



# Review Shedding the Light on the Natural History of Intracranial Aneurysms: An Updated Overview

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**Abstract**: The exact molecular pathways underlying the multifactorial natural history of intracranial aneurysms (IAs) are still largely unknown, to the point that their understanding represents an imperative challenge in neurovascular research. Wall shear stress (WSS) promotes the genesis of IAs through an endothelial dysfunction causing an inflammatory cascade, vessel remodeling, phenotypic switching of the smooth muscle cells, and myointimal hyperplasia. Aneurysm growth is supported by endothelial oxidative stress and inflammatory mediators, whereas low and high WSS determine the rupture in sidewall and endwall IAs, respectively. Angioarchitecture, age older than 60 years, female gender, hypertension, cigarette smoking, alcohol abuse, and hypercholesterolemia also contribute to growth and rupture. The improvements of aneurysm wall imaging techniques and the implementation of target therapies targeted against inflammatory cascade may contribute to significantly modify the natural history of IAs. This narrative review strives to summarize the recent advances in the comprehension of the mechanisms underlying the genesis, growth, and rupture of IAs.

**Keywords:** endothelial disfunction; inflammatory cascade; intra-aneurysmal flow; intracranial aneurysm; subarachnoid hemorrhage; wall shear stress

# 1. Introduction

Intracranial aneurysms (IAs) are life-threatening cerebrovascular pathologies with an incidence and prevalence of 1-6% and 3.2%, respectively, in the adult population [1].

IAs are focal enlargements of the arterial wall owing to the destruction of internal elastic lamina and tunica media. The sudden rupture of these thin-wall regions causes a subarachnoid hemorrhage (SAH), the latter accounting for more than 25% of strokes [2–4].

The occurrence of the IAs is related to genetic, hemodynamic, and inflammatory factors, which are responsible for the bulging of the arterial wall and, ultimately, its rupture. The precise molecular mechanisms underlying these events, however, remain unclear, although their understanding may theoretically lead to counteract this cascade.

In this study, we overview the most recent advances in the comprehension of the pathogenesis of IAs, focusing on the implications of gene mutations, inflammation pathways, and hemodynamic stress in the growth and rupture of the aneurysm. The role of



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**Copyright:** © 2021 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (https:// creativecommons.org/licenses/by/ 4.0/). neuroimaging in detecting aneurysm wall inflammation as well as that of target therapies in preventing rupture are also discussed.

#### 2. Genesis of Intracranial Aneurysms

#### 2.1. Genetic and Extrinsic Risk Factors

Over the last decade, several studies have delineated the structural differences between sporadic aneurysms and hereditary ones. Among these is the Familial Intracranial Aneurysm (FIA) study, a multicentric retrospective and prospective trial focused on the genome reconstruction of 475 families harboring IAs [5]. Results revealed that familial IAs is a dissimilar entity, with a higher incidence at a young age and a predilection for the middle cerebral artery (MCA). They are frequently large and multiple, and the patterns of inheritance include autosomal dominant transmission and anticipation [6–10].

Furthermore, many genetic syndromes were found to be susceptible to IA growth. Ehlers–Danlos syndrome type IV is a connective tissue disorder caused by the mutation of the COL3A1 gene, which transcripts for type III procollagen. The abnormal synthesis of collagen results in vascular fragility and then IA formation [11,12]. In patients affected by adult polycystic kidney disease, the risk of IAs increased by 40%, of which 25% were multiple [13,14]. The Floating-Harbor, an autosomal dominant genetic syndrome, is characterized by the SRCAP mutation. SCARP is an SNF2-related chromatin-remodeling ATPase protein, involved in endothelial integrity and repairing mechanisms [15]. Other rare hereditary diseases are strongly related to IA growth, such as neurofibromatosis type I, Marfan's syndrome, hereditary hemorrhagic telangiectasia, and alpha-1-antitrypsin deficiency [16–20].

Conversely, the sporadic aneurysms occurred more frequently in females who were older than 50 years. The higher occurrence during the postmenopausal period is due to the loss of the anti-inflammatory role of estrogens. The main extrinsic risk factors for IAs are cigarette smoking, arterial hypertension, hypercholesterolemia, alcohol consumption, drug abuse, and oral contraceptives. All of these increase the oxidative stress and activation of the inflammatory cascade, resulting in chronic arterial damage [21].

In addition, abnormalities of specific chromosomal loci are linked to IA genesis. Genome-wide linkage analysis reported a higher incidence of IAs currently within the mutation of TP53, antisense inhibitor gene/chromosome 9, cyclin-dependent kinase inhibitor 2B, G572C gene of IL-6, and mutation of SOX17/chromosome 8 [22–26].

Particularly, the SOX17 factor is involved in physiological vascular remodeling and arterial homeostasis. SOX17-deficient rat models treated with angiotensin II (Ang-II) infusion were proven to be more susceptible to IAs formation [27]. Conversely, in 2014, Peña Silva et al. reported that the expression of angiotensin 1–7 (Ang-1–7), which binds Mas receptors and acts against the hypertensive effects of Ang-II, inhibits the aneurysmal growth. They induced IAs in wild-type and Mas receptor-deficient rats through Ang-II infusion and intracranial elastase injection. The study group was also treated with Ang-1–7. Results showed a decreased IAs formation and rupture in the study group. These data support the protective role of Ang-1–7 with a Mas receptor-dependent mechanism [28].

Moreover, the T786C polymorphism of the nitric oxide synthase (NOs) gene reduces the vasoprotection mediated by nitric oxide (NO), and the mutation of the endothelin receptor A gene leads to a dysregulation of the arterial tone. Both of these conditions significantly increase the risk of IAs [29,30].

In 2007, Pimiento and colleagues investigated the role of endothelial NOS (eNOS) in IAs formation and growth. They compared aneurysmatic diameters in wild-type and eNOS-knockout mice and determined that the eNOS expression is related to the enlargement of the IAs wall [31].

The same year, Abruzzo et al. conducted a study on rat brain circulation specimens genetically modified with NOS-2, NOS-3, and plasminogen-activator inhibitor (PAI)-1 knockout and treated with left common carotid artery ligation. After histological analysis,

two IAs were found in NOS-3-knockout rats and none in NOS-2-knockout or PAI-1-knockout mice, demonstrating that only the former is a potential predisposing factor [32].

#### 2.2. Wall Shear Stress, Endothelial Dysfunction, and Role of Inflammation

Among the hemodynamic mechanisms of brain blood circulation, the wall shear stress (WSS) is the result of the tangential force impressed on the arterial walls by the pulsatile blood flow, influenced by the fluid viscosity and velocity. The endothelial function is regulated by mechanoreceptors, sensitive to the decrease or increase in WSS gradient. Both low and high WSS cause functional and morphological dysregulation in endothelial cells, especially at arterial bifurcation sites, triggering the formation of aneurysms [33–36].

The so-called endothelial dysfunction, due to the hemodynamic stress, results in matrix protein degradation, the production of oxidant and vasoconstrictive agents, inflammatory activation, leukocyte rolling, and adhesion, followed by vessel remodeling [37–39].

