



# Article The Impact of Matrix Metalloproteinase-11 Polymorphisms on Colorectal Cancer Progression and Clinicopathological Characteristics

Hsien-Cheng Huang <sup>1,2,†</sup>, Bei-Hao Shiu <sup>1,3,4,†</sup>, Shih-Chi Su <sup>5,6</sup>, Chi-Chou Huang <sup>3,4</sup>, Wen-Chien Ting <sup>3,4</sup>, Lun-Ching Chang <sup>7</sup>, Shun-Fa Yang <sup>1,8,\*</sup> and Ying-Erh Chou <sup>4,8,\*</sup>

- <sup>1</sup> Institute of Medicine, Chung Shan Medical University, Taichung 402, Taiwan; asiantcumed@gmail.com (H.-C.H.); shiubeihao@gmail.com (B.-H.S.)
- <sup>2</sup> Department of Emergency Medicine, Kuang Tien General Hospital, Taichung 433, Taiwan
- <sup>3</sup> Department of Surgery, Chung Shan Medical University Hospital, Taichung 402, Taiwan; hcjy341@gmail.com (C.-C.H.); tingwenchien@gmail.com (W.-C.T.)
- <sup>4</sup> School of Medicine, Chung Shan Medical University, Taichung 402, Taiwan
- <sup>5</sup> Whole-Genome Research Core Laboratory of Human Diseases, Chang Gung Memorial Hospital, Keelung 204, Taiwan; ssu1@cgmh.org.tw
- <sup>5</sup> Department of Dermatology, Drug Hypersensitivity Clinical and Research Center, Chang Gung Memorial Hospital, Linkou 333, Taiwan
- <sup>7</sup> Department of Mathematical Sciences, Florida Atlantic University, Boca Raton, FL 33431, USA; changl@fau.edu
- <sup>8</sup> Department of Medical Research, Chung Shan Medical University Hospital, Taichung 402, Taiwan
- Correspondence: ysf@csmu.edu.tw (S.-F.Y.); intointo814@gmail.com (Y.-E.C.)
- + These authors contributed equally to this work.

Abstract: Colorectal cancer (CRC) is the third most common cause of cancer mortality worldwide and the most prevalent cancer in Taiwan. The matrix metalloproteinase (MMP)-11 is a proteolytic enzyme of the MMP family which is involved in extracellular matrix degradation and tissue remodeling. In this study, we focused on the associations of MMP-11 single-nucleotide polymorphisms (SNPs) with CRC susceptibility and clinicopathological characteristics. The MMP-11 SNPs rs131451, rs738791, rs2267029, rs738792, and rs28382575 in 479 controls and 479 patients with CRC were analyzed with real-time polymerase chain reaction. We found that the MMP-11 SNP rs738792 "TC + CC" genotype was significantly associated with perineural invasion in colon cancer patients after controlling for clinical parameters [OR (95% CI) = 1.783 (1.074–2.960); *p* = 0.025]. The *MMP-11* rs131451 "TC + CC" genotypic variants were correlated with greater tumor T status [OR (95% CI):1.254 (1.025-1.534); p = 0.028] and perineural invasion [OR (95% CI):1.773 (1.027–3.062); p = 0.040) in male CRC patients. Furthermore, analyses of The Cancer Genome Atlas (TCGA) revealed that MMP-11 levels were upregulated in colorectal carcinoma tissue compared with normal tissues and were correlated with advanced stage, larger tumor sizes, and lymph node metastasis. Moreover, the data from the Genotype-Tissue Expression (GTEx) database exhibited that the MMP-11 rs738792 "CC" and "CT" genotypic variants have higher MMP-11 expression than the "TT" genotype. In conclusion, our results have demonstrated that the MMP-11 SNPs rs738792 and rs131451 may have potential to provide biomarkers to evaluate CRC disease progression, and the MMP-11 rs131451 polymorphism may shed light on sex discrepancy in CRC development and prognosis.

Keywords: colorectal cancer; MMP-11; polymorphism



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## 1. Introduction

Colorectal cancer (CRC), which is the third leading cause of cancer death worldwide, ranks as the third most common adult cancer in men the second most common in women [1–4]. In Taiwan, the incidence of CRC is the highest among all cancers, and CRC is responsible for the third highest cancer mortality [5]. Risk factors such as age, male sex, family history of colorectal cancer, race and ethnicity, cigarette smoking, excessive alcohol consumption, high consumption of red and processed meat, diabetes, obesity, and inflammatory bowel disease were suggested to be associated with CRC carcinogenesis [6–17].

The matrix metalloproteinases (MMPs) are a zinc-dependent endopeptidases family which is involved in extracellular matrix (ECM) degradation and tissue remodeling [18–21]. MMP-11, or stromelysin 3, is a proteolytic enzyme which belongs to the MMP family [22–24]. Previous studies have suggested that the MMP-11 may play a regulatory role in cancer cell growth, tumor migration, invasion, and metastasis in various cancers [22,24–28]. In colorectal cancer, it was suggested that MMP-11 is highly expressed in colonic carcinoma [29], and the elevated serum levels and mRNA expression of MMP-11 were correlated with poor prognosis in colon cancer patients [30,31]. Single-nucleotide polymorphisms (SNPs) are the most common type of genetic variations which may lead to amino acid substitution and alteration of protein function [32,33]. The MMP-11 SNPs were found to be associated with increased risk of cancer progression and development, metastasis, recurrence, or poor prognosis in many cancers including oral squamous cell carcinoma (OSCC) [34], hepatocellular carcinoma (HCC) [35,36], prostate cancer [37], and urothelial cell carcinoma (UCC) [38]. However, the associations and influences of MMP-11 polymorphisms regarding CRC tumor progression and clinicopathologic characteristics remained uninvestigated. In this study, we focused on five SNPs of MMP-11 rs131451, rs738791, rs2267029, rs738792, and rs28382575, and try to unveil their correlations to CRC susceptibility and clinicopathologic characteristics.

### 2. Materials and Methods

# 2.1. Subjects

In this study, we enrolled 479 CRC patients and 479 cancer-free controls. All participants were recruited from 2016 to 2020 at Chung Shan Medical University Hospital in Taichung, Taiwan. According to the American Joint Committee on Cancer (AJCC) [39], the TNM staging of the CRC patients who enrolled in our study were staged clinically at the time of diagnosis. The tumor differentiation was examined and rated under the AJCC classification by a pathologist. The demographic data of age and gender were reported by each participant and recorded. Individuals with neither a history of cancer of any sites nor any self-reported diseases, such as asthma, diabetes, or cardiovascular, autoimmune, and neurological diseases, were enrolled in the control group. This project was approved by the institutional review board of Chung Shan Medical University Hospital (IRB number CS1-20111), and informed written consent was provided by each participant at enrollment.

