REVIEW

The challenges of clinical trials in fragile X syndrome

Sébastien Jacquemont · Elizabeth Berry-Kravis · Randi Hagerman · Florian von Raison · Fabrizio Gasparini · George Apostol · Mike Ufer · Vincent Des Portes · Baltazar Gomez-Mancilla

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Abstract

Rationale Advances in understanding the underlying mechanisms of conditions such as fragile X syndrome (FXS) and autism spectrum disorders have revealed heterogeneous populations. Recent trials of novel FXS therapies have highlighted several challenges including subpopulations with possibly differential therapeutic responses, the lack of specific outcome measures capturing the full range of improvements of patients with FXS, and a lack of biomarkers that can track whether a specific mechanism is responsive to a new drug and whether the response correlates with clinical improvement.

Objectives We review the phenotypic heterogeneity of FXS and the implications for clinical research in FXS and other neurodevelopmental disorders.

S. Jacquemont

Service de Génétique Médicale, Centre Hospitalier Universitaire Vaudois, 1011 Lausanne, Switzerland

E. Berry-Kravis

Departments of Pediatrics, Neurological Sciences and Biochemistry, Rush University Medical Center, Chicago, IL 60612, USA

R. Hagerman

MIND Institute and Department of Pediatrics, UC Davis Health System, Sacramento, CA 95817, USA

F. von Raison · G. Apostol Neuroscience Development, Novartis Pharma AG, 4056 Basel, Switzerland

F. Gasparini · M. Ufer · B. Gomez-Mancilla (⊠) Novartis Institutes for BioMedical Research Basel, Forum 1, Novartis Campus, 4056 Basel, Switzerland e-mail: baltazar.gomezmancilla@novartis.com

V. Des Portes

National Reference Center for Fragile X and Other XLMR, Hospices Civils de Lyon, Université de Lyon and CNRS UMR 5304 (L2C2), Bron, France *Results* Residual levels of fragile X mental retardation protein (FMRP) expression explain in part the heterogeneity in the FXS phenotype; studies indicate a correlation with both cognitive and behavioral deficits. However, this does not fully explain the extent of phenotypic variance observed or the variability of drug response. Post hoc analyses of studies involving the selective mGluR5 antagonist mavoglurant and the GABA_B agonist arbaclofen have uncovered significant therapeutic responses following patient stratification according to *FMR1* promoter methylation patterns or baseline severity of social withdrawal, respectively. Future studies designed to quantify disease modification will need to develop new strategies to track changes effectively over time and in multiple symptom domains.

Conclusion Appropriate selection of patients and outcome measures is central to optimizing future clinical investigations of these complex disorders.

Keywords Fragile X syndrome \cdot Autism spectrum disorder \cdot FMRP \cdot mGluR5 \cdot FMR1 \cdot Disease modification \cdot Mavoglurant \cdot AFQ056 \cdot GABA \cdot Arbaclofen

Introduction

Based on advances in the understanding of the neurobiology of fragile X syndrome (FXS), targeted therapeutic agents designed to correct the underlying mechanisms of neural dysfunction have been assessed in patients. Investigating these new therapies for FXS in clinical trials has led to an increased understanding of the challenges involved in evaluating treatment efficacy in this complex condition. In particular, these early trials have highlighted several challenges: (a) heterogeneity, with the existence of subpopulations based on differing therapeutic response; (b) the lack of reliable biomarkers; and (c) the issue of specific and sensitive outcome measures, particularly in the context of future disease modification trials.

FXS is among the most common known inherited causes of intellectual disability and autism, typically caused by expansion of a cytosine-guanine-guanine (CGG) triplet repeat in the 5' untranslated region of the fragile mental retardation 1 (FMR1) gene. The presence of >200 CGG repeats (full mutation), combined with extensive methylation of the FMR1 promoter, containing the repeat sequence and upstream CpG islands, leads to transcriptional silencing of FMR1 and a complete or partial absence of fragile mental retardation protein (FMRP) (Bell et al. 1991; Sutcliffe et al. 1992). FMRP is a dendritic RNA binding protein that modulates local translation of mRNA at the synapse and postsynaptic density. Dendritic translation, in turn, influences the morphology and functionality of the synapse (synaptic plasticity). Loss of FMRP leads to dysregulation of translation and abnormal neuronal signaling in specific pathways, culminating in the morphological defects and aberrant synaptic plasticity observed in Fmr1-knockout animal models and post mortem brain tissue from individuals with FXS.

In addition to cognitive deficits, individuals with FXS typically present behavioral problems which include a range of anxiety symptoms, attention deficits, hyperarousal, irritability, and autism or autistic-like symptoms including social deficits (Garber et al. 2008). Despite a common genetic etiology, there is a wide-ranging variability in clinical presentation of FXS.

Many of these symptoms are shared with autism spectrum disorders (ASD), and approximately 2–6 % of all cases of autism in males are caused by FXS (Hagerman et al. 2010), dependent on the intelligence quotient (IQ) level of the autism cohort. The prevalence of autism among individuals with FXS is approximately 30 % when using the Autism Diagnostic Observation Schedule/Autism Diagnostic Interview—Revised (Hall et al. 2008; Harris et al. 2008; Kaufmann et al. 2004; Rogers et al. 2001).

