REVIEW Open Access



Colony-stimulating factor 1 receptor (CSF1R) inhibitors in cancer therapy

Michael A. Cannarile*†, Martin Weisser†, Wolfgang Jacob, Anna-Maria Jegg, Carola H. Ries and Dominik Rüttinger

Abstract

The tumor-permissive and immunosuppressive characteristics of tumor-associated macrophages (TAM) have fueled interest in therapeutically targeting these cells. In this context, the colony-stimulating factor 1 (CSF1)/ colony-stimulating factor 1 receptor (CSF1R) axis has gained the most attention, and various approaches targeting either the ligands or the receptor are currently in clinical development. Emerging data on the tolerability of CSF1/ CSF1R-targeting agents suggest a favorable safety profile, making them attractive combination partners for both standard treatment modalities and immunotherapeutic agents. The specificity of these agents and their potent blocking activity has been substantiated by impressive response rates in diffuse-type tenosynovial giant cell tumors, a benign connective tissue disorder driven by CSF1 in an autocrine fashion. In the malignant disease setting, data on the clinical activity of immunotherapy combinations with CSF1/CSF1R-targeting agents are pending. As our knowledge of macrophage biology expands, it becomes apparent that the complex phenotypic and functional properties of macrophages are heavily influenced by a continuum of survival, differentiation, recruitment, and polarization signals within their specific tissue environment. Thus, the role of macrophages in regulating tumorigenesis and the impact of depleting and/or reprogramming TAM as therapeutic approaches for cancer patients may vary greatly depending on organ-specific characteristics of these cells. We review the currently available clinical safety and efficacy data with CSF1/CSF1R-targeting agents and provide a comprehensive overview of ongoing clinical studies. Furthermore, we discuss the local tissue macrophage and tumor-type specificities and their potential impact on CSF1/CSF1R-targeting treatment strategies for the future.

Keywords: CSF1, CSF1R, Tumor-associated macrophage, Clinical trial, Cancer therapy, PVNS, Dt-GCT

Background

Macrophages are known to be a highly plastic cell type that adapts to the particular stromal environment present in malignant tumors, characterized by tissue necrosis, low oxygen pressure, and high concentrations of lactate and pyruvate [1]. Macrophages have been described as responding to this micromilieu with either a pro-inflammatory or an anti-inflammatory phenotype (also referred to as "fight" versus "fix" macrophages, respectively) [2]. In early stage as well as metastatic cancer, the dominant tumor-associated macrophage (TAM) phenotype is reported to be anti-inflammatory, immune-regulatory, and therefore tumor-promoting (also termed alternatively activated or M2 macrophages) as opposed to pro-inflammatory and

tumoricidal (classically activated or M1 macrophages). We and others believe that the continuum of different macrophage phenotypes present within the tumor microenvironment (TME) is difficult to capture solely with the M1/M2 dichotomy. However, for simplicity reasons, we use the term M1 or M2 macrophage/TAM to differentiate two extreme functional phenotypes in this review. M2 macrophages/TAM have been reported to promote tumor growth, angiogenesis, invasion, and metastasis as well as resistance to therapy [3, 4]. In addition, TAM infiltration has been shown to have a negative prognostic relevance in most tumor types [5]. This phenotype is a consequence of the continuous presence of growth factors such as colony-stimulating factor-1 (CSF1; or macrophage colony-stimulating factor [MCSF]) as well as the cluster of differentiation (CD)-4⁺ type 2 helper T-cell-derived (T_h2) cytokines interleukin (IL)-4, IL-13, and IL-10 in the TME. In

Roche Pharmaceutical Research and Early Development, Roche Innovation Center Munich, Nonnenwald 2, Penzberg 82377, Germany



^{*} Correspondence: Michael.cannarile@roche.com

[†]Equal contributors

contrast, M1 macrophages are ascribed tumoricidal functions and are generated in the presence of granulocytemacrophage colony-stimulating factor (GM-CSF or CSF2) and pro-inflammatory stimuli such as interferon (IFN)- γ , lipopolysaccharide, or tumor necrosis factor α [6] (Fig. 1).

CSF1 receptor (CSF1R)-mediated signaling is crucial for the differentiation and survival of the mononuclear phagocyte system and macrophages in particular [7]. CSF1R belongs to the type III protein tyrosine kinase receptor family, and binding of CSF1 or the more recently identified ligand, IL-34, induces homodimerization of the receptor and subsequent activation of receptor signaling [8]. As the intratumoral presence of CSF1R⁺ macrophages correlates with poor survival in various tumor types [5, 9], targeting CSF1R signaling in tumor-promoting TAM represents an attractive strategy to eliminate or repolarize these cells.