#### 2.2. Sample Preparation and DNA Extraction

For genomic DNA extraction, the peripheral blood specimens from normal controls and CRC patients who enrolled in our study were collected. The EDTA containing tubes were used to preserve the samples of peripheral whole blood. The blood samples were centrifuged with the settings of 3000 rpm, 10 min, and the buffy coats from centrifuged whole blood specimens were extracted and further used for the DNA extraction [40,41]. The genomic DNA extraction assay was performed with QIAamp DNA blood mini kits following the protocols of the manufacturer's manual to collect the DNA. The DNA elution was completed with the Tris-EDTA (TE) buffer, which was used to dissolve the DNA. Extracted DNA was further used as a DNA template in the real-time polymerase chain reactions (PCRs).

#### 2.3. Selection of MMP-11 SNPs

In the current study, a total of five SNPs of *MMP-11* rs131451, rs738791, rs2267029, rs738792, and rs28382575 were selected based on the International HapMap Project database [33]. The *MMP-11* rs131451 SNP was selected because the *MMP-11* rs131451 polymorphisms were suggested to be associated with late-stage tumors and high-risk D'Amico classification in prostate cancer patients with biochemical recurrence [37]. The *MMP-11* rs738791 was selected because patients with the rs738791 polymorphic variant were observed to have greater risk of HCC compared with the wild-type (CC) carriers [35]. The MMP-11 SNP rs738792 was selected because the OSCC patients who carried at least one polymorphic C allele of *MMP-11* rs738792 showed an increased incidence of lymph node metastasis compared with those patients with homozygous T/T [34], and carriers who have at least one C allele of the *MMP-11* SNP rs738792 are likely to progress to Child–Pugh B or C grade in HCC patients [35]. The *MMP-11* SNP rs28382575 was selected because the HCC patients with at least one C allele of the *MMP-11* SNP rs28382575 are suggested to have a higher risk of developing stage III/IV disease, large tumors, or lymph node metastasis [35].

#### 2.4. MMP-11 SNPs Genotyping

Assessment of allelic discrimination for the MMP-11 rs131451 (assay IDs: C\_\_2213679\_30), rs738791 (assay IDs: C\_\_2448099\_30), rs2267029 (assay IDs: C\_\_15871447\_20), rs738792 (assay IDs: C\_\_2213764\_20), and rs28382575 (assay IDs: C\_\_61238655\_10) SNPs was performed with an ABI StepOne Software v2.3 Real-Time PCR System. The TaqMan assay was used for the analysis of genotyping. The analysis and calculation of the final data of genotyping was processed with the SDS 7000 series software (Applied Biosystems, Foster City, CA, USA).

#### 2.5. Statistical Analysis

To compare the age (years), gender, tumor location, tumor stage, tumor T status, lymph node status, metastasis, lymphovascular invasion, perineural invasion, and pathologic grading, the Chi-squared test or Student's *t* test was performed between the patients with CRC and the controls. A statistical significance was suggested if p < 0.05. To compare the odds ratio (ORs) with their 95% confidence intervals (CIs) of the association between the genotypic frequencies and CRC risk, and the clinical pathological characteristics, the data was analyzed and assessed by the logistic regression models. All the data analysis in the current study was calculated and evaluated with SAS statistical software (Version 9.1, 2005; SAS Institute, Cary, NC, USA).

#### 3. Results

#### 3.1. Demographic and Clinical Characteristics of Study Cohorts

The distribution of demographical characteristics in 479 controls and 479 patients with CRC is demonstrated in Table 1. In our current study, we observed that the distribution of age (years) < 65 was 278 (58.0%) in controls and 251 (52.4%) in CRC patients, and the age  $\geq 65$  in controls and CRC patients was 201 (42.0%) and 228 (47.6%), respectively. For the distributions of gender, the male controls and CRC patients were 294 (61.4%) and 282 (58.9%), whereas the female controls and patients were 185 (38.6%) and 197 (41.1%), respectively. However, no statistically significant differences were found for age and gender between the CRC patients and the controls (Table 1).

Variable	Controls (N = 479) n (%)	Patients (N = 479) n (%)	p Value
Age (yrs)			
<65	278 (58.0%)	251 (52.4%)	0.079
$\geq 65$	201 (42.0%)	228 (47.6%)	
Gender	. ,		
Male	294 (61.4%)	282 (58.9%)	0.428
Female	185 (38.6%)	197 (41.1%)	
Tumor location			
Rectum		110 (23.0%)	
Left colon		222 (46.3%)	
Right colon		147 (30.7%)	
Stage			
I + II		229 (47.8%)	
III + IV		250 (52.2%)	
Tumor T status			
T1-T2		116 (24.2%)	
T3–T4		363 (75.8%)	
Lymph node status			
N0		239 (49.9%)	
N1 + N2		240(50.1%)	
Metastasis			
M0		402 (83.9%)	
M1		77 (16.1%)	
Lymphovascular			
invasion			
No		267 (55.7%)	
Yes		212 (44.3%)	
Perineural invasion			
No		272 (56.8%)	
Yes		207 (43.2%)	
Pathologic grading			
Well		6 (1.3%)	
Moderately		437 (91.2%)	
Poorly		36 (7.5%)	

**Table 1.** The distributions of demographical and clinical characteristics in 479 controls and 479 patients with CRC.

3.2. MMP-11 Gene Polymorphisms were Associated with the Clinicopathological Characteristics of CRC

The genotype distributions of *MMP-11* gene polymorphisms in 479 controls and 479 patients with CRC are listed in Table 2. The highest distribution frequencies in patients with CRC of *MMP-11* genetic polymorphisms rs131451, rs738791, rs2267029, rs738792, and rs28382575 were heterozygous for TC, homozygous for CC, homozygous for GG, homozygous for TT, and homozygous for TT, respectively. Logistic regression models were adopted to estimate the odds ratios (ORs) and their 95% confidence intervals (CIs). After adjustment for the effects of age and gender, no significant associations were found between the CRC patients and the controls (Table 2).