The mechanistic link underlying the high prevalence of autism in FXS may be mediated via FMRP and its role in regulating translation of a large number of proteins associated with autism: mGluR5, *N*-methyl-D-aspartate receptor subunits, mammalian target of rapamycin (mTOR), tuberous sclerosis complex 2, fragile-X-related protein 2, and neuroligin-3 (Ascano et al. 2012; Darnell et al. 2011; Iossifov et al. 2012). Many of these genes are associated with synaptic plasticity and function. The molecular pathways underlying the overlap between FXS and autism are complicated and poorly understood. A comprehensive review of the overlap between autism and FXS is beyond the scope of this paper, and several excellent reviews have been published on the topic.

In this review, we will discuss the clinical and molecular heterogeneity of FXS and the implications for clinical research. We will also highlight how future trials will need to carefully consider selection of the most appropriate patients and outcome measures, particularly in the context of evaluating disease modification. We will also consider the wider implications for other neurodevelopmental disorders, and whether what is learned from translational research in FXS can be extrapolated to other etiologies of ASD.

Fragile X syndrome—a clinically heterogeneous patient population

The significant heterogeneity in behavioral and cognitive deficits observed among individuals with FXS is explained in part by variations in residual levels of FMRP. The latter is determined by mosaicism of the CGG expansion size, methylation levels, and X chromosome inactivation. In females, inactivation of one of the two X chromosomes is random from one cell to the next, leading to a differential pattern of FMRP expression within tissues. As a result, the ratio of active normal and full mutation X chromosomes (X-activation ratio) significantly influences the extent of an individual's symptoms; all females with FXS are mosaic by definition. Similarly, in males, variations in the pattern of methylation or size of CGG expansion can result in mosaicism, in which transcriptional silencing of *FMR1* occurs in some but not all cells.

Several studies in both females and males have correlated the severity of intellectual disability with FMR1 activity and FMRP levels (Dyer-Friedman et al. 2002; Loesch et al. 2004; Reiss et al. 1995; Tassone et al. 1999). However, the variance reported in cognitive outcomes for males is also substantially influenced by the home environment, which is not the case for females (Dyer-Friedman et al. 2002; Hessl et al. 2001). Examination of the relationship between cognition and the molecular pathology of FXS has been limited by the sensitivity and floor effects of cognitive testing methods in very low functioning individuals. Only by developing new testing methods and/or algorithms for interpreting results from currently available cognitive measures can the extent of the link with FMRP levels be established. Table 1 provides details of the studies reviewed here. A study in 2009 sought to minimize the limitations associated with standard approaches to cognitive testing in the FXS population, by normalizing scores on the Wechsler Intelligence Scale for Children (WISC-III) (Hessl et al. 2009). Normalized WISC-III scores were based on raw score descriptive statistics from the publisher of the WISC-III (Psychological Corporation, San Antonio, TX, USA) and calculated using an age-dependent z-score transformation. The new scores and usual standardized scores were each correlated with the Vineland Adaptive Behavior Scales, and FMRP levels and the results compared. This analysis reported an enhanced correlation between cognition and residual FMRP expression for the new normalized scoring in contrast to standardized scoring, providing a more accurate assessment of the contribution of FMRP levels to cognitive

| Study | Population | Cognitive measures | FMRP levels/FMR1 activity | Citation |
|--|--|--|--|---|
| Pedigree analysis of children with FXS and unaffected siblings | Aged 6–17 years FXS: 80 M; 40 F Mosaicism: 9 M; 5 F | WISC-III | FMRP levels in peripheral blood determined by Immunocytochemistry | Dyer-Friedman et al. (2002); Hessl et al. (2001) |
| | Non-FXS siblings: 58 M; 62 F | | as % of FMRP-positive lymphocytes | |
| Pedigree analysis of 144 families with individuals affected by FXS | Aged 4–76 years Full mutation ^a : 87 M; 58 F | WAIS-III; WISC-III; WISC-R; WCST; RCFT; BDS | FMRP levels in peripheral blood determined by Immunocytochemistry | Loesch et al. (2004) |
| | Premutation ^b : 32 M; 142 F | | as % of FMRP-positive lymphocytes | |
| | Non-FXS relatives: 114 M; 57 F | | | |
| To specify and measure the relative contributions of genetics and epigenetic characteristics to variance in intellectual functioning | Aged 6–17 years Full mutation: 29 F Non-FXS: 50 F | WISC-R | <i>FMR1</i> activation ratio determined from southern blots of DNA extracted from peripheral lymphocytes | Reiss et al. (1995) |
| Investigation of the relationship between degree of FMRP expression and deficits associated with FXS | Aged 2–60 Full mutation: 19 F Completely methylated: 36 M | Leiter scale; WISC-R; WISC-III; WAIS-R; K-ABC; S-B; BSID; | FMRP levels in peripheral blood determined by Immunocytochemistry as % of FMRP-positive | Tassone et al. (1999) |
| | Partially methylated: 13 M | MDI; VABS ^c | lymphocytes | |
| | Repeat size mosaicism: 12 M | | | |
| Examination of the sensitivity of the WISC-III in FXS | Aged 6–17 years Full mutation: 134 M; 83 F | WISC-III | FMRP levels in peripheral blood determined by Immunocytochemistry | Hessl et al. (2009) |
| | Repeat size mosaicism: 44 M; 12 F | | as % of FMRP-positive lymphocytes | |
| | Methylation mosaicism: 13 M; 1 F | | | |

| Table 1 | Description | of studies | linking | cognition | to FMRP | levels |
|---------|-------------|------------|---------|-----------|---------|--------|
|---------|-------------|------------|---------|-----------|---------|--------|

