.16  | 32.83 ± 5.58  | −15.40%    | <i>t</i> = −2.1, n.s.              |
| 74 | Medulla spinalis                                   | 21.87 ± 4.04  | 26.61 ± 5.57  | −17.80%    | <i>t</i> = −1.8, n.s.              |
| 75 | Mesencephalon                                      | 45.48 ± 5.12  | 44.95 ± 6.65  | 1.20%      | <i>t</i> = 0.1, n.s.               |
| 76 | Diencephalon                                       | 55.70 ± 7.70  | 48.68 ± 9.75  | 14.40%     | <i>t</i> = 1.2, n.s.               |
| 77 | Nervus opticus                                     | 38.74 ± 12.28 | 31.93 ± 8.79  | 21.30%     | <i>t</i> = 1.2, n.s.               |
| 78 | Hemispherium cerebelli L                           | 35.54 ± 6.17  | 32.37 ± 5.65  | 9.80%      | <i>t</i> = 1.3, n.s.               |
| 79 | Hemispherium cerebelli R                           | 36.75 ± 4.89  | 31.20 ± 4.28  | 17.80%     | <i>t</i> = 3.1, <i>p</i> = 0.013*  |
| 80 | Commissura rostralis                               | 71.08 ± 10.63 | 58.07 ± 14.46 | 22.40%     | <i>t</i> = 1.4, n.s.               |
| 81 | Pedunculus olfactorius L                           | 31.04 ± 8.41  | 30.13 ± 14.68 | 3.00%      | <i>t</i> = 0.2, n.s.               |
| 82 | Pedunculus olfactorius R                           | 31.34 ± 11.36 | 30.75 ± 10.45 | 1.90%      | <i>t</i> = 0.2, n.s.               |
| 83 | Area septalis L                                    | 77.09 ± 13.20 | 49.41 ± 10.64 | 56.00%     | <i>t</i> = 3.5, <i>p</i> = 0.009** |
| 84 | Area septalis R                                    | 77.54 ± 13.71 | 51.78 ± 12.53 | 49.70%     | <i>t</i> = 3.2, <i>p</i> = 0.012*  |
| 85 | Nucleus et tractus spinalis nervi trigemini L      | 27.28 ± 3.49  | 32.52 ± 7.18  | −16.10%    | <i>t</i> = −2.1, n.s.              |
| 86 | Nucleus et tractus spinalis nervi trigemini R      | 28.66 ± 5.24  | 32.53 ± 6.39  | −11.90%    | <i>t</i> = −1.4, n.s.              |
| 87 | Nucleus ventralis caudalis thalami pars medialis L | 61.75 ± 9.00  | 58.64 ± 13.84 | 5.30%      | <i>t</i> = 0.4, n.s.               |
| 88 | Nucleus ventralis caudalis thalami pars medialis R | 59.68 ± 9.03  | 55.57 ± 14.06 | 7.40%      | <i>t</i> = 0.5, n.s.               |
| 89 | Amygdala L                                         | 47.08 ± 7.24  | 49.34 ± 11.89 | −4.60%     | <i>t</i> = −0.4, n.s.              |
| 90 | Amygdala R                                         | 48.58 ± 8.94  | 46.20 ± 9.44  | 5.10%      | <i>t</i> = 0.5, n.s.               |

*p* values are uncorrected for multiple comparisons and should be regarded as descriptive. L = Left, R = Right. n.s. = not significant. \**p* < 0.05, \*\**p* < 0.01, \*\*\**p* < 0.001.

Furthermore, we looked at individual changes in *z* scores in each voxel in all the nine dogs included in the analysis of the visual flickering checkerboard. Most, but not all, dogs' signal improved with the K9 coil (see Fig. 7), and increases in *z* scores were mostly larger than decreases.

### Movement correlation

Degree of correlation between movement and signal did not differ systematically between the coils (all *p* > 0.05), and neither did raw framewise displacement (all *p* > 0.2).

