

# *Aminopeptidase-N/CD13 (EC 3.4.11.2) Inhibitors: Chemistry, Biological Evaluations, and Therapeutic Prospects*

*Brigitte Bauvois,<sup>1</sup> Daniel Dauzonne<sup>2</sup>*

<sup>1</sup>Unité INSERM 507, Hôpital Necker, Université René Descartes Paris V, Bâtiment Lavoisier,  
161 rue de Sèvres, 75015 Paris, France

<sup>2</sup>UMR 176 Institut Curie-CNRS, Institut Curie, Section Recherche, 26 rue d'Ulm, 75248  
Paris CEDEX 05, France

Published online 7 October 2005 in Wiley InterScience (www.interscience.wiley.com).  
DOI 10.1002/med.20044



**Abstract:** Aminopeptidase N (APN)/CD13 (EC 3.4.11.2) is a transmembrane protease present in a wide variety of human tissues and cell types (endothelial, epithelial, fibroblast, leukocyte). APN/CD13 expression is dysregulated in inflammatory diseases and in cancers (solid and hematologic tumors). APN/CD13 serves as a receptor for coronaviruses. Natural and synthetic inhibitors of APN activity have been characterized. These inhibitors have revealed that APN is able to modulate bioactive peptide responses (pain management, vasopressin release) and to influence immune functions and major biological events (cell proliferation, secretion, invasion, angiogenesis). Therefore, inhibition of APN/CD13 may lead to the development of anti-cancer and anti-inflammatory drugs. This review provides an update on the biological and pharmacological profiles of known natural and synthetic APN inhibitors. Current status on their potential use as therapeutic agents is discussed with regard to toxicity and specificity. © 2005 Wiley Periodicals, Inc. *Med Res Rev*, 26, No. 1, 88–130, 2006

**Key words:** aminopeptidase; ectoenzyme; natural inhibitor; synthetic inhibitor; bestatin; cancer; inflammation

---

*Contract grant sponsor:* The Institut National de la Santé et de la Recherche Médicale (INSERM); *Contract grant sponsor:* the Centre National de La Recherche Scientifique (CNRS); *Contract grant sponsor:* the Institut Curie, the Association pour la Recherche sur le Cancer (ARC number 3473); *Contract grant sponsor:* la Fondation pour la Recherche Médicale; *Contract grant sponsor:* La Ligue Nationale Contre le Cancer.

*Correspondence to:* Daniel Dauzonne, UMR176 Institut Curie-CNRS, Institut Curie, Section Recherche, 26 rue d'Ulm, 75248 Paris CEDEX 05, France. E-mail: daniel.dauzonne@curie.fr

## 1. INTRODUCTION

Aminopeptidase N (EC 3.4.11.2, APN) is a metallo-dependent integral membrane protease.<sup>1</sup> The enzyme belongs to the M1 family of the MA clan of peptidases<sup>2</sup> also called gluzincins.<sup>3</sup> Aminopeptidase N consists of 967 amino acids with a short N-terminal cytoplasmic domain, a single transmembrane part, and a large cellular ectodomain containing the active site.<sup>4</sup> This enzyme was first isolated in 1963 by Pfeleiderer and Celliers from pig kidney<sup>5</sup> and is known under several different names (alanine aminopeptidase; microsomal aminopeptidase; microsomal leucine aminopeptidase aminopeptidase M; amino oligopeptidase; GP 150). In the last few years, certain surface molecules identified as cluster differentiation (CD) antigens were found to be identical to some membrane proteins. Thus, CD13 is identical to APN.<sup>6,7</sup> Soluble APN is detectable in plasma/serum and urine<sup>8–11</sup> but the mechanism of release of membrane APN remains unknown.

Membrane-bound APN/CD13 is widely distributed outside the hematopoietic system (epithelial-, endothelial-, fibroblast-cell types) with main sources being brush border membranes of kidney proximal tubule cells and enterocytes, and in the hematopoietic compartment is not confined to a particular lineage.<sup>1,12,13</sup> APN/CD13 is predominantly expressed on stem cells and on cells of the granulocytic and monocytic lineages at distinct stages of differentiation and is therefore considered as a marker of differentiation.<sup>14,15</sup>

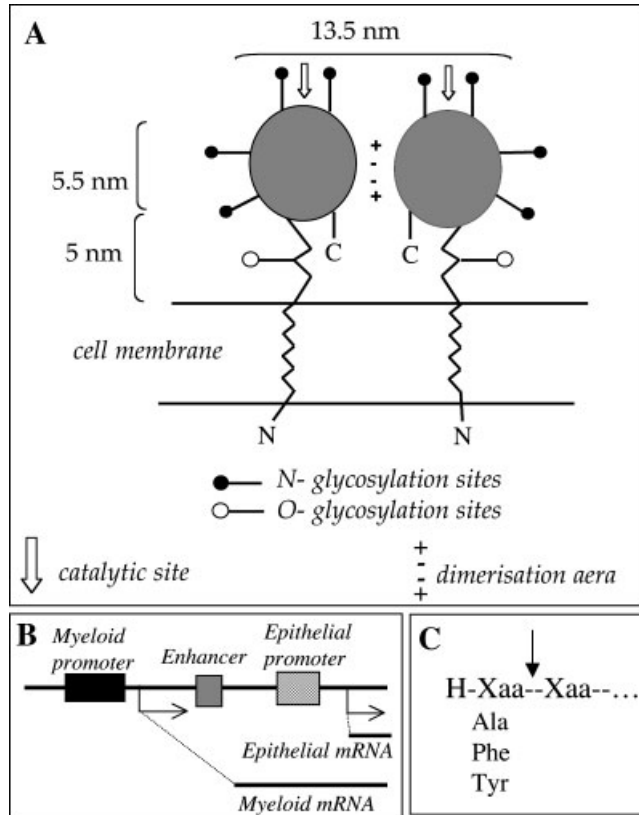
Dysregulated expression of membrane and/or soluble forms of APN/CD13 is observed in many diseases. Compiled observations indicate enhanced APN levels in tumor cells such as melanoma,<sup>16,17</sup> renal,<sup>18</sup> pancreas,<sup>19</sup> colon,<sup>20</sup> prostate,<sup>21</sup> gastric,<sup>22</sup> and thyroid<sup>23</sup> cancers. Tumor-infiltrating T cells in renal and lung cancers are CD13-positive.<sup>24,25</sup> APN activity is elevated in plasma and effusions of cancer patients.<sup>11</sup> APN activity on neutrophils from patients affected by a rare adrenal gland tumor, adrenal pheochromocytoma, is significantly increased as compared with healthy controls.<sup>26</sup> CD13 is overexpressed in acute and chronic myeloid leukemias<sup>1,12,27–29</sup> and in anaplastic large cell lymphomas.<sup>30,31</sup> Overexpression of APN/CD13 in T lymphocytes or neutrophils occurs in several inflammatory diseases (chronic pain, various forms of joint effusions, rheumatoid arthritis, multiple sclerosis, systemic sclerosis, systemic lupus erythematosus, polymyositis/dermatomyositis, pulmonary sarcoidosis).<sup>32–39</sup>

APN/CD13 may be therefore considered as a useful clinical marker. Whether this protease critically contributes to the pathological behavior remains however unknown. In this review, we briefly summarise knowledge on the structure and the mechanisms of cleavage of APN/CD13 to integrate current knowledge in natural and synthetic APN inhibitors. The reader is referred to excellent reviews for the characteristics of APN/CD13 and substrate specificity.<sup>25,40–45</sup> Various aspects on the roles of APN/CD13 are reviewed here in the context of the *in vitro* and *in vivo* use of certain APN inhibitors.

## 2. AMINOPEPTIDASE N/CD13

APN is anchored to the plasma membrane, *via* an uncleaved signal sequence, by the C-terminus (type II) facing extracellularly.<sup>1</sup> Membrane APN/CD13 is found as a dimer of two non covalently associated subunits with a relative molecular mass of 160 kDa (Fig. 1A).<sup>40,41,43</sup> The human CD13 gene, cloned in 1989<sup>6</sup> and subsequently mapped to chromosome 15 q25-26,<sup>46</sup> possesses two promoters (Fig. 1B).<sup>46–51</sup>

The cDNA sequence reveals the presence of the amino acid sequence His-Glu-Xaa-Xaa-His which is a Zn<sup>++</sup> binding motif found in one class of metallo-peptidases.<sup>3</sup> Site-directed mutagenesis indicates that extracellular cysteines in the molecule confer correct structure and consequently enzymatic activity and surface expression of APN.<sup>52</sup> Mutation of glutamic acid 355 in an aminopeptidase conserved region (the GAMEN motif) leads to an inactive enzyme<sup>53</sup> indicating that



**Figure 1.** Schematic diagrams showing protein (A) and promoter (B) structures of APN/CD13. The enzyme is a dimer of two non covalently associated monomers. The gene is controlled by two promoters, with an epithelial promoter and a myeloid promoter (in the hematopoietic system). (C) substrate specificity. The arrow indicates the bond cleaved.

this glutamic acid belongs to the anionic binding site in APN and interacts with the N-terminal  $\alpha$ -amino group of the substrate. APN/CD13 cleaves preferentially neutral amino acids (with the exception of proline) (Fig. 1C) from the unsubstituted N-terminus of oligopeptides.<sup>1,12</sup> Biologically active peptide substrates cleaved by APN/CD13 are neuropeptides (Met- and Leu-enkephalins, neurokinin A, Met-lys-bradykinin, and endorphins such as spinorphin),<sup>41,54-59</sup> vasoactive peptides (kallidin, somatostatin, and angiotensins)<sup>60-67</sup> and chemotactic peptides (monocyte chemotactic protein/MCP-1 and N-formyl methionine leucine phenylalanine/f-MLP).<sup>40,68</sup>

Apart from its hydrolytic ability, APN serves as a receptor for coronaviruses.<sup>69-72</sup> In humans, the 229E corona virus uses APN to enter alveolar cells and establish an upper respiratory tract infection.<sup>72</sup>

### 3. APN/CD13 INHIBITORS

Bradykinin (Arg-Pro-Pro-Gly-Phe-Ser-Pro-Phe-Arg) and substance P (Arg-Pro-Lys-Pro-Gln-Gln-Phe-Phe-Gly-Leu-Met-NH<sub>2</sub>) are natural peptides capable of inhibiting APN in micromolar concentrations.<sup>73</sup> Similarly, elevated concentrations of leucine, proline, L-alanine, L-arginine, L-glutamine, L-methionine, as well as divalent cations (Co<sup>2+</sup>, Zn<sup>2+</sup>, Mn<sup>2+</sup>, Ca<sup>2+</sup>, Ni<sup>2+</sup>) inhibit APN activity.<sup>40</sup> (for review) Moreover, molecules with a broad spectrum of action such as KCN, NaN<sub>3</sub>,

ammonium oxalate, *N*-ethyl-maleimide, and 8-hydroxyquinoline inhibit APN/CD13.<sup>40</sup> (for review) APN activity is also inhibited by puromycin (**1**),<sup>74,75</sup> lapstatin (**2**),<sup>76</sup> some *N*-phenylphthalimide derivatives such as compound **3**,<sup>77–80</sup> several *N*-phenylhomophthalimide derivatives like PIQ-22 (**4**)<sup>77,78</sup> which has later been described as a rather puromycin-sensitive aminopeptidase (PSA) inhibitor by the same group,<sup>80–83</sup> phosphinate dipeptide analogues illustrated by hPheP[CH<sub>2</sub>]Tyr (**5**),<sup>84</sup> pseudoglutamyl aminophosphinic peptides such as GluΨ(PO<sub>2</sub>CH<sub>2</sub>)Leu-Ala (**6**),<sup>85</sup> several variously substituted 3-amino-2-oxobutyramide exemplified by compound **7**,<sup>86</sup> α-aminoboronic derivatives such as the benzyl derivative **8**,<sup>87</sup> or α-aminobenzaldehydes illustrated by (*S*) 2-amino-5-methylpentanal (**9**).<sup>88</sup> An eclectic set of compounds has been described and used for the biochemical characterization or/and inhibition of other proteases—e.g.: urokinase-type plasminogen activator, dipeptidylpeptidase IV (DPPIV/CD26), or other different aminopeptidases including human enkephalin degrading aminopeptidase (HEDA), cytosolic leucine aminopeptidase (LAPc), glutamyl aminopeptidase (APA), and arginyl aminopeptidase (AP-B). In this context, it is also worth mentioning two systematic studies devoted to hydroxylated naturally occurring flavonoids such as baicalein (**10**), apigenin (**11**), or myricetin (**12**) and related compounds which, aside their activity on neutral endopeptidase (NEP/CD10) or angiotensin-converting enzyme (ACE/CD143), exhibited a significant *in vitro* inhibitory effect toward APN.<sup>89,90</sup> Formulas, *K<sub>i</sub>*, *IC*<sub>50</sub> or inhibition percentages of enzymes for compounds **1–12** are depicted in Figure 2. Two recent publications describing either the irreversible inhibition of both APN/CD13 and DPP IV/CD26 enzymatic activities by aqueous extracts of a *Cistus incanus* L.<sup>91</sup> or ACE, NEP, and APN inhibition by extracts of *Epilobium angustifolium*<sup>92</sup> deserve also quotation.

Although the borderline is not easy to position, leaving out the above-mentioned studies dealing with non-specific compounds targeting other enzymes and, incidentally, revealing an inhibitory activity on APN, we have chosen to focus the present review on the data tightly dedicated to natural and synthetic inhibitors of APN/CD13 itself.

### A. Naturally Occurring APN/CD13 Inhibitors

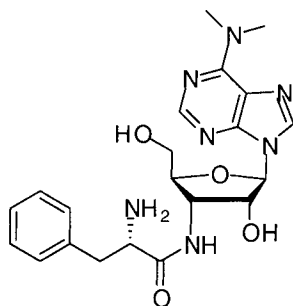
The most widely used among the naturally occurring APN/CD13 inhibitors are microorganism-produced and have been purified from microbial culture filtrates. A large part of them are generated by bacteria belonging to the order *Actinomycetales*, especially of the genera *Streptomyces*:

#### 1. Actinonin

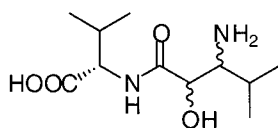
(2*R*)-*N*<sup>1</sup>-hydroxy-*N*<sup>1</sup>-[(1*S*)-1-[[2*S*]-2-(hydroxymethyl)-1-pyrrolidinyl]carbonyl]-2-methylpropyl]-2-pentylbutanediamide (**13**) was first isolated by R. Green and R. Bhagwan Singh from a Malayan strain of *Actinomycetes*. This compound was then listed as *Streptomyces* Cutter C/2 (N.C.I.B. 8845).<sup>93</sup> About 20 years later, actinonin was also obtained from another strain referenced MG848-hF6 and its inhibition against APN was found to be competitive with the substrate.<sup>94</sup> The structural study and the chemical synthesis of **13** and some analogues have aroused numerous works<sup>95–102</sup> completed by a structure–activity relationship investigation dealing with anti bacterial properties observed in this actinonin series.<sup>103</sup>

#### 2. AHPA-Val

(2*S*,3*R*)-3-amino-2-hydroxy-4-phenylbutanoyl-L-valine (**14**) and two closely related derivatives: AHPA-Val-Pro-Hyp (2*S*,3*R*)-3-amino-2-hydroxy-4-phenylbutanoyl-L-valyl-L-prolyl-(*trans*-4-hydroxy-L-proline) (MR387A) (**15**) and AHPA-Val-Pro-Pro (2*S*,3*R*)-3-amino-2-hydroxy-4-phenylbutanoyl-L-valyl-L-prolyl-L-proline) (MR387B) (**16**) were obtained from the culture broth of *Streptomyces neyagawaensis* SL-387.<sup>104–106</sup> The preparation of several novel synthetic AHPA derivatives (exemplified by **17**) bearing, for most of them, heterocyclic moieties and exhibiting

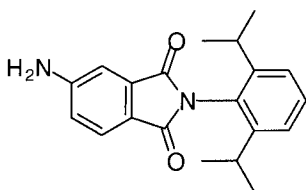


**Puromycin (1)**

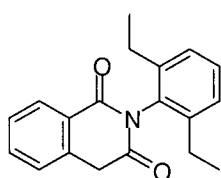


**Lapstatin (2)**

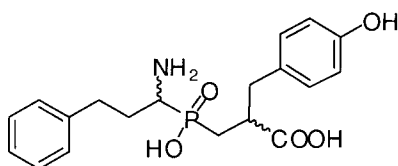
(The absolute configuration is not completely elucidated)



**3**



**PIQ-22 (4)**



**hPheP[CH<sub>2</sub>]Tyr (5)**

(mixture of four diastereoisomers)

IC<sub>50</sub> APM from rat blood plasma : 60 μM<sup>67</sup>

IC<sub>50</sub> APN : 50 μM, IC<sub>50</sub> AAP-S : 0.6 μM μM<sup>75</sup>

67% inhibition of APN from human seminal plasma at 100 μM<sup>74</sup>

IC<sub>50</sub> APN : 4.8 μM, IC<sub>50</sub> PSA : 0.6 μM<sup>83</sup>

IC<sub>50</sub> APM from hog kidney : 203 μM<sup>76</sup>

IC<sub>50</sub> APN from egg white : > 122 μM<sup>76</sup>

IC<sub>50</sub> LAP from porcine kidney : 41 μM<sup>76</sup>

IC<sub>50</sub> LAP from *Streptomyces rimosus* : 2.4 μM<sup>76</sup>

IC<sub>50</sub> LAP from *Aeromonas proteolytica* : 0.3 μM<sup>76</sup>

IC<sub>50</sub> API from *Streptomyces griseus* : 0.85 μM<sup>76</sup>

IC<sub>50</sub> APN : 5.4 μg/mL, IC<sub>50</sub> DPP-IV : 14.1 μg/mL, WI-38 : 17.4 μg/mL<sup>77,80</sup>

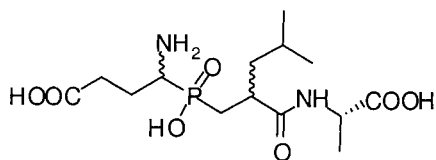
IC<sub>50</sub> APN : 16.8 μM, IC<sub>50</sub> DPP-IV : 251.2 μM<sup>79</sup>

IC<sub>50</sub> APN : 0.12 μg/mL, IC<sub>50</sub> DPP-IV : 14.1 μg/mL<sup>77,80</sup>

Inactive towards LAP, DPP-IV, Trypsin and Chymotrypsin<sup>80,82</sup>

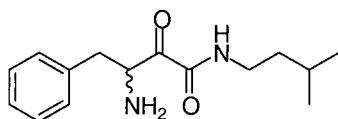
K<sub>i</sub> APN : 36 nM, K<sub>i</sub> LAP : 67nM<sup>84</sup>

**Figure 2.** Miscellaneous inhibitors of APN/CD13.



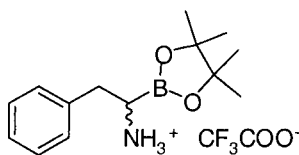
**GluΨ(PO<sub>2</sub>CH<sub>2</sub>)Leu-Ala (6)**  
(mixture of four diastereoisomers)

$K_i$  APN : 31  $\mu$ M,  $K_i$  APA : 0.8nM,  $K_i$  NEP : 62 nM,  $K_i$  ACE : 53  $\mu$ M<sup>85</sup>



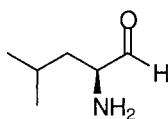
**7 (racemate)**

$K_i$  APN : 2.5  $\mu$ M,  $K_i$  LAP<sub>c</sub> : 1  $\mu$ M,  $K_i$  APB : 1.5  $\mu$ M<sup>86</sup>



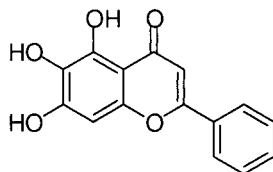
**8 (racemate)**

$IC_{50}$  APN : 20 nM,  $IC_{50}$  HEDA : 50 nM<sup>87</sup>



**9**

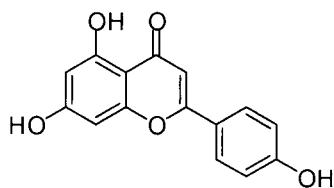
$K_i$  APN : 760 nM,  $K_i$  LAP<sub>c</sub> : 60 nM<sup>88</sup>



**Baicalein (10)**

% inhibition (at maximal concentration of 300  $\mu$ M)

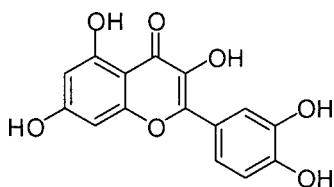
APN : 57 %, ACE : 10 %, NEP : 36 %<sup>90</sup>



**Apigenin (11)**

% inhibition (at 100  $\mu$ M) : APN : 26 %, inactive on LAP<sup>89</sup>

% inhibition (at maximal concentration of 300  $\mu$ M) : APN : 42 %, ACE : 18 %, NEP : 31 %<sup>90</sup>

**Myricetin (12)**

% inhibition (at 100  $\mu\text{M}$ ) : APN : 25 %, inactive on LAP<sup>89</sup>

% inhibition (at maximal concentration of 300  $\mu\text{M}$ ) : APN :

48 %, ACE : 26 %, NEP : 68 %<sup>90</sup>

**Figure 2.** (Continued)

interesting *in vivo* antitumor potencies (30–40% inhibitory rate on S180 sarcoma) has been recently reported.<sup>107</sup>

### 3. Amastatin

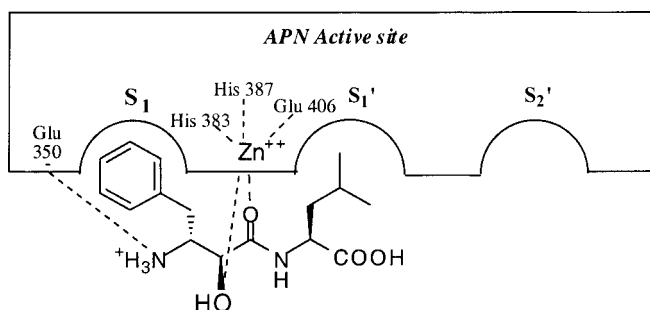
(2*S*,3*R*)-3-amino-2-hydroxy-5-methylhexanoyl-L-valine-L-valine-L-aspartic acid (**18**) has been reported to be a slow-binding competitive inhibitor of APN.<sup>108</sup> It was first isolated from the culture filtrate of *Streptomyces* sp. ME98-M3.<sup>109</sup> and its structure has been unambiguously determined.<sup>110</sup> Several enantioselective syntheses of this tetrapeptide have been reported,<sup>110,111</sup> and some of its analogues have also been prepared in the context of a SAR study.<sup>112</sup>

### 4. Bestatin

(2*S*,3*R*)-3-amino-2-hydroxy-4-phenylbutanoyl-L-leucine (Ubenimex<sup>®</sup>) (**19**) is an inhibitor of various leucine and arginine aminopeptidases,<sup>113</sup> and an efficient inhibitor of LTA<sub>4</sub> hydrolase.<sup>114–117</sup> However, in spite of its marked toxicity and of its relative lack of selectivity toward exopeptidases, it is one of the most used compound for its APN/CD13 inhibitory effects.<sup>118</sup> Bestatin has been described as a slow-binding competitive inhibitor of APN,<sup>108</sup> and a schematic representation of **19** within the active site of APN<sup>53,84</sup> is depicted in Figure 3. Bestatin was first isolated from a culture filtrate of *Streptomyces olivoreticuli* (MD976-C7)<sup>119</sup> and its chemical structure has been subsequently ascertained.<sup>120</sup> Several stereoselective total syntheses of **19** have been reported,<sup>121–130</sup> the preparation of its stereoisomers has been performed<sup>131</sup> and some ubenimex derivatives or analogues such as the *para*-hydroxybestatin (**20**),<sup>132</sup> the 2-thiolbestatin (**21**),<sup>133,134</sup> the bestatin thioamide (**22**),<sup>133,135</sup> or the reduced bestatin **23**<sup>136</sup> have also been prepared.