The primum movens of the inflammatory cascade is the transcription of NF- $\kappa$ B, which induces the expression of COX2, prostaglandin E2, and proinflammatory genes [40].

The proinflammatory signals prompt the secretion of chemokines, such as the monocyte chemoattractant protein-1 (MCP-1) and vascular cell adhesion molecule-1 (VCAM-1), which recruit leukocytes in the arterial wall. Endothelial-infiltrating lymphocytes and macrophages amplify the expression of inflammatory mediators, such as matrix metalloproteinases (MMPs), interleukins, cytokines, and the tumor necrosis factor- $\alpha$  (TNF- $\alpha$ ) [41–43].

The importance of inflammation in IAs progression is also demonstrated by experiments on animal models. In 2017, Aoki and colleagues studied the timing of NF-kB, PGE2, and prostaglandin E receptor subtype 2 (EP2) activation, and the consequent macrophage infiltration in the IAs wall. EP2 or NF-kB-knockout mice have shown less immune activation along the vessel walls and reduced IAs growth [44].

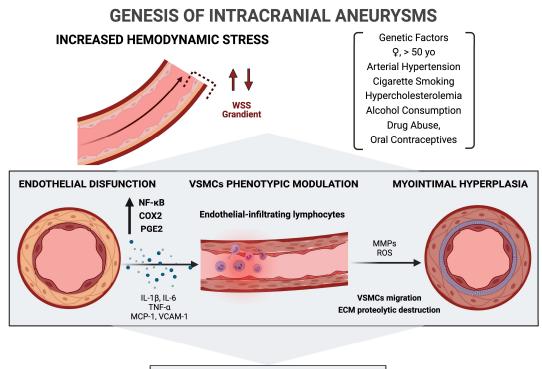
In 2014, Starke highlighted the involvement of TNF- $\alpha$  in IAs progression and rupture. They induced IAs in TNF- $\alpha$ -knockout mice subsequently treated with TNF- $\alpha$  inhibitor 3,6' dithiothalidomide (DTH), and results showed a decreased incidence of IAs formation [45].

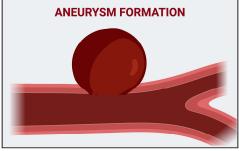
Furthermore, toll-like receptors 4 (TLR4) contribute to the maintenance of immune response. In 2020, Mitsui et al. demonstrated that TLR4-knockout mice, after aneurysmal induction, have a lower incidence of aneurysmal rupture than the control littermates [46].

These data support the role of the inflammatory signaling pathway in IAs pathophysiology.

The stochastic amplification of the inflammatory cascade provokes a phenotypic switching of the vascular smooth muscle cells (VSMCs) from a contractile phenotype to a secretory one [47–50]. Modified VSMCs are spider-like shaped, split, and incapable of excreting collagen for the extracellular matrix (ECM). Proinflammatory VSMCs also upregulate the NF- $\kappa\beta$ , IL-1 $\beta$ , TNF- $\alpha$ , and MMPs, leading to ECM transformation, proteolytic destruction, and thinning of the tunica media [51–54]. VSMCs move to the intima layer and proliferate, resulting in intimal degeneration, namely myointimal hyperplasia [55,56].

The oscillatory gradient of WSS, endothelial dysfunction, and VSMCs phenotypic modulation progressively stretch and destroy the arterial wall, contributing to IA genesis (Figure 1).

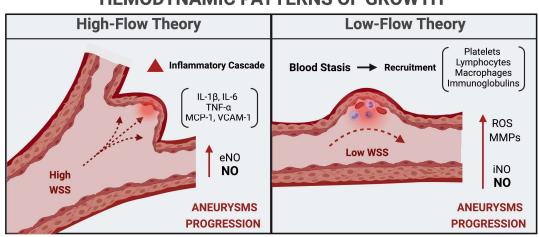




**Figure 1.** Hemodynamic and molecular mechanisms of intracranial aneurysms formation. COX2, cyclooxygenase-2; ECM, extracellular matrix; IL:, interleukin; MCP-1, monocyte chemoattractant protein-1; MMP, matrix metalloproteinase; PGE2, prostaglandin E2; ROS, reactive oxygen species; TNF- $\alpha$ , tumor necrosis factor- $\alpha$ ; VCAM-1, vascular cell adhesion molecule-1; VSMC, vascular smooth muscle cell; WS, wall shear stress.

## 3. Hemodynamic Phenotypes and Patterns of Growth

Two hemodynamic models support the role of WSS in IA progression, namely the high-flow theory and low-flow one [35]. According to the high-flow theory, the abnormal increase in WSS causes focal endothelial injuries and structural changes [57]. Prolonged biomechanical stimulation stretches collagen and elastic fibers, leading to gradual dilatation and enlargement of the IA wall. Moreover, the pressure load activates the eNOS, promoting a local overproduction of NO that acts as a strong vasodilator [58–60]. The low-WSS theory hypothesizes the occurrence of a slowing of the blood flow in the arterial lumen, which facilitates the recruitment of platelets, lymphocytes, macrophages, and immunoglobulins [61]. The degranulation of mast cells and macrophages stimulates the inducible nitric oxide synthase (iNOS) to activate NO and induce the upregulation of MMPs and reactive oxygen species (ROS) [62–64]. The oxidative stress and tissue-infiltrating inflammatory mediators promote endothelial damage, VSMC phenotypic change, and thrombosis, all of them creating the assumption of cell apoptosis and rupture of the aneurysm wall [35,65–67] (Figure 2).



**Figure 2.** Hemodynamic patterns of intracranial aneurysms growth. eNOS, endothelial nitric oxide synthase; IL, interleukin; iNOS, inducible nitric oxide synthase; MCP-1, monocyte chemoattractant protein-1; MMP, matrix metalloproteinase; NO, nitric oxide; ROS, reactive oxygen species; TNF- $\alpha$ , tumor necrosis factor- $\alpha$ ; VCAM-1, vascular cell adhesion molecule-1; WSS, wall shear stress.

The aforementioned flow patterns lead to two different IA hemodynamic phenotypes [68]. The progressive positive WSS creates a continuous increase in transmural pressure, driving the initiation of small, thin-walled, and translucent IAs. Otherwise, the aberrant blood stagnation over-activates inflammatory pathways and atherosclerotic-like processes at the arterial wall, which result in the formation of large, thick-walled, and thrombosed IAs.

### 4. Risk of Rupture

### 4.1. Histopathological Findings

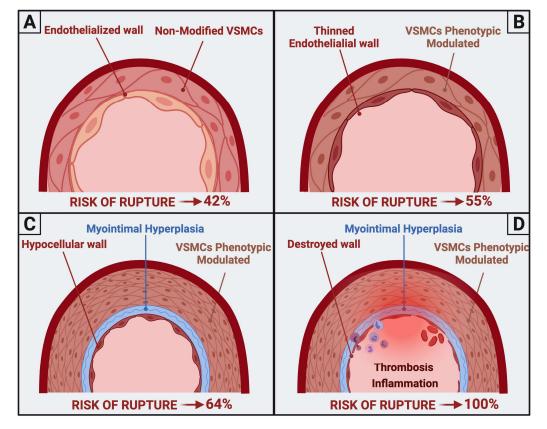
Specific histological hallmarks have been reported for the IA wall: the disruption of the tunica media and their replacement with modified non-contractile VSMCs, loss of the fibers of the internal elastic lamina, myointimal hyperplasia, thinned and shattered ECM, and proinflammatory endothelial cell dysfunction [69–71].