We further analyzed the distribution frequency of the clinical status and *MMP-11* genotype frequencies. In 369 of a total 479 CRC patients, a significant association was found in those individuals who carried the *MMP-11* rs738792 "TC + CC" genotypic variants, with a higher risk of perineural invasion of colon cancer patients after controlling for stages, tumor T status, lymph node status, metastasis, lymphovascular invasion, and cell differentiation (p = 0.025) (Table 3). We further analyzed the *MMP-11* polymorphisms of the clinical status in male and female CRC patients. The results demonstrated that the male CRC patients who carried the *MMP-11* rs131451 "TC + CC" genotype were associated with greater tumor T status (p = 0.028) and perineural invasion (p = 0.040) (Table 4). We further analyzed correlations of MMP-11 levels and their clinical parameters in CRC from The

Cancer Genome Atlas (TCGA) dataset. We observed that MMP-11 expression was prone to be upregulated in colorectal carcinoma tissue compared with normal tissues (Figure 1A). Moreover, MMP-11 levels were also correlated with late stage, larger tumor sizes, and lymph node metastasis (Figure 1B–D). We further used the Genotype-Tissue Expression (GTEx) database to evaluate the correlations of *MMP-11* rs738792 and rs131451 SNPs to MMP-11 expression. The results of the GTEx database exhibited that individuals with the "C" allele of the *MMP-11* rs738792 (CC and CT) genotype were associated with higher MMP-11 expression in the sigmoid colon compared with the "TT" carriers (p = 0.00048) (Figure 1). For *MMP-11* rs131451 SNPs, individuals with the rs131451 polymorphisms "CC" and "CT" genotype were associated with higher MMP-11 expression in the sigmoid colon (p = 0.031), respectively, compared with the wild-type "TT" carriers (Figure 2).

Variable	Controls (N = 479) n (%)	Patients (N = 479) n (%)	OR (95% CI)	AOR (95% CI)		
rs131451						
TT	162 (33.8%)	161 (33.6%)	1.000 (reference)	1.000 (reference)		
TC	234 (48.9%)	246 (51.4%)	1.058 (0.798-1.403)	1.065 (0.802-1.413)		
CC	83 (17.3%)	72 (15.0%)	0.873 (0.595-1.281)	0.889 (0.605-1.306)		
TC + CC	317 (66.2%)	318 (66.4%)	1.009 (0.772-1.319)	1.019 (0.779–1.333)		
rs738791		· · · ·	, , , , , , , , , , , , , , , , , , ,	· · · · ·		
CC	234 (48.9%)	213 (44.5%)	1.000 (reference)	1.000 (reference)		
СТ	196 (40.9%)	214 (44.7%)	1.199 (0.917–1.569)	1.204 (0.920-1.575)		
TT	49 (10.2%)	52 (10.9%)	1.166 (0.757-1.796)	1.202 (0.778-1.856)		
CT + TT	245 (51.1%)	266 (55.5%)	1.193 (0.925–1.538)	1.203 (0.933-1.553)		
rs2267029	· · · ·	· · ·	· · · ·	· · · · ·		
GG	266 (55.5%)	263 (54.9%)	1.000 (reference)	1.000 (reference)		
GA	185 (38.6%)	188 (39.2%)	1.028 (0.789-1.340)	1.022 (0.784-1.333)		
AA	28 (5.9%)	28 (5.9%)	1.011 (0.583-1.755)	1.033 (0.594-1.794)		
GA + AA	213 (44.5%)	216 (45.1%)	1.026 (0.795-1.323)	1.024 (0.793-1.321)		
rs738792						
TT	246 (51.4%)	241 (50.3%)	1.000 (reference)	1.000 (reference)		
TC	195 (40.7%)	203 (42.4%)	1.063 (0.815-1.385)	1.070 (0.820-1.396)		
CC	38 (7.9%)	35 (7.3%)	0.940 (0.575-1.538)	0.955 (0.583-1.564)		
TC + CC	233 (48.6%)	238 (49.7%)	1.043 (0.809-1.343)	1.051 (0.815-1.355)		
rs28382575						
TT	457 (95.4%)	446 (93.1%)	1.000 (reference)	1.000 (reference)		
TC	22 (4.6%)	33 (6.9%)	1.537 (0.882–2.677)	1.596 (0.914-2.787)		
CC	0 (0%)	0 (0.0%)				
TC + CC	22 (4.6%)	33 (6.9%)	1.537 (0.882–2.677)	1.596 (0.914–2.787)		

**Table 2.** Genotype distributions of *MMP-11* gene polymorphisms in 479 controls and 479 patients with CRC.

The odds ratios (ORs) with their 95% confidence intervals were estimated by logistic regression models.

The adjusted odds ratios (AORs) with their 95% confidence intervals were estimated by multiple logistic regression models after controlling for age and gender.

Variable	All (N = 479)				Rectum (N = 110)			Colon (N = 369)		
	TT (N = 241)	TC + CC (N = 238)	p Value	TT (N = 60)	TC + CC (N = 50)	p Value	TT (N = 181)	TC + CC (N = 188)	p Value	
Stages										
I + II	115 (47.7%)	114 (47.9%)	p = 0.892	31 (51.7%)	29 (58.0%)	p = 0.482	84 (46.4%)	85 (45.2%)	p = 0.945	
III + IV	126 (52.3%)	124 (52.1%)	,	29 (48.3%)	21 (42.0%)	,	97 (53.6%)	103 (54.8%)	,	
Tumor T status										
T1 + T2	62 (25.7%)	54 (22.7%)	p = 0.805	20 (33.3%)	16 (32.0%)	p = 0.569	42 (23.2%)	38 (20.2%)	p = 0.885	
T3 + T4	179 (74.3%)	184 (77.3%)		40 (66.7%)	34 (68.0%)		139 (76.8%)	150 (79.8%)		
Lymph node status										
Negative	119 (49.4%)	120 (50.4%)	p = 0.630	32 (53.3%)	30 (60.0%)	p = 0.411	87 (48.1%)	90 (47.9%)	p = 0.643	
Positive	122 (50.6%)	118 (49.6%)		28 (46.7%)	20 (40.0%)		94 (51.9%)	98 (52.1%)		
Metastasis										
Negative	204 (84.6%)	198 (83.2%)	p = 0.955	46 (76.7%)	45 (90.0%)	p = 0.090	158 (87.3%)	153 (81.4%)	p = 0.212	
Positive	37 (15.4%)	40 (16.8%)	,	14 (23.3%)	5 (10.0%)	,	23 (12.7%)	35 (18.6%)	,	
Lymphovascular invasion										
No	136 (56.4%)	131 (55.0%)	p = 0.457	38 (63.3%)	33 (66.0%)	p = 0.830	98 (54.1%)	98 (52.1%)	p = 0.426	
Yes	105 (43.6%)	107 (45.0%)		22 (36.7%)	17 (34.0%)		83 (45.9%)	90 (47.9%)		
Perineural invasion										
No	147 (61.0%)	125 (52.5%)	p = 0.051	39 (65.0%)	34 (68.0%)	p = 0.998	108 (59.7%)	91 (48.4%)	$p = 0.025^{a}$	
Yes	94 (39.0%)	113 (47.5%)		21 (35.0%)	16 (32.0%)		73 (40.3%)	97 (51.6%)		
Cell differentiation	. ,	. ,		. ,	. ,		. ,	. ,		
Well/Moderately	227 (94.2%)	216 (90.8%)	p = 0.164	60 (100%)	49 (98.0%)		167 (92.3%)	167 (88.8%)	p = 0.323	
Poorly	14 (5.8%)	22 (9.2%)	,	0 (0.0%)	1 (2.0%)		14 (7.7%)	21 (11.2%)	,	

Table 3. Distribution frequency of the clinical status and MMP-11 rs738792 genotype frequencies in 479 CRC patients.
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<sup>a</sup> AOR (95% CI):1.783 (1.074–2.960). The adjusted odds ratios (AORs) with their 95% confidence intervals were estimated by multiple logistic regression models after controlling for stages, tumor T status, lymph node status, metastasis, lymphovascular invasion, perineural invasion, and cell differentiation.