### 5. Phebestin

(2*S*,3*R*)-3-amino-2-hydroxy-4-phenylbutanoyl-L-valyl-L-phenylalanine (**24**) is a tripeptide produced by *Streptomyces* sp. MJ716-m3.<sup>137</sup> Some stereoselective syntheses of **24** have been recently reported.<sup>125,128,129</sup>

**Figure 3.** Binding of Bestatin to the active site of APN.

## 6. Probestin

(2*S*,3*R*)-3-amino-2-hydroxy-4-phenylbutanoyl-L-valyl-L-prolyl-L-proline) (**25**) is a tetrapeptide isolated from the culture of *Streptomyces azureus* (MH663-2F6)<sup>138</sup> and its structure has been unambiguously established.<sup>139</sup> Probestin has been described as a competitive inhibitor of APN<sup>138</sup> and, here also, some total syntheses have been lately described.<sup>125,128,129</sup>

An overview of the formulas of compounds **14–25** reveals that, except the synthetic analogue **23** prepared in its racemic form, they all possess the absolute configuration (2*S*,3*R*) which appears crucial for activity.<sup>136</sup> A comparable chiral framework is also existent in the side chain of the pharmacologically important series constituted by taxoids and, in this context, it is worth pointing out that numerous and various synthetic approaches to building blocks liable to lead to enantiomerically pure (2*S*,3*R*)-3-amino-2-hydroxyalkanoic structures and/or their diastereomers have attracted considerable attention.<sup>140–185</sup>

## 7. Leuhistin

(2*R*,3*S*)-3-amino-2-hydroxy-2-1*H*-(imidazol-4-ylmethyl)-5-methylhexanoic acid (**26**) has been isolated in 1991 by Takeuchi and co-workers from the culture broth of a bacteria belonging to the phylum *Firmicutes*: *Bacillus laterosporus* BM156-14F1.<sup>186,187</sup> This compound inhibits APN in a competitive manner with the substrate.<sup>186</sup> The structure of **26** and its absolute configuration have been thereafter ascertained by the same group.<sup>188</sup>

Several naturally occurring APN inhibitors are of vegetal origin:

## 8. Benzo[*c*]phenanthridines

Benzo[*c*]phenanthridines such as 1,2-Dimethoxy-12-methyl 1,3-dioxolo[4',5':4,5]benzo[1,2-*c*]phenanthridin-12-ium chloride or Chelerythrine (**27**) and some closely related alkaloids have recently been isolated from extracts of the Papaveraceae *Macleania cordata* (Wild.) R. Br. Some of these compounds showed an efficacy against APN similar to that of amastatin (**18**) or bestatin (**19**). A weaker inhibitory effect on DPP-IV has also been reported.<sup>189</sup>

## 9. Curcumin

(*E,E*-1,7-bis-(4-hydroxy-3-methoxyphenyl)-1,6-heptadiene-3,5-dione) (**28**) is a yellow natural phenolic compound isolated from the rhizomes of asian perennial herbs extensively cultivated in tropical areas and belonging to the Zingiberaceae family. All these plants are of the genera *Curcuma*. The most exploited representative is *Curcuma longa* L., whose dried rhizome is the source of the spice turmeric which is widely employed in food and has a long tradition of use in folk medicine. In addition to its irreversible APN/CD13 inhibition potencies,<sup>190</sup> curcumin is now considered by oncologists as a potential cancer chemopreventive agent,<sup>191,192</sup> and clinical trials in this context are carried out in several laboratories.<sup>193</sup> Furthermore, curcumin possesses anti-inflammatory activity and is a potent inhibitor of reactive oxygen-generating enzymes (e.g. lipooxygenase/cyclooxygenase-2, xanthine dehydrogenase/oxidase and inducible nitric oxide synthase).<sup>194</sup> Curcumin hinders also the initiation of carcinogenesis by inhibiting the cytochrome P-450 enzyme activity and increasing the levels of glutathione-S-transferase. Its anti-tumor effect in the promotion and progression stages has been attributed, in part, to the arrest of cancer cells in S, G2/M cycle phase, and induction of apoptosis.<sup>195</sup> It has also been proposed that curcumin may suppress tumor promotion by blocking signal transduction pathways in the target cells.<sup>196</sup> Curcumin is a potent inhibitor of protein kinase C, EGF-receptor tyrosine kinase and I- $\kappa$ B kinase. In addition, curcumin inhibits the activation of NF- $\kappa$ B and the expression of c-jun, c-fos, c-myc.<sup>194,197</sup> Last, curcumin has been



proposed as a HIV-1 or HIV-2 protease inhibitor,<sup>198</sup> as a HIV-1 integrase inhibitor,<sup>199</sup> and proved to be radioprotectant.<sup>200,201</sup> Several chemical synthesis of **28**, involving 2,4-pentanedione and vanillin, have been reported<sup>202–206</sup> as well as the preparations of some of its analogues designed as angiogenesis inhibitors<sup>207</sup> through their ability to inhibit endothelial cell proliferation.<sup>208</sup>

### 10. Betulinic Acid

(3 $\beta$ -hydroxylup-20(29)-en-28-oic acid) (**29**) is a pentacyclic compound widely present in the plant kingdom. This oxidized derivative of betulin owes its trivial name to the fact that this class of lupane type triterpenes was first isolated from *Betula* ssp. (birch trees). Afterwards, betulinic acid has been obtained from various other vegetal species including *Ancistrocladus* ssp., *Arbutus* ssp., *Diospyros* ssp., *Paeonia* ssp., *Picramnia* ssp., *Syzygium* ssp., *Tetracera* ssp., *Tryphillum* spp., *Zizyphus* ssp. One of the main current sources of betulinic acid from natural origin is the bark of plane trees (e.g. *Platanus acerifolia*) by employing a patented procedure.<sup>209</sup> In addition to its APN inhibitory activity in a dose-dependent manner,<sup>210</sup> and possibly as a partial consequence of this inhibitory potency, betulinic acid has been shown to modulate the immune response, to exhibit anti-inflammatory properties and to block HIV-1 entry into cells. It has also been reported to be a selective inhibitor of DNA polymerase  $\beta$  and to induce apoptosis in tumor cells. The wide range of biological properties linked to betulinic acid have recently been recapitulated and analyzed in three excellent reviews.<sup>211–213</sup> Several hemisynthesis of **28** starting from betulin via betulonic acid<sup>214–217</sup> or from various naturally occurring betulinic acid derivatives such as glycosides,<sup>218–221</sup> sulfates,<sup>222</sup> or dihydroxycinnamic esters<sup>223</sup> have been reported.

To our knowledge, only one naturally occurring APN inhibitor originates from animal kingdom:

### 11. Psammalin A

((*E,E*)-*N,N'*-Bis[(3-(3'-Bromo-4'-hydroxyphenyl)-2-oximidopropionyl)cystamine] (**30**) is a symmetrical disulfide compound bearing two hydroxyimino functional groups. This bis-bromotyrosine derivative was, almost simultaneously, first isolated in 1987 by three groups: from an unidentified marine sponge (probably of the Verongidae family) collected in Guam,<sup>224</sup> from a *Psammaplysilla* sp.,<sup>225</sup> and from *Thorectopsamma xana*.<sup>226</sup> Its structure has been unambiguously and independently established by these different authors. Thereafter, psammalin A has also been extracted from other sponges: *Psammaplysilla purpurea*,<sup>225,227</sup> *Dysidea* spp. (in this case, the authors have erroneously named «bisprasin»—the misspelled name of the psammalin A dimer—a compound which is obviously the psammalin A itself as judged by the reported formula)<sup>228</sup> *Aplysinella rhax*,<sup>229–231</sup> *Pseudoceratina purpurea*,<sup>232</sup> or from a two-sponge association: *Poecillastra wondoensis* and *Jaspis wondoensis*.<sup>233,234</sup> A biosynthetic pathway has been proposed for the formation of **30** involving modified cysteine and bromotyrosine<sup>227,232</sup> and, to our knowledge, only one laboratory preparation of psammalin A has been carried out starting from L-tyrosine through its *N,N'*-bis-(tetrahydropyran-2-yl)oxime derivative.<sup>235</sup> It is also worth pointing out that a library comprising about two hundred psammalin A type derivatives has recently been prepared by Nicolaou and his co-workers by using solution phase combinatorial synthesis with the aim to evaluate their antibacterial activity.<sup>235,236</sup>

In addition to its very recently reported ability to inhibit APN in a non-competitive manner thus inducing a suppression of *in vitro* angiogenesis,<sup>237</sup> **30** has been shown to induce a variety of biological effects: (i) a significant *in vitro* antibacterial activity against *Staphylococcus aureus*<sup>226</sup> and methicillin-resistant *Staphylococcus aureus*<sup>235,236,238</sup> which is assumed to be due to its ability to inhibit DNA gyrase,<sup>238</sup> (ii) a cytotoxicity against various human tumor cell lines,<sup>229,231–233</sup> (iii) an increase in Ca<sup>2+</sup> release from the heavy fraction of skeletal muscle sarcoplasmic reticulum,<sup>228</sup> (iv) an inhibition of topoisomerase II,<sup>239</sup> Leucine aminopeptidase and farnesyl protein transferase,<sup>229</sup> mycothiol-S-conjugate amidase,<sup>240</sup> chitinase,<sup>231</sup> histone deacetylase and DNA methyltransferase,<sup>232</sup>

and DNA replication by targeting polymerase  $\alpha$ -primase.<sup>241</sup> Some antifungal and insecticidal activities have been further reported.<sup>231</sup>

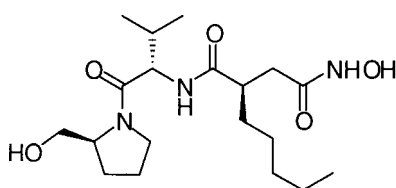
Chemical structures of APN inhibitors **13**–**30**, and enzyme inhibition values are depicted in Figure 4.

### B. Synthetic APN/CD13 Inhibitors

Several synthetic small molecules belonging to various chemical families have been reported to inhibit APN activity.

#### 1. $\alpha$ -Aminomethylketones

$\alpha$ -Aminomethylketones such as (*S*)-3-Amino-4-methylpentan-2-one hydrochloride (valine methyl ketone hydrochloride) (**31**)<sup>242</sup> have been described as potent competitive inhibitors of APN.<sup>243</sup>



**Actinonin (13)**

IC<sub>50</sub> APN : 0.4  $\mu$ g/mL, IC<sub>50</sub> LAP : 1  $\mu$ g/mL,

Inactive towards APA, APB, Methionine aminopeptidase and formylmethionine aminopeptidase<sup>94</sup>

IC<sub>50</sub> APN : 1.99  $\mu$ M, IC<sub>50</sub> APA : >100  $\mu$ M<sup>355</sup>

IC<sub>50</sub> APN : 0.4  $\mu$ g/mL, IC<sub>50</sub> APA : >100  $\mu$ g/mL, IC<sub>50</sub> APB : >100  $\mu$ g/mL<sup>137</sup>

IC<sub>50</sub> APN from *Bombyx mori* : 148  $\mu$ M<sup>368</sup>

IC<sub>50</sub> APN : 0.32  $\mu$ g/mL<sup>77</sup>

IC<sub>50</sub> Enkephalinaminopeptidase : 0.39  $\mu$ M,

IC<sub>50</sub> dipeptidylaminopeptidase : 1.1  $\mu$ M,

IC<sub>50</sub> Enkephalinase A : 5.6  $\mu$ M<sup>336</sup>

91% inhibition of APN, 16.5% inhibition of APA and 13% inhibition of MDP at 100  $\mu$ M, inactive towards APW<sup>369</sup>

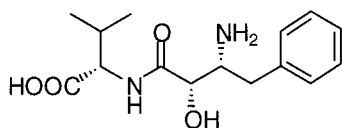
100% inhibition of APN from human seminal plasma at 100  $\mu$ M<sup>74</sup>

IC<sub>50</sub> Bacterial PDF 0.8-90 nM depending on the metal cation form of the enzyme<sup>370</sup>

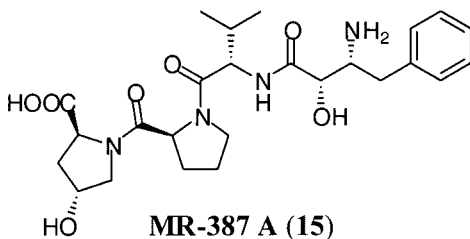
IC<sub>50</sub> Human PDF : 43 nM<sup>358</sup>

K<sub>i</sub> Meprin  $\alpha$  : 20 nM, K<sub>i</sub> Meprin  $\beta$  : 2  $\mu$ M<sup>360</sup>

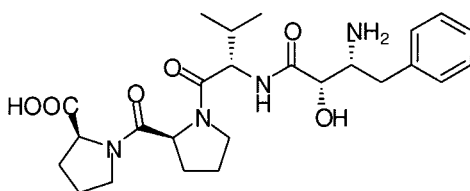
**Figure 4.** Natural inhibitors of APN/CD13.

**AHPA-Val (SL-387) (14)**

IC<sub>50</sub> APN from porcine kidney microsome :  
1.2 µg/mL, IC<sub>50</sub> APN from human  
fibrosarcoma HT1080 : 5.6 µg/mL, IC<sub>50</sub>  
APN from human myelogenous leukemia  
K562 : 7.8 µg/mL<sup>104</sup>

**MR-387 A (15)**

IC<sub>50</sub> APN from porcine kidney microsome :  
198 nM, IC<sub>50</sub> APN from human  
fibrosarcoma HT1080 : 218 nM, IC<sub>50</sub> APN  
from human myelogenous leukemia K562 :  
17 µM, APB from human myelogenous  
leukemia K562 : 651 nM<sup>105</sup>

**MR-387 B (16)**

IC<sub>50</sub> APN from porcine kidney microsome :  
164 nM, IC<sub>50</sub> APN from human  
fibrosarcoma HT1080 : 201 nM, IC<sub>50</sub> APN  
from human myelogenous leukemia K562 :  
4.6 µM, APB from human myelogenous  
leukemia K562 : 260 nM<sup>105</sup>

**Figure 4.** (Continued)

## 2. Alkyl D-Cysteines

Alkyl D-cysteines display also efficient competitive APN inhibitions. Among the five esters tested, an optimal inhibitory activity has been observed with the *n*-butyl derivative (**32**).<sup>244</sup>

## 3. 3-Amino-2-Tetralone Derivatives

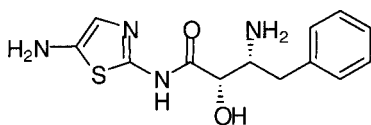
3-amino-2-tetralone derivatives such as the 2-amino-1,4-dihydro-2*H*-phenanthren-3-one hydrochloride (**33**) have been reported to be efficient and selective competitive inhibitors of APN. These compounds do not affect AP-A or AP-B and poorly inhibit LAPc.<sup>245</sup>

## 4. 3-Amino-2-Hydroxypropionaldehyde and 3-Amino-1-Hydroxypropan-2-One Derivatives

3-Amino-2-hydroxypropionaldehyde and 3-amino-1-hydroxypropan-2-one derivatives such as **34** and **35**, respectively. These competitive inhibitors of APN are very moderately active on LAPc or APB.<sup>246</sup>

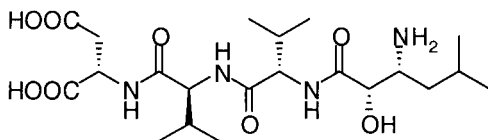
## 5. Flavone-8-Acetic Acid Derivatives

Flavone-8-acetic acid derivatives constitute a class of products whose the parent compound showed antiangiogenic properties.<sup>247</sup> In this series, products bearing a nitro group in the 2-position such as the 2',3-dinitroflavone-8 acetic acid (**36**) proved the most potent APN inhibitors and act by reversibly binding to the catalytic site of the enzyme. These compounds present the advantage to exhibit no



(17)

Inhibits the growth of mouse  $S_{180}$  tumors by 38%<sup>107</sup>

**Amastatin (18)**

$IC_{50}$  APN from pig kidney :  $0.5 \mu M$ <sup>371</sup>

$IC_{50}$  APN :  $3.16 \mu M$ ,  $IC_{50}$  APA :  $2 \mu M$ ,

$IC_{50}$  APW :  $1.58 \mu M$ <sup>355</sup>

$IC_{50}$  APN :  $150 nM$ <sup>372</sup>

$K_i$  APN :  $50 nM$ ,  $K_i$  LAP<sub>c</sub> :  $30 nM$ ,  $K_i$

APA :  $0.15 \mu M$ <sup>245</sup>

$IC_{50}$  APN :  $0.58 \mu g/mL$ ,  $IC_{50}$  APA :

$0.54 \mu g/mL$ ,  $IC_{50}$  APB :  $>100 \mu g/mL$ <sup>137</sup>

$IC_{50}$  APN from *Bombyx mori* :  $7.6 \mu M$ <sup>368</sup>

100% inhibition of APN, 100% inhibition of APA, 100% inhibition of APW and 18% inhibition of MDP at  $100 \mu M$ <sup>369</sup>

100% inhibition of APN from human seminal plasma at  $100 \mu M$ <sup>74</sup>

$K_i$  APN :  $52 nM$ ,  $K_i$  LAP<sub>c</sub> :  $30 nM$ ,  $K_i$

*Aeromonas* AP :  $0.26 nM$ <sup>357</sup>

$K_i^*$  APN :  $20 nM$ ,  $K_i^*$  LAP :  $0.2 \mu M$ <sup>108</sup>

$IC_{50}$  APM from rabbit kidney cortex :

$0.4 \mu M$ <sup>373</sup>

$IC_{50}$  APM from rat blood plasma :  $0.2 \mu M$ ,

$IC_{50}$  APA from rat blood plasma :  $8 \mu M$ <sup>67</sup>

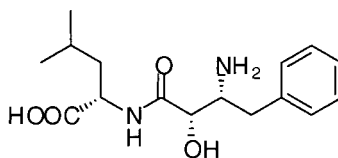
$IC_{50}$  APA :  $0.54 \mu g/mL$ ,  $IC_{50}$  APL :  $0.5$

$\mu g/mL$ ,  $IC_{50}$  APB :  $>250 \mu g/mL$ <sup>109</sup>

$IC_{50}$  APA :  $1.1 \mu M$ ,  $IC_{50}$  LAP :  $1.1 \mu M$ <sup>112,374</sup>

$IC_{50}$  APW :  $2 \mu M$ <sup>375</sup>

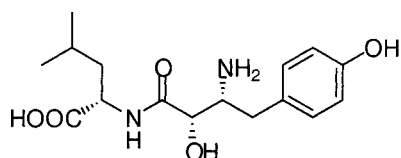
**Figure 4.** (Continued)



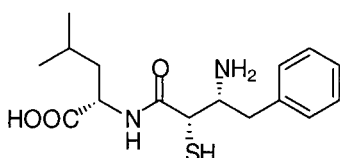
### Bestatin (19)

Ki APN : 3.03  $\mu\text{M}$ , Ki LAP<sub>c</sub> : 9 nM<sup>362</sup>  
 Ki<sup>\*</sup> APN : 4.1  $\mu\text{M}$ <sup>108</sup>  
 IC<sub>50</sub> APN from pig kidney : 16  $\mu\text{M}$ <sup>371</sup>  
 Ki APN : 1.45  $\mu\text{M}$ , Ki LAP<sub>c</sub> : 0.4 nM,  
 Ki *Aeromonas* AP : 18 nM<sup>357</sup>  
 Ki APN : 4.1  $\mu\text{M}$ , Ki LAP<sub>c</sub> : 20 nM,  
 Ki APB: 14 nM<sup>136</sup>  
 IC<sub>50</sub> APM from rabbit kidney cortex :  
 6  $\mu\text{M}$ <sup>373</sup>  
 IC<sub>50</sub> APN : 89.1  $\mu\text{M}$ , IC<sub>50</sub> APW : 7.9  $\mu\text{M}$ <sup>355</sup>  
 IC<sub>50</sub> APN : 43  $\mu\text{M}$ <sup>372</sup>  
 Ki APN : 3.5  $\mu\text{M}$ , Ki LAP<sub>c</sub> : 0.6 nM,  
 Ki APB : 6  $\mu\text{M}$ <sup>245</sup>  
 Ki APN : 3.5  $\mu\text{M}$ , Ki LAP<sub>c</sub> : 0.6 nM, Ki AP  
*Aeromonas* P. : 20 nM, Ki APB: 6  $\mu\text{M}$ <sup>246</sup>  
 IC<sub>50</sub> APN : 6.2  $\mu\text{g/mL}$ , IC<sub>50</sub> APA :  
 >100  $\mu\text{g/mL}$ , IC<sub>50</sub> APB : 0.05  $\mu\text{g/mL}$ <sup>137</sup>  
 IC<sub>50</sub> APN from *Bombyx mori* : 3.25 mM<sup>368</sup>  
 96% inhibition of APN from human seminal  
 plasma at 100  $\mu\text{M}$ <sup>74</sup>  
 52% inhibition of APN, 13.4% inhibition of  
 APA, 89% inhibition of APW and 29%  
 inhibition of MDP at 100  $\mu\text{M}$ <sup>369</sup>  
 IC<sub>50</sub> APN from rat blood plasma : 30  $\mu\text{M}$ <sup>67</sup>  
 IC<sub>50</sub> APN : 16.9  $\mu\text{M}$ <sup>210</sup>  
 IC<sub>50</sub> APN : 2.5  $\mu\text{M}$ <sup>190</sup>  
 IC<sub>50</sub> APN : 3.9  $\mu\text{M}$ , IC<sub>50</sub> A-LAP :  
 11.2  $\mu\text{M}$ <sup>249</sup>  
 IC<sub>50</sub> APB : 0.05  $\mu\text{g/mL}$ , IC<sub>50</sub> LAP :  
 0.01  $\mu\text{g/mL}$ <sup>119</sup>  
 Ki APB : 60 nM, Ki LAP : 20 nM<sup>353,356</sup>  
 IC<sub>50</sub> Enkephalin aminopeptidase : 1.1  $\mu\text{M}$ <sup>336</sup>  
 IC<sub>50</sub> APB : 0.05  $\mu\text{g/mL}$ , IC<sub>50</sub> LAP :  
 0.003  $\mu\text{g/mL}$ <sup>132</sup>  
 IC<sub>50</sub> APW : 6  $\mu\text{M}$ <sup>375</sup>