In addition to TAM, CSF1R expression can be detected on other myeloid cells within the tumor microenvironment such as dendritic cells, neutrophils, and myeloid-derived suppressor cells (MDSCs)

For the latter, Holmgaard and colleagues provided evidence for MDSC reprogramming towards a proinflammatory, tumoricidal phenotype upon treatment with a CSF1R

small-molecule inhibitor, PLX3397 [10]. However, a clear interpretation of the role of MDSCs in inflammatory responses remains challenging because of the phenotypic, morphological, and functional heterogeneity of these cells in mice and humans [11]. As our understanding of the influence of CSF1/CSF1R-mediated signaling on human myeloid-derived cells other than macrophages is just emerging, the focus of this review is on TAM and current clinical efforts to specifically target CSF1/CSF1R in cancer therapy. We also highlight the importance of site/organ and tumor-type specificities of TAM, which are now recognized as an important new frontier in cancer immunotherapy. Early clinical data suggest good tolerability of CSF1/CSF1R-targeting compounds; however, available efficacy data are still limited, with the exception of compelling anti-tumor activity observed in diffuse-type tenosynovial giant cell tumors (dt-GCT), a benign connective tissue disorder driven by CSF1 in an autocrine fashion [12]. The individual CSF1R inhibitors and their different drug-targeting properties have recently been reviewed [13]. Only two clinical-stage programs are currently targeting CSF1. No molecules targeting IL-34, the secondknown ligand for CSF1R, are in clinical development thus far. We use "CSF1R inhibitor" as a general term for both receptor- and ligand-targeting compounds.

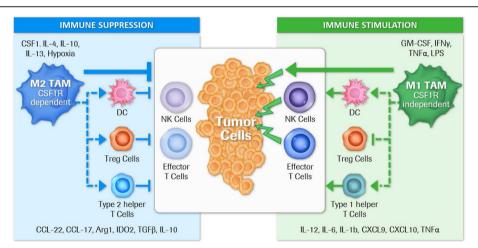


Fig. 1 Direct and indirect regulation of immune suppression or stimulation by tumor associated macrophage subtypes. Macrophage polarization within the tumor microenvironment is highly dependent on the local cytokine milieu which originates either from tumor cells, other stromal cells such as immune cells or fibroblasts, as well as macrophages themselves. The M2 TAM phenotype is a consequence of the continuous presence of growth factors such as colony-stimulating factor-1 (CSF1) as well as CD4+ T cell-derived Th2 cytokines interleukin (IL)-4, IL-13 and IL-10 (5). Besides the direct tumor growth promoting abilities of M2 TAM (not illustrated here), these macrophages efficiently suppress immune effector functions that are able to contribute to tumor cell elimination (3,4). This silencing of immune effector cells is achieved by producing cytokines and enzymes that may directly suppress effector cells or indirectly via other immune cell types such as intratumoral dendritic cells (DC), T regulatory cells (Treg cells) and Type 2 helper T cells. In contrast, M1 TAM are attributed with tumoricidal functions and are generated in the presence of GM-CSF and pro-inflammatory stimuli like IFNy, LPS or TNFα (5). Tumoricidal function can either be achieved through direct killing of tumor cells or by producing cytokines/chemokines that are activating/recruiting other immune stimulatory immune cells and inhibiting immune suppressive cells like Treg cells. Eventually a predominant M1 TAM phenotype may result in an anti-tumor immune effector cell activation. Published data suggest that tumor promoting and immune suppressive M2 macrophages/TAM are dependent on CSF1R mediated signals (31) making this receptor an attractive target to eliminate or repolarize these cells

Clinical activity with CSF1R inhibitor monotherapy

A variety of small molecules and monoclonal antibodies (mAbs) directed at CSF1R or its ligand CSF1 are in clinical development both as monotherapy and in combination with standard treatment modalities such as chemotherapy as well as other cancer-immunotherapy approaches (Tables 1–3).

Among the class of small molecules, pexidartinib (PLX3397), an oral tyrosine kinase inhibitor of CSF1R, cKIT, mutant fms-like tyrosine kinase 3 (FLT3), and platelet-derived growth factor receptor

(PDGFR)-β, is the subject of the broadest clinical development program in monotherapy, with completed or ongoing studies in c-kit-mutated melanoma, prostate cancer, glioblastoma (GBM), classical Hodgkin lymphoma (cHL), neurofibroma, sarcoma, and leukemias. Additional CSF1R-targeting small molecules, including ARRY-382, PLX7486, BLZ945, and JNJ-40346527, are currently being investigated in solid tumors and cHL. mAbs in clinical development include emactuzumab, AMG820, IMC-CS4, cabiralizumab, MCS110, and PD-0360324, with the latter two being the only compounds targeting the ligand CSF1.

