Brief Report

Others

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A Potential Issue with Screening Prediabetes or Diabetes Using Serum Glucose: A Delay in Diagnosis

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The aim of this study was to compare the fasting serum glucose level with the fasting plasma glucose level for diagnosing hyperglycemic states in real-life clinical situations. Additionally, we investigated a usual delay in sample processing and how such delays can impact the diagnosis of hyperglycemic states. Among 1,254 participants who had normoglycemia or impaired fasting glucose (IFG) assessed by the fasting serum glucose level, 20.9% were newly diagnosed with diabetes based on the plasma fasting glucose level. Of the participants with normoglycemia, 62.1% and 14.2% were newly diagnosed with IFG and diabetes, respectively, according to the plasma fasting glucose level. In our clinical laboratory for performing health examinations, the time delay from blood sampling to glycemic testing averaged 78 ± 52 minutes. These findings show that the ordinary time delay for sample processing of the serum glucose for screening hyperglycemic states may be an important reason for these diagnoses to be underestimated in Korea.

Keywords: Diagnosing diabetes; Plasma glucose; Serum glucose

INTRODUCTION

Fasting plasma glucose (FPG) should be measured for the screening or diagnosis of impaired fasting glucose and diabetes (hyperglycemic states) [1,2]. The standard guidelines for blood sample handling indicate that plasma or serum must be separated from cells as soon as possible and within a maximum of 2 hours [3]. Therefore, the serum glucose concentrations are often measured with automatic analyzers in health examinations, and these are used as a screening test for glycemic states [4]. This may be one of the main reasons for the diagnosis of hyperglycemic states to be underestimated in Korea.

It is well known that the glucose concentrations decrease over time in whole blood *ex vivo* as a result of glycolysis [5]. However, no recent reports have addressed the level of the difference between the fasting serum glucose and FPG in Korea. The purpose of this study was to compare the fasting serum glucose level with the FPG for diagnosing hyperglycemic states in real-life clinical situations. We also evaluated an ordinary time delay in sample processing using our clinical laboratory tests to assess health examinations at the National Health Insurance Corp. for a period of a month to determine how time delays impact the diagnoses of hyperglycemic states.

METHODS

We recruited 1,254 participants who were diagnosed with normoglycemia or IFG using the serum blood glucose level; these were from 2,028 participants enrolled in Hallym University Sacred Heart Hospital and Kangbuk Samsung Hospital for

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evaluating metabolic abnormalities, including IFG or dyslipidemia according to health examinations performed by the National Health Insurance Corp. or by personal medical checkup between 2006 and 2009.

Fasting blood samples were collected from all 1,254 participants at our centers in the morning after a 12-hour overnight fast. In two consecutive samples, 3 mL of blood was drawn and placed in common tubes for the glucose measurement. Serum samples were commonly assembled at room temperature and centrifuged within 1 hour. The fasting serum glucose level was measured within 2 hours with an automated analyzer using the hexokinase/glucose-6-phosphate dehydrogenase method (Modular DPE; Hitachi Hitechnologies, Tokyo, Japan). The

Table 1. Baseline characteristics of participants with normal glucose tolerance or impaired fasting glucose (n=1,254)

Characteristic	Mean±SD	Range
Male sex, %	71.9	
Age, yr	48.1±9.6	18-88
Height, cm	167.1 ± 8.1	143-185
Weight, kg	69.7±10.6	34-103.2
Body mass index, kg/m ²	24.9 ± 2.8	17.1-34.9
Waist, cm	84.2 ± 8.1	58-117
Systolic blood pressure, mm Hg	120.3 ± 15.5	80-178
Diastolic blood pressure, mm Hg	77.6±9.2	54-110
Serum glucose, mg/dL	108.5 ± 6.5	100-125
FPS, mg/dL ^a	119.4 ± 9.9	100-183
PP2, mg/dL	156.9 ± 45.7	63-388
Glycosylated hemoglobin	5.8 ± 0.5	4.4-8.8
Fasting insulin	9.6±3.9	1.4-33.2
Fasting C-peptide	3.1±1.5	0.7-28.2
Total cholesterol, mg/dL	201.1 ± 36.6	111-417
Triglyceride, mg/dL	165.5 ± 113.7	33-1,235
HDL-C, mg/dL	47.6 ± 10.8	28-100
LDL-C, mg/dL	120.7 ± 32.1	33-289
ALT, mg/dL	28.6±19.7	4-293
AST, mg/dL	25.6 ± 12.0	9–239
γGT, mg/dL	45.2±68.9	3-1,744

FPS, fasting plasma glucose; PP2, 2-hour postprandial serum glucose; HDL-C, high density lipoprotein cholesterol; LDL-C, low density lipoprotein cholesterol; ALT, alanine transaminase; AST, aspartate transaminase; γ GT, γ -glutamyl transpeptidase.