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Variable	A11 (N = 479)			Male (N = 282)			Female (N = 197)		
	TT (N = 161)	TC + CC (N = 318)	p Value	TT (N = 96)	TC + CC (N = 186)	<i>p</i> value	TT (N = 65)	TC + CC (N = 132)	p Value
Stages									
I + II	80 (49.7%)	149 (46.9%)	p = 0.317	51 (53.1%)	94 (50.5%)	p = 0.812	29 (44.6%)	55 (41.7%)	p = 0.134
III + IV	81 (50.3%)	169 (53.1%)		45 (46.9%)	92 (49.5%)		36 (55.4%)	77 (58.3%)	
Tumor T status									
T1 + T2	47 (29.2%)	69 (21.7%)	p = 0.216	34 (35.4%)	43 (23.1%)	p = 0.028 <sup>a</sup>	13 (20.0%)	26 (19.7%)	p = 0.999
T3 + T4	114 (70.8%)	249 (78.3%)		62 (64.6%)	143 (76.9%)		52 (80.0%)	106 (80.3%)	
Lymph node status									
Negative	81 (50.3%)	158 (49.7%)	p = 0.238	52 (54.2%)	99 (53.2%)	p = 0.545	29 (44.6%)	59 (44.7%)	p = 0.172
Positive	80 (49.7%)	160 (50.3%)		44 (45.8%)	87 (46.8%)		36 (55.4%)	73 (55.3%)	
Metastasis									
Negative	135 (83.9%)	267 (84.0%)	p = 0.380	84 (87.5%)	157 (84.4%)	p = 0.663	51 (78.5%)	110 (83.3%)	p = 0.152
Positive	26 (16.1%)	51 (16.0%)	,	12 (12.5%)	29 (15.6%)	,	14 (21.5%)	22 (16.7%)	
Lymphovascular invasion									
No	95 (59.0%)	172 (54.1%)	p = 0.942	59 (61.5%)	104 (55.9%)	p = 0.697	36 (55.4%)	68 (51.5%)	p = 0.554
Yes	66 (41.0%)	146 (45.9%)		37 (38.5%)	82 (44.1%)		29 (44.6%)	64 (48.5%)	
Perineural invasion									
No	99 (61.5%)	173 (54.4%)	p = 0.341	66 (68.8%)	104 (55.9%)	p = 0.040 b	33 (50.8%)	69 (52.3%)	p = 0.849
Yes	62 (38.5%)	145 (45.6%)	,	30 (31.2%)	82 (44.1%)		32 (49.2%)	63 (47.7%)	
Cell differentiation	. ,	. ,		. ,	. ,		. ,	. ,	
Well/Moderately	154 (95.7%)	289 (90.9%)	p = 0.096	91 (94.8%)	165 (88.7%)	p = 0.129	63 (96.9%)	124 (93.9%)	p = 0.371
Poorly	7 (4.3%)	29 (9.1%)		5 (5.2%)	21 (11.3%)		2 (3.1%)	8 (6.1%)	

Table 4. Distribution frequency of the clinical status and MMP-11 rs131451 genotype frequencies in 479 CRC patients with different genders.

<sup>a</sup> AOR (95% CI):1.254 (1.025–1.534); <sup>b</sup> AOR (95% CI):1.773 (1.027–3.062). The adjusted odds ratios (AORs) with their 95% confidence intervals were estimated by multiple logistic regression models after controlling for stages, tumor T status, lymph node status, metastasis, lymphovascular invasion, perineural invasion, and cell differentiation.



**Figure 1.** MMP-11 level of colorectal cancer patients from TCGA database. (**A**) MMP-11 levels were compared between the colorectal cancer tumor tissues and normal tissue. (**B**) MMP-11 levels were compared between stage I + II and stage III + IV. (**C**) MMP-11 levels were compared between the T1 + T2 stage and T3 + T4 stage. (**D**) MMP-11 levels were compared between the N0 stage and N1 stage.



**Figure 2.** Distribution of MMP-11 expression in whole blood and colon (sigmoid, transverse) of *MMP-11* SNPs rs738792, rs131451 from Genotype-Tissue Expression (GTEx) database.

#### 4. Discussion

In this study, we demonstrated the associations between the *MMP-11* SNPs and CRC. The incidence of CRC is typically low at ages younger than 50 years but strongly increases with age [6,42]. Although the incidence of early-onset CRC (EOCRC) patients, referring to those individuals younger than 50 years, has been rapidly rising over the last 20 years [43–45], the median age at diagnosis of CRC is about 70 years in developed countries [6,46]. Previous study has suggested that the *MMP-11* polymorphisms and environmental carcinogens were associated with an increased risk for the development of OSCC [34]. Moreover, carriers of the CT + TT allele of the *MMP-11* rs738791 variant were suggested to possess greater risk of HCC compared with the wild-type (CC) carriers [35],

and those with the *MMP-11* rs28382575 polymorphic "CT" genotype were found to be susceptible to UCC [38]. However, after we analyzed the genotype distributions of *MMP-11* polymorphisms in 479 controls and 479 patients with CRC, no statistically significant association was found between these two groups (Table 2), suggesting a limited effect of *MMP-11* polymorphisms for the susceptibility of CRC carcinogenesis.

We further analyzed the correlations between the *MMP-11* SNPs and clinical status of CRC, and we found that in 369 of a total 479 CRC patients, individuals who carried the *MMP-11* rs738792 "TC + CC" polymorphic variants were associated with higher risk of perineural invasion in colon cancer patients after controlling for clinical parameters (p = 0.025) (Table 3). Moreover, after we analyzed the *MMP-11* polymorphisms of the clinical status in CRC patients with different genders, we found that the male CRC patients who carried the *MMP-11* rs131451 "TC + CC" genotypic variants were correlated with greater tumor T status (p = 0.028) and perineural invasion (p = 0.040) (Table 4). Furthermore, numerous studies reported that perineurial invasion is associated with poor prognosis in CRC patients [47–49]. Therefore, the correlations among CRC prognosis and *MMP-11* polymorphism will be further investigated in our future work.