In 2004, Frosen and colleagues described four aneurysmal wall histological subtypes with a progressively increased risk of rupture. Type A is marked by an endothelialized wall and non-modified VSMCs; type B harbors modified VSMCs and a thin endothelial wall; type C is characterized by the hypocellular wall, modified VSMCs, and myointimal hyperplasia; type D involves hypocellular and destroyed wall with traces of thrombosis [72] (Figure 3). The reported average risk of rupture is 42%, 55%, 64%, and 100% for type 1, type 2, type 3, and type 4, respectively [72].

#### 4.2. Size, Angioarchitectural Features, and Intra-Aneurysmal Flow

In 1998, the International Study of Unruptured Intracranial Aneurysms (ISUIA) analyzed the natural history of 2621 IAs, describing a rupture rate of 0.05%/year for IAs smaller than 10 mm and 1%/year for larger ones [62]. In 2003, the prospective part of the ISUIA trial established the cut-off size at 7 mm [3]. Later, Morita et al. confirmed that the risk of rupture progressively increases based on the size, and it becomes relevant over 7 mm [73]. They also found a different likelihood of rupture according to the site, with a risk of 2%, 1.7%, 1.3%, 0.65%, and 0.2% for the basilar tip, posterior communicating, anterior communicating, MCA, and paraclinoid internal carotid artery, respectively [73].

# HEMODYNAMIC PATTERNS OF GROWTH



# HISTOLOGICAL SUBTYPES OF INTRACRANIAL ANEURYSMS

**Figure 3.** Histological subtypes of intracranial aneurysms and risk of rupture. (**A**) Type A; (**B**) type B; (**C**) type C; (**D**) type D.

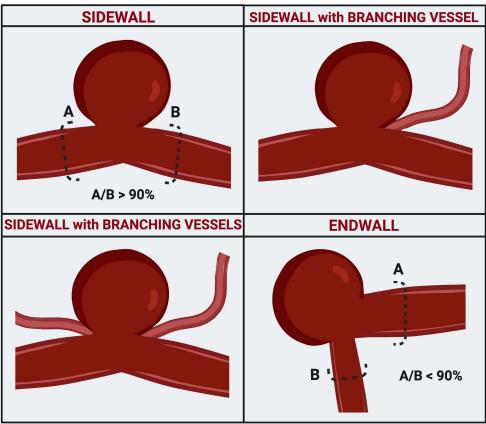
The explanation of these data lies in the distribution of hemodynamic forces in the different locations of IAs. Based on rheologic properties and relationships with neighboring arteries, the saccular aneurysms were classified as sidewall (SWA), sidewall with branching vessels (SWBVA), and endwall (EWA) [74]. The SWAs are the focal outpouching of one artery where the percentage ratio between the pre- and post-aneurysmal segment is >90%. The SWBVAs are similar but hold smaller side branches. In EWAs, the so-called bifurcation aneurysms, the pre- and post-aneurysmal segments, have a different diameter at the neck, and their percentage ratio is <90% (Figure 4).

Angiographic computational flow techniques allows us to comprehend the intraaneurysmal hemodynamic patterns and flow velocity in the different regions of the IA.

The low WSS proved to be responsible for the rupture of SWAs at the neck or dome [75–77]. Blood stasis has been identified as the major input to the atherosclerotic-like wall remodeling and formation of intraluminal thrombi via the recruitment of platelets and inflammatory mediators [76,78–80].

Conversely, high-WSS regions represent the breaking points for EWAs and SWBVAs. Depending on the angioarchitecture, the points of pressure impingement may mainly involve bifurcations, narrow necks, or the take-off of collateral branches [81,82].

These findings demonstrated a close relationship between the risk of rupture and angioarchitecture. The diameter of the neck, aspect ratio (dome/neck ratio), size ratio, height-width ratio, flow angle, and presence of one or more collateral branches all affect the risk [75,83,84].



# GEOMETRICAL FEATURES OF INTRACRANIAL ANEURYSMS

Figure 4. Angio-architectural classification of intracranial aneurysms.

In addition, age older than 60 years, female gender, hypertension, cigarette smoking, alcohol abuse, and hypercholesterolemia seem to contribute to the growth and rupture of the aneurysm [85–87]. In 2012, Greving et al. introduced a grading system with the aim to predict the five-year risk of rupture for IAs. The PHASES score includes patients' demographics, clinical data, size and site of aneurysms, and the occurrence of previous bleeding. A PHASES score rating >3 is highly suggestive of rupture and provides an indication for treatment [88,89].

### 5. Future Perspectives: Imaging of Inflammation and Target Therapies

In recent decades, many efforts have been focused on novel diagnostic tools for the rupture risk stratification and treatment choice. In 2012, Hasan et al. proposed ferumoxytolenhanced magnetic resonance imaging (FE-MRI) to predict aneurysm instability. The ferumoxytol, an intravenous iron oxide employed in anemia and renal failure, is uptaken by macrophages and lymphocytes with a peak of 24 h. The IA wall enhancement allows the estimation of the inflammatory process at the arterial wall, which reflects the active progression of aneurysmal remodeling and demolition. Therefore, the FE-MRI can predict the IA instability and improve the identification of rupture-prone aneurysms [90–92]. The identification of endangered IAs permits us to choose the most appropriate, tailored management.

As reported by our group, the recent advances in translational medicine, epigenetics, and immunogenomics in many neurosurgical fields have led to the proposal of new therapies targeting the inflammatory cascade [93–99].

The most promising approach involves the chronic use of acetylic acid (ASA) as the primary prevention for IA rupture. ASA not only irreversibly blocks the cyclooxygenase (COX)-1 and -2 but also the ROS formation, resulting in a decrease in the inflammatory cascade. In 2011, the same group of Hasan and colleagues reported the results of a

comparative study on patients enrolled in the ISUIA trial and treated with ASA three times per weekly or daily. The overall data suggested a lower risk of SAH in the ASA group [100].

Statins were tested in preventing IA rupture, exploiting their ability to block upstream the inflammatory cascade. They inhibit the NF- $\kappa$ B activation, MMPs, and iNoS expression. In 2012, Tada and colleagues proved the dose-dependent effects of statins in the treatment of IAs. Particularly, a low dose of Pravastatin (5 mg/kg per day) was effective in avoiding the IA rupture, while a high dose of Pravastatin (50 mg/kg per day) may increase cell apoptosis and arterial wall damage [101].

COX-2 inhibitors, angiotensin receptor blockers, and mast-cell degranulation inhibitors are still under evaluation as tailored, preventive therapies to reverse the process of growth and rupture [102–106].

#### 6. Conclusions

The hemodynamic imbalance and focal variations of the WSS were proven to be responsible for the genesis of IA.