^aFPS indicates the fasting plasma glucose or serum glucose measured by immediate sample separation.

plasma samples were immediately centrifuged and the FPG was measured with an enzymatic assay (BIOSEN C-line; EKF Diagnostics GmbH, Berlin, Germany) within 15 minutes. We also investigated a time delay from blood sampling to glycemic testing using the order communication system (RefoMax, Seoul, Korea) at Hallym University Sacred Heart Hospital as part of health examinations performed by the National Health Insurance Corp. for a period of month because we could not directly measure the real serum-clot contact time for clinical chemistry laboratory results. We conducted a cross-sectional study, which was approved by the Institutional Review Board of Hallym University Sacred Heart Hospital.

RESULTS

A total of 2,028 participants were initially included in this study. Of these, 717 with newly diagnosed diabetes and 57 with no biochemical data were excluded; as a result, 1,254 subjects were finally included in this analysis.

The characteristics of the study population are presented in Table 1. Their mean glucose concentrations were 119.4 ± 9.9 mg/dL for plasma and 108.5 ± 6.5 mg/dL for serum (mean difference, 10.9 ± 7.4 mg/dL). Overall, 1,254 subjects were diagnosed with normoglycemia (n=169, 13.5%) or IFG (n=1,085, 86.5%) through measuring the fasting serum glucose at our centers. In this study population (n=1,254), 20.9% had newly diagnosed diabetes according to their FPG levels. Of the participants with normoglycemia (n=169), 105 (62.1%) and 24 (14.2%) were newly diagnosed with IFG and diabetes, respectively, based on their FPG levels (Table 2).

The time delay from blood sampling to glycemic testing at our clinical laboratory for health examinations of the National Health Insurance Corp. averaged 78 ± 52 minutes for 729 subjects for the fasting serum glucose level of the 844 subjects who

Table 2. Plasma glycemic status in subjects with impaired fasting glucose or normoglycemia diagnosed with serum

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Serum (<i>n</i> =1,254)	Plasma
Normoglycemia	Normoglycemia (23.7%, <i>n</i> =40)
(13.5%, <i>n</i> =169)	Impaired fasting glucose (62.1%, <i>n</i> =105)
	Diabetes mellitus (14.2%, $n=24$)
Impaired fasting glucose (86.5%, <i>n</i> =1,085)	Impaired fasting glucose (77.9%, <i>n</i> =846)
	Diabetes mellitus (22.1%, $n=239$)

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were examined over a period of 1 month.

DISCUSSION

The results of this study showed that the serum glucose measurement and ordinary time delay in the sample processing before getting the result of that measurement may underestimate the diagnosis of hyperglycemic states in real-life clinical situations. Decreases in the glucose concentrations in whole blood ex vivo have been reported to average 5% to 7% (10 mg/ dL) per hour [5]. We performed a preliminary study of the serum-clot contact effect in 33 tests. The blood samples were kept at room temperature before serum-clot separation, and glucose concentrations were measured for 3 hours at 30-minute intervals. The serum glucose level with each processing delay of 30 minutes decreased by an average of 7.4±2.2 mg/dL per hour, and the largest decrease (4.2±1.1 mg/dL) occurred in the first 30 minutes. This decrease in the glucose concentration may result in the missed diagnosis of hyperglycemic states for a large proportion of the population with glucose levels near the cutoffs for these diagnoses [6]. The importance of early diagnosis for diabetes, which facilitates prevention of diabetic complications, has been emphasized in Korea because its rate of diabetes-related deaths is highest among the Organization for Economic Co-operation and Development (OECD) nations even though significant progress has been made in reducing the cardiovascular risk factors of Korean patients with diagnosed diabetes [7].

Several studies have demonstrated that the glucose concentrations are slightly higher in plasma than in serum [8-10], while others have found no significant difference between these levels [11,12]. However, these results do not seem to reflect the real clinical settings because the studies were conducted according to study protocols. We have confirmed that time delays (average of 78±52 minutes) occurred in the process of measuring the serum glucose level at our clinical laboratory for National Health Examinations, and this delay is considered to be one of the important reasons for the diagnosis of hyperglycemic states to be underestimated in Korea. A previous study from the Korea National Health and Nutrition Examination Survey also showed that serum separation and refrigeration within 30 minutes of venous sampling is recommended over the NaF (non-adjacent form) method to minimize the pre-analytical impact on diabetes detection [13,14].