### Discussion

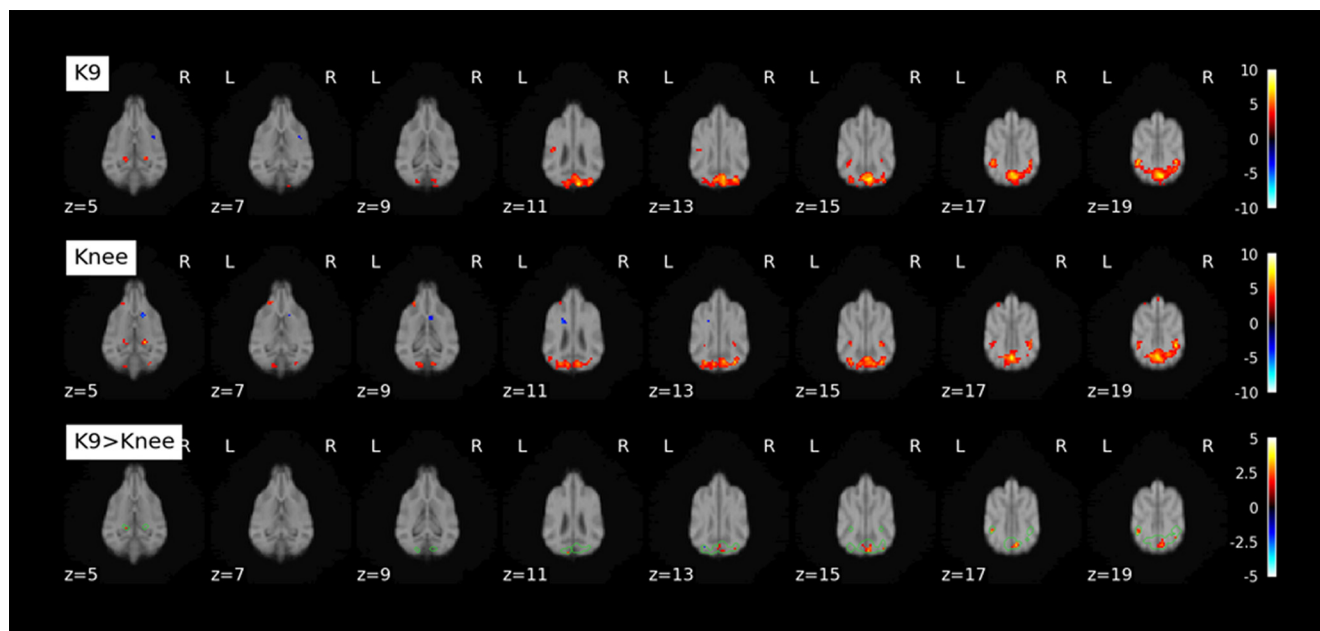
The aim of this study was to validate the new K9 coil across various MRI modalities. To this end, we compared results from the K9 coil with results from a human knee coil, commonly used for dog fMRI. Data were compared in terms of data quality as expressed in SNR, and second-level results in a classical GLM fMRI analysis. Since the design of the K9 coil was tailored to dog cranial anatomy, we expected the K9 coil data quality to outperform the knee coil, and possibly lead to more robust results.

The comparison of the standard human knee coil for dog brain imaging with our inhouse K9 coil has produced a range of evidence that the K9 coil indeed offers higher sensitivity compared with the knee coil. In particular, spatial and temporal signal-to-noise ratios were increased with the K9 coil, across all imaging modalities. In the structural data, we noted an increase of roughly 45%

across gray and white matter. Of note, since the K9 coil came into use later, dogs might have been more trained but also older. The expected increase in SNR (and tSNR) because of better training should however be more than mitigated by increases in signal noise that are observable with increased age (in humans; Yao et al., 2013; McIntosh et al., 2014). When conducting parallel imaging with array coils, nonuniform noise amplification occurs (as was the case only in our structural scans), which can be accounted for in array SNR calculation using the pseudo multiple replica method (Robson et al., 2008) together with gradient echo-based proton-density weighted images and noise-only scans. We were, however, not able to perform these additional scans because of the general constraints of data collection in dogs. Therefore, the reported SNR values for the structural scan comparisons should be regarded with some caution. Future research will be performed to quantify SNR more accurately.

In functional imaging modalities, the differences were also very noticeable. With regard to the resting-state scans, both the knee and the K9 coil can be used for dog fMRI, however the K9 coil was much more sensitive in terms of both SNR and tSNR across the canine brain in our small sample of dogs (*n* = 6 for resting-state). All dorsal brain areas exhibited increases in tSNR in the K9 coil, and this is of particular interest for the investigation of convergent evolution of higher cognition, such as social cognition, since dorsal areas are more likely to contribute to these operations (Rushworth et al., 2013). While Figure