Abnormal blood flow pulses lead to endothelial dysfunction, which causes a chronic inflammatory reaction. Leukocyte wall infiltration and the overproduction of proinflammatory mediators result in progressive remodeling of the wall, phenotypic switching of the VSMCs, myointimal hyperplasia, degradation of ECM, and aneurysmal progression.

Familial and genetic susceptibility, morphological features, and angioarchitecture significantly affect the risk of rupture.

Further advances in imaging techniques for wall instability and target therapies specifically targeted against the inflammatory pathways may significantly contribute to the improvement of the management of IA.

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#### References

- 1. Juvela, S. Prevalence of and risk factors for intracranial aneurysms. *Lancet Neurol.* **2011**, *10*, 595–597. [CrossRef]
- Johnston, S.C.; Selvin, S.; Gress, D.R. The burden, trends, and demographics of mortality from subarachnoid hemorrhage. *Neurology* 1998, 50, 1413–1418. [CrossRef] [PubMed]
- Wiebers, D.O.; Whisnant, J.P.; Huston, J., 3rd; Meissner, I.; Brown, R.D., Jr.; Piepgras, D.G.; Forbes, G.S.; Thielen, K.; Nichols, D.; O'Fallon, W.M.; et al. Unruptured intracranial aneurysms: Natural history, clinical outcome, and risks of surgical and endovascular treatment. *Lancet* 2003, 362, 103–110. [CrossRef]
- Serrone, J.C.; Maekawa, H.; Tjahjadi, M.; Hernesniemi, J. Aneurysmal subarachnoid hemorrhage: Pathobiology, current treatment and future directions. *Expert Rev. Neurother.* 2015, 15, 367–380. [CrossRef] [PubMed]
- 5. Broderick, J.P.; Sauerbeck, L.R.; Foroud, T.; Huston, J., 3rd; Pankratz, N.; Meissner, I.; Brown, R.D., Jr. The Familial Intracranial Aneurysm (FIA) study protocol. *BMC Med. Genet.* **2005**, *6*, 17. [CrossRef]
- 6. Bromberg, J.E.; Rinkel, G.J.; Algra, A.; van Duyn, C.M.; Greebe, P.; Ramos, L.M.; van Gijn, J. Familial subarachnoid hemorrhage: Distinctive features and patterns of inheritance. *Ann. Neurol.* **1995**, *38*, 929–934. [CrossRef] [PubMed]
- Struycken, P.M.; Pals, G.; Limburg, M.; Pronk, J.C.; Wijmenga, C.; Pearson, P.L.; Luijten, J.A.; van den Berg, J.S.; Vermeulen, M.; Rinkel, G.J.; et al. Anticipation in familial intracranial aneurysms in consecutive generations. *Eur. J. Hum. Genet.* 2003, 11, 737–743. [CrossRef] [PubMed]