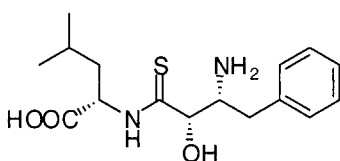
Figure 4. (Continued)

***para*-hydroxybestatin (20)**

IC<sub>50</sub> APB : 0.007 μg/mL, IC<sub>50</sub> LAP :  
0.02 μg/mL<sup>132</sup>

**α-Thiolbestatin (21)**

Ki APN : 4.4 μM, Ki LAP : 0.55 μM,  
Ki APB : 4.6 nM<sup>133</sup>

**Bestatin thioamide (22)**

Ki APN : 40.3 μM, Ki LAP : 0.33 μM,  
Ki APB : 2.4 μM<sup>133</sup>

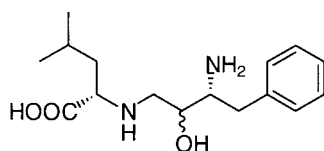
**Figure 4.** (Continued)

toxicity towards cultured human cells, to induce no apoptosis, and to be inactive on other proteases such as MMP-9, ACE, NEP, γ-glutamyl transpeptidase, cathepsin G, or DPPIV.<sup>248</sup>

#### 6. *N*-Hydroxy-2-(naphthalene-2-ylsulfanyl)Acetamide

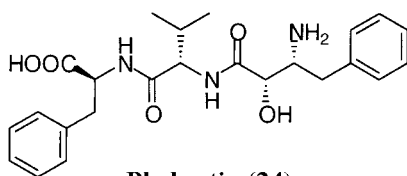
*N*-Hydroxy-2-(naphthalene-2-ylsulfanyl)acetamide (**37**) has recently been identified as a potent APN inhibitor. It acts in a dose-dependent manner and is inactive on metalloenzymes MMP-2, MMP-9, MMP-14, or A-LAP.<sup>249</sup>

The design of synthetic APN inhibitors has often been relied to structure–activity studies based on active site models derived from structural data obtained on the zinc-dependent protease thermolysin crystallized with a variety of inhibitors.<sup>250</sup> Molecules capable of interacting with at least the S<sub>1</sub> subsite of APN and which have a strong zinc-chelating group<sup>251,252</sup> were designed. According to these criteria, some α-aminophosphinic acids and derivatives such as **38** or **39**<sup>253</sup> have been prepared and proved to be very potent APN inhibitors. According to the patterns of these models, synthesis of analogs such as the iodo derivative **40** (*RB 129*) have next been performed to give rise to the radiolabelled (<sup>125</sup>I)RB 129<sup>254</sup> which represents a useful probe to investigate the physiological role of APN.<sup>13,255,256</sup> In the same context, several β-aminothiols exemplified by **41**<sup>257</sup> or **42**<sup>251</sup> have been conceived and synthesized. The research in this field has then been extended to more elaborated series by Roques and co-workers, and novel sulfur-containing molecules capable of inhibiting APN such as **43**, **44**,<sup>258</sup> **45**, **46**<sup>259</sup> or **47**<sup>253</sup> were prepared. From these works on β-aminothiols, two products emerged: *PC 18* (*S*)(2-amino-4-methylthiobutanethiol) (**48**)<sup>253</sup> and *EC 27* (*S*)(2-aminopentan-1,5-dithiol) (**49**).<sup>259</sup> These products have essentially aroused deeper studies because they are able to induce vasopressin release by acting on the half-life of angiotensin III.<sup>61,66,260,261</sup>



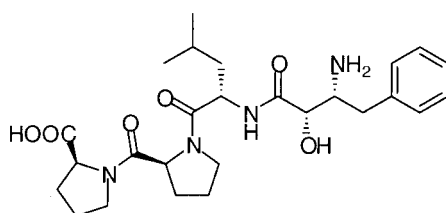
**23 (racemate)**

Ki APN : 2.1 mM, Ki LAP<sub>0</sub> : > 1 mM,  
Ki APB: 14 nM<sup>136</sup>



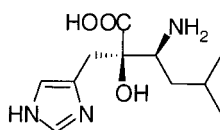
**Phebestin (24)**

IC<sub>50</sub> APN : 0.18 µg/mL, IC<sub>50</sub> APA :  
9 µg/mL, IC<sub>50</sub> APB : 9 µg/mL<sup>137</sup>



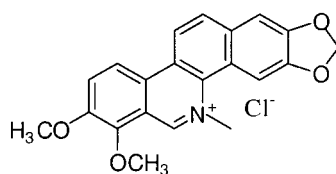
**Probestin (25)**

Ki APN : 19 nM<sup>138</sup>  
IC<sub>50</sub> APN : 50 nM, IC<sub>50</sub> APA : 19.9 µM,  
IC<sub>50</sub> APW : 5 µM<sup>355</sup>  
IC<sub>50</sub> APN : < 10 nM<sup>372</sup>  
IC<sub>50</sub> APN : 0.03 µg/mL, IC<sub>50</sub> APA : >100  
µg/mL, IC<sub>50</sub> APB : 37 µg/mL<sup>137</sup>  
IC<sub>50</sub> APN from *Bombyx mori* : 74 µM<sup>368</sup>



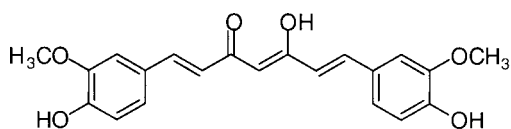
**Leuhistin (26)**

Ki APN : 0.23 µM<sup>186</sup>  
IC<sub>50</sub> APN : 0.2 µg/mL, IC<sub>50</sub> APA :  
10 µg/mL, IC<sub>50</sub> APB : 13 µg/mL<sup>137</sup>  
100% inhibition of APN from human  
seminal plasma at 100 µM<sup>74</sup>  
IC<sub>50</sub> APN from *Bombyx mori* : 0.89  
mM : 089 mM<sup>368</sup>



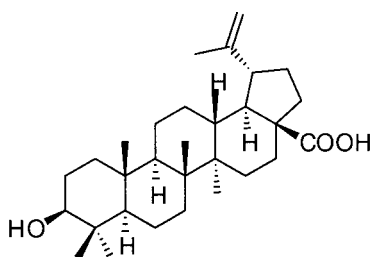
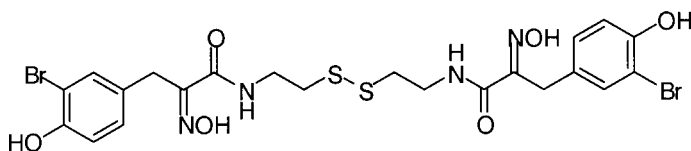
**Chelerythrine (27)**

82% inhibition of APN and 38% inhibition  
of DPP IV at 50 µM<sup>189</sup>



**Curcumin (28)**

Ki APN : 11.2 µM<sup>190</sup>

IC<sub>50</sub> APN : 7.3 μM<sup>210</sup>**Betulinic acid (29)**IC<sub>50</sub> APN : 18 μM<sup>237</sup>**Psammaplin A (30)***Figure 4. (Continued)*

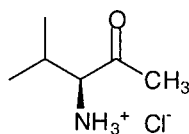
Formulas of the synthetic APN inhibitors **31–49**, and enzyme inhibition values are outlined in Figure 5.

### **C. Synthetic Dual APN/CD13 and E-24.11/CD10 (NEP) Inhibitors**

The similarities between the active sites of APN and the membrane-bound protease neutral endopeptidase 24.11 (EC3.4.24.11, CD10, NEP) led to the idea that mixed inhibitors could be developed by selecting frameworks bearing a strong zinc-chelating group and a residue able to interact with at least one subsite (S<sub>1</sub>, S<sub>1</sub>', and S<sub>2</sub>') of each peptidase.<sup>65,251,262–265</sup> The first dual E24.11/APN inhibitors developed were hydroxamate-containing molecules such as *Kelatorphan* (**50**) or *RB 38A* (**51**)<sup>262,266,267</sup> whose several analogs have been synthesized and found to be also potent inhibitors of leukotriene A<sub>4</sub> hydrolase.<sup>268</sup> However, the important water solubility of these compounds is an impediment for crossing the blood–brain barrier and, consequently, for obtaining a good bioavailability. Another strategy, involving more lipophilic derivatives, led to the synthesis of *RB 101* (*N*-((*R,S*)-2-benzyl-3((*S*)-2-amino-4-methylthio)butyldithio)-1-oxopropyl)-*L*-phenylalanine benzyl ester (**52**) and *RB 120* (*N*-((*S*)-2-benzyl-3((*S*)-2-amino-4-methylthio)butyldithio)-1-oxopropyl)-*L*-alanine benzyl ester (**53**), two dual inhibitors in which a disulfide bridge links the APN inhibitor PC 18 with analogs (the phenylalanine analog (ST 43) in the case of **52**, or the alanine analogue in the case of **53**) of the benzyl ester of Thiorphan, a specific NEP inhibitor<sup>269</sup> (Fig. 6).<sup>251,263,270</sup> Such mixed inhibitors present the advantage to possess the above-mentioned disulfide bond which is relatively stable in plasma, in contrast to its rapid cleavage in brain, thus allowing the delivery of the NEP and APN inhibitors in their active form toward their respective target.<sup>263</sup> The development of such mixed inhibitors has constituted an important advance in the research of new antihypertensives and novel antinociceptive drugs devoid of opioid side effects.<sup>264,271</sup> (for reviews) More recently, a new generation of phosphinic acid derivatives have been prepared as NEP/APN dual inhibitors, and compounds such as **54** have been successfully tested in this context.<sup>252,272</sup>

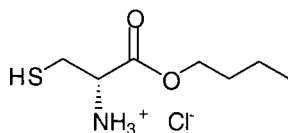
Chemical structures of APN inhibitors **50–54**, and enzyme inhibition values are outlined in Figure 7.





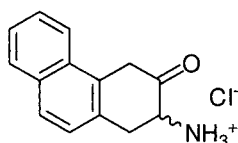
**31**

Ki APN : 0.55  $\mu\text{M}$ <sup>243</sup>



**32**

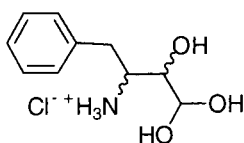
Ki APN : 0.18  $\mu\text{M}$ <sup>244</sup>



**33**

Ki APN : 0.5  $\mu\text{M}$ , Ki LAP<sub>c</sub> : 120  $\mu\text{M}$ , Ki APA : > 1 mM,  
Ki APB : > 1 mM<sup>245</sup>

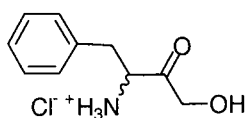
(absolute stereochemistry not specified)



**34**

Ki APN : 3  $\mu\text{M}$ , Ki LAP<sub>c</sub> : 0.1 mM, Ki AP *Aeromonas p.* :  
30  $\mu\text{M}$ <sup>246</sup>

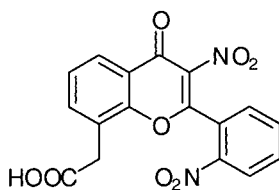
(absolute stereochemistry not specified)



**35**

Ki APN : 1  $\mu\text{M}$ <sup>246</sup>

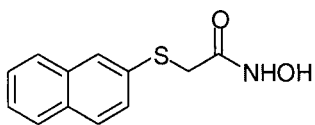
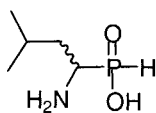
(absolute stereochemistry not specified)



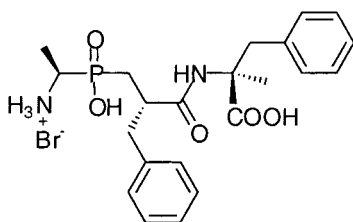
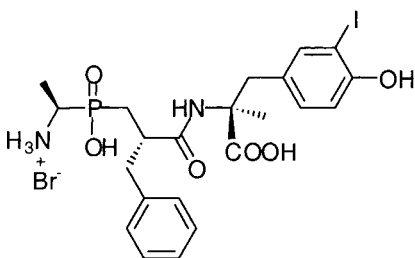
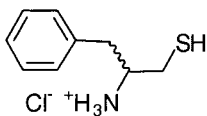
**36**

IC<sub>50</sub> APN : 25  $\mu\text{M}$ <sup>248</sup>

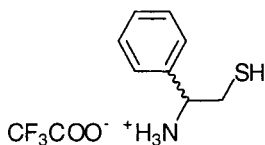
**Figure 5.** Synthetic inhibitors of APN/CD13.

**37**IC<sub>50</sub> APN : 3.4 μM<sup>249</sup>**38**

(mixture of R + S isomers)

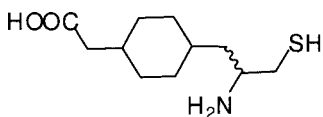
K<sub>i</sub> APN : 1.2 μM<sup>253</sup>**39**K<sub>i</sub> APN : 0.6 nM, K<sub>i</sub> APA : 0.13 μM, K<sub>i</sub> APB : >10 μM<sup>253</sup>**RB 129 (40)**K<sub>i</sub> APN : 0.95 nM<sup>254</sup>**Phetiol (41)**

(absolute stereochemistry not specified)

K<sub>i</sub> APN : 5 nM, K<sub>i</sub> PSA : 10 nM<sup>257</sup>**42**

(absolute stereochemistry not specified)

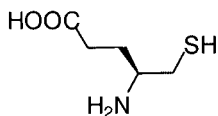
IC<sub>50</sub> APN : 25 nM<sup>251</sup>



**43**

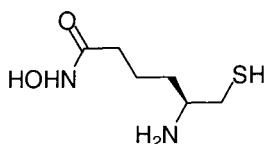
(mixture of R + S isomers)

Ki APN : 35 nM, Ki APA : 2.8  $\mu\text{M}$ <sup>258</sup>



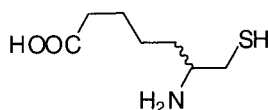
**44**

Ki APN : 0.12  $\mu\text{M}$ , Ki APA : 0.14  $\mu\text{M}$ <sup>258</sup>



**45**

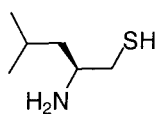
Ki APN : 37 nM, Ki APA : 2.0  $\mu\text{M}$ <sup>259</sup>



**46**

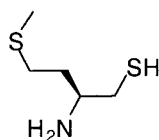
(mixture of R + S isomers)

Ki APN : 0.28  $\mu\text{M}$ , Ki APA : 1.6  $\mu\text{M}$ <sup>259</sup>



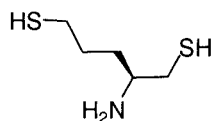
**47**

Ki APN : 22 nM<sup>253</sup>



**PC 18 (48)**

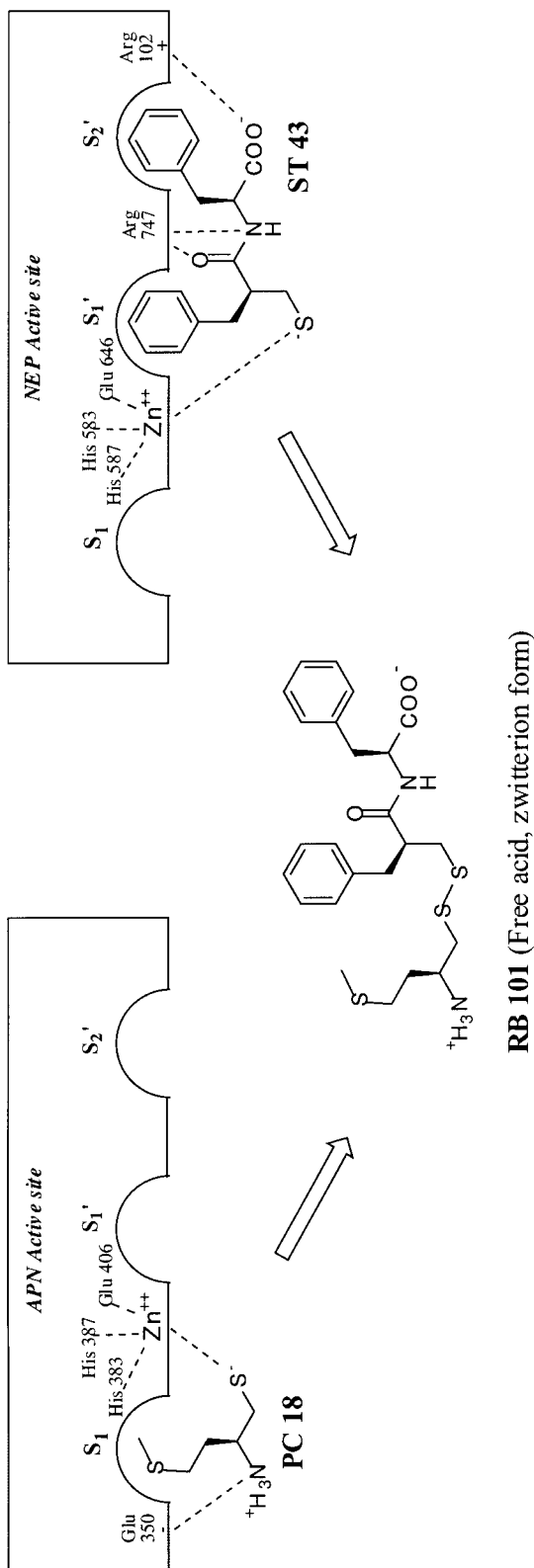
Ki APN : 8 nM, Ki APA : 17.2  $\mu\text{M}$ , Ki APB : 1  $\mu\text{M}$ <sup>260</sup>  
Ki APN : 11 nM<sup>339</sup>



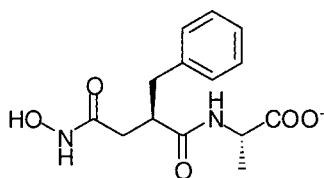
**EC 27 (49)**

Ki APN : 32 nM, Ki APA : 2.4  $\mu\text{M}$ <sup>259,261</sup>

*Figure 5. (Continued)*



**Figure 6.** Conceptualization of APN/NEP dual inhibitors (example of **RB 101**).



**Kelatorphan (50)**

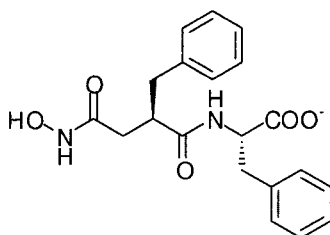
Ki APN : 0.38  $\mu$ M, Ki NEP : 1.8 nM<sup>264</sup>

Ki APN : 7  $\mu$ M, Ki NEP : 1.7 nM<sup>339</sup>

Ki APN : 0.38  $\mu$ M, Ki DAP : 0.9 nM, Ki NEP : 2.0 nM<sup>376</sup>

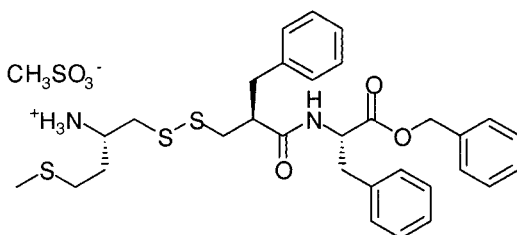
IC<sub>50</sub> NEP : 46 nM, IC<sub>50</sub> LTA<sub>4</sub> hydrolase : 5 nM, IC<sub>50</sub>

Aminopeptidase : 7 nM, IC<sub>50</sub> ACE : >10 $\mu$ M<sup>268</sup>



**RB 38A (51)**

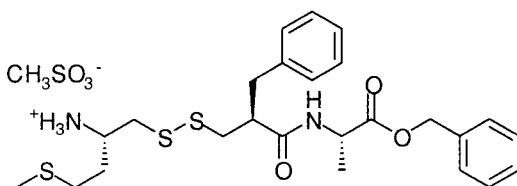
Ki APN : 0.12  $\mu$ M, Ki NEP : 0.9 nM<sup>264</sup>



**RB 101 (52)**

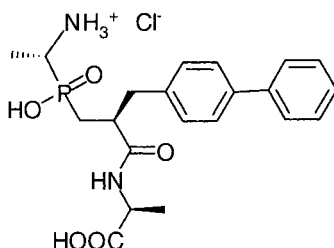
IC<sub>50</sub> APN : 16 $\mu$ M, IC<sub>50</sub> NEP : >100 $\mu$ M<sup>267</sup>

After 15 min of preincubation with whole rat brain membrane, IC<sub>50</sub> APN : 11 nM, IC<sub>50</sub> NEP : 4 nM<sup>267</sup>



**RB 120 (53)**

To our knowledge, no quantitative specific value dealing with APN or NEP inhibition has been published yet.



**54**

Ki APN : 2.9 nM, Ki NEP : 1.2 nM<sup>272</sup>

Ki APN : 2.9 nM, Ki NEP : 1.2 nM, Ki ACE : 0.12  $\mu$ M<sup>252</sup>

Ki APN : 2.9 nM, Ki NEP : 1.2 nM<sup>339</sup>

Ki APN : 2.9 nM, Ki APA : >1 $\mu$ M, Ki NEP : 1.2 nM,

Ki ACE : 0.12  $\mu$ M<sup>376</sup>

**Figure 7.** Dual inhibitors of APN/CD13.

#### 4. APN/CD13 INHIBITORS IN MODULATION OF CELL FUNCTIONS

The effects of some of these above described inhibitors on cell behavior have been assayed in *in vitro* approaches. Table I provides a summary of most relevant studies in the human system.