BDS Behavior Dyscontrol Scale, *F* females, *M* males, *RCFT* Rey Complex Figure Test, *WAIS-III* Weschler Adult Intelligence Scale—Third Edition, *WCST* Wisconsin Card Sorting Test, *WISC-III* Weschler Intelligence Scale for Children—Third Edition, *WISC-R* WISC—Revised, *K-ABC* Kaufman Assessment Battery for Children, *S-B* Stanford–Binet Intelligence test, *MDI* Mental Developmental Index, *BSID* Bayley Scales of Infant Development, *VABS* Vineland Adaptive Behavior Scale

^a Included individuals with repeat size mosaicism, unmethylated full mutation

^b Included two individuals with 40-49 CGG repeats

^c Used when standard IQ test could not be obtained

skills in lower functioning individuals with FXS. Hessl et al. (2001) were among the first to show that behavioral effects correlated with FMRP expression levels. Correlations were the strongest for internalizing problems in females, including withdrawn and anxious/depressed behavior. In contrast, environmental factors were shown to play a significant role in behavioral problems for males (Hessl et al. 2001). In particular, non-pharmacological interventional therapy and the quality of the home environment were linked to fewer behavioral and autistic symptoms (Hessl et al. 2001).

However, much of the variance in FXS phenotype remains unexplained, and most available studies are compounded by the methodology used to determine FMRP levels (Iwahashi et al. 2009; Willemsen et al. 1995, 1997). In the past, FMRP expression has been estimated indirectly using an immunocy-tochemical approach to calculate the percentage of lymphocytes expressing any FMRP, without quantifying the amount of FMRP expressed by the cell (Iwahashi et al. 2009; Willemsen et al. 1995, 1997). More quantitative methods capable of analyzing FMRP levels are required for improving our understanding of the relationship between the molecular pathology of FXS and the clinical phenotype. Furthermore, recent observations report a cross-reaction between the anti-FMRP monoclonal 7G1-1 and the RNA binding protein

Caprin 1 (El et al. 2012), suggesting that anti-FMRP antibodies may not be as exclusive as previously thought. Other *FMR1*-related measures, such as DNA-methylation patterns and the characterization of additional genetic variants at the exome level, using next generation sequencing tools, may provide other means of explaining phenotypic heterogeneity in FXS.

Subgroups of FXS

Although FXS is usually associated with moderate to severe intellectual disability, the syndrome can present as learning difficulties in an individual with an IQ within the low normal or borderline range (70–90); this is most often observed in females. In one study, 50 % of females were placed in the normal or borderline range (i.e., IQ>70) (de Vries et al. 1996). Residual FMRP levels are in part related to the X-activation ratio, potentially influencing the level of intellectual disability in females (Dyer-Friedman et al. 2002; Loesch et al. 2004; Reiss et al. 1995; Tassone et al. 1999). Clinical correlation of FMRP expression levels in lymphocytes and brain cells may be hampered by individual differences in the X-activation ratios for brain and blood.

Higher-functioning (IQ>70) males with FXS also express greater levels of FMRP than those individuals with more pronounced deficits, due to mosaicism. Some of these mosaic males have a proportion of cells containing a premutation expansion (55–200 CGG repeats) and a proportion of cells with a full mutation (size mosaicism). Other individuals have the full mutation in both unmethylated and methylated forms (methylation mosaicism) (Hagerman et al. 1994; Tassone et al. 1999).

Further evidence of clinical subgroups is provided by the characterization of the Prader–Willi phenotype (PWP) (McLennan et al. 2011). PWP occurs in <10 % of individuals with FXS who present with hyperphagia, lack of satiation after meals, and hypogonadism or delayed puberty, but test negative for the 15q11–q13 deletion or uniparental maternal disomy associated with Prader–Willi syndrome. Instead, a study suggests that this subgroup has lowered expression of a gene located on chromosome 15 in the 15q11–q13 region, cytoplasmic FMR1 interacting protein 1 (*CYFIP1*) (Nowicki et al. 2007). CYFIP1 interacts with FMRP and Rac1, linking two processes which underlie synaptic remodeling—cytoskel-etal reorganization and protein translation (Bardoni and Mandel 2002; Schenck et al. 2003; Zarnescu et al. 2005).

FXS has been described in association with other etiologies of intellectual disability, including autism, Down syndrome, Klinefelter syndrome, Turner syndrome, and trisomy X (Hagerman and Hagerman 2002). With the availability of comparative genomic hybridization and whole exome analyses, additional mutations may be uncovered in many more patients with a primary diagnosis of FXS. The heterogeneity in the FXS population and the varying treatment responses observed in recent trials now require new paradigms to design and implement future clinical trials for FXS.

Patient stratification for clinical trials

The appropriate selection of patients is crucial for any clinical research. In the case of FXS, patients have typically been selected based on a positive diagnostic test confirming the expansion of >200 CGG repeats in the *FMR1* promoter. Treatments developed to target the underlying pathophysiology of the syndrome may address symptoms directly associated with FMRP deficits, and their therapeutic efficacy may be dependent largely on the extent of FMRP expression within the patient population. Conversely, with molecular stratification techniques, it may be possible to identify subpopulations of patients with FXS who are more likely to respond to certain treatments with specific molecular targets.