- 8. Schievink, W.I. Genetics and aneurysm formation. Neurosurg. Clin. N. Am. 1998, 9, 485–495. [CrossRef]
- 9. Schievink, W.I. Genetics of intracranial aneurysms. *Neurosurgery* 1997, 40, 651–662, discussion 662–653. [CrossRef] [PubMed]
- 10. Ronkainen, A.; Miettinen, H.; Karkola, K.; Papinaho, S.; Vanninen, R.; Puranen, M.; Hernesniemi, J. Risk of harboring an unruptured intracranial aneurysm. *Stroke* **1998**, *29*, 359–362. [CrossRef]
- 11. Kato, T.; Hattori, H.; Yorifuji, T.; Tashiro, Y.; Nakahata, T. Intracranial aneurysms in Ehlers-Danlos syndrome type IV in early childhood. *Pediatr. Neurol.* 2001, 25, 336–339. [CrossRef]
- 12. Olubajo, F.; Kaliaperumal, C.; Choudhari, K.A. Vascular Ehlers-Danlos Syndrome: Literature review and surgical management of intracranial vascular complications. *Clin. Neurol. Neurosurg.* **2020**, *193*, 105775. [CrossRef]
- 13. Masoumi, A.; Reed-Gitomer, B.; Kelleher, C.; Bekheirnia, M.R.; Schrier, R.W. Developments in the management of autosomal dominant polycystic kidney disease. *Ther. Clin. Risk Manag.* **2008**, *4*, 393–407. [CrossRef]
- Sanchis, I.M.; Shukoor, S.; Irazabal, M.V.; Madsen, C.D.; Chebib, F.T.; Hogan, M.C.; El-Zoghby, Z.; Harris, P.C.; Huston, J.; Brown, R.D.; et al. Presymptomatic Screening for Intracranial Aneurysms in Patients with Autosomal Dominant Polycystic Kidney Disease. *Clin. J. Am. Soc. Nephrol.* 2019, 14, 1151–1160. [CrossRef]
- Menzies, L.; D'Arco, F.; Ganesan, V.; Hurst, J.A. Intracranial vascular pathology in two further patients with Floating-Harbor syndrome: Proposals for cerebrovascular disease risk management. *Eur. J. Med. Genet.* 2020, 63, 103785. [CrossRef] [PubMed]
- 16. Takeshima, Y.; Kaku, Y.; Nishi, T.; Mukasa, A.; Yamashiro, S. Multiple Cerebral Aneurysms Associated with Neurofibromatosis Type 1. *J. Stroke Cerebrovasc. Dis.* **2019**, *28*, e83–e91. [CrossRef] [PubMed]
- 17. Kim, S.T.; Brinjikji, W.; Kallmes, D.F. Prevalence of Intracranial Aneurysms in Patients with Connective Tissue Diseases: A Retrospective Study. *AJNR Am. J. Neuroradiol.* **2016**, *37*, 1422–1426. [CrossRef]
- 18. Finney, L.H.; Roberts, T.S.; Anderson, R.E. Giant intracranial aneurysm associated with Marfan's syndrome. Case report. *J. Neurosurg.* **1976**, *45*, 342–347. [CrossRef]
- 19. Román, G.; Fisher, M.; Perl, D.P.; Poser, C.M. Neurological manifestations of hereditary hemorrhagic telangiectasia (Rendu-Osler-Weber disease): Report of 2 cases and review of the literature. *Ann. Neurol.* **1978**, *4*, 130–144. [CrossRef]
- Schievink, W.I.; Katzmann, J.A.; Piepgras, D.G.; Schaid, D.J. Alpha-1-antitrypsin phenotypes among patients with intracranial aneurysms. J. Neurosurg. 1996, 84, 781–784. [CrossRef] [PubMed]
- 21. Oyesiku, N.M.; Colohan, A.R.; Barrow, D.L.; Reisner, A. Cocaine-induced aneurysmal rupture: An emergent negative factor in the natural history of intracranial aneurysms? *Neurosurgery* **1993**, *32*, 518–525, discussion 525–516. [CrossRef]
- 22. Sun, H.; Zhang, D.; Zhao, J. The interleukin-6 gene -572G>C promoter polymorphism is related to intracranial aneurysms in Chinese Han nationality. *Neurosci. Lett.* **2008**, 440, 1–3. [CrossRef]
- 23. Liu, Y.; Sun, J.; Wu, C.; Cao, X.; He, M.; You, C. The interleukin-6-572G/C gene polymorphism and the risk of intracranial aneurysms in a Chinese population. *Genet. Test. Mol. Biomark.* 2012, *16*, 822–826. [CrossRef] [PubMed]
- 24. Li, L.; Sima, X.; Bai, P.; Zhang, L.; Sun, H.; Liang, W.; Liu, J.; Zhang, L.; Gao, L. Interactions of miR-34b/c and TP53 polymorphisms on the risk of intracranial aneurysm. *Clin. Dev. Immunol.* **2012**, 2012, 567586. [CrossRef]
- Alg, V.S.; Sofat, R.; Houlden, H.; Werring, D.J. Genetic risk factors for intracranial aneurysms: A meta-analysis in more than 116,000 individuals. *Neurology* 2013, *80*, 2154–2165. [CrossRef] [PubMed]
- 26. Hong, E.P.; Kim, B.J.; Kim, C.; Choi, H.J.; Jeon, J.P. Association of SOX17 Gene Polymorphisms and Intracranial Aneurysm: A Case-Control Study and Meta-Analysis. *World Neurosurg.* **2018**, *110*, e823–e829. [CrossRef] [PubMed]
- Lee, S.; Kim, I.K.; Ahn, J.S.; Woo, D.C.; Kim, S.T.; Song, S.; Koh, G.Y.; Kim, H.S.; Jeon, B.H.; Kim, I. Deficiency of endotheliumspecific transcription factor Sox17 induces intracranial aneurysm. *Circulation* 2015, 131, 995–1005. [CrossRef] [PubMed]
- Peña Silva, R.A.; Kung, D.K.; Mitchell, I.J.; Alenina, N.; Bader, M.; Santos, R.A.; Faraci, F.M.; Heistad, D.D.; Hasan, D.M. Angiotensin 1-7 reduces mortality and rupture of intracranial aneurysms in mice. *Hypertension* 2014, 64, 362–368. [CrossRef] [PubMed]
- 29. Caranci, F.; Briganti, F.; Cirillo, L.; Leonardi, M.; Muto, M. Epidemiology and genetics of intracranial aneurysms. *Eur. J. Radiol.* **2013**, *82*, 1598–1605. [CrossRef]
- 30. Marchese, E.; Vignati, A.; Albanese, A.; Nucci, C.G.; Sabatino, G.; Tirpakova, B.; Lofrese, G.; Zelano, G.; Maira, G. Comparative evaluation of genome-wide gene expression profiles in ruptured and unruptured human intracranial aneurysms. *J. Biol. Regul. Homeost. Agents* **2010**, *24*, 185–195.
- Pimiento, J.M.; Maloney, S.P.; Tang, P.C.; Muto, A.; Westvik, T.S.; Fitzgerald, T.N.; Fancher, T.T.; Tellides, G.; Dardik, A. Endothelial nitric oxide synthase stimulates aneurysm growth in aged mice. J. Vasc. Res. 2008, 45, 251–258. [CrossRef]
- 32. Abruzzo, T.; Kendler, A.; Apkarian, R.; Workman, M.; Khoury, J.C.; Cloft, H.J. Cerebral aneurysm formation in nitric oxide synthase-3 knockout mice. *Curr. Neurovasc. Res.* 2007, *4*, 161–169. [CrossRef]
- 33. Katritsis, D.; Kaiktsis, L.; Chaniotis, A.; Pantos, J.; Efstathopoulos, E.P.; Marmarelis, V. Wall shear stress: Theoretical considerations and methods of measurement. *Prog. Cardiovasc. Dis.* 2007, 49, 307–329. [CrossRef]
- 34. Staarmann, B.; Smith, M.; Prestigiacomo, C.J. Shear stress and aneurysms: A review. Neurosurg. Focus 2019, 47, E2. [CrossRef]
- 35. Sforza, D.M.; Putman, C.M.; Cebral, J.R. Hemodynamics of Cerebral Aneurysms. *Annu. Rev. Fluid Mech.* 2009, 41, 91–107. [CrossRef] [PubMed]
- 36. Frösen, J.; Tulamo, R.; Paetau, A.; Laaksamo, E.; Korja, M.; Laakso, A.; Niemelä, M.; Hernesniemi, J. Saccular intracranial aneurysm: Pathology and mechanisms. *Acta Neuropathol.* **2012**, *123*, 773–786. [CrossRef] [PubMed]