Generally, the colorectal cancer screening guidelines do not distinguish females from males, and sex specificity was not considered for interpretation in CRC despite the differences in tumor location between women and men [8,50,51]. Consistent with these results, the sex specificity was not significant between the CRC patients and controls in our study (p = 0.428) (Table 1). However, for the clinical status, results of the *MMP-11* rs131451 polymorphic variants showed discrepancy between male and female CRC patients (Table 4). Intriguingly, similar results of MMP-11 rs131451 expression with sex differences were observed in our previous studies. The MMP-11 SNPs, including the rs131451, have shown no impact on uterine cervical cancer in Taiwanese women [52], whereas the MMP-11 rs131451 "TC + CC" polymorphic variants were correlated with advanced clinical stage (T stage; p = 0.007) and high-risk D'Amico classification (p = 0.015) in prostate cancer patients with biochemical recurrence [37]. Of note, analyses of TCGA revealed that MMP-11 levels were correlated with larger tumor sizes (Figure 1). Moreover, according to the data of the GTEx database, it was suggested that both the MMP-11 rs131451 "CC + CT" genotypic variants expressed in the sigmoid colon or transverse colon were associated with higher MMP-11 expression compared with the wild-type "TT" carriers (Figure 2). However, in CRC, not only the male but also the female CRC patients with rs131451 "TC + CC" genotype were significantly associated with advanced tumor T status and perineural invasion (Table 4). In contrast, in the aspect of CRC prognostic biomarkers, it was suggested that there are clear sex differences in CRC characteristics, and sex-specific CRC prognostic biomarkers including ESM1, GUCA2A, and VWA2 for males and CLDN1 and FUT1 for female CRC patients were proposed [53]. One possible mechanism to explain this phenomenon was the interaction of sex hormones and their regulators in CRC. The sex hormones were suggested as contributors for gender disparity in incidence and mortality of CRC [54–57]. Notably, despite its ambiguous and contradictory role in CRC, testosterone was suggested to be involved in CRC development and prognosis [56,58], and the androgen was suggested to regulate MMPs (MMP-2) and the cellular processes of intimal hyperplasia [59]. Moreover, in the aspect of androgen receptor (AR), it was suggested that the expression of MMP-2 and MMP-9 was associated with the presence of AR in epithelial ovarian tumors [60], and the expressions of the MMP-11 and AR were significantly higher in cancer-associated fibroblasts (CAFs) from castration-resistant prostate tumors (CRPC) [61], suggesting a possible mechanism: that the higher MMP-11 expression resulting from MMP-11 rs131451 polymorphisms might be associated with the induction of AR presence in CRC (Table 4) (Figures 1 and 2). Men have a 20- to 25-fold higher testosterone production when compared with women [62,63], and the testosterone levels in women prior to menopause decline approximately 50% compared with their third decade [62,64]. Therefore, although the correlations and interactions of sex hormones with MMP-11 expression have remained unclear to date, it can be proposed that it is the AR presence induced by higher levels

of MMP-11 expression which result from MMP-11 rs131451 "CC + CT" polymorphisms, but also the direct interaction of testosterone and MMP-11, which is responsible for the discrepancy of sex specificity and poor prognosis in prostate cancer [61] and CRC [30,31] (Table 4) (Figure 2). In addition, for the application of MMPs as biomarkers in CRC detection, a previous study led by Koga et al. [65] demonstrated that the messenger RNA (mRNA) expression of MMP-7 in the colonocytes isolated from feces was significantly higher in CRC patients than in healthy volunteers [65,66]. Moreover, numerous studies reported that expressions of MMP-14, MMP-17, and MMP-19 may be used as prognostic markers in CRC [67-69]. Therefore, a multi-SNP analysis for MMP-11, MMP-14, MMP-17, and MMP-19 will be investigated in our future work. Taken together, although the exact mechanisms and regulations remained incompletely understood, the MMP-11 rs738792 and rs131451 SNPs may provide potential candidates for CRC biomarkers since these polymorphic variants were both linked with perineural invasion (Tables 3 and 4) and higher expression of MMP-11 (Figure 1). Of note, the MMP-11 rs131451 "TC + CC" genotype was further associated with greater tumor T status (p = 0.028) and perineural invasion (p = 0.040) in male CRC patients (Table 4), thereby providing a possible more detailed mechanism to explain the reason why the sex discrepancy of MMP-11 expression in CRC exists, and result in CRC disease development and prognosis with sex differences [31]. However, future well-designed studies are required to elucidate the exact mechanisms of MMP-11 polymorphisms in CRC progression considering sex specificity, especially the detailed influences of sex hormones such as the decreasing levels of androgen and testosterone with age to MMP-11 expression in CRC disease progression and prognosis.

#### 5. Conclusions

In conclusion, our results have demonstrated that despite the fact that *MMP-11* SNPs were not associated with CRC susceptibility, CRC patients who carried the *MMP-11* rs738792 "TC + CC" polymorphic variants were associated with higher risk of perineural invasion of the colon, and the male CRC patients who carried the *MMP-11* rs131451 "TC + CC" genotypic variants were associated with greater tumor T status and perineural invasion. The *MMP-11* rs738792 and rs131451 polymorphisms were also associated with higher MMP-11 expression either in the sigmoid colon or transverse colon. The *MMP-11* SNPs rs738792 and rs131451 may have potential to provide biomarkers to evaluate CRC disease progression, and the *MMP-11* rs131451 polymorphisms may shed light on sex discrepancy in CRC development and prognosis.