##### A. APN Inhibitors as Modulators of Cell Growth and Maturation

Actinonin, bestatin, probestin, and psammaplin A (at 1–100  $\mu$ M concentrations) were shown to reduce the growth of human T/B lymphocytes, dendritic and cord blood CD34<sup>+</sup> cells<sup>273–276</sup> and human myeloid and lymphoid cell lines,<sup>273,274,276–282</sup> as well as the proliferation of

**Table I.** Effects of APN/CD13 Inhibitors in *in vitro*, in Animal and Clinical Approaches

INHIBITOR	IN VITRO	IN ANIMALS	IN CLINICAL TRIALS
Actinonin	growth ↓ apoptosis ↑ migration ↑ invasion ↓	tumor growth ↓  neovascularization ↓ pain management ↓	
Amastatin	chemotaxis ↑ angiogenesis ↓	blood pressure ↓	
Bestatin (ubemimex)	growth ↓ differentiation ↑ apoptosis ↑ migration ↑↓ invasion ↓ angiogenesis ↑↓	tumor growth ↓ fetal growth ↓ monocyte activation ↑ placental apoptosis ↑ neutrophil migration ↓ neovascularization ↓ inflammation ↓	Remission in : AML lymphoma ( <i>monocyte &amp; lymphocyte activation</i> ) lung carcinoma
Betulinic acid	apoptosis ↑ angiogenesis ↓	tumor growth ↓	
Curcumin	angiogenesis ↓		
Leuhistin	invasion ↓	neutrophil migration ↓ inflammation ↓	
Probestin	growth ↓		
Psammaplin A	growth ↓ angiogenesis ↓		
PC 18 & EC 27 RB 101& RB 120		blood pressure ↓ pain management ↓	

AML, acute myeloid leukemia.

↓ Decrease ; ↑ Increase.

keratinocytes<sup>283,284</sup> and various tumor and endothelial cell lines.<sup>237,285–287</sup> A question central to APN inhibition studies is how cell growth can be turned off by APN inhibitors. APN inhibitors may alter the processing of (unknown) growth factors directly involved in the regulation of growth. In addition, several studies indicate that inhibitors like actinonin and probestin may transmit intracellular-transduction signals by interfering with the MAP kinase signaling pathway.<sup>25,279,288,289</sup> A second cell signaling pathway involving the Wnt-5a proto-oncogene appears also affected by inhibition of APN by actinonin.<sup>290</sup>

It has to be pointed out that actinonin (at a 10  $\mu\text{M}$  concentration) inhibited the growth of both CD13-positive myeloid and CD13-negative lymphoma cell lines<sup>287</sup> suggesting that the effects induced by actinonin are not likely to be mediated by CD13. Moreover, amastatin at a concentration which inhibits APN activity was found without any effect on the growth of human myeloid cell lines<sup>274,291</sup>.

Bestatin-mediated cell growth arrest is associated with an induction of cell maturation of clonogenic GM-CFU (granulocyte-macrophage colony forming unit) cells from human immature derived-bone marrow cells.<sup>292,293</sup> Similarly, treatment of human myeloid U937 and NB4 cell lines with bestatin induced phenotypic changes characteristic of macrophage (U937) or neutrophil (NB4) maturation.<sup>280,293,294</sup>

### ***B. Effects of APN Inhibitors on Cell Secretion***

Cell growth arrest induced by APN inhibitors correlates with alternated secretion of proinflammatory and immunosuppressive cytokines involved in pathophysiological processes. Bestatin (2.9  $\mu\text{M}$ ) increased the levels of IL-8 secreted by endothelial cells,<sup>295</sup> and of IL-1 release from mouse peritoneal macrophages and IL-2 release from concanavalin-stimulated T cells.<sup>296</sup> Probestin induces the synthesis and release of TGF- $\beta$ 1.<sup>41,297</sup>

### ***C. Effects of APN Inhibitors on Apoptosis***

Recent observations point to the involvement of APN in the process of apoptosis (programmed cell death). Bestatin and actinonin (starting 30  $\mu\text{M}$ ) induce apoptosis in a large variety of cell lines, i.e. myeloid (P39/TSU, HL-60, U937, NB4) and lymphoid (Jurkat, BJAB, NALM6, BOE) cells, and carcinoma (fibrosarcoma, cervical, and lung carcinoma).<sup>274,282,287,291,298,299</sup> Betulinic acid induces apoptosis in the HT29 colon cancer cell line (26  $\mu\text{M}$ )<sup>84</sup> and in acute leukemia cells (50  $\mu\text{M}$ ).<sup>300</sup>

### ***D. Effects of APN Inhibitors on Cell Motility***

In a general way, cell motility (migration and invasion) may be influenced by the processing of chemokines and/or degradation of the extracellular matrix (ECM). The two small proteins with chemotactic activity, MCP-1 and f-MLP, are *in vitro* hydrolyzed by APN/CD13. With regard to MCP-1, there is no current data reporting the potential action of APN inhibitors on the MCP-1-mediated migration. Actinonin and amastatin were able to enhance the chemotactic response of human neutrophils toward f-MLP.<sup>301</sup> One explanation of the effects of actinonin or amastatin would be that both inhibitors prevent the inactivation of f-MLP by APN, to further enhance the f-MLP-mediated chemotactic response. It has however to underline that both inhibitors weakly inhibited APN enzymatic activity over the range from  $10^{-8}$  to  $10^{-4}$  M, concentrations that are effective on neutrophil migration.<sup>301</sup>

APN inhibition by actinonin or bestatin significantly enhanced the *in vitro* migration of eosinophils across HUVEC monolayers.<sup>302</sup> Moreover, actinonin, bestatin as well as leuhistin (50–150  $\mu\text{M}$ ) significantly blocked the invasion of various human metastatic tumor cells into reconstituted basement membranes<sup>303,304</sup> or into Matrigel.<sup>21,305–307</sup> These latter data suggested that

APN could be indirectly involved in type IV collagen degradation by activating type IV procollagenase/proMMP-9.<sup>17,303,304</sup> Recent studies demonstrated that soluble APN/CD13 induces *in vitro* chemotactic migration of T lymphocytes, and that bestatin at high concentration (580  $\mu\text{M}$ ) abolishes this process, suggesting that the enzymatic activity of APN was responsible for the chemotactic activity.<sup>34,36,304,308</sup>

## 5. APN INHIBITORS AND ANGIOGENESIS

The demonstration of the participation of APN in angiogenesis has come from recent studies in which blocking APN activity by APN inhibitors resulted in the perturbation of “angiogenic” assays (Table I).

### A. *In Vitro* Assays

APN/CD13 is expressed on the human umbilical vein endothelial cells (HUVECs) of angiogenic, but not normal, vasculature.<sup>309</sup> Bestatin, betulinic acid, amastatin, curcumin, and psammaplin A (10–250  $\mu\text{M}$ ) abrogate the ability of the HUVECs cultured on matrigel to organize a capillary network<sup>20,190,237,310–312</sup> without altering their proliferation rates.<sup>310</sup> In contrast, one study underlines the proangiogenic effect of bestatin (8–250  $\mu\text{M}$ ) which instead causes matrix degradation and stimulates the invasion of microvascular endothelial cells into a fibrin matrix.<sup>313</sup>

### B. *In Vivo* Assays

In the chorioallantoic membrane (CAM) assay, the angiogenic response is determined by measuring the number of avian extraembryonic capillary vessels that grow within a matrix polymer (containing an angiogenic molecule such as fibroblast growth factor-2/FGF-2) placed on the yolk sac membrane of a 4 day embryo in culture.<sup>314</sup> The chick vasculature expresses a phenotype APN/CD13, and subsequent treatment with bestatin or actinonin (200  $\mu\text{g}$ ) inhibited FGF-2-induced angiogenesis.<sup>309</sup> In the mouse retinal neovascularization model, bestatin (200  $\mu\text{g}/\text{mouse}$ ) leads to the blockade of hypoxia-induced retinal neovascularization in mice.<sup>309</sup> The intraperitoneal administration of bestatin (50–100 mg/kg/day) after the orthotopic implantation of B16-BL6 melanoma cells into mice reduces the number of vessels oriented toward the established primary tumor mass on the dorsal side of mice.<sup>311</sup>

## 6. EFFECTS OF APN/CD13 INHIBITORS IN ANIMAL MODELS

Compiled data documenting the involvement of APN/CD13 in pathophysiological events (cancer, inflammation, infection, pain suppression) have come from studies which blocked APN activity in rodent models (Table I).

Studies in rats indicate that administration of bestatin leads to the inhibition of fetal growth and the induction of placental apoptosis.<sup>315,316</sup> The *in vivo* anti-cancer activities of bestatin and betulinic acid have been reported through their capacities to inhibit the growth of syngeneic tumor (leukemia/melanoma/ovarian/hepatoma/gastric carcinoma) cells implanted in mice<sup>16,213,309,310,317–325</sup> and rats.<sup>319,326,327</sup> Doses as low as 0.5 mg/kg for bestatin and 5 mg/kg for betulinic acid were used in these studies. Moreover, high doses (up to 500 mg/kg) did not lead to any cytotoxic effect in mice.

Bestatin, leuhistin, and betulinic acid have been investigated for anti-inflammatory properties. Betulinic acid possessed moderate ant-inflammatory abilities at relatively high concentrations



(100 mg/kg/mouse, i.v.).<sup>213</sup> In contrast, bestatin and leuhistin inhibit acute inflammation associated with the accumulation of polymorphonuclear neutrophils in a mouse model (2 mg/kg, i.v.).<sup>57,328</sup> Moreover, oral administration of bestatin (5 mg/kg) in carcinoma-bearing mice induces generation of cytotoxic T cells and NK (natural killer) cells.<sup>317</sup>

Angiotensins II and III are two peptide effectors of the brain rennin–angiotensin system that participate in the control of blood pressure, increase water consumption and vasopressin release. In hypertensive rats, infusion of amastatin (16 nmol/min i.v.) prevents degradation of angiotensins associated with blood pressure decrease.<sup>67,329</sup> In the mouse brain, APN inhibition by PC18 or EC27 (10–300 µg injected intracerebroventricularly) increases the half life of angiotensin III, resulting in enhanced vasopressin release.<sup>61,66,260</sup>

Several studies report that bestatin exerts anti-infectious properties by augmenting host resistance to bacterial, viral or fungal experimental infections in mice by inducing neutrophil and macrophage activation<sup>330,331</sup> and enhancing antibody production.<sup>330–335</sup>

Finally, in the central nervous system, enkephalins which modulate responses to painful stimuli, are inactivated by APN and the membrane-bound protease neutral endopeptidase 24.11 (EC3.4.24.11, CD10). This led to the idea that inhibition of these enzymes (alone or in combination) could achieve clinically efficient analgesia. Actinonin as well as the dual inhibitors RB101 and RB120 (9 mg/kg, i.v.; 80 mg/kg, i.p.) exhibited analgesic properties against chronic pain in rats and mice.<sup>261,263,267,336,341</sup>

## 7. EFFECTS OF BESTATIN IN CLINICAL TRIALS

In first clinical trials, bestatin (30 mg/daily) has been used to treat patients with acute and chronic myeloid leukemias (AML, CML) and lymphomas.<sup>342–346</sup> Therapeutic efficacy was demonstrated by a prolongation of survival in patients with AML<sup>345,346</sup> and lymphomas,<sup>342,343,347</sup> and in promoting graft versus leukemia effects in patients following allogeneic bone marrow transplant.<sup>348</sup>

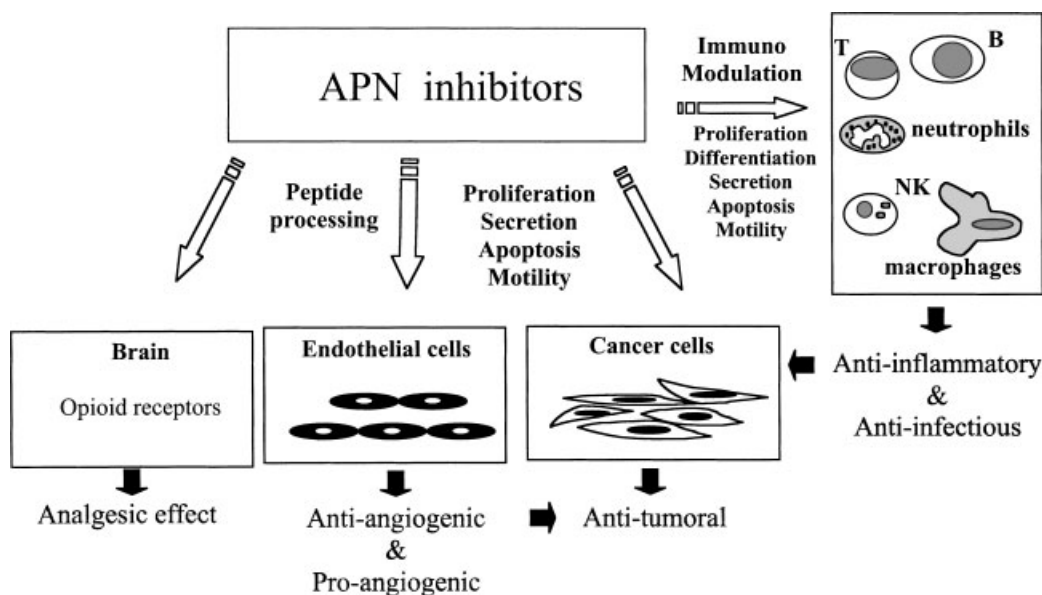
In a phase Ib trial, activation of blood monocytes and increase in the CD4/CD8 lymphocyte ratio were observed in Hodgkin's and non-Hodgkin's lymphoma patients treated orally with high doses of bestatin (90–180 mg/daily/60 days) following autologous bone marrow transplantation.<sup>330,334,349</sup>

In phase III trials in resected stage I squamous cell lung carcinoma, survival was statistically better for patients who were treated with bestatin (30 mg/daily/2 years) as a post-operative adjuvant therapy than those who received a placebo.<sup>350,351</sup>

## 8. CONCLUDING COMMENTS AND PERSPECTIVES

APN/CD13, is useful in defining clinical subgroups of patients with various malignancies or inflammatory diseases. The use of natural and synthetic APN inhibitors has revealed that APN/CD13 participates to the control of major biological processes such as proliferation, secretion and apoptosis. Dysregulation of APN/CD13 in tumors is often linked to tumor invasion and angiogenesis. Studies on non-hematopoietic cells suggest that APN/CD13 may influence cell migration and invasion. APN/CD13 inhibitors have been shown to alter angiogenesis in *in vitro* and *in vivo* assays. Documented evidence underlines both the antiangiogenic and proangiogenic effects of bestatin.<sup>309,310,313</sup> Figure 8 summarizes our current understanding of the involvement of APN inhibitors in the modulation of these events. The detailed molecular mechanisms underlying these effects are however yet unclear.

Importantly, the requirement for APN in these processes has been mostly confirmed with studies in which APN/CD13 expression was blocked by neutralizing CD13 antibodies<sup>20,285,303,309,310</sup> or antisense CD13 oligonucleotides,<sup>20,41,352</sup> or enhanced by the use of CD13 transfectants.<sup>17</sup>



**Figure 8.** Biological effects of APN/CD13 inhibitors. The actions of APN inhibitors *in vitro* and *in vivo* (animal models) are diverse; they may directly target cancer cells, or act indirectly against targets by activation of immune cells (T, B, neutrophils, natural killer/NK cells, macrophages) or alteration of angiogenesis (endothelial cells). In the brain, APN inhibitors exhibit analgesic properties by increasing the levels of Met-enkephalin.

It has however to be pointed out that most of APN inhibitors lack tight specificity by inhibiting other membrane-bound metalloproteases or secreted matrix metalloproteinases (MMPs) (Table I). For example, bestatin interacts with leucyl-aminopeptidase (EC3.4.11.1, oxytocinase, Leu-AP), aminopeptidase B (EC 3.4.11.6, AP-B) and aminopeptidase W (EC 3.4.11.16, AP-W)<sup>136,353–357</sup> thus suggesting that some of the observed chemotherapeutic actions of bestatin may be due to inhibition of other cell surface peptidases. Actinonin was recently shown to interact with human peptide deformylase,<sup>358,359</sup> meprin  $\alpha$  (EC 3.4.24.18, endopeptidase 24.18),<sup>360</sup> and MMP-2.<sup>361</sup> Amastatin and probestin in the low micromolar range (1.5–20  $\mu\text{M}$ ) inhibit aminopeptidase A (EC 3.4.11.2, AP-A) and AP-W.<sup>109,355,362</sup> Leuhistin inhibits AP-A and AP-B to the same degree than APN.<sup>186</sup> Curcumin and betulinic acid block MMP-9 expression and collagenase activity through inhibition of NF- $\kappa$ B activation.<sup>363–367</sup> In addition, the use of available APN inhibitors in some experimental situations has revealed complex effects on cell behavior. As mentioned in paragraph 4.A, CD13-positive and CD13-negative cell lines are equally sensitive to the growth-inhibitory effect of actinonin (50–260  $\mu\text{M}$ )<sup>287</sup> thus emphasizing that actinonin may induce unspecific cytotoxic side-effects. Moreover, betulinic acid inhibits tube formation of bovine aortic endothelial cells at a concentration which had no effect on the cell viability and *in vivo* APN activity of endothelial cells, thus indicating an APN-independent mode of action of betulinic acid.<sup>312</sup>

Together, these observations emphasize the need for more specific and targeted APN inhibitors to (re)evaluate the actions of APN/CD13 in pathophysiological processes. Future consideration has to be given to the obtention of the three-dimensional structure of APN determined by NMR spectroscopy to help APN inhibitor design strategy. Further *in vitro* and *in vivo* studies with promising non cytotoxic APN inhibitors (such as psammaphin A, phosphonic derivatives, flavone-8-acetic acid derivatives) are also required before clinically prescribing an APN inhibitor as an anti-cancer or anti-inflammatory agent.

**ACKNOWLEDGMENTS**

This work was supported by grants from the Institut National de la Santé et de la Recherche Médicale (I.N.S.E.R.M.), the Centre National de La Recherche Scientifique (C.N.R.S.), the Institut Curie, the Association pour la Recherche sur le Cancer (ARC number 3473), la Fondation pour la Recherche Médicale and La Ligue Nationale Contre le Cancer.

**REFERENCES**

1. Antczak C, De Meester I, Bauvois B. Ecto-peptidases in pathophysiology. *Bioessays* 2001;23(3):251–260.
2. Rawlings ND, Barrett AJ. MEROPS: The peptidase database. *Nucleic Acids Res* 1999;27(1):325–331.
3. Hooper NM. Families of zinc metalloproteases. *FEBS Lett* 1994;354(1):1–6.
4. Olsen J, Cowell GM, Konigshofer E, Danielsen EM, Moller J, Laustsen L, Hansen OC, Welinder KG, Engberg J, Hunziker W, et al. Complete amino acid sequence of human intestinal aminopeptidase N as deduced from cloned cDNA. *FEBS Lett* 1988;238(2):307–314.
5. Pfeleiderer G, Celliers PG. Isolation of an aminopeptidase from kidney particles. *Biochem Z* 1963;339:186–189.
6. Look AT, Ashmun RA, Shapiro LH, Peiper SC. Human myeloid plasma membrane glycoprotein CD13 (gp150) is identical to aminopeptidase N. *J Clin Invest* 1989;83(4):1299–1307.
7. Shipp MA, Look AT. Hematopoietic differentiation antigens that are membrane-associated enzymes: Cutting is the key! *Blood* 1993;82(4):1052–1070.
8. Jung K, Pergande M, Wischke UW. Characterization of particulate and soluble variants of the brush-border enzymes alanine aminopeptidase, alkaline phosphatase and gamma-glutamyltransferase in human urine. *Biomed Biochim Acta* 1984;43(12):1357–1364.
9. Favaloro EJ, Browning T, Facey D. CD13 (GP150; aminopeptidase-N): Predominant functional activity in blood is localized to plasma and is not cell-surface associated. *Exp Hematol* 1993;21(13):1695–1701.
10. Kawai M, Otake Y, Hara Y. High-molecular-mass isoform of aminopeptidase N/CD13 in serum from cholestatic patients. *Clin Chim Acta* 2003;330(1-2):141–149.
11. van Hensbergen Y, Broxterman HJ, Hanemaaijer R, Jorna AS, van Lent NA, Verheul HM, Pinedo HM, Hoekman K. Soluble aminopeptidase N/CD13 in malignant and nonmalignant effusions and intratumoral fluid. *Clin Cancer Res* 2002;8(12):3747–3754.
12. Antczak C, De Meester I, Bauvois B. Transmembrane proteases as disease markers and targets for therapy. *J Biol Regul Homeost Agents* 2001;15(2):130–139.
13. Jardinaud F, Banisadr G, Noble F, Melik-Parsadaniantz S, Chen H, Dugave C, Laplace H, Rostene W, Fournie-Zaluski MC, Roques BP, Popovici T. Ontogenic and adult whole body distribution of aminopeptidase N in rat investigated by in vitro autoradiography. *Biochimie* 2004;86(2):105–113.
14. Razak K, Newland AC. The significance of aminopeptidases and haematopoietic cell differentiation. *Blood Rev* 1992;6(4):243–250.
15. Razak K, Newland AC. Induction of CD13 expression on fresh myeloid leukaemia: Correlation of CD13 expression with aminopeptidase-N activity. *Leuk Res* 1992;16(6-7):625–630.
16. Menrad A, Speicher D, Wacker J, Herlyn M. Biochemical and functional characterization of aminopeptidase N expressed by human melanoma cells. *Cancer Res* 1993;53(6):1450–1455.
17. Fujii H, Nakajima M, Saiki I, Yoneda J, Azuma I, Tsuruo T. Human melanoma invasion and metastasis enhancement by high expression of aminopeptidase N/CD13. *Clin Exp Metastasis* 1995;13(5):337–344.
18. Kitamura Y, Watanabe M, Komatsubara S, Sakata Y. [Urinary excretion of glycine, proline dipeptide, aminopeptidase, N-acetyl-beta-D-glucosaminidase, alanine aminopeptidase, and low molecular protein in patients with renal cell carcinoma]. *Hinyokika Kyo* 1990;36(5):535–539.
19. Ikeda N, Nakajima Y, Tokuhara T, Hattori N, Sho M, Kanehiro H, Miyake M. Clinical significance of aminopeptidase N/CD13 expression in human pancreatic carcinoma. *Clin Cancer Res* 2003;9(4):1503–1508.
20. Hashida H, Takabayashi A, Kanai M, Adachi M, Kondo K, Kohno N, Yamaoka Y, Miyake M. Aminopeptidase N is involved in cell motility and angiogenesis: Its clinical significance in human colon cancer. *Gastroenterology* 2002;122(2):376–386.
21. Ishii K, Usui S, Sugimura Y, Yamamoto H, Yoshikawa K, Hirano K. Inhibition of aminopeptidase N (AP-N) and urokinase-type plasminogen activator (uPA) by zinc suppresses the invasion activity in human urological cancer cells. *Biol Pharm Bull* 2001;24(3):226–230.