- Cebral, J.R.; Detmer, F.; Chung, B.J.; Choque-Velasquez, J.; Rezai, B.; Lehto, H.; Tulamo, R.; Hernesniemi, J.; Niemela, M.; Yu, A.; et al. Local Hemodynamic Conditions Associated with Focal Changes in the Intracranial Aneurysm Wall. *AJNR Am. J. Neuroradiol.* 2019, 40, 510–516. [CrossRef]
- Boussel, L.; Rayz, V.; McCulloch, C.; Martin, A.; Acevedo-Bolton, G.; Lawton, M.; Higashida, R.; Smith, W.S.; Young, W.L.; Saloner, D. Aneurysm growth occurs at region of low wall shear stress: Patient-specific correlation of hemodynamics and growth in a longitudinal study. *Stroke* 2008, *39*, 2997–3002. [CrossRef] [PubMed]
- 39. Sakamoto, N.; Saito, N.; Han, X.; Ohashi, T.; Sato, M. Effect of spatial gradient in fluid shear stress on morphological changes in endothelial cells in response to flow. *Biochem. Biophys. Res. Commun.* **2010**, *395*, 264–269. [CrossRef] [PubMed]
- 40. Turjman, A.S.; Turjman, F.; Edelman, E.R. Role of fluid dynamics and inflammation in intracranial aneurysm formation. *Circulation* **2014**, *129*, 373–382. [CrossRef]
- 41. Hasan, D.; Chalouhi, N.; Jabbour, P.; Hashimoto, T. Macrophage imbalance (M1 vs. M2) and upregulation of mast cells in wall of ruptured human cerebral aneurysms: Preliminary results. *J. Neuroinflamm.* **2012**, *9*, 222. [CrossRef]
- 42. Moriwaki, T.; Takagi, Y.; Sadamasa, N.; Aoki, T.; Nozaki, K.; Hashimoto, N. Impaired progression of cerebral aneurysms in interleukin-1beta-deficient mice. *Stroke* 2006, *37*, 900–905. [CrossRef]
- 43. Aoki, T.; Kataoka, H.; Ishibashi, R.; Nozaki, K.; Morishita, R.; Hashimoto, N. Reduced collagen biosynthesis is the hallmark of cerebral aneurysm: Contribution of interleukin-1beta and nuclear factor-kappaB. *Arterioscler. Thromb. Vasc. Biol.* **2009**, *29*, 1080–1086. [CrossRef]
- Aoki, T.; Frösen, J.; Fukuda, M.; Bando, K.; Shioi, G.; Tsuji, K.; Ollikainen, E.; Nozaki, K.; Laakkonen, J.; Narumiya, S. Prostaglandin E2-EP2-NF-κB signaling in macrophages as a potential therapeutic target for intracranial aneurysms. *Sci. Signal.* 2017, 10. [CrossRef]
- Starke, R.M.; Chalouhi, N.; Jabbour, P.M.; Tjoumakaris, S.I.; Gonzalez, L.F.; Rosenwasser, R.H.; Wada, K.; Shimada, K.; Hasan, D.M.; Greig, N.H.; et al. Critical role of TNF-α in cerebral aneurysm formation and progression to rupture. *J. Neuroinflamm.* 2014, 11, 77. [CrossRef]
- 46. Mitsui, K.; Ikedo, T.; Kamio, Y.; Furukawa, H.; Lawton, M.T.; Hashimoto, T. TLR4 (Toll-Like Receptor 4) Mediates the Development of Intracranial Aneurysm Rupture. *Hypertension* 2020, *75*, 468–476. [CrossRef] [PubMed]
- 47. Ali, M.S.; Starke, R.M.; Jabbour, P.M.; Tjoumakaris, S.I.; Gonzalez, L.F.; Rosenwasser, R.H.; Owens, G.K.; Koch, W.J.; Greig, N.H.; Dumont, A.S. TNF-α induces phenotypic modulation in cerebral vascular smooth muscle cells: Implications for cerebral aneurysm pathology. *J. Cereb. Blood Flow Metab.* 2013, 33, 1564–1573. [CrossRef] [PubMed]
- Frösen, J. Smooth muscle cells and the formation, degeneration, and rupture of saccular intracranial aneurysm wall—A review of current pathophysiological knowledge. *Transl. Stroke Res.* 2014, *5*, 347–356. [CrossRef]
- 49. Hao, H.; Gabbiani, G.; Bochaton-Piallat, M.L. Arterial smooth muscle cell heterogeneity: Implications for atherosclerosis and restenosis development. *Arterioscler. Thromb. Vasc. Biol.* **2003**, *23*, 1510–1520. [CrossRef] [PubMed]
- Tulamo, R.; Frösen, J.; Hernesniemi, J.; Niemelä, M. Inflammatory changes in the aneurysm wall: A review. J. Neurointerv. Surg. 2010, 2, 120–130. [CrossRef]
- 51. Song, Y.; Liu, P.; Li, Z.; Shi, Y.; Huang, J.; Li, S.; Liu, Y.; Zhang, Z.; Wang, Y.; Zhu, W.; et al. The Effect of Myosin Light Chain Kinase on the Occurrence and Development of Intracranial Aneurysm. *Front. Cell. Neurosci.* **2018**, *12*, 416. [CrossRef] [PubMed]
- 52. Appelboom, G.; Chapel, D.; Connolly, E.; Goodman, A.; Lopresti, M.; Taylor, B.; Zilinyi, R. Role of the complement cascade in cerebral aneurysm formation, growth, and rupture. *Neuroimmunol. Neuroinflamm.* **2015**, *2*, 93–101. [CrossRef]
- Chalouhi, N.; Ali, M.S.; Jabbour, P.M.; Tjoumakaris, S.I.; Gonzalez, L.F.; Rosenwasser, R.H.; Koch, W.J.; Dumont, A.S. Biology of intracranial aneurysms: Role of inflammation. J. Cereb. Blood Flow Metab. 2012, 32, 1659–1676. [CrossRef]
- Guo, F.; Li, Z.; Song, L.; Han, T.; Feng, Q.; Guo, Y.; Xu, J.; He, M.; You, C. Increased apoptosis and cysteinyl aspartate specific protease-3 gene expression in human intracranial aneurysm. *J. Clin. Neurosci.* 2007, 14, 550–555. [CrossRef]
- 55. Fennell, V.S.; Kalani, M.Y.; Atwal, G.; Martirosyan, N.L.; Spetzler, R.F. Biology of Saccular Cerebral Aneurysms: A Review of Current Understanding and Future Directions. *Front. Surg.* **2016**, *3*, 43. [CrossRef]
- Xu, Z.; Rui, Y.N.; Hagan, J.P.; Kim, D.H. Intracranial Aneurysms: Pathology, Genetics, and Molecular Mechanisms. *Neuromol. Med.* 2019, 21, 325–343. [CrossRef] [PubMed]
- 57. Nakatani, H.; Hashimoto, N.; Kang, Y.; Yamazoe, N.; Kikuchi, H.; Yamaguchi, S.; Niimi, H. Cerebral blood flow patterns at major vessel bifurcations and aneurysms in rats. *J. Neurosurg.* **1991**, *74*, 258–262. [CrossRef]
- 58. Fukuda, S.; Hashimoto, N.; Naritomi, H.; Nagata, I.; Nozaki, K.; Kondo, S.; Kurino, M.; Kikuchi, H. Prevention of rat cerebral aneurysm formation by inhibition of nitric oxide synthase. *Circulation* **2000**, *101*, 2532–2538. [CrossRef]
- 59. Sho, E.; Sho, M.; Singh, T.M.; Xu, C.; Zarins, C.K.; Masuda, H. Blood flow decrease induces apoptosis of endothelial cells in previously dilated arteries resulting from chronic high blood flow. *Arterioscler. Thromb. Vasc. Biol.* 2001, 21, 1139–1145. [CrossRef] [PubMed]
- 60. Palumbo, P.; Lombardi, F.; Siragusa, G.; Dehcordi, S.R.; Luzzi, S.; Cimini, A.; Cifone, M.G.; Cinque, B. Involvement of NOS2 Activity on Human Glioma Cell Growth, Clonogenic Potential, and Neurosphere Generation. *Int. J. Mol. Sci* 2018, *19*, 2801. [CrossRef]
- 61. Tulamo, R.; Frösen, J.; Junnikkala, S.; Paetau, A.; Pitkäniemi, J.; Kangasniemi, M.; Niemelä, M.; Jääskeläinen, J.; Jokitalo, E.; Karatas, A.; et al. Complement activation associates with saccular cerebral artery aneurysm wall degeneration and rupture. *Neurosurgery* **2006**, *59*, 1069–1076, discussion 1076–1067. [CrossRef]