Molecular stratification of patients has been used in the clinical development of the selective mGluR5 antagonist, AFQ056 (mavoglurant). In a proof-of-concept crossover-design study, 30 male patients with FXS, aged 18–35 years, were treated with mavoglurant. A subgroup of patients with a completely methylated *FMR1* promoter region were identified using a bisulfatesequencing based method, more sensitive than the widely used Southern blot analysis. In a post hoc analysis, these individuals showed significant improvements in Aberrant Behavior Checklist—Community Edition (ABC-C) total score (-27.8 vs placebo; *p* <0.001), despite no significant improvements in the overall population (Jacquemont et al. 2011).

Following these results, efficacy studies have been initiated in male and female adults and adolescents with FXS. Molecular profiling is used to enrich the population with completely methylated patients. Trials are designed with two strata: patients who have completely methylated *FMR1* promoter regions and patients with a partially methylated promoter, as assessed using a newly developed DNA methylation assay based on DNA-methylation-specific restriction enzymes and real-time PCR. These trials are ongoing and no data are currently available.

The reasons for the significant response seen in the completely methylated population and the variable response among those in the partially methylated population in the initial study are unknown. Possible factors include the relationship between methylation status and *FMR1* mRNA and FMRP expression (Jacquemont et al. 2011), resulting in a range of severity and varied behavioral and cognitive dysfunction exhibited by the completely methylated groups, relative to the partially methylated group.

As we understand and identify the additional genetic and environmental factors mediating phenotypic variability and therapeutic response to treatments (for example, using techniques such as array-comparative genomic hybridization, imaging genetics, whole genome, and exome sequencing), it may be possible to identify reliable biomarkers and use these to select patients most likely to benefit from specific treatments. This should lead to better clinical trial designs and lower attrition rates for FXS therapies.

Evaluating disease modification

Disease modification can be defined as a normalization or partial normalization of the core mechanism underlying FXS, which translates into a stabilization or improvement in symptoms. Therefore, improvements across multiple symptom domains in FXS (e.g., cognitive, behavioral, and neurological) could be considered as disease modifications. A key challenge to assessing disease modification is identifying an appropriate outcome measure, given the wide range of symptoms observed in this patient population. This is a critical factor that needs to be considered when designing future clinical trials evaluating disease modification.

Currently, it is unclear which measures are the most relevant for these studies. A report by the National Institutes of Health Working Outcome measures group concluded that there is currently no single measure that can effectively evaluate treatment for FXS, based on a review of published data (Berry-Kravis et al. 2013). They emphasized the need for greater consistency in the selection of outcome measures in clinical trials and the identification of a set of core measures for FXS. The group favored a single-composite approach grouping core features of FXS within each symptom domain. Although this would be useful in studies of drugs which target related symptoms, it might not detect improvements in a single sub-domain. Other challenges facing a singlecomposite approach include the variation in symptom presentation associated with age and level of impairment, plus the selection of features to be included in the measure. The ABC-C, expressive language sampling, prepulse inhibition, and neuroimaging were highlighted as promising measures in need of further development. Also, the group recommended testing outcome measures currently validated for other conditions that share core symptoms with FXS for their feasibility and validity in FXS.

As discussed in a recent review (Gross et al. 2012), any outcome measure selected for a clinical trial in FXS must be able to test a broad ability range, overcome problems of cooperation and variable performance, be reproducible and quantifiable, and show improvement in quality of life and function.

Recent trials of both mavoglurant (Jacquemont et al. 2011) and STX209 (arbaclofen) (Berry-Kravis et al. 2012) have looked at changes in the ABC-C and its subscales as clinical endpoints. The ABC-C was developed to assess problem behaviors in children and adults with intellectual disability (Aman et al. 1985) and has been effectively employed in trials for ASD treatments. However, it was unknown if the ABC-C is sensitive enough to detect disease modification in patients with FXS. A version of the ABC-C has been developed with enhanced specificity and sensitivity for FXS, ABC-C for FXS (Sansone et al. 2012) (Fig. 1). In post hoc analyses of the mavoglurant FXS trial, the responder subgroup (patients with a completely methylated *FMR1* promoter region) showed similar significant improvements in both the ABC-C (-27.8 vs placebo; p < 0.001) (Jacquemont et al. 2011) and ABC-C for FXS (-25.61; p < 0.001) (Jaecklin et al., presented at the 13th International Fragile X Conference, July 25–29, 2012, Miami, FL, USA).

Post hoc analyses of the arbaclofen phase II trial in children and adults with FXS using the ABC-C for FXS scale reported significant improvements in the Social Avoidance subscale (-1.2 vs placebo; p=0.01), despite no significant improvement in the other subscales of the ABC-C for FXS (Berry-Kravis et al. 2012). Additional benefit was found in a subgroup of patients with more severe social impairment at baseline (ABC-C Lethargy/Social Withdrawal \geq 8). In these patients, there was a significant improvement in the average Social Avoidance subscale score (-2.2 vs placebo; p=0.04).