22. Carl-McGrath S, Lendeckel U, Ebert M, Wolter AB, Roessner A, Rocken C. The ectopeptidases CD10, CD13, CD26, and CD143 are upregulated in gastric cancer. *Int J Oncol* 2004;25(5):1223–1232.
23. Kehlen A, Lendeckel U, Dralle H, Langner J, Hoang-Vu C. Biological significance of aminopeptidase N/CD13 in thyroid carcinomas. *Cancer Res* 2003;63(23):8500–8506.
24. Riemann D, Gohring B, Langner J. Expression of aminopeptidase N/CD13 in tumour-infiltrating lymphocytes from human renal cell carcinoma. *Immunol Lett* 1994;42(1–2):19–23.
25. Riemann D, Kehlen A, Langner J. CD13—not just a marker in leukemia typing. *Immunol Today* 1999;20(2):83–88.
26. Balog T, Marotti T, Sverko V, Marotti M, Krolo I, Rocic B, Karapanda N. Enkephalin degrading enzymes in pheochromocytoma patients. *Oncol Rep* 2003;10(1):253–258.
27. Boldt DH, Kopecky KJ, Head D, Gehly G, Radich JP, Appelbaum FR. Expression of myeloid antigens by blast cells in acute lymphoblastic leukemia of adults. The Southwest Oncology Group experience. *Leukemia* 1994;8(12):2118–2126.
28. Tatsumi E. A mini-review of CD13 antigen in AML: Easy induction or enhancement of expression in vitro culture and necessary consideration for assessment. *Southeast Asian J Trop Med Public Health* 2002;33(Suppl 2):155–157.
29. Shao Z, Chen G, Lin Z, Zhang Y, Hao Y, Chu Y, Zheng Y, Qian L, Yang T, Yang C, Feng B. Immunophenotype of myeloid cells in myelodysplastic syndromes and its clinical implications. *Chin Med J (Engl)* 1998;111(1):28–31.
30. Popnikolov NK, Payne DA, Hudnall SD, Hawkins HK, Kumar M, Norris BA, Elghetany MT. CD13-positive anaplastic large cell lymphoma of T-cell origin—a diagnostic and histogenetic problem. *Arch Pathol Lab Med* 2000;124(12):1804–1808.
31. Dunphy CH, Gardner LJ, Manes JL, Bee CS, Taysi K. CD30+ anaplastic large-cell lymphoma with aberrant expression of CD13: Case report and review of the literature. *J Clin Lab Anal* 2000;14(6):299–304.
32. Dan H, Tani K, Hase K, Shimizu T, Tamiya H, Biraa Y, Huang L, Yanagawa H, Sone S. CD13/aminopeptidase N in collagen vascular diseases. *Rheumatol Int* 2003;23(6):271–276.
33. Abe T, Yamamoto Y, Hazato T. [Changes in aminopeptidase N located on neutrophils derived from patients with chronic pain]. *Masui* 1998;47(2):151–155.
34. Shimizu T, Tani K, Hase K, Ogawa H, Huang L, Shinomiya F, Sone S. CD13/aminopeptidase N-induced lymphocyte involvement in inflamed joints of patients with rheumatoid arthritis. *Arthritis Rheum* 2002;46(9):2330–2338.
35. Ziaber J, Baj Z, Pasniki J, Chmielewski H, Tchorzewski H. Expression of aminopeptidase N (APN) on peripheral blood mononuclear cells' surface as a marker of these cells' transendothelial migration properties in the course of multiple sclerosis. *Mediators Inflamm* 2000;9(1):45–48.
36. Tani K, Ogushi F, Huang L, Kawano T, Tada H, Hariguchi N, Sone S. CD13/aminopeptidase N, a novel chemoattractant for T lymphocytes in pulmonary sarcoidosis. *Am J Respir Crit Care Med* 2000;161(5):1636–1642.
37. Riemann D, Schwachula A, Hentschel M, Langner J. Demonstration of CD13/aminopeptidase N on synovial fluid T cells from patients with different forms of joint effusions. *Immunobiology* 1993;187(1–2):24–35.
38. Hafler DA, Hemler ME, Christenson L, Williams JM, Shapiro HM, Strom TB, Strominger JL, Weiner HL. Investigation of in vivo activated T cells in multiple sclerosis and inflammatory central nervous system diseases. *Clin Immunol Immunopathol* 1985;37(2):163–171.
39. Riemann D, Wollert HG, Menschikowski J, Mittenzwei S, Langner J. Immunophenotype of lymphocytes in pericardial fluid from patients with different forms of heart disease. *Int Arch Allergy Immunol* 1994;104(1):48–56.
40. Lendeckel U, Kahne T, Riemann D, Neubert K, Arndt M, Reinhold D. Review: The role of membrane peptidases in immune functions. *Adv Exp Med Biol* 2000;477:1–24.
41. Lendeckel U, Arndt M, Frank K, Wex T, Ansoerge S. Role of alanyl aminopeptidase in growth and function of human T cells (review). *Int J Mol Med* 1999;4(1):17–27.
42. Goette A, Arndt M, Rocken C, Spiess A, Staack T, Geller JC, Huth C, Ansoerge S, Klein HU, Lendeckel U. Regulation of angiotensin II receptor subtypes during atrial fibrillation in humans. *Circulation* 2000;101(23):2678–2681.
43. Sjoström H, Noren O, Olsen J. Structure and function of aminopeptidase N. *Adv Exp Med Biol* 2000;477:25–34.
44. Vlahovic P, Stefanovic V. Kidney ectopeptidases. Structure, functions, and clinical significance. *Pathol Biol (Paris)* 1998;46(10):779–786.

45. Breljak D, Gabrilovac J, Boranic M. Aminopeptidase N/CD13 and haematopoietic cells. *Haema* 2003;6(4):453–461.
46. Watt VM, Willard HF. The human aminopeptidase N gene: Isolation, chromosome localization, and DNA polymorphism analysis. *Hum Genet* 1990;85(6):651–654.
47. Shapiro LH. Myb and Ets proteins cooperate to transactivate an early myeloid gene. *J Biol Chem* 1995;270(15):8763–8771.
48. Olsen J, Kokholm K, Troelsen JT, Laustsen L. An enhancer with cell-type dependent activity is located between the myeloid and epithelial aminopeptidase N (CD 13) promoters. *Biochem J* 1997;322(Pt 3):899–908.
49. Lendeckel U, Wex T, Arndt M, Frank K, Franke A, Ansorge S. Identification of point mutations in the aminopeptidase N gene by SSCP analysis and sequencing. *Hum Mutat* 1998; Suppl 1:S158–S160.
50. Shapiro LH, Ashmun RA, Roberts WM, Look AT. Separate promoters control transcription of the human aminopeptidase N gene in myeloid and intestinal epithelial cells. *J Biol Chem* 1991;266(18):11999–12007.
51. Hedge SP, Kumar A, Kurschner C, Shapiro LH. c-Maf interacts with c-Myb to regulate transcription of an early myeloid gene during differentiation. *Mol Cell Biol* 1998;18(5):2729–2737.
52. Firla B, Arndt M, Frank K, Thiel U, Ansorge S, Tager M, Lendeckel U. Extracellular cysteines define ectopeptidase (APN, CD13) expression and function. *Free Radic Biol Med* 2002;32(7):584–595.
53. Luciani N, Marie-Claire C, Ruffet E, Beaumont A, Roques BP, Fournie-Zaluski MC. Characterization of Glu350 as a critical residue involved in the N-terminal amine binding site of aminopeptidase N (EC 3.4.11.2): Insights into its mechanism of action. *Biochemistry* 1998;37(2):686–692.
54. Giros B, Gros C, Solhonne B, Schwartz JC. Characterization of aminopeptidases responsible for inactivating endogenous (Met5)enkephalin in brain slices using peptidase inhibitors and anti-aminopeptidase M antibodies. *Mol Pharmacol* 1986;29(3):281–287.
55. Furuhashi M, Mizutani S, Kurauchi O, Kasugai M, Narita O, Tomoda Y. In vitro degradation of opioid peptides by human placental aminopeptidase M. *Exp Clin Endocrinol* 1988;92(2):235–237.
56. Miller BC, Thiele DL, Hersh LB, Cottam GL. Methionine enkephalin is hydrolyzed by aminopeptidase N on CD4+ and CD8+ spleen T cells. *Arch Biochem Biophys* 1994;311(1):174–179.
57. Yamamoto Y, Kanazawa H, Shimamura M, Ueki M, Hazato T. Inhibitory action of spinorphin, an endogenous regulator of enkephalin-degrading enzymes, on carrageenan-induced polymorphonuclear neutrophil accumulation in mouse air-pouches. *Life Sci* 1998;62(19):1767–1773.
58. Wang LH, Ahmad S, Benter IF, Chow A, Mizutani S, Ward PE. Differential processing of substance P and neurokinin A by plasma dipeptidyl(amino)peptidase IV, aminopeptidase M and angiotensin converting enzyme. *Peptides* 1991;12(6):1357–1364.
59. Lucius R, Sievers J, Mentlein R. Enkephalin metabolism by microglial aminopeptidase N (CD13). *J Neurochem* 1995;64(4):1841–1847.
60. Robertson MJ, Cunoosamy MP, Clark KL. Effects of peptidase inhibition on angiotensin receptor agonist and antagonist potency in rabbit isolated thoracic aorta. *Br J Pharmacol* 1992;106(1):166–172.
61. Zini S, Fournie-Zaluski MC, Chauvel E, Roques BP, Corvol P, Llorens-Cortes C. Identification of metabolic pathways of brain angiotensin II and III using specific aminopeptidase inhibitors: Predominant role of angiotensin III in the control of vasopressin release. *Proc Natl Acad Sci USA* 1996;93(21):11968–11973.
62. Chansel D, Czekalski S, Vandermeersch S, Ruffet E, Fournie-Zaluski MC, Ardaillou R. Characterization of angiotensin IV-degrading enzymes and receptors on rat mesangial cells. *Am J Physiol* 1998;275(4 Pt 2):F535–F542.
63. Mizutani S, Taira H, Kurauchi O, Ito Y, Imaizumi H, Furuhashi M, Narita O, Tomoda Y. Effect of microsomal leucine aminopeptidase from human placenta (microsomal P-LAP) on pressor response to infused angiotensin II (A-II) in rat. *Exp Clin Endocrinol* 1987;90(2):206–212.
64. Montiel JL, Cornille F, Roques BP, Noble F. Nociceptin/orphanin FQ metabolism: Role of aminopeptidase and endopeptidase 24.15. *J Neurochem* 1997;68(1):354–361.
65. Waksman G, Hamel E, Bouboutou R, Besselievre R, Fournie-Zaluski MC, Roques BP. Regional distribution of enkephalinase in the rat brain by autoradiography. *C R Acad Sci III* 1984;299(14):613–615.
66. Llorens-Cortes C. Identification of metabolic pathways of brain angiotensin II and angiotensin III: Predominant role of angiotensin III in the control of vasopressin secretion. *C R Seances Soc Biol Fil* 1998;192(4):607–618.
67. Ahmad S, Ward PE. Role of aminopeptidase activity in the regulation of the pressor activity of circulating angiotensins. *J Pharmacol Exp Ther* 1990;252(2):643–650.
68. Ahmad S, Wang L, Ward PE. Dipeptidyl(amino)peptidase IV and aminopeptidase M metabolize circulating substance P in vivo. *J Pharmacol Exp Ther* 1992;260(3):1257–1261.

69. Delmas B, Gelfi J, Kut E, Sjostrom H, Noren O, Laude H. Determinants essential for the transmissible gastroenteritis virus-receptor interaction reside within a domain of aminopeptidase-N that is distinct from the enzymatic site. *J Virol* 1994;68(8):5216–5224.
70. Delmas B, Gelfi J, L'Haridon R, Vogel LK, Sjostrom H, Noren O, Laude H. Aminopeptidase N is a major receptor for the entero-pathogenic coronavirus TGEV. *Nature* 1992;357(6377):417–420.
71. Delmas B, Gelfi J, Sjostrom H, Noren O, Laude H. Further characterization of aminopeptidase-N as a receptor for coronaviruses. *Adv Exp Med Biol* 1993;342:293–298.
72. Yeager CL, Ashmun RA, Williams RK, Cardellicchio CB, Shapiro LH, Look AT, Holmes KV. Human aminopeptidase N is a receptor for human coronavirus 229E. *Nature* 1992;357(6377):420–422.
73. Xu Y, Wellner D, Scheinberg DA. Substance P and bradykinin are natural inhibitors of CD13/aminopeptidase N. *Biochem Biophys Res Commun* 1995;208(2):664–674.
74. Huang K, Takahara S, Kinouchi T, Takeyama M, Ishida T, Ueyama H, Nishi K, Ohkubo I. Alanyl aminopeptidase from human seminal plasma: Purification, characterization, and immunohistochemical localization in the male genital tract. *J Biochem (Tokyo)* 1997;122(4):779–787.
75. Yamamoto Y, Li YH, Ushiyama I, Nishimura A, Ohkubo I, Nishi K. Puromycin-sensitive alanyl aminopeptidase from human liver cytosol: Purification and characterization. *Forensic Sci Int* 2000;113(1–3):143–146.
76. Repic Lampret B, Kidric J, Kralj B, Vitale L, Pokorny M, Renko M. Lapstatin, a new aminopeptidase inhibitor produced by *Streptomyces rimosus*, inhibits autogenous aminopeptidases. *Arch Microbiol* 1999;171(6):397–404.
77. Miyachi H, Kato M, Kato F, Hashimoto Y. Novel potent nonpeptide aminopeptidase N inhibitors with a cyclic imide skeleton. *J Med Chem* 1998;41(3):263–265.
78. Shimazawa R, Takayama H, Fujimoto Y, Komoda M, Dodo K, Yamasaki R, Shirai R, Koiso Y, Miyata K, Kato F, Kato M, Miyachi H, Hashimoto Y. Novel small molecule nonpeptide aminopeptidase n inhibitors with a cyclic imide skeleton. *J Enzyme Inhib* 1999;14(4):259–275.
79. Shimazawa R, Takayama H, Kato F, Kato M, Hashimoto Y. Nonpeptide small-molecular inhibitors of dipeptidyl peptidase IV: *N*-phenylphthalimide analogs. *Bioorg Med Chem Lett* 1999;9(4):559–562.
80. Takahashi H, Komoda M, Kakuta H, Hashimoto Y. Preparation of novel specific aminopeptidase inhibitors with a cyclic imide skeleton. *Yakugaku Zasshi* 2000;120(10):909–921.
81. Kagechika H, Komoda M, Fujimoto Y, Koiso Y, Takayama H, Kadoya S, Miyata K, Kato F, Kato M, Hashimoto Y. Potent homophthalimide-type inhibitors of B16F10/L5 mouse melanoma cell invasion. *Biol Pharm Bull* 1999;22(9):1010–1012.
82. Komoda M, Kakuta H, Takahashi H, Fujimoto Y, Kadoya S, Kato F, Hashimoto Y. Specific inhibitor of puromycin-sensitive aminopeptidase with a homophthalimide skeleton: Identification of the target molecule and a structure–activity relationship study. *Bioorg Med Chem* 2001;9(1):121–131.
83. Kakuta H, Tanatani A, Nagasawa K, Hashimoto Y. Specific nonpeptide inhibitors of puromycin-sensitive aminopeptidase with a 2,4(1H,3H)-quinazolinedione skeleton. *Chem Pharm Bull (Tokyo)* 2003;51(11):1273–1282.
84. Grembecka J, Mucha A, Cierpicki T, Kafarski P. The most potent organophosphorus inhibitors of leucine aminopeptidase. Structure-based design, chemistry, and activity. *J Med Chem* 2003;46(13):2641–2655.
85. Georgiadis D, Vazeux G, Llorens-Cortes C, Yiotakis A, Dive V. Potent and selective inhibition of zinc aminopeptidase A (EC 3.4.11.7, APA) by glutamyl aminophosphinic peptides: Importance of glutamyl aminophosphinic residue in the P1 position. *Biochemistry* 2000;39(5):1152–1155.
86. Ocain TD, Rich DH. alpha-Keto amide inhibitors of aminopeptidases. *J Med Chem* 1992;35(3):451–456.
87. Shenvi AB. alpha-Aminoboronic acid derivatives: Effective inhibitors of aminopeptidases. *Biochemistry* 1986;25(6):1286–1291.
88. Andersson L, Isley TC, Wolfenden R. alpha-aminoaldehydes: Transition state analogue inhibitors of leucine aminopeptidase. *Biochemistry* 1982;21(17):4177–4180.
89. Parellada J, Suarez G, Guinea M. Inhibition of zinc metallopeptidases by flavonoids and related phenolic compounds: Structure–activity relationships. *J Enzyme Inhib* 1998;13(5):347–359.
90. Bormann H, Melzig MF. Inhibition of metallopeptidases by flavonoids and related compounds. *Pharmazie* 2000;55(2):129–132.
91. Lendeckel U, Arndt M, Wolke C, Reinhold D, Kahne T, Ansorge S. Inhibition of human leukocyte function, alanyl aminopeptidase (APN, CD13) and dipeptidylpeptidase IV (DP IV, CD26) enzymatic activities by aqueous extracts of *Cistus incanus* L. ssp. *incanus*. *J Ethnopharmacol* 2002;79(2):221–227.
92. Kiss A, Kowalski J, Melzig MF. Compounds from *Epilobium angustifolium* inhibit the specific metallopeptidases ACE, NEP, and APN. *Planta Med* 2004;70(10):919–923.

93. Gordon JJ, Kelly BK, Miller GA. Actinonin: An antibiotic substance produced by an actinomycete. *Nature* 1962;195:701–702.
94. Umezawa H, Aoyagi T, Tanaka T, Suda H, Okuyama A, Naganawa H, Hamada M, Takeuchi T. Production of actinonin, an inhibitor of aminopeptidase M, by actinomycetes. *J Antibiot (Tokyo)* 1985;38(11):1629–1630.
95. Gordon JJ, Devlin JP, East AJ, Ollis WD, Wright DE, Ninet L. Studies concerning the antibiotic actinonin. Part I. The constitution of actinonin. A natural hydroxamic acid with antibiotic activity. *J Chem Soc [Perkin 1]* 1975;(9):819–825.
96. Anderson NH, Ollis WD, Thorpe JE, Ward AD. Studies concerning the antibiotic actinonin. Part II. Total synthesis of actinonin and some structural analogues by the isomaleimide method. *J Chem Soc [Perkin 1]* 1975;(9):825–830.
97. Devlin JP, Ollis WD, Thorpe JE, Wood RJ, Broughton BJ, Warren PJ, Wooldridge KRH, Wright DE. Studies concerning the antibiotic actinonin. Part III. Synthesis of structural analogues of actinonin by the anhydride–imide method. *J Chem Soc [Perkin 1]* 1975;(9):830–842.
98. Broughton BJ, Warren PJ, Wooldridge KRH, Wright DE, Ollis WD, Wood RJ. Studies concerning the antibiotic actinonin. Part IV. Synthesis of structural analogues of actinonin by the mixed anhydride method. *J Chem Soc [Perkin 1]* 1975;(9):842–846.
99. Devlin JP, Ollis WD, Thorpe JE. Studies concerning the antibiotic actinonin. Part V. Synthesis of structural analogues of actinonin by the anhydride–ester method. *J Chem Soc [Perkin 1]* 1975;(9):846–848.
100. Devlin JP, Ollis WD, Thorpe JE, Wright DE. Studies concerning the antibiotic actinonin. Part VI. Synthesis of structural analogues of actinonin by dicyclohexylcarbodiimide coupling reactions. *J Chem Soc [Perkin 1]* 1975;(9):848–852.
101. Anderson NH, Devlin JP, Jones S, Ollis WD, Thorpe JE. Studies concerning the antibiotic actinonin. Part VII. Mass spectra of actinonin and related compounds. *J Chem Soc [Perkin 1]* 1975;(9):852–857.
102. Bashiardes G, Bodwell GJ, Gxxxx DS. Asymmetric synthesis of (–)-actinonin and (–)-epi-actinonin. *J Chem Soc [Perkin Trans 1]* 1993;(4):459–469.
103. Broughton BJ, Chaplen P, Freeman WA, Warren PJ, Wooldridge KRH, Wright DE. Studies concerning the antibiotic actinonin. Part VIII. Structure–activity relationships in the actinonin series. *J Chem Soc [Perkin 1]* 1975;(9):857–860.
104. Chung MC, Lee HJ, Chun HK, Lee CH, Kim SI, Kho YH. Bestatin analogue from *Streptomyces neyagawaensis* SL-387. *Biosci Biotechnol Biochem* 1996;60(5):898–900.
105. Chung MC, Chun HK, Han KH, Lee HJ, Lee CH, Kho YH. MR-387A and B, new aminopeptidase N inhibitors, produced by *Streptomyces neyagawaensis* SL-387. *J Antibiot (Tokyo)* 1996;49(1):99–102.
106. Chung MC, Lee CH, Lee HJ, Kho YH, Chun HK. Biosynthesis of peptide inhibitor MR-387 by *Streptomyces neyagawaensis*. *Biotechnol Lett* 1997;19(7):607–610.
107. Ma T, Xu W-F, Wang J-L, Yuan Y-M. Design, synthesis and anti-cancer activity of AHPA derivatives. *Chinese J Med Chem* 2003;13(2):70–75.
108. Rich DH, Moon BJ, Harbeson S. Inhibition of aminopeptidases by amastatin and bestatin derivatives. Effect of inhibitor structure on slow-binding processes. *J Med Chem* 1984;27(4):417–422.
109. Aoyagi T, Tobe H, Kojima F, Hamada M, Takeuchi T, Umezawa H. Amastatin, an inhibitor of aminopeptidase A, produced by actinomycetes. *J Antibiot (Tokyo)* 1978;31(6):636–638.
110. Tobe H, Morishima H, Naganawa H, Takita T, Aoyagi T, Umezawa H. Structure and chemical synthesis of amastatin. *Agric Biol Chem* 1979;43(3):591–596.
111. Rich DH, Moon BJ, Boparai AS. Synthesis of (2S, 3R)-3-amino-2-hydroxy-5-methylhexanoic acid derivatives. Application to the synthesis of amastatin, an inhibitor of aminopeptidase. *J Org Chem* 1980;45(12):2288–2290.
112. Tobe H, Morishima H, Aoyagi T, Umezawa H, Ishiki K, Nakamura K, Yoshioka T, Shimauchi Y, Inui T. Synthesis and structure–activity relationships of amastatin analogues, inhibitors of aminopeptidase A. *Agric Biol Chem* 1982;46(7):1865–1872.
113. Burley SK, David PR, Lipscomb WN. Leucine aminopeptidase: Bestatin inhibition and a model for enzyme-catalyzed peptide hydrolysis. *Proc Natl Acad Sci USA* 1991;88(16):6916–6920.
114. Orning L, Krivi G, Fitzpatrick FA. Leukotriene A4 hydrolase. Inhibition by bestatin and intrinsic aminopeptidase activity establish its functional resemblance to metallohydrolase enzymes. *J Biol Chem* 1991;266(3):1375–1378.
115. Evans JF, Kargman S. Bestatin inhibits covalent coupling of [3H]LTA4 to human leukocyte LTA4 hydrolase. *FEBS Lett* 1992;297(1–2):139–142.
116. Baker JR, Kylstra TA, Bigby TD. Effects of metalloproteinase inhibitors on leukotriene A4 hydrolase in human airway epithelial cells. *Biochem Pharmacol* 1995;50(7):905–912.