- 62. Unruptured intracranial aneurysms—Risk of rupture and risks of surgical intervention. *N. Engl. J. Med.* **1998**, 339, 1725–1733. [CrossRef]
- 63. Starke, R.M.; Chalouhi, N.; Ali, M.S.; Jabbour, P.M.; Tjoumakaris, S.I.; Gonzalez, L.F.; Rosenwasser, R.H.; Koch, W.J.; Dumont, A.S. The role of oxidative stress in cerebral aneurysm formation and rupture. *Curr. Neurovasc. Res.* 2013, 10, 247–255. [CrossRef] [PubMed]
- 64. Aoki, T.; Nishimura, M.; Kataoka, H.; Ishibashi, R.; Nozaki, K.; Hashimoto, N. Reactive oxygen species modulate growth of cerebral aneurysms: A study using the free radical scavenger edaravone and p47phox(-/-) mice. *Lab. Investig.* **2009**, *89*, 730–741. [CrossRef] [PubMed]
- 65. Liepsch, D.W. Flow in tubes and arteries—A comparison. *Biorheology* **1986**, *23*, 395–433. [CrossRef] [PubMed]
- 66. Ziegler, T.; Bouzourène, K.; Harrison, V.J.; Brunner, H.R.; Hayoz, D. Influence of oscillatory and unidirectional flow environments on the expression of endothelin and nitric oxide synthase in cultured endothelial cells. *Arterioscler. Thromb. Vasc. Biol.* **1998**, *18*, 686–692. [CrossRef]
- Ollikainen, E.; Tulamo, R.; Kaitainen, S.; Honkanen, P.; Lehti, S.; Liimatainen, T.; Hernesniemi, J.; Niemelä, M.; Kovanen, P.T.; Frösen, J. Macrophage Infiltration in the Saccular Intracranial Aneurysm Wall as a Response to Locally Lysed Erythrocytes That Promote Degeneration. J. Neuropathol. Exp. Neurol. 2018, 77, 890–903. [CrossRef]
- Meng, H.; Tutino, V.M.; Xiang, J.; Siddiqui, A. High WSS or low WSS? Complex interactions of hemodynamics with intracranial aneurysm initiation, growth, and rupture: Toward a unifying hypothesis. AJNR Am. J. Neuroradiol. 2014, 35, 1254–1262. [CrossRef]
- Texakalidis, P.; Sweid, A.; Mouchtouris, N.; Peterson, E.C.; Sioka, C.; Rangel-Castilla, L.; Reavey-Cantwell, J.; Jabbour, P. Aneurysm Formation, Growth, and Rupture: The Biology and Physics of Cerebral Aneurysms. *World Neurosurg.* 2019, 130, 277–284. [CrossRef]
- 70. Brisman, J.L.; Song, J.K.; Newell, D.W. Cerebral aneurysms. N. Engl. J. Med. 2006, 355, 928–939. [CrossRef]
- 71. Niemann, A.; Weigand, S.; Hoffmann, T.; Skalej, M.; Tulamo, R.; Preim, B.; Saalfeld, S. Interactive exploration of a 3D intracranial aneurysm wall model extracted from histologic slices. *Int. J. Comput. Assist. Radiol. Surg.* **2020**, *15*, 99–107. [CrossRef]
- 72. Frösen, J.; Piippo, A.; Paetau, A.; Kangasniemi, M.; Niemelä, M.; Hernesniemi, J.; Jääskeläinen, J. Remodeling of saccular cerebral artery aneurysm wall is associated with rupture: Histological analysis of 24 unruptured and 42 ruptured cases. *Stroke* 2004, 35, 2287–2293. [CrossRef] [PubMed]
- 73. Morita, A.; Kirino, T.; Hashi, K.; Aoki, N.; Fukuhara, S.; Hashimoto, N.; Nakayama, T.; Sakai, M.; Teramoto, A.; Tominari, S.; et al. The natural course of unruptured cerebral aneurysms in a Japanese cohort. *N. Engl. J. Med.* **2012**, 366, 2474–2482. [CrossRef] [PubMed]
- 74. Hassan, T.; Timofeev, E.V.; Saito, T.; Shimizu, H.; Ezura, M.; Matsumoto, Y.; Takayama, K.; Tominaga, T.; Takahashi, A. A proposed parent vessel geometry-based categorization of saccular intracranial aneurysms: Computational flow dynamics analysis of the risk factors for lesion rupture. *J. Neurosurg.* **2005**, *103*, 662–680. [CrossRef]
- 75. Ujiie, H.; Tachibana, H.; Hiramatsu, O.; Hazel, A.L.; Matsumoto, T.; Ogasawara, Y.; Nakajima, H.; Hori, T.; Takakura, K.; Kajiya, F. Effects of size and shape (aspect ratio) on the hemodynamics of saccular aneurysms: A possible index for surgical treatment of intracranial aneurysms. *Neurosurgery* 1999, 45, 119–129, discussion 129–130. [CrossRef] [PubMed]
- 76. Kyriacou, S.K.; Humphrey, J.D. Influence of size, shape and properties on the mechanics of axisymmetric saccular aneurysms. *J. Biomech.* **1996**, *29*, 1015–1022. [CrossRef]
- 77. Weir, B.; Disney, L.; Karrison, T. Sizes of ruptured and unruptured aneurysms in relation to their sites and the ages of patients. *J. Neurosurg.* **2002**, *96*, 64–70. [CrossRef]
- Peiffer, V.; Sherwin, S.J.; Weinberg, P.D. Does low and oscillatory wall shear stress correlate spatially with early atherosclerosis? A systematic review. *Cardiovasc. Res.* 2013, 99, 242–250. [CrossRef]
- 79. Samaniego, E.A.; Roa, J.A.; Hasan, D. Vessel wall imaging in intracranial aneurysms. J. Neurointerv. Surg. 2019, 11, 1105–1112. [CrossRef]
- Niemelä, M.; Frösen, J.; Hernesniemi, J.; Dashti, R.; Palotie, A. Molecular pathology of aneurysms. Surg. Neurol. 2008, 70, 36–38. [CrossRef]
- Kataoka, K.; Taneda, M.; Asai, T.; Kinoshita, A.; Ito, M.; Kuroda, R. Structural fragility and inflammatory response of ruptured cerebral aneurysms. A comparative study between ruptured and unruptured cerebral aneurysms. *Stroke* 1999, 30, 1396–1401. [CrossRef] [PubMed]
- 82. Tateshima, S.; Viñuela, F.; Villablanca, J.P.; Murayama, Y.; Morino, T.; Nomura, K.; Tanishita, K. Three-dimensional blood flow analysis in a wide-necked internal carotid artery-ophthalmic artery aneurysm. *J. Neurosurg.* 2003, *99*, 526–533. [CrossRef]
- 83. Duan, Z.; Li, Y.; Guan, S.; Ma, C.; Han, Y.; Ren, X.; Wei, L.; Li, W.; Lou, J.; Yang, Z. Morphological parameters and anatomical locations associated with rupture status of small intracranial aneurysms. *Sci Rep.* **2018**, *8*, 6440. [CrossRef] [PubMed]
- 84. Ricci, A.; Di Vitantonio, H.; De Paulis, D.; Del Maestro, M.; Raysi, S.D.; Murrone, D.; Luzzi, S.; Galzio, R.J. Cortical aneurysms of the middle cerebral artery: A review of the literature. *Surg. Neurol. Int.* **2017**, *8*, 117. [CrossRef] [PubMed]
- 85. Juvela, S.; Poussa, K.; Lehto, H.; Porras, M. Natural history of unruptured intracranial aneurysms: A long-term follow-up study. *Stroke* 2013, 44, 2414–2421. [CrossRef] [PubMed]
- 86. Murayama, Y.; Takao, H.; Ishibashi, T.; Saguchi, T.; Ebara, M.; Yuki, I.; Arakawa, H.; Irie, K.; Urashima, M.; Molyneux, A.J. Risk Analysis of Unruptured Intracranial Aneurysms: Prospective 10-Year Cohort Study. *Stroke* 2016, 47, 365–371. [CrossRef]