Other behavioral rating scales which have been used to evaluate individuals with FXS and have shown sensitivity to change are summarized in Table 2. In the mavoglurant study, treatment benefits within the completely methylated population were captured using the following scales: Visual Analogue scale (VAS) ratings of parent-nominated behaviors, Clinical Global Impression-Severity (CGI-S) scale, Clinical Global Impression-Improvement (CGI-I), CGI efficacy index, Repetitive Behavior Scale-Revised, and Social Responsiveness Scale-Adult Research Version, despite no change in the primary endpoint (Jacquemont et al. 2011). Similarly, VAS ratings showed improvements following arbaclofen treatment in the entire per-protocol cohort, in the absence of an improvement in the primary endpoint, the ABC-C irritability subscale, or in other subscales of the ABC-C (Berry-Kravis et al. 2012). Generally, the scales listed in Table 2 are rarely used as primary outcome measures. However, a phase II trial of minocycline in children and adolescents with FXS used the CGI-I scale as the primary outcome measure, reporting a significant overall improvement (2.49± 0.13 vs 2.97 \pm 0.13 in placebo; p=0.02) (Leigh et al. 2013). Post hoc analyses of the VAS scores categorized according to behavior observed significant changes in VAS ratings of parentnominated anxiety and mood-related behaviors (5.26±0.46 vs 4.05 ± 0.46 in placebo; p=0.05).

The results of these studies suggest that rating scales can detect therapeutic responses in patients with FXS, and appropriate selection of patients may avoid obscuring a treatment response. They also highlight the importance of using methods such as the ABC-C for FXS scale, which have been validated in this patient population and are therefore more

| Irritability | Lethargy | Stereotypy | Hyperactivity | Inappropriate Speech | Social Avoidance |
|--|---|---|---|--|--|
| 2. Injures self | 12. Preoccupied, stares into space | 6. Meaningless, recurring movements | 1. Excessively active at home, school, work | 9. Talks excessively | 5. Seeks isolation |
| 4. Aggressive to others | 20. Fixed facial expression | 11. Stereotyped, repetitive behavior | 13. Impulsive | 22. Repetitive speech | 16. Withdrawn, prefers solitary activities |
| 7. Boisterous | 23. Only sits and watches others | 17. Bizarre in behavior | 15. Restless | 33. Talks to self loudly | 30. Isolates him/herself |
| 8. Screams inappropriately | 25. Depressed mood | 35. Repetitive hand, body, head movements | 28. No attention to instructions | 46. Repeats words/phrase over and over | 42. Prefers to be alone |
| 10. Temper tantrums | 32. Sits/stands in one position for a long time | 45. Waves/shakes extremities repeatedly | 31. Disrupts group activities | | |
| 14. Irritable and whiny | 37. Unresponsive to structured activities | 49. Rocks back and forth | 38. Does not stay in seat during lesson | | |
| 18. Disobedient | | 27. Moves head back and forth | 39. Will not sit still for any length of time | | |
| 19. Yells inappropriately | 43. No word or gesture | | 44. Easily distractible | | |
| 21. Disturbs others | 51. Pays no attention when spoken to | | 48. Constantly runs or jumps | | |
| 24. Uncooperative | 53. Inactive, never moves spontaneously | | 54. Excessively active | | |
| 29. Demands must be met immediately | 55. Responds negatively to affection | | | | |
| 34. Cries over minor annoyances | 56. Deliberately ignores directions | | | | |
| 36. Quick mood changes | 58. Shows few social reactions | Irritabili | - | | |
| 41. Cries/screams inappropriately | 3. Listless, sluggish, inactive | Letharg Stereot | | Original catego | ries |
| 47. Stamps feet/bangs | 26. Resists physical contact | Hypera | | from the ABC-C | |
| objects/slams doors 50. Deliberately | <u> </u> | Inappro | priate Speech | | |
| hurts him/herself 52. Does physical violence to self | | [_] АВС-С | scale factors not i | ncluded in the AB | C for FXS scale |
| 57. Has outburst when doesn't get | | | | | |

Fig. 1 ABC-C for FXS scale. Development of the ABC-C for FXS scale led to the addition of the new Social Avoidance subscale containing specific factors from the ABC-C lethargy subscale

way

sensitive to changes in specific FXS characteristics. Whether or not the ABC-C for FXS scale can detect changes indicative of disease modification is as yet unknown. Even though the ABC-C may not capture improvements associated with clinical treatment and despite the need to develop and use diseasespecific versions, the ABC-C remains the most accepted scale for trials involving patients with developmental delay and behavior issues.

The most appropriate measure for evaluating the extent of disease modification will likely vary according to the age of the patients. It is generally assumed that the younger the patient at the time of treatment onset, the greater the therapeutic benefit may be. This could potentially be assessed by tracking improvements in developmental milestones (e.g., walking, toilet training, and language). Improvements in cognitive function, using study designs investigating literacy and/ or numeracy skills after intensive non-pharmacological interventions (delivered with placebo-controlled trials of medications), may also provide an alternative approach to establish disease modification. However, current cognitive outcome measures (in particular standardized IQ tests) have not been validated or standardized for populations with intellectual disability. These measures have inherent problems, such as floor effects and learning effects (test retest effect), and alternative approaches need to be validated within this patient population. One approach, the Test of Attentional Performance for Children (Testbatterie zur Aufmerksamkeitsprüfung für Kinder, KiTAP), uses a computer-based approach to measure the core executivefunction deficits of attention and inhibition. In a recent pilot study, the KiTAP showed that it could provide reliable and clinically relevant scores in a population of individuals with FXS over a wide range of age and function (Knox et al. 2012).

The appropriate timing of any intervention aimed at disease modification is also important, as it is not yet clear if there are age limits beyond which disease modification is no longer possible. For example, preclinical data suggest that it is possible to rescue the FXS phenotype in adult *Fmr1*-knockout mice following chronic treatment with an mGluR5 antagonist (Michalon et al. 2012), and no age-related effects were reported in the recent trial evaluating arbaclofen in patients with FXS aged 6–39 years (Berry-Kravis et al. 2012). Finally, any study designed to quantify disease modification will need to be multidimensional and longitudinal in order to effectively track the rate of any changes that occur.