117. Andberg M, Wetterholm A, Medina JF, Haeggstrom JZ. Leukotriene A4 hydrolase: A critical role of glutamic acid-296 for the binding of bestatin. *Biochem J* 2000;345(Pt 3):621–625.
118. Scornik OA, Botbol V. Bestatin as an experimental tool in mammals. *Curr Drug Metab* 2001;2(1):67–85.
119. Umezawa H, Aoyagi T, Suda H, Hamada M, Takeuchi T. Bestatin, an inhibitor of aminopeptidase B, produced by actinomycetes. *J Antibiot (Tokyo)* 1976;29(1):97–99.
120. Suda H, Takita T, Aoyagi T, Umezawa H. The structure of bestatin. *J Antibiot (Tokyo)* 1976;29(1):100–101.
121. Suda H, Takita T, Aoyagi T, Umezawa H. The chemical synthesis of bestatin. *J Antibiot (Tokyo)* 1976;29(5):600–601.
122. Kobayashi S, Isobe T, Ohno M. A stereocontrolled synthesis of (–)-bestatin from an acyclic allylamine by iodocyclocarbamation. *Tetrahedron Lett* 1984;25(44):5079–5082.
123. Pearson WH, Hines JV. Synthesis of [beta]-amino-[alpha]-hydroxy acids via aldol condensation of a chiral glycolate enolate. Synthesis of (–)-bestatin. *J Org Chem* 1989;54(17):4235–4237.
124. Norman BH, Morris ML. A stereospecific synthesis of (–)-L-Bestatin from -malic acid. *Tetrahedron Lett* 1992;33(45):6803–6806.
125. Wasserman HH, Xia M, Petersen AK, Jorgensen MR, Curtis EA. Synthesis of the peptidic [alpha]-hydroxy amides phebestin, probestin, and bestatin from [alpha]-keto amide precursors. *Tetrahedron Lett* 1999;40(34):6163–6166.
126. Bergmeier SC, Stanchina DM. Acylnitrene route to vicinal amino alcohols. Application to the synthesis of (–)-bestatin and analogues. *J Org Chem* 1999;64(8):2852–2859.
127. Nemoto H, Ma R, Suzuki I, Shibuya M. A new one-pot method for the synthesis of [alpha]-siloxyamides from aldehydes or ketones and its application to the synthesis of (–)-bestatin. *Org Lett* 2000;2(26):4245–4247.
128. Righi G, D'Achille C, Pescatore G, Bonini C. New stereoselective synthesis of the peptidic aminopeptidase inhibitors bestatin, phebestin and probestin. *Tetrahedron Lett* 2003;44(37):6999–7002.
129. Wasserman HH, Petersen AK, Xia M. Application of acyl cyanophosphorane methodology to the synthesis of protease inhibitors: Poststatin, eurystatin, phebestin, probestin, and bestatin. *Tetrahedron* 2003;59(35):6771–6784.
130. Lee JH, Lee BW, Jang KC, Jeong I-Y, Yang MS, Lee SG, Park KH. Chiroselective synthesis of the (2*S*,3*R*)- and (2*S*,3*S*)-3-amino-2-hydroxy-4-phenylbutanoic acids from sugar: Application to (–)-bestatin. *Synthesis* 2003;(6):829–836.
131. Nishizawa R, Saino T, Takita T, Suda H, Aoyagi T. Synthesis and structure–activity relationships of bestatin analogues, inhibitors of aminopeptidase B. *J Med Chem* 1977;20(4):510–515.
132. Saino T, Seya K, Nishizawa R, Takita T, Aoyagi T, Umezawa H. Synthesis of *p*-hydroxyubanimex. *J Antibiot (Tokyo)* 1987;40(8):1165–1169.
133. Ocain TD, Rich DH. Synthesis of sulfur-containing analogues of bestatin. Inhibition of aminopeptidases by alpha-thiolbestatin analogues. *J Med Chem* 1988;31(11):2193–2199.
134. Gordon EM, Godfrey JD, Delaney NG, Asaad MM, Von Langen D, Cushman DW. Design of novel inhibitors of aminopeptidases. Synthesis of peptide-derived diamino thiols and sulfur replacement analogues of bestatin. *J Med Chem* 1988;31(11):2199–2211.
135. Yuan W, Miunoz B, Wong C-H, Haeggström JZ, Wetterholm A, Samuelsson B. Development of selective tight-binding inhibitors of leukotriene A4 hydrolase. *J Med Chem* 1993;36(2):211–220.
136. Harbeson SL, Rich DH. Inhibition of arginine aminopeptidase by bestatin and arphamenine analogues. Evidence for a new mode of binding to aminopeptidases. *Biochemistry* 1988;27(19):7301–7310.
137. Nagai M, Kojima F, Naganawa H, Hamada M, Aoyagi T, Takeuchi T. Phebestin, a new inhibitor of aminopeptidase N, produced by *Streptomyces* sp. MJ716-m3. *J Antibiot (Tokyo)* 1997;50(1):82–84.
138. Aoyagi T, Yoshida S, Nakamura Y, Shigihara Y, Hamada M, Takeuchi T. Probestin, a new inhibitor of aminopeptidase M, produced by *Streptomyces azureus* MH663-2F6. I. Taxonomy, production, isolation, physico-chemical properties, and biological activities. *J Antibiot (Tokyo)* 1990;43(2):143–148.
139. Yoshida S, Nakamura Y, Naganawa H, Aoyagi T, Takeuchi T. Probestin, a new inhibitor of aminopeptidase M, produced by *Streptomyces azureus* MH663-2F6. II. Structure determination of probestin. *J Antibiot (Tokyo)* 1990;43(2):149–153.
140. Nakamura H, Suda H, Takita T, Aoyagi T, Umezawa H. X-ray structure determination of (2*S*, 3*R*)-3-amino-2-hydroxy-4-phenylbutanoic acid, a new amino acid component of bestatin. *J Antibiot (Tokyo)* 1976;29(1):102–103.
141. Kato K, Saino T, Nishizawa R, Takita T, Umezawa H. Regio- and stereo-specific synthesis of threo-3-amino-2-hydroxy-acids, novelamino-acids contained in aminopeptidase inhibitors of microbial origin. *J Chem Soc Perkin I* 1980;(7):1618–1621.



142. Johnson RL. Renin inhibitors. Substitution of the leucyl residues of Leu-Leu-Val-Phe-OCH<sub>3</sub> with 3-amino-2-hydroxy-5-methylhexanoic acid. *J Med Chem* 1982;25(5):605–610.
143. Reetz MT, Drewes MW, Harms K, Reif W. Stereoselective cyanohydrin-forming reactions of chiral [alpha]-amino aldehydes. *Tetrahedron Lett* 1988;29(27):3295–3298.
144. Herranz R, Castro-Pichel J, Garcia-Lopez T. Tributyltin cyanide, a novel reagent for the stereoselective preparation of 3-amino-2-hydroxy acids via cyanohydrin intermediates. *Synthesis* 1989;(9) 703–706.
145. Angelastro RA, Peet NP, Bey P. An efficient synthesis of novel [alpha]-diketone and [alpha]-keto ester derivatives of N-protected amino acids and peptides. *J Org Chem* 1989;54(16):3913–3916.
146. Herranz R, Castro-Pichel J, Vinuesa S, Garcia-Lopez MT. Stereoselection in the synthesis of threo- and erythro-3-amino-2-hydroxy-4-phenylbutanoic acid using chiral acetal templates. *J Chem Soc Chem Commun* 1989;(14):938–939.
147. Palomo C, Arrieta A, Cossio FP, Aizpurua JM, Mielgo A, Aurrekoetxea N. Highly stereoselective synthesis of [alpha]-hydroxy [beta]-amino acids through [beta]-lactams: Application to the synthesis of the taxol and bestatin side chains and related systems. *Tetrahedron Lett* 1990;31(44):6429–6432.
148. Matsuda F, Matsumoto T, Ohsaki M, Ito Y, Terashima S. An expeditious synthesis of the (2*R*, 3*S*)- and (2*S*, 3*R*)-3-amino-2-hydroxycarboxylic acids. *Chem Lett* 1990; XX:723–724.
149. Kobayashi Y, Takemoto Y, Ito Y, Terashima S. A novel synthesis of the (2*R*,3*S*)- and (2*S*3*R*)-3-amino-2-hydroxycarboxylic acid derivatives, the key components of a renin inhibitor and bestatin, from methyl (*R*)- and (*S*)-mandelate. *Tetrahedron Lett* 1990;31(21):3031–3034.
150. Iizuka K, Kamijo T, Harada H, Akahane K, Kubota T, Umeyama H, Ishida T, Kiso Y. Orally potent human renin inhibitors derived from angiotensinogen transition state: Design, synthesis, and mode of interaction. *J Med Chem* 1990;33(10):2707–2714.
151. Herranz R, Castro-Pichel J, Vinuesa S, Garcia-Lopez MT. An improved one-pot method for the stereoselective synthesis of the (2*S*,3*R*)-3-amino-2-hydroxy acids: Key intermediates for bestatin and amastatin. *J Org Chem* 1990;55(7):2232–2234.
152. Inokuchi T, Tanigawa S, Kanazaki M, Torii S. A concise conversion of glucose to a chiral erythrose derivative and astereospesific synthesis of 3-amino-4-cyclohexyl-2-hydroxybutanoates. *Synlett* 1991;(10):707–708.
153. Ishibuchi A, Nagatani T, Ishizuka T, Kunieda T. Facile synthesis of (2*S*,3*R*)-3-amino-2-hydroxycarboxylic acids, the key components of amastatin and bestatin. *Nat Product Lett* 1992;1(1):21–24.
154. Kobayashi Y, Takemoto Y, Kamijo T, Harada H, Ito Y, Terashima S. A stereoselective synthesis of the (2*R*, 3*S*)- and (2*S*, 3*R*)-3-amino-2-hydroxybutyric acid derivatives, the key components of a renin inhibitor and bestatin. *Tetrahedron* 1992;48(10):1853–1868.
155. Ishizuka T, Ishibuchi S, Kunieda T. Chiral synthons for 2-amino alcohols. Facile preparation of optically active amino hydroxy acids of biological interest. *Tetrahedron* 1993;49(9):1841–1852.
156. Kawabata T, Kiryu Y, Sugiura Y, Fuji K. An enantiodivergent synthesis of threo [beta]-amino alcohols: Preparation of key intermediates for bestatin and the related peptides. *Tetrahedron Lett* 1993;34(32): 5127–5130.
157. Jefford CW, Jian Bo Wang, Zhi-Hui Lu. A concise diastereospesific synthesis of 3-amino-2-hydroxy acids. *Tetrahedron Lett* 1993;34(47):7557–7560.
158. Patel RN, Ramesh N, Banerjee A, Howell JM, McNamee CG, Brozozowski D, Mirfakhrae D, Nanduri V, Thottathil JK, Szarka LJ. Microbial synthesis of (2*R*,3*S*)-(-)-*N*-benzoyl-3-phenyl isoserine ethyl ester-a taxol side-chain synthon. *Tetrahedron Asymmetry* 1993;4(9):2069–2084.
159. Dondoni A, Perrone D. 2-Thiazolyl [alpha]-amino ketones: A new class of reactive intermediates for the stereocontrolled synthesis of unusual amino acids. *Synthesis* 1993;(11):1162–1176.
160. Bunnage ME, Davies SG, Goodwin CJ. Asymmetric synthesis of allophenylnorstatine. *Synlett* 1993;(10):731–732.
161. Kearns J, Kayser MM. Application of yeast-catalyzed reductions to synthesis of (2*R*,3*S*)-phenylisoserine. *Tetrahedron Lett* 1994;35(18):2845–2848.
162. Sasai H, Kim W-S, Suzuki T, Shibusaki M, Mitsuda M, Hasegawa J, Ohashi T. Diastereoselective catalytic asymmetric nitroaldol reaction utilizing rare earth-Li-(*R*)-BINOL complex. A highly efficient synthesis of norstatine. *Tetrahedron Lett* 1994;35(33):6123–6126.
163. Slee DH, Laslo KL, Elder JH, Ollmann IR, Gustchina JK, Zdanov A, Wlodawer A, Wong C-H. Selectivity in the inhibition of HIV and FIV protease: Inhibitory and mechanistic studies of pyrrolidine-containing [alpha]-keto amides and hydroxymethylamine core structures. *J Am Chem Soc* 1995;117(48):11867–11878.

164. Enders D, Reinhold U. Diastereo- and enantioselective synthesis of 1,2-amino alcohols from glycol aldehyde hydrazones; asymmetric synthesis of (*R,R*)-statin. *Angew Chem Int Ed Engl* 1995;34(11):1219–1222.
165. Kang SH, Ryu DH. Versatile synthetic routes to threo-[beta]-amino hydroxy carboxylic acids, statine and its analogues. *Bioorganic Med Chem Lett* 1995;5(24):2959–2962.
166. Dondoni A, Perrone D, Semola T. Synthesis of taxol and taxotere side chains by 2-(trimethylsilyl)thiazole based homologation of L-phenylglycine. *Synthesis* 1995;(2):181–186.
167. Pasto M, Casterjon P, Moyano A, Pericas MA, Riera A. A catalytic asymmetric synthesis of cyclohexylnorstatine. *J Org Chem* 1996;61(17):6033–6037.
168. Li G, Chang H-T, Sharpless KB. Catalytic asymmetric aminohydroxylation (AA) of olefins. *Angew Chem Int Ed Engl* 1996;35(4):451–454.
169. Li G, Angert HH, Sharpless KB. N-halocarbamate salts lead to more efficient catalytic asymmetric aminohydroxylation. *Angew Chem Int Ed Engl* 1996;35(23/24):2813–2817.
170. Jefford CW, McNulty J, Lu Z-H, Wang JB. The enantioselective synthesis of [beta]-aminoacids, their [alpha]-hydroxy derivatives, and the N-terminal components of bestatin and microginin. *Helv Chim Acta* 1996;79(IV):1203–1216.
171. Pasto M, Moyano A, Pericas MA, Riera A. An enantioselective, stereodivergent approach to anti- and syn-[alpha]-hydroxy-[beta]-amino acids from anti-3-amino-1,2-diols. Synthesis of the ready for coupling taxotere(*R*) side chain. *Tetrahedron Asymmetry* 1996;7(1):243–262.
172. Veerasha G, Datta A. Stereoselective synthesis of (–)-*N*-Boc-statine and (–)-*N*-Boc-Norstatine. *Tetrahedron Lett* 1997;38(29):5223–5224.
173. Righi G, Chionne A, D'Achille R, Bonini C. Metal halide-mediated opening of three membered rings: Enantioselective synthesis of (2*S*,3*R*)-3-amino-2-hydroxydecanoic acid and (3*R*)-3-aminodecanoic acid. *Tetrahedron Asymmetry* 1997;8(6):903–907.
174. Sugimura H, Miura M, Yamada N. Enantiospecific and diastereoselective synthesis of syn-[beta]-amino-[alpha]-hydroxy acids. *Tetrahedron Asymmetry* 1997;8(24):4089–4099.
175. Shibata N, Itoh E, Terashima S. Practical synthesis of (2*S*, 3*S*)-3-amino-2-hydroxy-4-phenylbutyric acid, a key component of HIV protease inhibitors. *Chem Pharm Bull (Tokyo)* 1998;46(4):733–735.
176. May BCH, Abell AD. A convenient preparation of (2*S*, 3*S*)-3-amino-2-hydroxy-4-phenylbutanoic acid; an important peptide bond isostere. *Synth Commun* 1999;29(14):2515–2525.
177. Seki M, Matsumoto K. A novel synthesis of allophenylnorstatine from (*R*)-aspartic acid. *Synthesis* 1999;(6):924–926.
178. Ha H-J, Ahn Y-G, Lee GS. Asymmetric synthesis of 3-amino-2-hydroxy-4-phenylbutanoate. *Tetrahedron: Asymmetry* 1999;10(12):2327–2336.
179. Audin P, Pothion C, Fehrentz J-A., Loffet A, Martinez J, Paris J. Diastereoselective synthesis of N-protected [beta]-amino-[alpha]-hydroxyacids (norstatines) from urethane N-carboxyanhydrides (UNCAs). *J Chem Res (S)* 1999(3):282–283.
180. Hinoue K, Furukawa Y, Yaegashi K. Daisow Co., LTD, assignee. Process for producing erythro-3-amino-2-hydroxybutyric acid derivatives. 2000.
181. Benaglia M, Cinquini M, Cozzi F. The *S*-thioester enolate/imine condensation: A shortcut to [beta]-lactams. *Eur J Org Chem* 2000;(4):563–593.
182. Semple JE, Owens TD, Nguyen K, Levy OE. New synthetic technology for efficient construction of [alpha]-hydroxy-[beta]-amino amides via the Passerini reaction. *Org Lett* 2000;2(18):2769–2772.
183. Hayashi Y, Kinoshita Y, Hidaka K, Kiso A, Uchibori H, Kimura T, Kiso Y. Analysis of amide bond formation with an [alpha]-hydroxy-[beta]-amino acid derivative, 3-amino-2-hydroxy-4-phenylbutanoic acid, as an acyl component: Byproduction of homobis lactone. *J Org Chem* 2001;66(16):5537–5544.
184. Phukan P. A short and enantioselective synthesis of N-terminal components of bestatin, amastatin, and microginin. *Indian J Chem* 2002;41B(5):1015–1018.
185. Tasic G, Matovic R, Saicic RN. Stereoselective synthesis of [alpha]-hydroxy-[beta]-amino acids: The chiral pool approach. *J Serb Chem Soc* 2004;69(11):981–990.
186. Aoyagi T, Yoshida S, Matsuda N, Ikeda T, Hamada M, Takeuchi T. Leuhistin, a new inhibitor of aminopeptidase M, produced by *Bacillus laterosporus* BMI156-14F1. I. Taxonomy, production, isolation, physico-chemical properties, and biological activities. *J Antibiot (Tokyo)* 1991;44(6):573–578.
187. Yoshida S, Aoyagi T, Takeuchi T. Biosynthetic study of leuhistin, a new inhibitor of aminopeptidase M. *J Antibiot (Tokyo)* 1991;44(6):683–684.
188. Yoshida S, Naganawa H, Aoyagi T, Takeuchi T, Takeuchi Y, Kodama Y. Leuhistin, a new inhibitor of aminopeptidase M, produced by *Bacillus laterosporus* BMI156-14F1. II. Structure determination of leuhistin. *J Antibiot (Tokyo)* 1991;44(6):579–581.

189. Sedo A, Vlasticova K, Bartak P, Vespalec R, Vicar J, Simanek V, Ulrichova J. Quaternary benzo[c]phenanthridine alkaloids as inhibitors of aminopeptidase N and dipeptidyl peptidase IV. *Phytother Res* 2002;16(1):84–87.
190. Shim JS, Kim JH, Cho HY, Yum YN, Kim SH, Park HJ, Shim BS, Choi SH, Kwon HJ. Irreversible inhibition of CD13/aminopeptidase N by the antiangiogenic agent curcumin. *Chem Biol* 2003;10(8):695–704.
191. Surh Y. Molecular mechanisms of chemopreventive effects of selected dietary and medicinal phenolic substances. *Mutat Res* 1999;428(1–2):305–327.
192. Dorai T, Aggarwal BB. Role of chemopreventive agents in cancer therapy. *Cancer Lett* 2004;215(2):129–140.
193. Aggarwal BB, Kumar A, Bharti AC. Anticancer potential of curcumin: Preclinical and clinical studies. *Anticancer Res* 2003;23(1A):363–398.
194. Surh YJ, Chun KS, Cha HH, Han SS, Keum YS, Park KK, Lee SS. Molecular mechanisms underlying chemopreventive activities of anti-inflammatory phytochemicals: Down-regulation of COX-2 and iNOS through suppression of NF-kappa B activation. *Mutat Res* 2001;480-481:243–268.
195. Chauhan DP. Chemotherapeutic potential of curcumin for colorectal cancer. *Curr Pharm Des* 2002;8(19):1695–1706.
196. Sarkar FH, Li Y. Cell signaling pathways altered by natural chemopreventive agents. *Mutat Res* 2004;555(1-2):53–64.
197. Leu TH, Maa MC. The molecular mechanisms for the antitumorigenic effect of curcumin. *Curr Med Chem Anti-Canc Agents* 2002;2(3):357–370.
198. Sui Z, Salto R, Li J, Craik C, Ortiz de Montellano PR. Inhibition of the HIV-1 and HIV-2 proteases by curcumin and curcumin boron complexes. *Bioorg Med Chem* 1993;1(6):415–422.
199. De Clercq E. Current lead natural products for the chemotherapy of human immunodeficiency virus (HIV) infection. *Med Res Rev* 2000;20(5):323–349.
200. Jagetia GC, Rajanikant GK. Effect of curcumin on radiation-impaired healing of excisional wounds in mice. *J Wound Care* 2004;13(3):107–109.
201. Jagetia GC, Rajanikant GK. Role of curcumin, a naturally occurring phenolic compound of turmeric in accelerating the repair of excision wound, in mice whole-body exposed to various doses of gamma-radiation. *J Surg Res* 2004;120(1):127–138.
202. Pabon HJJ. Synthesis of curcumin and related compounds. *Recl Trav Chim Pays-Bas* 1964;83:379–386.
203. Pedersen U, Rasmussen PB, Lawesson S-O. Synthesis of naturally occurring curcuminoids and related compounds. *Liebigs Ann Chem* 1985;(8):1557–1569.
204. Babu KVD, Rajasekharan KN. Simplified condition for synthesis of curcumin I and other curcuminoids. *Org Prep Proced Int* 1994;26(6):674–677.
205. Mazumder A, Neamati N, Sunder S, Schulz J, Pertz H, Eich E, Pommier Y. Curcumin analogs with altered potencies against HIV-1 integrase as probes for biochemical mechanisms of drug action. *J Med Chem* 1997;40(19):3057–3063.
206. Baar BLMv, Rozendal J, Goot Hvd. Electron ionization mass spectrometry of curcumin analogues: An olefin metathesis reaction in the fragmentation of radical cations. *J Mass Spectrom* 1998;33(4):319–327.
207. Robinson TP, Ehlers T, Hubbard IR, Bai X, Arbiser JL, Goldsmith DJ, Bowen JP. Design, synthesis, and biological evaluation of angiogenesis inhibitors: Aromatic enone and dienone analogues of curcumin. *Bioorg Med Chem Lett* 2003;13(1):115–117.
208. Shim JS, Kim DH, Jung HJ, Kim JH, Lim D, Lee SK, Kim KW, Ahn JW, Yoo JS, Rho JR, Shin J, Kwon HJ. Hydrazinocurcumin, a novel synthetic curcumin derivative, is a potent inhibitor of endothelial cell proliferation. *Bioorg Med Chem* 2002;10(8):2439–2444.
209. Dräger B, Galgon T, Neubert R, Wohlrab W. Method of producing betulonic acid. Jan. 16, 2001; US Patent No 6,175,035 B1
210. Melzig MF, Bormann H. Betulinic acid inhibits aminopeptidase N activity. *Planta Med* 1998;64(7):655–657.
211. Baglin I, Mitaine-Offer AC, Nour M, Tan K, Cave C, Lacaille-Dubois MA. A review of natural and modified betulinic, ursolic and echinocystic acid derivatives as potential antitumor and anti-HIV agents. *Mini Rev Med Chem* 2003;3(6):525–539.
212. Eiznhamer DA, Xu ZQ. Betulinic acid: A promising anticancer candidate. *IDrugs* 2004;7(4):359–373.
213. Cichewicz RH, Kouzi SA. Chemistry, biological activity, and chemotherapeutic potential of betulinic acid for the prevention and treatment of cancer and HIV infection. *Med Res Rev* 2004;24(1):90–114.
214. Kim DSHL, Chen Z, Nguyen VT, Pezzuto JM, Qiu S, Lu Z-Z. A concise semi-synthetic approach to betulinic acid from betulin. *Synth Commun* 1997;27(9):1607–1612.