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## References

- 1. Li, J.; Ma, X.; Chakravarti, D.; Shalapour, S.; DePinho, R.A. Genetic and biological hallmarks of colorectal cancer. *Genes Dev.* 2021, 35, 787–820. [CrossRef]
- 2. Bray, F.; Ferlay, J.; Soerjomataram, I.; Siegel, R.L.; Torre, L.A.; Jemal, A. Global cancer statistics 2018: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. *CA Cancer J. Clin.* **2018**, *68*, 394–424. [CrossRef] [PubMed]
- 3. Dekker, E.; Tanis, P.J.; Vleugels, J.L.A.; Kasi, P.M.; Wallace, M.B. Colorectal cancer. *Lancet* 2019, 394, 1467–1480. [CrossRef]
- Biller, L.H.; Schrag, D. Diagnosis and Treatment of Metastatic Colorectal Cancer: A Review. JAMA 2021, 325, 669–685. [CrossRef] [PubMed]
- 5. Chuang, J.P.; Lee, J.C.; Leu, T.H.; Hidajah, A.C.; Chang, Y.H.; Li, C.Y. Association of gout and colorectal cancer in Taiwan: A nationwide population-based cohort study. *BMJ Open* **2019**, *9*, e028892. [CrossRef] [PubMed]
- 6. Brenner, H.; Kloor, M.; Pox, C.P. Colorectal cancer. Lancet 2014, 383, 1490–1502. [CrossRef]
- Sninsky, J.A.; Shore, B.M.; Lupu, G.V.; Crockett, S.D. Risk Factors for Colorectal Polyps and Cancer. *Gastrointest. Endosc. Clin. N. Am.* 2022, 32, 195–213. [CrossRef]
- 8. Kim, S.E.; Paik, H.Y.; Yoon, H.; Lee, J.E.; Kim, N.; Sung, M.K. Sex- and gender-specific disparities in colorectal cancer risk. *World J. Gastroenterol.* 2015, 21, 5167–5175. [CrossRef]
- 9. Sankaranarayanan, R.; Swaminathan, R.; Brenner, H.; Chen, K.; Chia, K.S.; Chen, J.G.; Law, S.C.; Ahn, Y.O.; Xiang, Y.B.; Yeole, B.B.; et al. Cancer survival in Africa, Asia, and Central America: A population-based study. *Lancet Oncol.* 2010, *11*, 165–173. [CrossRef]
- 10. Taylor, D.P.; Burt, R.W.; Williams, M.S.; Haug, P.J.; Cannon-Albright, L.A. Population-based family history-specific risks for colorectal cancer: A constellation approach. *Gastroenterology* **2010**, *138*, 877–885. [CrossRef]
- 11. Jess, T.; Rungoe, C.; Peyrin-Biroulet, L. Risk of colorectal cancer in patients with ulcerative colitis: A meta-analysis of populationbased cohort studies. *Clin. Gastroenterol. Hepatol.* **2012**, *10*, 639–645. [CrossRef] [PubMed]
- 12. Liang, P.S.; Chen, T.Y.; Giovannucci, E. Cigarette smoking and colorectal cancer incidence and mortality: Systematic review and meta-analysis. *Int. J. Cancer* 2009, *124*, 2406–2415. [CrossRef] [PubMed]
- Fedirko, V.; Tramacere, I.; Bagnardi, V.; Rota, M.; Scotti, L.; Islami, F.; Negri, E.; Straif, K.; Romieu, I.; La Vecchia, C.; et al. Alcohol drinking and colorectal cancer risk: An overall and dose-response meta-analysis of published studies. *Ann. Oncol.* 2011, 22, 1958–1972. [CrossRef] [PubMed]
- 14. Chan, D.S.; Lau, R.; Aune, D.; Vieira, R.; Greenwood, D.C.; Kampman, E.; Norat, T. Red and processed meat and colorectal cancer incidence: Meta-analysis of prospective studies. *PLoS ONE* **2011**, *6*, e20456. [CrossRef] [PubMed]
- 15. Ma, Y.; Yang, Y.; Wang, F.; Zhang, P.; Shi, C.; Zou, Y.; Qin, H. Obesity and risk of colorectal cancer: A systematic review of prospective studies. *PLoS ONE* **2013**, *8*, e53916. [CrossRef] [PubMed]
- 16. Jiang, Y.; Ben, Q.; Shen, H.; Lu, W.; Zhang, Y.; Zhu, J. Diabetes mellitus and incidence and mortality of colorectal cancer: A systematic review and meta-analysis of cohort studies. *Eur. J. Epidemiol.* **2011**, *26*, 863–876. [CrossRef]
- 17. Keller, D.S.; Windsor, A.; Cohen, R.; Chand, M. Colorectal cancer in inflammatory bowel disease: Review of the evidence. *Tech. Coloproctol.* **2019**, *23*, 3–13. [CrossRef]
- 18. Hsiao, Y.H.; Su, S.C.; Lin, C.W.; Chao, Y.H.; Yang, W.E.; Yang, S.F. Pathological and therapeutic aspects of matrix metalloproteinases: Implications in childhood leukemia. *Cancer Metastasis Rev.* **2019**, *38*, 829–837. [CrossRef]
- 19. Yang, J.S.; Lin, C.W.; Su, S.C.; Yang, S.F. Pharmacodynamic considerations in the use of matrix metalloproteinase inhibitors in cancer treatment. *Expert Opin. Drug Metab. Toxicol.* **2016**, *12*, 191–200. [CrossRef]
- 20. Su, S.C.; Hsieh, M.J.; Yang, W.E.; Chung, W.H.; Reiter, R.J.; Yang, S.F. Cancer metastasis: Mechanisms of inhibition by melatonin. *J. Pineal Res.* 2017, 62, e12370. [CrossRef]
- 21. Su, C.W.; Lin, C.W.; Yang, W.E.; Yang, S.F. TIMP-3 as a therapeutic target for cancer. *Ther. Adv. Med. Oncol.* 2019, 11, 1758835919864247. [CrossRef]
- 22. Zhang, X.; Huang, S.; Guo, J.; Zhou, L.; You, L.; Zhang, T.; Zhao, Y. Insights into the distinct roles of MMP-11 in tumor biology and future therapeutics (Review). *Int. J. Oncol.* **2016**, *48*, 1783–1793. [CrossRef] [PubMed]
- Matziari, M.; Dive, V.; Yiotakis, A. Matrix metalloproteinase 11 (MMP-11; stromelysin-3) and synthetic inhibitors. *Med. Res. Rev.* 2007, 27, 528–552. [CrossRef] [PubMed]
- 24. Pittayapruek, P.; Meephansan, J.; Prapapan, O.; Komine, M.; Ohtsuki, M. Role of Matrix Metalloproteinases in Photoaging and Photocarcinogenesis. *Int. J. Mol. Sci.* **2016**, *17*, 868. [CrossRef] [PubMed]
- 25. Kossakowska, A.E.; Huchcroft, S.A.; Urbanski, S.J.; Edwards, D.R. Comparative analysis of the expression patterns of metalloproteinases and their inhibitors in breast neoplasia, sporadic colorectal neoplasia, pulmonary carcinomas and malignant non-Hodgkin's lymphomas in humans. *Br. J. Cancer* **1996**, *73*, 1401–1408. [CrossRef]
- Scheau, C.; Badarau, I.A.; Costache, R.; Caruntu, C.; Mihai, G.L.; Didilescu, A.C.; Constantin, C.; Neagu, M. The Role of Matrix Metalloproteinases in the Epithelial-Mesenchymal Transition of Hepatocellular Carcinoma. *Anal. Cell. Pathol.* 2019, 2019, 9423907. [CrossRef]
- 27. Greco, M.; Arcidiacono, B.; Chiefari, E.; Vitagliano, T.; Ciriaco, A.G.; Brunetti, F.S.; Cuda, G.; Brunetti, A. HMGA1 and MMP-11 Are Overexpressed in Human Non-melanoma Skin Cancer. *Anticancer Res.* **2018**, *38*, 771–778. [CrossRef]
- 28. Motrescu, E.R.; Rio, M.C. Cancer cells, adipocytes and matrix metalloproteinase 11: A vicious tumor progression cycle. *Biol. Chem.* **2008**, *389*, 1037–1041. [CrossRef]