Multiple treatments targeting different mechanisms involved in FXS

Mavoglurant was developed to therapeutically block mGluR5, targeting the excessive glutamatergic signaling associated with a lack of FMRP (Levenga et al. 2011). Loss of FMRP and the resultant mGluR5-dependent dysregulation of

synaptic plasticity are thought to contribute to the pathology and symptomatology of FXS (Darnell et al. 2011; Jacquemont et al. 2007; Levenga et al. 2010). This is referred to as the mGluR theory of FXS, which proposes that inhibition of group I mGluR signaling might be a potential therapeutic target in FXS (Bear et al. 2004).

Following the results of studies that showed treatment with mGluR5 antagonists, including mavoglurant, could rescue several synaptic phenotypes in animal models (Choi et al. 2010; de Vrij et al. 2008; Levenga et al. 2011; McBride et al. 2005; Tucker et al. 2006; Yan et al. 2005), it was hypothesized that mavoglurant had the potential to treat the underlying pathophysiology of FXS, thus differing from current pharmacotherapy for FXS which is symptom-driven.

Other agents targeting specific molecular pathways are in development for FXS (Table 3); these include RG7090 (RO4917523), another mGluR5 antagonist; STX209 (arbaclofen, R-baclofen), a γ -aminobutyric acid type B (GABA_B) receptor agonist (Berry-Kravis et al. 2012); and minocycline, a matrix metalloproteinase-9 antagonist (Bilousova et al. 2009; Leigh et al. 2013; Paribello et al. 2010; Utari et al. 2010). All of these agents aim to address the underlying pathology of FXS by targeting specific molecular pathways. However, there are currently no data linking these specific pathways to particular symptoms of FXS. Furthermore, there is a lack of biomarkers related to the mechanism targeted by these new treatments (e.g., synaptic plasticity rescued in animal models) preventing investigators from evaluating how effective these agents are at targeting a particular pathway.

What insights into ASD can be gained from studying FXS?

ASD encompasses an etiology and clinically heterogeneous population, which has posed challenges in the search to find effective treatments. However, recent research shows promise for the mapping of the multitude of aforementioned genetic variants in ASD onto shared pathways. Given the overwhelming degree of locus heterogeneity, finding convergence in specific molecular pathways will be required in order to perform targeted treatment trials with sufficient sample size. Understanding the molecular overlap between FXS and ASD, combined with lessons learned from planning and running clinical trials for FXS, may provide valuable insight into the most appropriate study designs for investigating potential therapies in ASD.

We have previously highlighted the link between FMRP and autism-candidate genes associated with synaptic plasticity (Ascano et al. 2012; Darnell et al. 2011; Iossifov et al. 2012). Lack of FMRP interferes with synaptic plasticity causing disruption of various pathways, for example, up-regulation of mGluR5 and mTOR signaling (Bear et al. 2004; Sharma

| Rating scales ^a | Study | | | | | |
|---|---|--|--|---|--|--|
| | Lithium | Minocycline Owen Jakel (# – 10) | Aripiprazole | Mavoglurant | Arbaclofen | Acamprosate |
| | Open label $(n = 16)$ | Open label $(n - 19)$ Phase II ^b $(n = 66)$ | Open label $(n=12)$ | Phase II $(n=30)$ | Phase II $(n=63)$ | Open label $(n = 12)$ |
| CGI (National Institute of Mental Health 1970) | B 4.7 (0.9) C 1.3 (1.1) N <u>e</u> improved 13 <i>n</i> =0 004 | Open label | B 4.5 (0.5) E 3.5 (0.5) p = 0.008 Effect size 2 | CM subgroup (n = 7) D -1.78 (-2.34 to -1.22) | CGI-S B 5.1 (0.13) E 4.5 (0.12) | CGI-I E 1.9 Ne improved 9 CGLS |
| | | B 4.7 (0.5) C -1.6 (0.8) Me improved 12 <i>p</i> =0.003 | | p < 0.001 | p = 0.09 SSI subgroup $(n = 27)$ CGI-I E 2.5 (0.24) $p = 0.02$ | E 3.33 (0.45) E 3.33 (0.5) p < 0.0001 Effect size 2 |
| | | Phase II CGI-I $(n = 55)$ 2.49 (0.13) p = 0.02 | | | CGI-S B 5.4 (0.22) E 4.4 (0.21) <i>p</i> =0.009 | |
| VAS (Facco et al. 2011) | B 18.3 (13.0) C 22.5 (21.8) Ne improved 12 p = 0.003 | Open label B 19.3 (7.67) C 15.6 (20.25) Ne improved 18 p < 0.001 Phase II ^c Severity of target behavior 2 ($n = 55$) B 2.62 (0.23) E 4.91 (0.31) p = 0.06 Anxiety/mood ($n = 26$) B 2.47 (0.25) E 5.26 (0.46) p = 0.05 Other ^d ($n = 12$) B 3.49 (0.54) E 5.84 (0.54) | | CM subgroup $(n = 7)$ D 31.84 (14.01–49.67) p = 0.006 | Problem behaviors B 2.2 (0.22) E 4.2 (0.32) p = 0.04 | |
| SRS (Constantino et al. 2003) | | 60000 - d | B 124.5 (26.7) E 90.1 (31.6) p < 0.001 | CM subgroup $(n=7)$ D -17.91 (-30.4 to -5.77) | | |