215. Son LB, Kaplun AP, Shpilevskii AA, Andiya-Pravdiviy YE, Alekseeva SG, Grigor'ev VB, Shvets VI. The synthesis of betulinic acid from betulin and its solubilization with liposomes. *Russ Rev Bioorg Chem* 1998;24(10):700–705.
216. Kim DS, Pezzuto JM, Pisha E. Synthesis of betulinic acid derivatives with activity against human melanoma. *Bioorg Med Chem Lett* 1998;8(13):1707–1712.
217. Baltina LA, Flekhter OB, Nigmatullina LR, Boreko EI, Pavlova NI, Nikolaeva SN, Savinova OV, Tolstikov GA. Lupane triterpenes and derivatives with antiviral activity. *Bioorg Med Chem Lett* 2003;13(20):3549–3552.
218. Tiwari KP, Minocha PK. Pavophylline, a new saponin from the stem of *Pavonia zeylanica*. *Phytochemistry* 1980;19:701–704.
219. Tiwari KP, Srivastava SD, Srivastava SK. [alpha]-l-rhamnopyranosyl-3[beta]-hydroxy-lup-20(29)-en-28-oic acid from the stem of *Dillenia pentagyna*. *Phytochemistry* 1980;19:980–981.
220. Mandloi D, Sant PG. A saponin from *Asparagus gonocladus*. *Phytochemistry* 1981;20(7):1687–1688.
221. Purohit MC, Pant G, Rawat MS. A betulinic acid glycoside from *Schefflera venulosa*. *Phytochemistry* 1991;30(7):2419.
222. Kitajima J, Shindo M, Tanaka Y. Two new triterpenoid sulfates from the leaves of *Schefflera octophylla*. *Chem Pharm Bull* 1990;38(3):714–716.
223. Otsuka H, Fujioka S, Komiya T, Goto M, Hiramatsu Y, Fujimura H. Studies on anti-inflammatory agents. V. A new anti-inflammatory constituent of *Pyraacantha crenulata* roem. *Chem Pharm Bull* 1981;29(11):3099–3104.
224. Arabshahi L, Schmitz FJ. Brominated tyrosine metabolites from an unidentified sponge. *J Org Chem* 1987;52(16):3584–3586.
225. Quinoà E, Crews P. Phenolic constituents of *Psammaphysilla*. *Tetrahedron Lett* 1987;28(28):3229–3232.
226. Rodriguez AD, Akee RK, Schuer PJ. Two bromotyrosine-cysteine derived metabolites from a sponge. *Tetrahedron Lett* 1987;28(42):4989–4992.
227. Jiménez C, Crews P. Novel marine sponge derived amino acids 13. Additional psammaphlin derivatives from *Psammaphysilla purpurea*. *Tetrahedron* 1991;47(12–13):2097–2102.
228. Suzuki A, Matsunaga K, Shin H, Tabudrav J, Shizuri Y, Ohizumi Y. Bisprasin, a novel Ca(2+) releaser with caffeine-like properties from a marine sponge, *Dysidea* spp., acts on Ca(2+)-induced Ca(2+) release channels of skeletal muscle sarcoplasmic reticulum. *J Pharmacol Exp Ther* 2000;292(2):725–730.
229. Shin J, Lee H-S, Seo Y, Rho JR, Cho KW, Paul VJ. New bromotyrosine metabolites from the sponge *Aplysinella rhax*. *Tetrahedron* 2000;56(46):9071–9077.
230. Pham NB, Butler MS, Quinn RJ. Isolation of psammaphlin A 11 $\epsilon$ -sulfate and bisprasin 11 $\epsilon$ -sulfate from the marine sponge *Aplysinella rhax*. *J Nat Prod* 2000;63(3):393–395.
231. Tabudravu JN, Eijssink VG, Gooday GW, Jaspars M, Komander D, Legg M, Synstad B, van Aalten DM. Psammaphlin A, a chitinase inhibitor isolated from the Fijian marine sponge *Aplysinella rhax*. *Bioorg Med Chem* 2002;10(4):1123–1128.
232. Pina IC, Gautschi JT, Wang GY, Sanders ML, Schmitz FJ, France D, Cornell-Kennon S, Sambucetti LC, Remiszewski SW, Perez LB, Bair KW, Crews P. Psammaphlins from the sponge *Pseudoceratina purpurea*: Inhibition of both histone deacetylase and DNA methyltransferase. *J Org Chem* 2003;68(10):3866–3873.
233. Jung JH, Sim CJ, Lee CO. Cytotoxic compounds from a two-sponge association. *J Nat Prod* 1995;58(11):1722–1726.
234. Park Y, Liu Y, Hong J, Lee CO, Cho H, Kim DK, Im KS, Jung JH. New bromotyrosine derivatives from an association of two sponges, *Jaspis wondoensis* and *Poecillastra wondoensis*. *J Nat Prod* 2003;66(11):1495–1498.
235. Nicolaou KC, Hughes R, Pfefferkorn JA, Barluenga S, Roecker AJ. Combinatorial synthesis through disulfide exchange: Discovery of potent psammaphlin A type antibacterial agents active against methicillin-resistant *Staphylococcus aureus* (MRSA). *Chemistry* 2001;7(19):4280–4295.
236. Nicolaou KC, Hughes R, Pfefferkorn JA, Barluenga S. Optimization and mechanistic studies of psammaphlin A type antibacterial agents active against methicillin-resistant *Staphylococcus aureus* (MRSA). *Chemistry* 2001;7(19):4296–4310.
237. Shim JS, Lee HS, Shin J, Kwon HJ. Psammaphlin A, a marine natural product, inhibits aminopeptidase N and suppresses angiogenesis in vitro. *Cancer Lett* 2004;203(2):163–169.
238. Kim D, Lee IS, Jung JH, Yang SI. Psammaphlin A, a natural bromotyrosine derivative from a sponge, possesses the antibacterial activity against methicillin-resistant *Staphylococcus aureus* and the DNA gyrase-inhibitory activity. *Arch Pharm Res* 1999;22(1):25–29.

239. Kim D, Lee IS, Jung JH, Lee CO, Choi SU. Psammaplin A, a natural phenolic compound, has inhibitory effect on human topoisomerase II and is cytotoxic to cancer cells. *Anticancer Res* 1999;19(5B):4085–4090.
240. Nicholas GM, Eckman LL, Ray S, Hughes RO, Pfeifferkorn JA, Barluenga S, Nicolaou KC, Bewley CA. Bromotyrosine-derived natural and synthetic products as inhibitors of mycothiol-S-conjugate amidase. *Bioorg Med Chem Lett* 2002;12(17):2487–2490.
241. Jiang Y, Ahn EY, Ryu SH, Kim DK, Park JS, Yoon HJ, You S, Lee BJ, Lee DS, Jung JH. Cytotoxicity of psammaplin A from a two-sponge association may correlate with the inhibition of DNA replication. *BMC Cancer* 2004;4(1):70.
242. Fittkau S, Jahreis G, Peters K. [alpha]-aminoketone-Ein beitrag zur synthese optisch aktiver derivate von aminosäuren und peptiden. *J Prakt Chem* 1986;328(4):529–538.
243. Jahreis G, Fittkau S, Aurich H. [alpha]-Aminomethylketones as inhibitors of a membrane-bound alanine aminopeptidase. *Biomed Biochim Acta* 1987;46(10):683–686.
244. Bergin JD, Clapp CH. Inhibition of aminopeptidase M by alkyl D-cysteines. *J Enzyme Inhib* 1989;3(2):127–131.
245. Schalk C, d'Orchymont H, Jauch MF, Tarnus C. 3-Amino-2-tetralone derivatives: Novel potent and selective inhibitors of aminopeptidase-M (EC 3.4.11.2). *Arch Biochem Biophys* 1994;311(1):42–46.
246. Tarnus C, Remy JM, d'Orchymont H. 3-Amino-2-hydroxy-propionaldehyde and 3-amino-1-hydroxypropan-2-one derivatives: New classes of aminopeptidase inhibitors. *Bioorg Med Chem* 1996;4(8):1287–1297.
247. Lindsay CK, Gomez DE, Thorgeirsson UP. Effect of flavone acetic acid on endothelial cell proliferation: Evidence for antiangiogenic properties. *Anticancer Res* 1996;16(1):425–431.
248. Bauvois B, Puiffe ML, Bongui JB, Paillat S, Monneret C, Dauzonne D. Synthesis and biological evaluation of novel flavone-8-acetic acid derivatives as reversible inhibitors of aminopeptidase N/CD13. *J Med Chem* 2003;46(18):3900–3913.
249. Lee J, Shim JS, Jung SA, Lee ST, Kwon HJ. *N*-hydroxy-2-(naphthalene-2-ylsulfanyl)-acetamide, a novel hydroxamic acid-based inhibitor of aminopeptidase N and its anti-angiogenic activity. *Bioorg Med Chem Lett* 2005;15(1):181–183.
250. Holden HM, Matthews BW. The binding of L-valyl-L-tryptophan to crystalline thermolysin illustrates the mode of interaction of a product of peptide hydrolysis. *J Biol Chem* 1988;263(7):3256–3260.
251. Fournie-Zaluski MC, Coric P, Turcaud S, Bruetsch L, Lucas E, Noble F, Roques BP. Potent and systemically active aminopeptidase N inhibitors designed from active-site investigation. *J Med Chem* 1992;35(7):1259–1266.
252. Chen H, Noble F, Mothe A, Meudal H, Coric P, Danascimento S, Roques BP, George P, Fournie-Zaluski MC. Phosphinic derivatives as new dual enkephalin-degrading enzyme inhibitors: Synthesis, biological properties, and antinociceptive activities. *J Med Chem* 2000;43(7):1398–1408.
253. Chen H, Roques BP, Fournie-Zaluski MC. Design of the first highly potent and selective aminopeptidase N (EC 3.4.11.2) inhibitor. *Bioorg Med Chem Lett* 1999;9(11):1511–1516.
254. Chen H, Bischoff L, Fournie-Zaluski MC, Roques BP. Synthesis of 2(S)-benzyl-3-[hydroxy(1*ϕ*(*R*)-aminoethyl)phosphinyl]propanoyl-L-3-[<sup>125</sup>I]-iodotyrosine: A radiolabelled inhibitor of aminopeptidase N. *J Labelled Compd Radiopharm* 2000;43:103–111.
255. Noble F, Luciani N, Da Nascimento S, Lai-Kuen R, Bischoff L, Chen H, Fournie-Zaluski MC, Roques BP. Binding properties of a highly potent and selective iodinated aminopeptidase N inhibitor appropriate for radioautography. *FEBS Lett* 2000;467(1):81–86.
256. Noble F, Banisadr G, Jardinaud F, Popovici T, Lai-Kuen R, Chen H, Bischoff L, Parsadaniantz SM, Fournie-Zaluski MC, Roques BP. First discrete autoradiographic distribution of aminopeptidase N in various structures of rat brain and spinal cord using the selective iodinated inhibitor [<sup>125</sup>I]RB 129. *Neuroscience* 2001;105(2):479–488.
257. Gros C, Giros B, Schwartz JC, Vlaiculescu A, Costentin J, Lecomte JM. Potent inhibition of cerebral aminopeptidases by carbaphethiol, a parenterally active compound. *Neuropeptides* 1988;12(3):111–118.
258. Chauvel EN, Coric P, Llorens-Cortes C, Wilk S, Roques BP, Fournie-Zaluski MC. Investigation of the active site of aminopeptidase A using a series of new thiol-containing inhibitors. *J Med Chem* 1994;37(9):1339–1346.
259. Chauvel EN, Llorens-Cortes C, Coric P, Wilk S, Roques BP, Fournie-Zaluski MC. Differential inhibition of aminopeptidase A and aminopeptidase N by new beta-amino thiols. *J Med Chem* 1994;37(18):2950–2957.
260. Reaux A, de Mota N, Zini S, Cadel S, Fournie-Zaluski MC, Roques BP, Corvol P, Llorens-Cortes C. PC18, a specific aminopeptidase N inhibitor, induces vasopressin release by increasing the half-life of brain angiotensin III. *Neuroendocrinology* 1999;69(5):370–376.

261. Reaux A, Iturrioz X, Vazeux G, Fournie-Zaluski MC, David C, Roques BP, Corvol P, Llorens-Cortes C. Aminopeptidase A, which generates one of the main effector peptides of the brain rennin-angiotensin system, angiotensin III, has a key role in central control of arterial blood pressure. *Biochem Soc Trans* 2000;28(4):435-440.
262. Fournie-Zaluski MC, Chaillet P, Bouboutou R, Coulaud A, Cherot P, Waksman G, Costentin J, Roques BP. Analgesic effects of kelatorphan, a new highly potent inhibitor of multiple enkephalin degrading enzymes. *Eur J Pharmacol* 1984;102(3-4):525-528.
263. Fournie-Zaluski MC, Coric P, Turcaud S, Lucas E, Noble F, Maldonado R, Roques BP. Mixed inhibitor-prodrug as a new approach toward systemically active inhibitors of enkephalin-degrading enzymes. *J Med Chem* 1992;35(13):2473-2481.
264. Roques BP, Noble F. Dual inhibitors of enkephalin-degrading enzymes (neutral endopeptidase 24.11 and aminopeptidase N) as potential new medications in the management of pain and opioid addiction. *NIDA Res Monogr* 1995;147:104-145.
265. Roques BP, Noble F, Crine P, Fournie-Zaluski MC. Inhibitors of neprilysin: Design, pharmacological and clinical applications. *Methods Enzymol* 1995;248:263-283.
266. Schmidt C, Peyroux J, Noble F, Fournie-Zaluski MC, Roques BP. Analgesic responses elicited by endogenous enkephalins (protected by mixed peptidase inhibitors) in a variety of morphine-sensitive noxious tests. *Eur J Pharmacol* 1991;192(2):253-262.
267. Noble F, Soleilhac JM, Soroca-Lucas E, Turcaud S, Fournie-Zaluski MC, Roques BP. Inhibition of the enkephalin-metabolizing enzymes by the first systemically active mixed inhibitor prodrug RB 101 induces potent analgesic responses in mice and rats. *J Pharmacol Exp Ther* 1992;261(1):181-190.
268. Penning TD, Askonas LJ, Djuric SW, Haak RA, Yu SS, Michener ML, Krivi GG, Pyla EY. Kelatorphan and related analogs: Potent and selective inhibitors of leukotriene A4 hydrolase. *Bioorg Med Chem Lett* 1995;5(21):2517-2522.
269. Turner AJ, Murphy LJ. Molecular pharmacology of endothelin converting enzymes. *Biochem Pharmacol* 1996;51(2):91-102.
270. Roques BP. Zinc metallopeptidases: Active site structure and design of selective and mixed inhibitors: new approaches in the search for analgesics and anti-hypertensives. *Biochem Soc Trans* 1993;21(Pt 3):678-685.
271. Roques BP. Peptidomimetics as receptors agonists or peptidase inhibitors: A structural approach in the field of enkephalins, ANP, and CCK. *Biopolymers* 1992;32(4):407-410.
272. Chen H, Noble F, Coric P, Fournie-Zaluski MC, Roques BP. Aminophosphinic inhibitors as transition state analogues of enkephalin-degrading enzymes: A class of central analgesics. *Proc Natl Acad Sci USA* 1998;95(20):12028-12033.
273. Sakurada K, Imamura M, Kobayashi M, Tachibana N, Abe K, Tanaka M, Okabe M, Morioka M, Kasai M, Sugiura T, Miyazaki T. Inhibitory effect of bestatin on the growth of human leukemic cells. *Acta Oncol* 1990;29(6):799-802.
274. Sekine K, Fujii H, Abe F. Induction of apoptosis by bestatin (ubenimex) in human leukemic cell lines. *Leukemia* 1999;13(5):729-734.
275. Rosenzweig M, Tailleux L, Gluckman JC. CD13/N-aminopeptidase is involved in the development of dendritic cells and macrophages from cord blood CD34(+) cells. *Blood* 2000;95(2):453-460.
276. Ino K, Isobe K, Goto S, Nakashima I, Tomoda Y. Inhibitory effect of bestatin on the growth of human lymphocytes. *Immunopharmacology* 1992;23(3):163-171.
277. Morikawa K, Morikawa S, Nakano A, Oseko F. Bestatin, an inhibitor of aminopeptidase B, suppresses the proliferation and differentiation of human B-cells in vitro. *Int J Immunopharmacol* 1989;11(8):905-913.
278. Lendeckel U, Scholz B, Arndt M, Frank K, Spiess A, Chen H, Roques BP, Ansorge S. Inhibition of alanyl-aminopeptidase suppresses the activation-dependent induction of glycogen synthase kinase-3beta (GSK-3beta) in human T cells. *Biochem Biophys Res Commun* 2000;273(1):62-65.
279. Lendeckel U, Kahne T, Arndt M, Frank K, Ansorge S. Inhibition of alanyl aminopeptidase induces MAP-kinase p42/ERK2 in the human T cell line KARPAS-299. *Biochem Biophys Res Commun* 1998;252(1):5-9.
280. Murata M, Kubota Y, Tanaka T, Iida-Tanaka K, Takahara J, Irino S. Effect of ubenimex on the proliferation and differentiation of U937 human histiocytic lymphoma cells. *Leukemia* 1994;8(12):2188-2193.
281. Lohn M, Mueller C, Langner J. Cell cycle retardation in monocytoid cells induced by aminopeptidase N (CD13). *Leuk Lymphoma* 2002;43(2):407-413.
282. Sawafuji K, Miyakawa Y, Weisberg E, Griffin JD, Ikeda Y, Kizaki M. Aminopeptidase inhibitors inhibit proliferation and induce apoptosis of K562 and STI571-resistant K562 cell lines through the MAPK and GSK-3beta pathways. *Leuk Lymphoma* 2003;44(11):1987-1996.

283. Gabrilovac J, Cupic B, Breljak D, Zekusic M, Boranic M. Expression of CD13/aminopeptidase N and CD10/neutral endopeptidase on cultured human keratinocytes. *Immunol Lett* 2004;91(1):39–47.
284. Thielitz A, Bukowska A, Wolke C, Vetter R, Lendeckel U, Wrenger S, Hashimoto Y, Ansorge S, Gollnick H, Reinhold D. Identification of extra- and intracellular alanyl aminopeptidases as new targets to modulate keratinocyte growth and differentiation. *Biochem Biophys Res Commun* 2004;321(4):795–801.
285. Ino K, Goto S, Okamoto T, Nomura S, Nawa A, Isobe K, Mizutani S, Tomoda Y. Expression of aminopeptidase N on human choriocarcinoma cells and cell growth suppression by the inhibition of aminopeptidase N activity. *Jpn J Cancer Res* 1994;85(9):927–933.
286. Ino K, Goto S, Kosaki A, Nomura S, Asada E, Misawa T, Furuhashi Y, Mizutani S, Tomoda Y. Growth inhibitory effect of bestatin on choriocarcinoma cell lines in vitro. *Biotherapy* 1991;3(4):351–357.
287. Xu Y, Lai LT, Gabrilove JL, Scheinberg DA. Antitumor activity of actinonin in vitro and in vivo. *Clin Cancer Res* 1998;4(1):171–176.
288. Santos AN, Langner J, Herrmann M, Riemann D. Aminopeptidase N/CD13 is directly linked to signal transduction pathways in monocytes. *Cell Immunol* 2000;201(1):22–32.
289. Lohn M, Mueller C, Thiele K, Kahne T, Riemann D, Langner J. Aminopeptidase N-mediated signal transduction and inhibition of proliferation of human myeloid cells. *Adv Exp Med Biol* 1997;421:85–91.
290. Lendeckel U, Arndt M, Frank K, Spiess A, Reinhold D, Ansorge S. Modulation of WNT-5A expression by actinonin: Linkage of APN to the WNT-pathway? *Adv Exp Med Biol* 2000;477:35–41.
291. Sekine K, Fujii H, Abe F, Nishikawa K. Augmentation of death ligand-induced apoptosis by aminopeptidase inhibitors in human solid tumor cell lines. *Int J Cancer* 2001;94(4):485–491.
292. Shibuya K, Chiba S, Hino M, Kitamura T, Miyagawa K, Takaku F, Miyazano K. Enhancing effect of ubenimex (bestatin) on proliferation and differentiation of hematopoietic progenitor cells, and the suppressive effect on proliferation of leukemic cell lines via peptidase regulation. *Biomed Pharmacother* 1991;45(2-3):71–80.
293. Blazsek I, Comisso M, Misset JL. Modulation of bone marrow cell functions in vitro by bestatin (ubenimex). *Biomed Pharmacother* 1991;45(2-3):81–86.
294. Hirano T, Kizaki M, Kato K, Abe F, Masuda N, Umezawa K. Enhancement of sensitivity by bestatin of acute promyelocytic leukemia NB4 cells to all-*trans* retinoic acid. *Leuk Res* 2002;26(12):1097–1103.
295. Mishima Y, Matsumoto-Mishima Y, Terui Y, Katsuyama M, Yamada M, Mori M, Ishizaka Y, Ikeda K, Watanabe J, Mizunuma N, Hayasawa H, Hatake K. Leukemic cell-surface CD13/aminopeptidase N and resistance to apoptosis mediated by endothelial cells. *J Natl Cancer Inst* 2002;94(13):1020–1028.
296. Shibuya K, Hayashi E, Abe F, Takahashi K, Horinishi H, Ishizuka M, Takeuchi T, Umezawa H. Enhancement of interleukin 1 and interleukin 2 releases by ubenimex. *J Antibiot (Tokyo)* 1987;40(3):363–369.
297. Lendeckel U, Arndt M, Bukowska A, Tadge J, Wolke C, Kahne T, Neubert K, Faust J, Ittenson A, Ansorge S, Reinhold D. Synergistic action of DPIV and APN in the regulation of T cell function. *Adv Exp Med Biol* 2003;524:123–131.
298. Lin M, He J, Cai Z, Qian W. Aminopeptidase inhibitor Bestatin induces HL-60 cell apoptosis through activating caspase 3. *Zhonghua Xue Ye Xue Za Zhi* 2001;22(7):348–350.
299. Ezawa K, Minato K, Dobashi K. Induction of apoptosis by ubenimex (Bestatin) in human non-small-cell lung cancer cell lines. *Biomed Pharmacother* 1996;50(6-7):283–289.
300. Ehrhardt H, Fulda S, Fuhrer M, Debatin KM, Jeremias I. Betulinic acid-induced apoptosis in leukemia cells. *Leukemia* 2004;18(8):1406–1412.
301. Matsuda N, Katsuragi Y, Saiga Y, Tanaka T, Nakamura M. Effects of aminopeptidase inhibitors actinonin and amastatin on chemotactic and phagocytic responses of human neutrophils. *Biochem Int* 1988;16(2):383–390.
302. Braun RK, Foerster M, Workalemahu G, Haefner D, Kroegel C, Walker C. Differential regulation of aminopeptidase N (CD13) by transendothelial migration and cytokines on human eosinophils. *Exp Lung Res* 2003;29(2):59–77.
303. Saiki I, Fujii H, Yoneda J, Abe F, Nakajima M, Tsuruo T, Azuma I. Role of aminopeptidase N (CD13) in tumor-cell invasion and extracellular matrix degradation. *Int J Cancer* 1993;54(1):137–143.
304. Yoneda J, Saiki I, Fujii H, Abe F, Kojima Y, Azuma I. Inhibition of tumor invasion and extracellular matrix degradation by ubenimex (bestatin). *Clin Exp Metastasis* 1992;10(1):49–59.
305. Ishii K, Usui S, Sugimura Y, Yoshida S, Hioki T, Tatematsu M, Yamamoto H, Hirano K. Aminopeptidase N regulated by zinc in human prostate participates in tumor cell invasion. *Int J Cancer* 2001;92(1):49–54.
306. Fujii H, Nakajima M, Aoyagi T, Tsuruo T. Inhibition of tumor cell invasion and matrix degradation by aminopeptidase inhibitors. *Biol Pharm Bull* 1996;19(1):6–10.