There are several limitations in our study. First, because all of

our recruited subjects had visited hospitals, there may have been a selection bias toward higher risk factors for diabetes. Second, our cross-sectional study results may be somewhat limited due to the lack of prospective follow-up data, limiting the clinical implications of our findings. Despite these limitations, the present study, to the best of our knowledge, is the first report to show the direct comparison between the fasting serum glucose and FPG levels measured at approximately the same time when diagnosing hyperglycemic states. Glucose concentrations should be measured in plasma or serum immediately after sample separation (or within a maximum of 30 minutes) for early diagnosis and timely intervention in hyperglycemic states. Further prospective studies are also necessary to compare the fasting serum glucose level with the FPG to diagnose hyperglycemic states and evaluate the ordinary delay in sample processing in the Korean population.

CONFLICTS OF INTEREST

No potential conflict of interest relevant to this article was reported.

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REFERENCES

- 1. Sacks DB, Arnold M, Bakris GL, Bruns DE, Horvath AR, Kirkman MS, Lernmark A, Metzger BE, Nathan DM; National Academy of Clinical Biochemistry. Position statement executive summary: guidelines and recommendations for laboratory analysis in the diagnosis and management of diabetes mellitus. Diabetes Care 2011;34:1419-23.
- Ko SH, Kim SR, Kim DJ, Oh SJ, Lee HJ, Shim KH, Woo MH, Kim JY, Kim NH, Kim JT, Kim CH, Kim HJ, Jeong IK, Hong EK, Cho JH, Mok JO, Yoon KH; Committee of Clinical Practice Guidelines, Korean Diabetes Association. 2011 Clinical practice guidelines for type 2 diabetes in Korea. Diabetes Metab J 2011;35:431-6.
- Tietz NW, Burtis CA, Ashwood ER, Brund DE. Tietz textbook of clinical chemistry and molecular diagnostics. 4th ed. Philadelphia: Elsevier Saunders; 2006. Chapter 25, Carbohydrate; p837-901.

- Jee SH, Suh I, Kim IS, Appel LJ. Smoking and atherosclerotic cardiovascular disease in men with low levels of serum cholesterol: the Korea Medical Insurance Corporation Study. JAMA 1999;282:2149-55.
- Chan AY, Swaminathan R, Cockram CS. Effectiveness of sodium fluoride as a preservative of glucose in blood. Clin Chem 1989;35:315-7.
- Sacks DB, Arnold M, Bakris GL, Bruns DE, Horvath AR, Kirkman MS, Lernmark A, Metzger BE, Nathan DM. Guidelines and recommendations for laboratory analysis in the diagnosis and management of diabetes mellitus. Clin Chem 2011;57:e1-47.
- Yu SH, Kang JG, Hwang YC, Ahn KJ, Yoo HJ, Ahn HY, Park SW, Park CY. Increasing achievement of the target goals for glycemic, blood pressure and lipid control for adults with diagnosed diabetes in Korea. J Diabetes Investig 2013;4:460-5.
- Gambino R, Piscitelli J, Ackattupathil TA, Theriault JL, Andrin RD, Sanfilippo ML, Etienne M. Acidification of blood is superior to sodium fluoride alone as an inhibitor of glycolysis. Clin Chem 2009;55:1019-21.
- 9. Stahl M, Jorgensen LG, Hyltoft Petersen P, Brandslund I, de Fine Olivarius N, Borch-Johnsen K. Optimization of preana-

lytical conditions and analysis of plasma glucose. 1. Impact of the new WHO and ADA recommendations on diagnosis of diabetes mellitus. Scand J Clin Lab Invest 2001;61:169-79.

- Carstensen B, Lindstrom J, Sundvall J, Borch-Johnsen K, Tuomilehto J; DPS Study Group. Measurement of blood glucose: comparison between different types of specimens. Ann Clin Biochem 2008;45(Pt 2):140-8.
- 11. Boyanton BL Jr, Blick KE. Stability studies of twenty-four analytes in human plasma and serum. Clin Chem 2002;48:2242-7.
- 12. Miles RR, Roberts RF, Putnam AR, Roberts WL. Comparison of serum and heparinized plasma samples for measurement of chemistry analytes. Clin Chem 2004;50:1704-6.
- 13. Lee YW, Cha YJ, Chae SL, Song J, Yun YM, Park HI, Seong MW, Whang DH, Kim HS, Kim JH, Lee BS, Hwang YS. Effectiveness of sodium fluoride as a glycolysis inhibitor on blood glucose measurement: comparison of blood glucose using specimens from the Korea National Health and Nutrition Examination Survey. Korean J Lab Med 2009;29:524-8.
- Kweon S, Kim Y, Jang MJ, Kim Y, Kim K, Choi S, Chun C, Khang YH, Oh K. Data resource profile: the Korea National Health and Nutrition Examination Survey (KNHANES). Int J Epidemiol 2014;43:69-77.