 Table 2
 Other clinician/parental rating scales assessing behavioral symptoms which have shown sensitivity to change in FXS studies

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| Table 2 (continued) | | | | | | |
|---|--|--|--|---|---|--|
| Rating scales ^a | Study | | | | | |
| | Lithium | Minocycline | Aripiprazole | Mavoglurant | Arbaclofen | Acamprosate |
| | Open label $(n=16)$ | Phase II ^b $(n = 15)$ | Open label $(n=12)$ | Phase II $(n=30)$ | Phase II $(n=63)$ | Open label $(n=12)$ |
| RBS-R (Lam and Aman 2007) | | | Effect size 1.3 | p = 0.031 D - 3.81 (-6.91 to -0.70) p = 0.046 CM subgroup $(n = 7)$ D - 9.81 (-16.57 to -3.05) | | |
| VABS-II (Sparrow et al. 1984) | VABS-MAB Baseline 22.9 (6.4) Change -4.4 (4.9) | | | 0000- <i>d</i> | SSI subgroup (<i>n</i> =27) VABS-S B 80.1 (8.1) | VABS-C B 63.4 (10.1) E 66.6 (11.2) |
| | Ne improved 12 $p=0.007$ | | | | E 99.6 (3.38) p = 0.03 | p = 0.03 Effect size 0.32 VABS-EC B 69.8 (23.0) E 78.9 (21.2) p = 0.003 Effect size 0.04 |
| Citation | Berry-Kravis et al. (2008) | Leigh et al. (2013); Paribello et al. (2010) | Erickson et al. (2011) | Jacquemont et al. (2011) | Berry-Kravis et al. (2012) | Erickson et al. (2013) |
| Data presented as either: mean baseline scores (SD) <i>B</i> baseline, <i>C</i> change, <i>E</i> endpoint, <i>D</i> difference, <i>C</i> Visual Analogue Scale of Behavior, <i>SRS</i> Social R Subscale, <i>VABS</i> Communication subsca ^a If just a subscale was sensitive to change the namo ^b Data presented as Least squares mean (SE) ^c Investigators performed an ad hoc analysis of VAS VAS scores for those specific behaviors/symptoms ^d Other category included the following behaviors: b away, noncompliance/defiance, and self injury | Data presented as either: mean baseline scores (SD), mean change (SD), mean endpoint score (SD), or mean difference (90 % CI) <i>B</i> baseline, <i>C</i> change, <i>E</i> endpoint, <i>D</i> difference, <i>CM</i> completely methylated, <i>SSI</i> severe social impairment, <i>CGI</i> Clinical Global Impression, <i>CGI-I</i> CGI of Improvement, <i>CGI-S</i> CGI of Severity, <i>VAS</i> Visual Analogue Scale of Behavior, <i>SRS</i> Social Responsiveness Scale, <i>RBS-R</i> Repetitive Behavior Scale—Revised, <i>VABS-MAB</i> VABS- <i>M</i> and Adaptive Behavior Scale—II, <i>VABSS</i> occialization Subscale, <i>VABSC</i> Ommunication subscale, <i>VABSExpressive</i> Communication Subscale, <i>VABSMAB</i> VABSMaladaptive Behavior Scale—II, <i>VABSS</i> occialization ^a If just a subscale was sensitive to change the name is given in parenthesis ^b Data presented as Least squares mean (SE) ^c Investigators performed an ad hoc analysis of VAS scores by behavior category; they categorized VAS symptoms and defined a combined symptom-specific VAS score as the average of the individual VAS scores for those specific behaviors/symptoms ^d Other category included the following behaviors: being organized, potty training, self-calming/self-soothing, verbal initiation of play, chewing objects, overstuffing, scratching stomach, belching, running away, noncompliance/defiance, and self injury | change (SD), mean endpoint oletely methylated, <i>SSI</i> severe veness Scale, <i>RBS-R</i> Repetit <i>S-EC</i> VABS—Expressive Cc n in parenthesis by behavior category; they ci by behavior category; they ci anized, potty training, self-ca | score (SD), or mean differe e social impairment, <i>CGI</i> C tive Behavior Scale—Revis ommunication Subscale, <i>VA</i> anegorized VAS symptoms a ategorized VAS symptoms a ategorized VAS symptoms a | nce (90 % CI) linical Global Impression, <i>CG</i> ed, <i>VABS-II</i> Vineland Adapti <i>BS-MAB</i> VABS—Maladaptiv <i>BS-MAB</i> VABS—Maladaptiv ind defined a combined sympt initiation of play, chewing obje | 1-1 CGI of Improvement, CGI- ve Behavior Scale—II, VABS- e Behavior Subscale om-specific VAS score as the a com-specific VAS score as the a cts, overstuffing, scratching stor | CGI of Severity, VAS VABS—Socialization verage of the individual mach, belching, running |