307. Kido A, Krueger S, Haeckel C, Roessner A. Possible contribution of aminopeptidase N (APN/CD13) to invasive potential enhanced by interleukin-6 and soluble interleukin-6 receptor in human osteosarcoma cell lines. *Clin Exp Metastasis* 1999;17(10):857–863.
308. Huang L, Tani K, Ogushi F, Ogawa H, Shimizu T, Motoki Y, Moriguchi H, Sone S. Role of CD13/aminopeptidase N in rat lymphocytic alveolitis caused by thoracic irradiation. *Radiat Res* 2002;157(2):191–198.
309. Pasqualini R, Koivunen E, Kain R, Lahdenranta J, Sakamoto M, Stryhn A, Ashmun RA, Shapiro LH, Arap W, Ruoslahti E. Aminopeptidase N is a receptor for tumor-homing peptides and a target for inhibiting angiogenesis. *Cancer Res* 2000;60(3):722–727.
310. Bhagwat SV, Lahdenranta J, Giordano R, Arap W, Pasqualini R, Shapiro LH. CD13/APN is activated by angiogenic signals and is essential for capillary tube formation. *Blood* 2001;97(3):652–659.
311. Aozuka Y, Koizumi K, Saitoh Y, Ueda Y, Sakurai H, Saiki I. Anti-tumor angiogenesis effect of aminopeptidase inhibitor bestatin against B16–BL6 melanoma cells orthotopically implanted into syngeneic mice. *Cancer Lett* 2004;216(1):35–42.
312. Kwon HJ, Shim JS, Kim JH, Cho HY, Yum YN, Kim SH, Yu J. Betulinic acid inhibits growth factor-induced in vitro angiogenesis via the modulation of mitochondrial function in endothelial cells. *Jpn J Cancer Res* 2002;93(4):417–425.
313. van Hensbergen Y, Broxterman HJ, Peters E, Rana S, Elderkamp YW, van Hinsbergh VW, Koolwijk P. Aminopeptidase inhibitor bestatin stimulates microvascular endothelial cell invasion in a fibrin matrix. *Thromb Haemost* 2003;90(5):921–929.
314. Ribatti D, Vacca A, Roncali L, Dammacco F. The chick embryo chorioallantoic membrane as a model for in vivo research on anti-angiogenesis. *Curr Pharm Biotechnol* 2000;1(1):73–82.
315. Murata Y, Ohno Y, Itakura A, Takeuchi M, Nakashima Y, Kuno N, Mizutani S. Bestatin results in pathophysiological changes similar to preeclampsia in rats via induction of placental apoptosis. *Horm Metab Res* 2003;35(6):343–348.
316. Furuhashi M, Mizutani S, Kurauchi O, Kasugai M, Tomoda Y. Effects of bestatin on intrauterine growth of rat fetuses. *Horm Metab Res* 1989;21(7):366–368.
317. Ishizuka M, Masuda T, Mizutani S, Takeuchi T, Umezawa H. Antitumor cells found in tumor-bearing mice given ubenimex. *J Antibiot (Tokyo)* 1987;40(5):697–701.
318. Abe F, Shibuya K, Uchida M, Takahashi K, Horinishi H, Matsuda A, Ishizuka M, Takeuchi T, Umezawa H. Effect of bestatin on syngeneic tumors in mice. *Gann* 1984;75(1):89–94.
319. Abe F, Yamashita T, Takahashi K, Matsuda A, Ichikawa T, Umezawa H. Antitumor effect of bestatin combined with bleomycin against hepatoma AH 66 subcutaneously transplanted in rats. *Jpn J Antibiot* 1984;37(4):589–592.
320. Talmadge JE. Preclinical approaches to the development of effective immunotherapeutic protocols for the treatment of metastasis. *Prog Clin Biol Res* 1986;212:197–215.
321. Tsuruo T, Naganuma K, Iida H, Yamori T, Tsukagoshi S, Sakurai Y. Inhibition of lymph node metastasis of P388 leukemia by bestatin in mice. *J Antibiot (Tokyo)* 1981;34(9):1206–1209.
322. Pisha E, Chai H, Lee IS, Chagwedera TE, Farnsworth NR, Cordell GA, Beecher CW, Fong HH, Kinghorn AD, Brown DM, et al. Discovery of betulinic acid as a selective inhibitor of human melanoma that functions by induction of apoptosis. *Nat Med* 1995;1(10):1046–1051.
323. Zuco V, Supino R, Righetti SC, Cleris L, Marchesi E, Gambacorti-Passerini C, Formelli F. Selective cytotoxicity of betulinic acid on tumor cell lines, but not on normal cells. *Cancer Lett* 2002;175(1):17–25.
324. Yasukawa K, Takido M, Matsumoto T, Takeuchi M, Nakagawa S. Sterol and triterpene derivatives from plants inhibit the effects of a tumor promoter, and sitosterol and betulinic acid inhibit tumor formation in mouse skin two-stage carcinogenesis. *Oncology* 1991;48(1):72–76.
325. Yasukawa K, Yu SY, Kakinuma S, Takido M. Inhibitory effect of rikkunshi-to, a traditional Chinese herbal prescription, on tumor promotion in two-stage carcinogenesis in mouse skin. *Biol Pharm Bull* 1995;18(5):730–733.
326. Abe F, Shibuya K, Ashizawa J, Takahashi K, Horinishi H, Matsuda A, Ishizuka M, Takeuchi T, Umezawa H. Enhancement of antitumor effect of cytotoxic agents by bestatin. *J Antibiot (Tokyo)* 1985;38(3):411–414.
327. Ebihara K, Abe F, Yamashita T, Shibuya K, Hayashi E, Takahashi K, Horinishi H, Enomoto M, Ishizuka M, Umezawa H. The effect of ubenimex on *N*-methyl-*N*-(4-nitro-*N*-nitrosoguanidine)-induced stomach tumor in rats. *J Antibiot (Tokyo)* 1986;39(7):966–970.
328. Yamamoto Y, Ono H, Ueda A, Shimamura M, Nishimura K, Hazato T. Spinorphin as an endogenous inhibitor of enkephalin-degrading enzymes: Roles in pain and inflammation. *Curr Protein Pept Sci* 2002;3(6):587–599.



329. Wright JW, Amir HZ, Murray CE, Roberts KA, Harding JW, Mizutani S, Ward PE. Use of aminopeptidase M as a hypotensive agent in spontaneously hypertensive rats. *Brain Res Bull* 1991;27(5):545–551.
330. Schorlemmer HU, Bosslet K, Dickneite G, Luben G, Sedlacek HH. Studies on the mechanisms of action of the immunomodulator bestatin in various screening test systems. *Behring Inst Mitt* 1984;(74):157–173.
331. Aoyagi K, Itoh N, Abe F, Abe S, Uchida K, Ishizuka M, Takeuchi T, Yamaguchi H. Enhancement by ubenimex (bestatin) of host resistance to *Candida albicans* infection. *J Antibiot (Tokyo)* 1992;45(11):1778–1784.
332. Knoblich A, Muller WE, Harle-Grupp V, Falke D. Enhancement of antibody formation against herpes simplex virus in mice by the T-cell mitogen bestatin. *J Gen Virol* 1984;65(Pt 10):1675–1686.
333. Dickneite G, Kaspareit F, Sedlacek HH. Stimulation of cell-mediated immunity by bestatin correlates with reduction of bacterial persistence in experimental chronic *Salmonella typhimurium* infection. *Infect Immunol* 1984;44(1):168–174.
334. Tanaka N, Kumamoto Y, Hirose T, Yokoo A. Study of the prophylactic effect of ubenimex on experimental pyelonephritis induced by *Pseudomonas* in neutropenic mice. *Kansenshogaku Zasshi* 1989;63(7):748–756.
335. Harada Y, Kajiki A, Higuchi K, Ishibashi T, Takamoto M. The mode of immunopotentiating action of bestatin: enhanced resistance to *Listeria monocytogenes* infection. *J Antibiot (Tokyo)* 1983;36(10):1411–1414.
336. Hachisu M, Hiranuma T, Murata S, Aoyagi T, Umezawa H. Analgesic effect of actinonin, a new potent inhibitor of multiple enkephalin degrading enzymes. *Life Sci* 1987;41(2):235–240.
337. Noble F, Turcaud S, Fournie-Zaluski MC, Roques BP. Repeated systemic administration of the mixed inhibitor of enkephalin-degrading enzymes, RB101, does not induce either antinociceptive tolerance or cross-tolerance with morphine. *Eur J Pharmacol* 1992;223(1):83–89.
338. Noble F, Smadja C, Valverde O, Maldonado R, Coric P, Turcaud S, Fournie-Zaluski MC, Roques BP. Pain-suppressive effects on various nociceptive stimuli (thermal, chemical, electrical and inflammatory) of the first orally active enkephalin-metabolizing enzyme inhibitor RB 120. *Pain* 1997;73(3):383–391.
339. Chen H, Noble F, Roques BP, Fournie-Zaluski MC. Long lasting antinociceptive properties of enkephalin degrading enzyme (NEP and APN) inhibitor prodrugs. *J Med Chem* 2001;44(21):3523–3530.
340. Benoist JM, Keime F, Montagne J, Noble F, Fournie-Zaluski MC, Roques BP, Willer JC, Le Bars D. Depressant effect on a C-fibre reflex in the rat, of RB101, a dual inhibitor of enkephalin-degrading enzymes. *Eur J Pharmacol* 2002;445(3):201–210.
341. Nieto MM, Wilson J, Walker J, Benavides J, Fournie-Zaluski MC, Roques BP, Noble F. Facilitation of enkephalins catabolism inhibitor-induced antinociception by drugs classically used in pain management. *Neuropharmacology* 2001;41(4):496–506.
342. Ota K, Kurita S, Yamada K, Masaoka T, Uzuka Y, Ogawa N. Immunotherapy with bestatin for acute nonlymphocytic leukemia in adults. *Cancer Immunol Immunother* 1986;23(1):5–10.
343. Ota K, Kurita S, Yamada K, Masaoka T, Uzuka Y, Ogawa N. Results of follow-up studies on prognosis after immunotherapy with bestatin in acute nonlymphocytic leukemia. *Gan To Kagaku Ryoho* 1986;13(4 Pt 1):1017–1025.
344. Ino K, Bierman PJ, Varney ML, Heimann DG, Kuszynski CA, Walker SA, Talmadge JE. Monocyte activation by an oral immunomodulator (bestatin) in lymphoma patients following autologous bone marrow transplantation. *Cancer Immunol Immunother* 1996;43(4):206–212.
345. Arimori S, Nagao T, Shimizu Y, Watanabe K, Komatsuda M. The effect of bestatin on patients with acute and chronic leukemia and malignant lymphoma. *Tokai J Exp Clin Med* 1980;5(1):63–71.
346. Usuka Y, Saito Y. Bestatin treatment of myelodysplastic syndromes and chronic myelogenous leukemia. *Biomed Pharmacother* 1991;45(2-3):87–93.
347. Kurita S, Ota K, Yamada K, Masaoka T, Uzuka Y, Ogawa N. [Immunotherapy with bestatin for acute non-lymphocytic leukemia (ANLL) in adults]. *Gan To Kagaku Ryoho* 1984;11(12 Pt 2):2742–2750.
348. Hiraoka A, Shibata H, Masaoka T. Immunopotentiation with Ubenimex for prevention of leukemia relapse after allogeneic BMT. The Study Group of Ubenimex for BMT. *Transplant Proc* 1992;24(6):3047–3048.
349. Bierman PJ, Abe F, Buyukberber S, Ino K, Talmadge JE. Partial review of immunotherapeutic pharmacology in stem cell transplantation. *In Vivo* 2000;14(1):221–236.
350. Ichinose Y, Genka K, Koike T, Kato H, Watanabe Y, Mori T, Iioka S, Sakuma A, Ohta M. Randomized double-blind placebo-controlled trial of bestatin in patients with resected stage I squamous-cell lung carcinoma. *J Natl Cancer Inst* 2003;95(8):605–610.

351. Yasumitsu T, Ohshima S, Nakano N, Kotake Y, Tominaga S. Bestatin in resected lung cancer. A randomized clinical trial. *Acta Oncol* 1990;29(6):827–831.
352. Wex T, Lendeckel U, Reinhold D, Kahne T, Arndt M, Frank K, Ansorge S. Antisense-mediated inhibition of aminopeptidase N (CD13) markedly decreases growth rates of hematopoietic tumour cells. *Adv Exp Med Biol* 1997;421:67–73.
353. Suda H, Aoyagi T, Takeuchi T, Umezawa H. Inhibition of aminopeptidase B and leucine aminopeptidase by bestatin and its stereoisomer. *Arch Biochem Biophys* 1976;177(1):196–200.
354. Kuramochi H, Motegi A, Iwabuchi M, Takahashi K, Horinishi H, Umezawa H. Action of ubenimex on aminopeptidase activities in spleen cells and peritoneal macrophages from mice. *J Antibiot (Tokyo)* 1987;40(11):1605–1611.
355. Tiekou S, Hooper NM. Inhibition of aminopeptidases N, A and W. A re-evaluation of the actions of bestatin and inhibitors of angiotensin converting enzyme. *Biochem Pharmacol* 1992;44(9):1725–1730.
356. Umezawa H, Ishizuka M, Aoyagi T, Takeuchi T. Enhancement of delayed-type hypersensitivity by bestatin, an inhibitor of aminopeptidase B and leucine aminopeptidase. *J Antibiot (Tokyo)* 1976;29(8):857–859.
357. Wilkes SH, Prescott JM. The slow, tight binding of bestatin and amastatin to aminopeptidases. *J Biol Chem* 1985;260(24):13154–13162.
358. Lee MD, Antczak C, Li Y, Sirotnak FM, Bornmann WG, Scheinberg DA. A new human peptide deformylase inhibitable by actinonin. *Biochem Biophys Res Commun* 2003;312(2):309–315.
359. Lee MD, She Y, Soskis MJ, Borella CP, Gardner JR, Hayes PA, Dy BM, Heaney ML, Philips MR, Bornmann WG, Sirotnak FM, Scheinberg DA. Human mitochondrial peptide deformylase, a new anticancer target of actinonin-based antibiotics. *J Clin Invest* 2004;114(8):1107–1116.
360. Kruse MN, Becker C, Lottaz D, Kohler D, Yiallouris I, Krell HW, Sterchi EE, Stocker W. Human meprin alpha and beta homo-oligomers: Cleavage of basement membrane proteins and sensitivity to metalloprotease inhibitors. *Biochem J* 2004;378(Pt 2):383–389.
361. Lee SJ, Jang JW, Kim YM, Lee HI, Jeon JY, Kwon YG, Lee ST. Endostatin binds to the catalytic domain of matrix metalloproteinase-2. *FEBS Lett* 2002;519(1–3):147–152.
362. Leyhausen G, Schuster DK, Vaith P, Zahn RK, Umezawa H, Falke D, Muller WE. Identification and properties of the cell membrane bound leucine aminopeptidase interacting with the potential immunostimulant and chemotherapeutic agent bestatin. *Biochem Pharmacol* 1983;32(6):1051–1057.
363. Ray S, Chattopadhyay N, Mitra A, Siddiqi M, Chatterjee A. Curcumin exhibits antimetastatic properties by modulating integrin receptors, collagenase activity, and expression of Nm23 and E-cadherin. *J Environ Pathol Toxicol Oncol* 2003;22(1):49–58.
364. Shishodia S, Potdar P, Gairola CG, Aggarwal BB. Curcumin (diferuloylmethane) down-regulates cigarette smoke-induced NF-kappaB activation through inhibition of I kappa B kinase in human lung epithelial cells: Correlation with suppression of COX-2, MMP-9, and cyclin D1. *Carcinogenesis* 2003;24(7):1269–1279.
365. Takada Y, Aggarwal BB. Betulinic acid suppresses carcinogen-induced NF-kappa B activation through inhibition of I kappa B kinase and p65 phosphorylation: Abrogation of cyclooxygenase-2 and matrix metalloprotease-9. *J Immunol* 2003;171(6):3278–3286.
366. Aggarwal S, Takada Y, Singh S, Myers JN, Aggarwal BB. Inhibition of growth and survival of human head and neck squamous cell carcinoma cells by curcumin via modulation of nuclear factor-kappaB signaling. *Int J Cancer* 2004;111(5):679–692.
367. Mohan R, Sivak J, Ashton P, Russo LA, Pham BQ, Kasahara N, Raizman MB, Fini ME. Curcuminoids inhibit the angiogenic response stimulated by fibroblast growth factor-2, including expression of matrix metalloproteinase gelatinase B. *J Biol Chem* 2000;275(14):10405–10412.
368. Hua G, Tsukamoto K, Taguchi R, Tomita M, Miyajima S, Ikezawa H. Characterization of aminopeptidase N from the brush border membrane of the larvae midgut of silkworm, *Bombyx mori* as a zinc enzyme. *Biochim Biophys Acta* 1998;1383(2):301–310.
369. Hooper NM, Hesp RJ, Tiekou S. Metabolism of aspartame by human and pig intestinal microvillar peptidases. *Biochem J* 1994;298(Pt 3):635–639.
370. Chen DZ, Patel DV, Hackbarth CJ, Wang W, Dreyer G, Young DC, Margolis PS, Wu C, Ni ZJ, Trias J, White RJ, Yuan Z. Actinonin, a naturally occurring antibacterial agent, is a potent deformylase inhibitor. *Biochemistry* 2000;39(6):1256–1262.
371. Matsas R, Stephenson SL, Hryszko J, Kenny AJ, Turner AJ. The metabolism of neuropeptides. Phase separation of synaptic membrane preparations with Triton X-114 reveals the presence of aminopeptidase N. *Biochem J* 1985;231(2):445–449.
372. Murray H, Turner AJ, Kenny AJ. The aminopeptidase activity in the human T-cell lymphoma line (Jurkat) is not at the cell surface and is not aminopeptidase N (CD-13). *Biochem J* 1994;298(Pt 2):353–360.

373. Ward PE, Benter IF, Dick L, Wilk S. Metabolism of vasoactive peptides by plasma and purified renal aminopeptidase M. *Biochem Pharmacol* 1990;40(8):1725–1732.
374. Tobe H, Kojima F, Aoyagi T, Umezawa H. Purification by affinity chromatography using amastatin and properties of aminopeptidase A from pig kidney. *Biochim Biophys Acta* 1980;613(2):459–468.
375. Gee NS, Kenny AJ. Proteins of the kidney microvillar membrane. Enzymic and molecular properties of aminopeptidase W. *Biochem J* 1987;246(1):97–102.
376. Roques BP. Insights into peptide and protein function: A convergent approach. *J Pept Sci* 2001;7(2):63–73.

---

**Daniel Dauzonne** did his undergraduate studies at the Paris VI University and received his PhD in 1981. After a post-doctoral stay with Professor A. G. M. Barrett at the Imperial college in London, he joined the CNRS (Centre National de la Recherche Scientifique) at Institut Curie (Paris), being appointed «Attaché de Recherche», then «Directeur de Recherche» in 1996. His research focuses on the synthesis of new anticancer agents, particularly in the fields of podophyllotoxin and flavonoid derivatives.

**Brigitte Bauvois** received a PhD from the University of Lille (France) in 1982 and did postdoctoral research in the laboratory of Dr S. Roth at University of Pennsylvania in Philadelphia (USA). She joined the INSERM (Institut National de la Santé et de la Recherche Médicale) laboratory of Prof. J. Caen (Paris, France) as “Attaché de Recherche” and became “Directeur de Recherche” in 1993. She is appointed to her current position in the INSERM laboratory of Prof. P. Lesavre (Paris, France). Research interests are in inflammation and cancers, and in mechanisms of biological processes involving proteases and integrins.