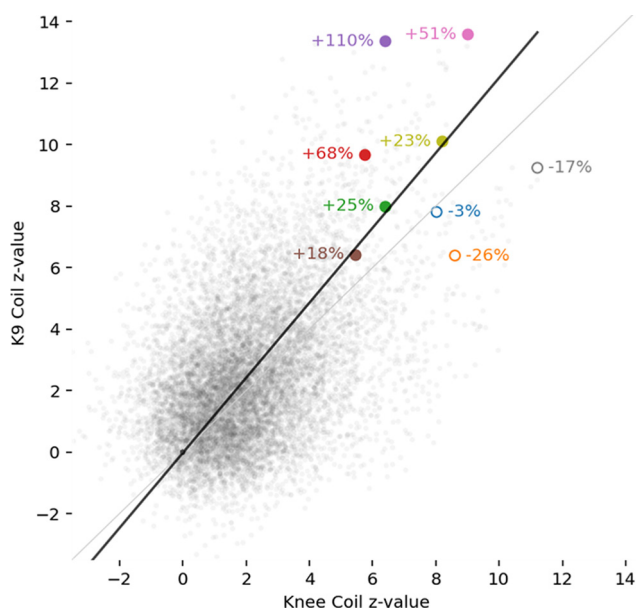
**Figure 6.** K9, Top row, Activation found contrasting Checkerboard > Baseline (fixation cross) with the K9 coil. Knee, Middle row, Activation found contrasting Checkerboard > Baseline (fixation cross) with the knee coil. K9 > Knee, Bottom row, Paired  $t$  test. Green outlines show conjunction of K9 and Knee coil activation. Second-level analysis was performed on the single-subject contrasts and thresholded at  $p < 0.001$  ( $k \geq 5$  voxels for display purposes). Colorbar represents  $t$  values.

5 shows also decreases of tSNR in the K9 coil as compared with the knee coil, most notably in the olfactory lobe, no voxel was found to have statistically better tSNR in the knee coil as compared with the K9 coil. Finally, tSNR in the resting-state modality saw a similar increase as in the structural modality, of roughly 46% from knee to K9 coil. Please note that some of the decreases noted

could also stem from changes in the field of view settings: we had issues with wrapovers in the temporal lobe, and fixed this by tilting the field of view, so that in some dogs, parts of the olfactory bulb might have been cut.

For the visual flickering checkerboard, we found robust activation in the primary visual cortex of dogs with both coils. However, with the K9 coil, a few additional clusters were identified in the paired  $t$  test, in particular in the occipital lobe, as can be seen in Figure 6. On an individual level, not all dogs benefitted from the K9 coil equally, some even had decreases in voxelwise  $z$  scores (Fig. 7). However, fewer individuals exhibited decreases, and the decreases were generally smaller than the increases found in the other dogs. The tSNR increases seen across modalities thus also translated into more activation being detected in highly plausible areas (occipital lobe, primary visual cortex) in a standard second-level GLM analysis of fMRI data in a robust paradigm.

Overall, the strongest evidence in favor of the K9 coil comes from the raw SNR and tSNR increases. These clearly demonstrate that data quality is much improved in the K9 coil. Based on the lack of differences in raw framewise displacement between the coils, this difference does not solely come from a reduction in motion artifacts, on the contrary, dogs seemed to move more in the K9 coil (albeit not significantly more). The improvement in data quality thus rather comes directly from the coil's data acquisition properties, and may be even larger if the added movement (which may as well be a consequence of higher ages, see above) is better accounted for in future work (e.g., by dedicated training). The substantial improvements in SNR do also lead to improvements on the second-level analysis of the functional visual flickering checkerboard



**Figure 7.** Individual  $z$ -values for each voxel in each dog in the data from the visual flickering checkerboard paradigm. Largest individual increases (6 dogs) or decreases (3 dogs) are labeled.

data. Despite the visual flickering checkerboard paradigm being very robust, we were still able to find a multitude of small clusters of brain areas involved in visual processing that were only detected by the K9 coil. The increase in tSNR in all dorsal and most ventral regions with the K9 coil, our new hardware offers the opportunity to investigate smaller effects of interest, which is particularly relevant for the investigation of higher order cognition, as well as social cognition, in dogs and potentially other canines.

The main aim of this study was to examine possible benefits of a dog-tailored MRI coil. We find compelling evidence that the K9 coil will lead to significant improvements in data quality and dog MR imaging. It should be noted though that the K9 coil comes with its own shortcomings because of its high specificity: it is limited to usage in dogs, not all canines, and tailored to a reduced range of breeds in particular. Some larger-skulled dogs will not fit, and for very small dogs the distance to the coil elements might also be too great. But the same would hold true and even more so for the human knee coil.

The K9 coil yields an almost 50% increase in SNR compared with the knee coil, in particular in dorsal cortical areas, across all investigated modalities. With canine neuroimaging as an emerging field, key constraints of small samples and short functional runs emphasize the need for tailored hardware. While existing human imaging hardware will certainly lend valid results as well, especially when robust effects can be expected, the K9 coil offers improved data quality, better subject fit and comfort, and we thus expect it to be a key contribution to the ongoing advancement of dog and canine neuroimaging.

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