| Class | Drug | Action | Study phase | | Published data |
|---------------------------|--------------------------------|---|---------------|--------------|---|
| | | | Completed | Ongoing | |
| Glutamatergic | Mavoglurant (AFQ056) | mGluR5 antagonist | II | II; IIb; III | Jacquemont et al. (2011) |
| | RG7090 (RO4917523) | mGluR5 antagonist | Па | Π | |
| | STX107 | mGluR5 antagonist | None | | |
| | Fenobam (NPL-2009) | mGluR5 antagonist | Open label | None | Berry-Kravis et al. (2009) |
| | Memantine ^a | NMDA receptor antagonist | Open label | None | Erickson et al. (2009) |
| GABAergic | Arbaclofen (STX209/R-baclofen) | GABA _B receptor agonist | Π | III | Berry-Kravis et al. (2012) |
| | Ganaxolone | GABA _A receptor agonist | None | Π | |
| | Acamprosate ^{a,b} | GABA _A receptor agonist, NMDA receptor antagonist; anti-oxidant | Open label | III/II | Erickson et al. (2010); Erickson et al. (2013) |
| Atypical antipsychotics | Aripiprazole ^a | Partial dopamine D2 receptor agonist; serotonin 5-HT _{1A} agonist; SSRI (serotonin 5-HT _{2A} antasonist) | Open label | None | Erickson et al. (2011) |
| Antidepressant/anxiolytic | Sertraline ^a | SSRI (serotonin 5-HTA antagonist) | Open label | II | Indah Winami et al. (2012) |
| Mood stabilizer | Lithium ^a | Phospholipase C inhibitor; GSK-3 inhibitor | Open label | None | Berry-Kravis et al. (2008) |
| Antibiotic | Minocycline ^a | MMP9 inhibitor | Open label II | None | Paribello et al. (2010); Leigh et al. (2013) |

Table 3 Drugs in clinical development for treating FXS; drug class, action, study phase, and published data

mGluR metabotropic glutamate receptor, GABA_B γ -aminobutyric acid type B, MMP9 metalloproteinase 9, NMDA N-methyl-D-aspartate, 5-HT_{1A} 5-hydroxytryptamine 1A, GSK-3 glycogen synthase-3, SSRI selective serotonin uptake inhibitor

^a Drug already approved by FDA for other indications

^b Mode of action still unclear

et al. 2010) and down regulation of the GABA and dopamine systems (D'Hulst and Kooy 2007; Wang et al. 2008).

The glutamate and GABA pathways have both been implicated in ASD by the use of Fmr1 KO mice as a model system for studying autistic behaviors (Rogers et al. 2013) and the increasing evidence of altered expression of mGluR5, FMRP, and GABA receptors in individuals with autism (Fatemi et al. 2010, 2011; Fatemi and Folsom 2011). Agents targeting these two pathways in FXS (Table 3) may therefore be useful in ASD. Research using models of FXS to elucidate converging molecular pathways behind autistic behaviors might identify novel therapeutic targets for ASD. However, treatments targeting these pathways could have different outcomes for individuals across the ASD spectrum despite similarities in clinical presentation. To illustrate, tuberous sclerosis complex (TSC) and FXS share clinical characteristics (including ASD) and are caused by haploinsufficiency of TSC1 and TSC2 associated with the regulation of protein synthesis at the synapse (for review, see Orlova and Crino 2010). In vivo data indicate that the synaptic dysfunction observed in mouse models of FXS and TSC falls at opposite ends of the physiological spectrum (Auerbach et al. 2011; Bateup et al. 2011) and can be rescued with agents that modulate mGluR5 in opposite directions or by crossing the mouse strains (Auerbach et al. 2011).

Given the wide range of symptoms observed in the ASD patient population, clinical trials in ASD face similar problems to trials in FXS in regard to patient selection and choice of outcome measures. The development of biomarkers stratification and endophenotyping methodologies in FXS trials could be adapted for ASD trials. Likewise, new or modified measures (e.g., ABC-C for FXS) that can effectively track improvements in symptoms may also be transferable to studies in ASD, but their feasibility across the ASD spectrum will need to be tested. Because of the prevalence of autism in FXS

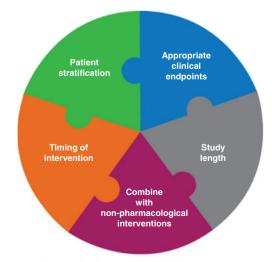


Fig. 2 Key elements for future clinical trials in FXS

and shared neurophysiology with ASD, FXS can be viewed as a model for autism.

Conclusions

Advances in understanding the neurobiology of "monogenic" syndromes such as FXS have revealed heterogeneity at the level of phenotype, manifestations of the causative mutation, and drug response. Complex disorders such as ASD are further complicated by increasing evidence for a heterogeneous etiology and mechanism of disease, unlike FXS which is caused by a mutation within a single genetic locus, the *FMR1* gene. Current research is focused on identifying common therapeutic targets among patients with different molecular etiologies. The development of novel treatments for specific molecular targets for these disorders has the potential to rescue specific phenotypes and may result in what can be classified as disease modification.

Developing biomarkers may aid patient stratification for clinical trials and predict response to treatments. In addition to patient stratification methods, future clinical trial designs will also need to consider appropriate endpoints and length and timing of interventions (Fig. 2). Ultimately, it may be possible to stratify patients for genetic risk factors associated with neurodevelopmental disorders such as FXS and ASD, to enable early implementation of customized therapeutic interventions that could normalize brain development and optimize clinical outcomes.

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ongoing clinical trials with Novartis Pharma AG and F. Hoffman-La Roche. He also has acted as a consultant for Novartis Pharma AG and received reimbursement for travel expenses. B Gomez-Mancilla, F von Raison, F Gasparini, M Ufer, and G Apostol are employees of Novartis Pharma AG and hold shares with Novartis Pharma AG. F Gasparini and B Gomez-Mancilla have also received reimbursement from Novartis Pharma AG for travel expenses, and the spouse of G Apostol is an employee of Novartis International AG.

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