- 87. Roessler, K.; Cejna, M.; Zachenhofer, I. Aneurysmatic subarachnoidal haemorrhage: Incidence and location of small ruptured cerebral aneurysms—A retrospective population-based study. *Wien. Klin. Wochenschr.* **2011**, *123*, 444–449. [CrossRef]
- Greving, J.P.; Wermer, M.J.; Brown, R.D., Jr.; Morita, A.; Juvela, S.; Yonekura, M.; Ishibashi, T.; Torner, J.C.; Nakayama, T.; Rinkel, G.J.; et al. Development of the PHASES score for prediction of risk of rupture of intracranial aneurysms: A pooled analysis of six prospective cohort studies. *Lancet Neurol.* 2014, 13, 59–66. [CrossRef]
- 89. Neyazi, B.; Sandalcioglu, I.E.; Maslehaty, H. Evaluation of the risk of rupture of intracranial aneurysms in patients with aneurysmal subarachnoid hemorrhage according to the PHASES score. *Neurosurg. Rev.* 2019, 42, 489–492. [CrossRef]
- 90. Texakalidis, P.; Hilditch, C.A.; Lehman, V.; Lanzino, G.; Pereira, V.M.; Brinjikji, W. Vessel Wall Imaging of Intracranial Aneurysms: Systematic Review and Meta-analysis. *World Neurosurg.* **2018**, *117*, 453–458.e451. [CrossRef]
- Hasan, D.M.; Mahaney, K.B.; Magnotta, V.A.; Kung, D.K.; Lawton, M.T.; Hashimoto, T.; Winn, H.R.; Saloner, D.; Martin, A.; Gahramanov, S.; et al. Macrophage imaging within human cerebral aneurysms wall using ferumoxytol-enhanced MRI: A pilot study. *Arterioscler. Thromb. Vasc. Biol.* 2012, *32*, 1032–1038. [CrossRef]
- Vasanawala, S.S.; Nguyen, K.L.; Hope, M.D.; Bridges, M.D.; Hope, T.A.; Reeder, S.B.; Bashir, M.R. Safety and technique of ferumoxytol administration for MRI. *Magn. Reason. Med.* 2016, 75, 2107–2111. [CrossRef]
- 93. Campanella, R.; Guarnaccia, L.; Caroli, M.; Zarino, B.; Carrabba, G.; La Verde, N.; Gaudino, C.; Rampini, A.; Luzzi, S.; Riboni, L.; et al. Personalized and translational approach for malignant brain tumors in the era of precision medicine: The strategic contribution of an experienced neurosurgery laboratory in a modern neurosurgery and neuro-oncology department. *J. Neurol. Sci.* 2020, *417*, 117083. [CrossRef]
- Luzzi, S.; Crovace, A.M.; Del Maestro, M.; Giotta Lucifero, A.; Elbabaa, S.K.; Cinque, B.; Palumbo, P.; Lombardi, F.; Cimini, A.; Cifone, M.G.; et al. The cell-based approach in neurosurgery: Ongoing trends and future perspectives. *Heliyon* 2019, 5, e02818. [CrossRef]
- 95. Giotta Lucifero, A.; Luzzi, S.; Brambilla, I.; Guarracino, C.; Mosconi, M.; Foiadelli, T.; Savasta, S. Gene therapies for high-grade gliomas: From the bench to the bedside. *Acta Biomed.* **2020**, *91*, 32–50. [CrossRef]
- 96. Giotta Lucifero, A.; Luzzi, S.; Brambilla, I.; Schena, L.; Mosconi, M.; Foiadelli, T.; Savasta, S. Potential roads for reaching the summit: An overview on target therapies for high-grade gliomas. *Acta Biomed.* **2020**, *91*, 61–78. [CrossRef] [PubMed]
- 97. Giotta Lucifero, A.; Luzzi, S.; Brambilla, I.; Trabatti, C.; Mosconi, M.; Savasta, S.; Foiadelli, T. Innovative therapies for malignant brain tumors: The road to a tailored cure. *Acta Biomed.* 2020, *91*, 5–17. [CrossRef] [PubMed]
- Luzzi, S.; Giotta Lucifero, A.; Brambilla, I.; Magistrali, M.; Mosconi, M.; Savasta, S.; Foiadelli, T. Adoptive immunotherapies in neuro-oncology: Classification, recent advances, and translational challenges. *Acta Biomed.* 2020, *91*, 18–31. [CrossRef]
- Giotta Lucifero, A.; Luzzi, S. Against the Resilience of High-Grade Gliomas: The Immunotherapeutic Approach (Part I). *Brain Sci.* 2021, 11, 386. [CrossRef] [PubMed]
- Hasan, D.M.; Mahaney, K.B.; Brown, R.D., Jr.; Meissner, I.; Piepgras, D.G.; Huston, J.; Capuano, A.W.; Torner, J.C. Aspirin as a promising agent for decreasing incidence of cerebral aneurysm rupture. *Stroke* 2011, 42, 3156–3162. [CrossRef] [PubMed]
- Tada, Y.; Kitazato, K.T.; Yagi, K.; Shimada, K.; Matsushita, N.; Kinouchi, T.; Kanematsu, Y.; Satomi, J.; Kageji, T.; Nagahiro, S. Statins promote the growth of experimentally induced cerebral aneurysms in estrogen-deficient rats. *Stroke* 2011, 42, 2286–2293. [CrossRef]
- 102. Aoki, T.; Nishimura, M.; Kataoka, H.; Ishibashi, R.; Miyake, T.; Takagi, Y.; Morishita, R.; Hashimoto, N. Role of angiotensin II type 1 receptor in cerebral aneurysm formation in rats. *Int. J. Mol. Med.* **2009**, *24*, 353–359. [CrossRef]
- 103. Fisher, C.L.; Demel, S.L. Nonsteroidal Anti-Inflammatory Drugs: A Potential Pharmacological Treatment for Intracranial Aneurysm. *Cerebrovasc. Dis. Extra* 2019, 9, 31–45. [CrossRef]
- Suzuki, T.; Kamio, Y.; Makino, H.; Hokamura, K.; Kimura, T.; Yamasaki, T.; Hiramatsu, H.; Umemura, K.; Namba, H. Prevention Effect of Antiplatelets on Aneurysm Rupture in a Mouse Intracranial Aneurysm Model. *Cerebrovasc. Dis.* 2018, 45, 180–186. [CrossRef]
- 105. Fujiwara, Y.; Shiraya, S.; Miyake, T.; Yamakawa, S.; Aoki, M.; Makino, H.; Nishimura, M.; Morishita, R. Inhibition of experimental abdominal aortic aneurysm in a rat model by the angiotensin receptor blocker valsartan. *Int. J. Mol. Med.* **2008**, *22*, 703–708.
- 106. Ishibashi, R.; Aoki, T.; Nishimura, M.; Hashimoto, N.; Miyamoto, S. Contribution of mast cells to cerebral aneurysm formation. *Curr. Neurovasc. Res.* 2010, 7, 113–124. [CrossRef] [PubMed]