- 29. Johnson, L.D.; Hunt, D.M.; Kim, K.; Nachtigal, M. Amplification of stromelysin-3 transcripts from carcinomas of the colon. *Hum. Pathol.* **1996**, 27, 964–968. [CrossRef]
- 30. Pang, L.; Wang, D.W.; Zhang, N.; Xu, D.H.; Meng, X.W. Elevated serum levels of MMP-11 correlate with poor prognosis in colon cancer patients. *Cancer Biomark.* 2016, 16, 599–607. [CrossRef]
- 31. Morini, S.R.; Denadai, M.V.; Waisberg, J.; Lopes Filho, G.J.; Matos, D.; Saad, S.S. Metalloproteinases and colorectal cancer. Correlation of gene expression and clinical-pathological parameters. *Acta Cir. Bras.* **2020**, *35*, e202000707. [CrossRef]
- 32. Koberle, B.; Koch, B.; Fischer, B.M.; Hartwig, A. Single nucleotide polymorphisms in DNA repair genes and putative cancer risk. *Arch. Toxicol.* **2016**, *90*, 2369–2388. [CrossRef]
- 33. International HapMap, C. The International HapMap Project. Nature 2003, 426, 789–796. [CrossRef] [PubMed]
- Lin, C.W.; Yang, S.F.; Chuang, C.Y.; Lin, H.P.; Hsin, C.H. Association of matrix metalloproteinase-11 polymorphisms with susceptibility and clinicopathologic characteristics for oral squamous cell carcinoma. *Head Neck* 2015, 37, 1425–1431. [CrossRef] [PubMed]
- Wang, B.; Hsu, C.J.; Lee, H.L.; Chou, C.H.; Su, C.M.; Yang, S.F.; Tang, C.H. Impact of matrix metalloproteinase-11 gene polymorphisms upon the development and progression of hepatocellular carcinoma. *Int. J. Med. Sci.* 2018, 15, 653–658. [CrossRef] [PubMed]
- Saad, H.; Zahran, M.A.; Hendy, O.; Abdel-Samiee, M.; Bedair, H.M.; Abdelsameea, E. Matrix Metalloproteinase-11 Gene Polymorphisms as a Risk for Hepatocellular Carcinoma Development in Egyptian Patients. *Asian Pac. J. Cancer Prev.* 2020, 21, 3725–3734. [CrossRef] [PubMed]
- Hsieh, C.Y.; Chou, Y.E.; Lin, C.Y.; Wang, S.S.; Chien, M.H.; Tang, C.H.; Lin, J.C.; Wen, Y.C.; Yang, S.F. Impact of Matrix Metalloproteinase-11 Gene Polymorphisms on Biochemical Recurrence and Clinicopathological Characteristics of Prostate Cancer. *Int. J. Environ. Res. Public Health* 2020, 17, 8603. [CrossRef]
- Li, C.C.; Hsieh, M.J.; Wang, S.S.; Hung, S.C.; Lin, C.Y.; Kuo, C.W.; Yang, S.F.; Chou, Y.E. Impact of Matrix Metalloproteinases 11 Gene Variants on Urothelial Cell Carcinoma Development and Clinical Characteristics. *Int. J. Environ. Res. Public Health* 2020, 17, 475. [CrossRef]
- 39. Edge, S.B.; Compton, C.C. The American Joint Committee on Cancer: The 7th edition of the AJCC cancer staging manual and the future of TNM. *Ann. Surg. Oncol.* 2010, *17*, 1471–1474. [CrossRef]
- 40. Chung, T.T.; Pan, M.S.; Kuo, C.L.; Wong, R.H.; Lin, C.W.; Chen, M.K.; Yang, S.F. Impact of RECK gene polymorphisms and environmental factors on oral cancer susceptibility and clinicopathologic characteristics in Taiwan. *Carcinogenesis* **2011**, *32*, 1063–1068. [CrossRef]
- Hsiao, P.C.; Chen, M.K.; Su, S.C.; Ueng, K.C.; Chen, Y.C.; Hsieh, Y.H.; Liu, Y.F.; Tsai, H.T.; Yang, S.F. Hypoxia inducible factor-1alpha gene polymorphism G1790A and its interaction with tobacco and alcohol consumptions increase susceptibility to hepatocellular carcinoma. *J. Surg. Oncol.* 2010, *102*, 163–169. [CrossRef] [PubMed]
- 42. The Lancet Oncology. Colorectal cancer: A disease of the young? Lancet Oncol. 2017, 18, 413. [CrossRef]
- 43. Patel, S.G.; Ahnen, D.J. Colorectal Cancer in the Young. Curr. Gastroenterol. Rep. 2018, 20, 15. [CrossRef] [PubMed]
- 44. Mauri, G.; Sartore-Bianchi, A.; Russo, A.G.; Marsoni, S.; Bardelli, A.; Siena, S. Early-onset colorectal cancer in young individuals. *Mol. Oncol.* **2019**, *13*, 109–131. [CrossRef]
- 45. Patel, S.G.; Murphy, C.C.; Lieu, C.H.; Hampel, H. Early age onset colorectal cancer. Adv. Cancer Res. 2021, 151, 1–37. [CrossRef]
- 46. Siegel, R.; DeSantis, C.; Virgo, K.; Stein, K.; Mariotto, A.; Smith, T.; Cooper, D.; Gansler, T.; Lerro, C.; Fedewa, S.; et al. Cancer treatment and survivorship statistics, 2012. *CA Cancer J. Clin.* **2012**, *62*, 220–241. [CrossRef]
- Baxter, N.N.; Kennedy, E.B.; Bergsland, E.; Berlin, J.; George, T.J.; Gill, S.; Gold, P.J.; Hantel, A.; Jones, L.; Lieu, C.; et al. Adjuvant Therapy for Stage II Colon Cancer: ASCO Guideline Update. J. Clin. Oncol. Off. J. Am. Soc. Clin. Oncol. 2022, 40, 892–910. [CrossRef]
- 48. Leijssen, L.G.J.; Dinaux, A.M.; Taylor, M.S.; Deshpande, V.; Kunitake, H.; Bordeianou, L.G.; Berger, D.L. Perineural Invasion Is a Prognostic but not a Predictive Factor in Nonmetastatic Colon Cancer. *Dis. Colon Rectum* **2019**, *62*, 1212–1221. [CrossRef]
- Skancke, M.; Arnott, S.M.; Amdur, R.L.; Siegel, R.S.; Obias, V.J.; Umapathi, B.A. Lymphovascular Invasion and Perineural Invasion Negatively Impact Overall Survival for Stage II Adenocarcinoma of the Colon. *Dis. Colon Rectum* 2019, 62, 181–188. [CrossRef]
- 50. Hansen, I.O.; Jess, P. Possible better long-term survival in left versus right-sided colon cancer-a systematic review. *Dan. Med. J.* **2012**, *59*, A4444.
- 51. Pal, S.K.; Hurria, A. Impact of age, sex, and comorbidity on cancer therapy and disease progression. *J. Clin. Oncol.* **2010**, *28*, 4086–4093. [CrossRef] [PubMed]
- Ng, S.C.; Wang, P.H.; Lee, Y.C.; Lee, C.Y.; Yang, S.F.; Shen, H.P.; Hsiao, Y.H. Impact of matrix metalloproteinase-11 gene polymorphisms on development and clinicopathologcial variables of uterine cervical cancer in Taiwanese women. *Int. J. Med. Sci.* 2019, *16*, 774–782. [CrossRef] [PubMed]
- Hases, L.; Ibrahim, A.; Chen, X.; Liu, Y.; Hartman, J.; Williams, C. The Importance of Sex in the Discovery of Colorectal Cancer Prognostic Biomarkers. *Int. J. Mol. Sci.* 2021, 22, 1354. [CrossRef] [PubMed]
- 54. Hang, D.; Shen, H. Sex Hormone and Colorectal Cancer: The Knowns and Unknowns. *Cancer Epidemiol. Biomark. Prev.* 2021, 30, 1302–1304. [CrossRef]

- 55. Hang, D.; He, X.; Kvaerner, A.S.; Chan, A.T.; Wu, K.; Ogino, S.; Hu, Z.; Shen, H.; Giovannucci, E.L.; Song, M. Plasma sex hormones and risk of conventional and serrated precursors of colorectal cancer in postmenopausal women. *BMC Med.* **2021**, *19*, 18. [CrossRef]
- Yang, W.; Giovannucci, E.L.; Hankinson, S.E.; Chan, A.T.; Ma, Y.; Wu, K.; Fuchs, C.S.; Lee, I.M.; Sesso, H.D.; Lin, J.H.; et al. Endogenous sex hormones and colorectal cancer survival among men and women. *Int. J. Cancer* 2020, 147, 920–930. [CrossRef]
- 57. Lin, J.H.; Zhang, S.M.; Rexrode, K.M.; Manson, J.E.; Chan, A.T.; Wu, K.; Tworoger, S.S.; Hankinson, S.E.; Fuchs, C.; Gaziano, J.M.; et al. Association between sex hormones and colorectal cancer risk in men and women. *Clin. Gastroenterol. Hepatol.* **2013**, *11*, 419–424.e1. [CrossRef]
- 58. Bouras, E.; Papandreou, C.; Tzoulaki, I.; Tsilidis, K.K. Endogenous sex steroid hormones and colorectal cancer risk: A systematic review and meta-analysis. *Discov. Oncol.* **2021**, *12*, 8. [CrossRef]
- Mountain, D.J.; Freeman, B.M.; Kirkpatrick, S.S.; Beddies, J.W.; Arnold, J.D.; Freeman, M.B.; Goldman, M.H.; Stevens, S.L.; Klein, F.A.; Grandas, O.H. Androgens regulate MMPs and the cellular processes of intimal hyperplasia. *J. Surg. Res.* 2013, 184, 619–627. [CrossRef]
- 60. Morales-Vasquez, F.; Castillo-Sanchez, R.; Gomora, M.J.; Almaraz, M.A.; Pedernera, E.; Perez-Montiel, D.; Rendon, E.; Lopez-Basave, H.N.; Roman-Basaure, E.; Cuevas-Covarrubias, S.; et al. Expression of metalloproteinases MMP-2 and MMP-9 is associated to the presence of androgen receptor in epithelial ovarian tumors. *J. Ovarian Res.* **2020**, *13*, 86. [CrossRef]
- Eiro, N.; Fernandez-Gomez, J.; Sacristan, R.; Fernandez-Garcia, B.; Lobo, B.; Gonzalez-Suarez, J.; Quintas, A.; Escaf, S.; Vizoso, F.J. Stromal factors involved in human prostate cancer development, progression and castration resistance. *J. Cancer Res. Clin. Oncol.* 2017, 143, 351–359. [CrossRef] [PubMed]
- 62. Horstman, A.M.; Dillon, E.L.; Urban, R.J.; Sheffield-Moore, M. The role of androgens and estrogens on healthy aging and longevity. *J. Gerontol. Ser. A Biol. Sci. Med. Sci.* **2012**, *67*, 1140–1152. [CrossRef] [PubMed]
- 63. Goodman-Gruen, D.; Barrett-Connor, E. Sex differences in the association of endogenous sex hormone levels and glucose tolerance status in older men and women. *Diabetes Care* 2000, 23, 912–918. [CrossRef] [PubMed]
- 64. Zumoff, B.; Strain, G.W.; Miller, L.K.; Rosner, W. Twenty-four-hour mean plasma testosterone concentration declines with age in normal premenopausal women. *J. Clin. Endocrinol. Metab.* **1995**, *80*, 1429–1430. [CrossRef] [PubMed]
- Koga, Y.; Yasunaga, M.; Moriya, Y.; Akasu, T.; Fujita, S.; Yamamoto, S.; Kozu, T.; Baba, H.; Matsumura, Y. Detection of colorectal cancer cells from feces using quantitative real-time RT-PCR for colorectal cancer diagnosis. *Cancer Sci.* 2008, 99, 1977–1983. [CrossRef]
- Lech, G.; Slotwinski, R.; Slodkowski, M.; Krasnodebski, I.W. Colorectal cancer tumour markers and biomarkers: Recent therapeutic advances. World J. Gastroenterol. 2016, 22, 1745–1755. [CrossRef]
- 67. Yang, B.; Gao, J.; Rao, Z.; Shen, Q. Clinicopathological and prognostic significance of α5β1-integrin and MMP-14 expressions in colorectal cancer. *Neoplasma* **2013**, *60*, 254–261. [CrossRef]
- 68. Chen, Z.; Wu, G.; Ye, F.; Chen, G.; Fan, Q.; Dong, H.; Zhu, X.; Wu, C. High expression of MMP19 is associated with poor prognosis in patients with colorectal cancer. *BMC Cancer* **2019**, *19*, 448. [CrossRef]
- 69. Nimri, L.; Barak, H.; Graeve, L.; Schwartz, B. Restoration of caveolin-1 expression suppresses growth, membrane-type-4 metalloproteinase expression and metastasis-associated activities in colon cancer cells. *Mol. Carcinog.* **2013**, *52*, 859–870. [CrossRef]