Table 1 CSF1/CSF1R inhibitors as monotherapy in current clinical development

Class	Target	Compound	Clinical Phase	Sponsor	Indication	ClinicalTrials.gov identifier	Status/Results	Reference
Small molecules	CSF1R (and cKIT, Flt3)	Pexidartinib (PLX3397,	2	The Christie NHS Foundation Trust	KIT-mutated advanced acral and mucosal melanoma	NCT02071940	Ongoing	-
		PLX108-01)	1/2	Plexxikon	Unresectable or metastatic KIT-mutated melanoma	NCT02975700	Ongoing	-
			2	Plexxikon	Advanced castration-resistant prostate cancer with bone metastasis and high circulating tumor cell counts	NCT01499043	Not yet reported	-
			2	Plexxikon	Recurrent GBM	NCT01349036	ORR: 0% CBR: 7/38 (18%)	[14, 26]
			1/2	NCI	Refractory leukemias and refractory solid tumors, including neurofibromatosis type 1-associated plexiform neurofibromas	NCT02390752	Ongoing	-
			2	Plexxikon	Relapsed or refractory cHL	NCT01217229	ORR: 1/20 (5%)	[15]
			1/2	Plexxikon	Relapsed or refractory FLT3-ITD-positive acute myeloid leukemia	NCT01349049	Ongoing	-
			1	Plexxikon	Advanced, incurable, solid tumors in which the target kinases are linked to disease pathophysiology	NCT01004861	Ongoing	-
	CSF1R (and Trk)	PLX7486	1	Plexxikon	Solid tumors	NCT01804530	Ongoing	-
	CSF1R	ARRY-382	1	Array BioPharma	Solid tumors	NCT01316822	ORR: 0% CBR: 4/26 (15%)	[17]
	CSF1R	JNJ-40346527	1/2	Johnson& Johnson	cHL	NCT01572519	ORR: 1/21 (5%) CBR: 11/21 (52%)	[16]
	CSF1R	BLZ945	1/2	Novartis	Solid tumors	NCT02829723	Ongoing	-
Monoclonal antibodies	CSF1R	Emactuzumab (RG7155)	1	Roche	Solid tumors	NCT01494688	PMR: 5/44 (11%) ORR: 0% CBR: 6/40 (24%)	[18]
	CSF1R	AMG820	1	Amgen	Solid tumors	NCT01444404	ORR: 1/25 (4%) CBR: 6/25 (24%)	[19]
	CSF1R	IMC-CS4	1	Eli Lilly	Solid tumors	NCT01346358	Ongoing	-
		(LY3022855)	1	Eli Lilly	Breast and prostate cancer	NCT02265536	Ongoing	-
	CSF1	MCS110	1/2	Novartis	Prostate cancer	NCT00757757	Terminated	-

CBR clinical benefit rate, cHL classical Hodgkin lymphoma, CSF1 colony-stimulating factor 1, CSF1R colony-stimulating factor 1 receptor, GBM glioblastoma, NCI National Cancer Institute, NHS National Health Service, ORR objective response rate, PMR partial metabolic response

A phase 2 study in 38 patients with recurrent GBM treated with pexidartinib did not show significant improvement in 6-month progression-free survival (PFS) compared to historical control data. Of 38 patients, seven (18%) experienced stable disease; no partial or complete responses were observed [14]. An objective response rate (ORR) of 5% was reported with single agent PLX3397 in 20 heavily pre-treated patients with cHL [15]. Comparable efficacy in relapsed or refractory cHL was demonstrated with JNJ-40346527 in a phase 1/2 clinical study. Out of 21 patients enrolled, one showed a complete response (ORR 5%) and 11 (52%) experienced stable disease [16].

Results from a phase 1 study investigating ARRY-382 in advanced solid tumors were recently reported by Bendell et al. Out of 26 patients, four (15%) had stable disease, and no objective responses were observed [17]. A phase 1/2 study with BLZ945 in solid tumors is ongoing.

Data from a phase 1 dose-escalation and expansion study investigating emactuzumab showed partial metabolic responses in fluorodeoxyglucose-positron emission tomography in 5/44 (11%) patients and stable disease by Response Evaluation Criteria in Solid Tumors (RECIST) in 6/40 (15%) patients [18]. In addition, the study provided proof of mechanism, demonstrating significant

TAM reduction with emactuzumab in paired pre- and on-treatment tumor biopsies (Fig. 2).

Papadopoulos et al. reported that 6/25 patients (24%) treated with AMG820 had a best overall response of stable disease, and one paraganglioma patient (4%) had a partial response, with a 40% reduction in tumor burden [19].

Results from two ongoing single-agent phase 1 studies of IMC-CS4 in solid tumors and breast and prostate cancer are pending. A phase 1/2 study in prostate cancer of the only anti-CSF1 antibody, MSC110, has been terminated; however, several clinical trials are underway with MSC110 in combination with chemotherapy or immune checkpoint inhibitor therapy (Tables 2 and 3).

Anti-tumor activity of CSF1R inhibitors in diffusetype tenosynovial giant cell tumor (dt-GCT)

dt-GCT of the soft tissue (alternatively known as pigmented villonodular synovitis [PVNS]) is an orphan disease characterized by overexpression of CSF1 and is usually caused by chromosomal translocations involving chromosome 1p13 where the CSF1 gene is located. CSF1R activation leads to the recruitment of CSF1R-expressing macrophages that constitute a large part of the tumor mass in dt-GCT, thus making this pathway an ideal therapeutic target for compounds interfering with the CSF1/

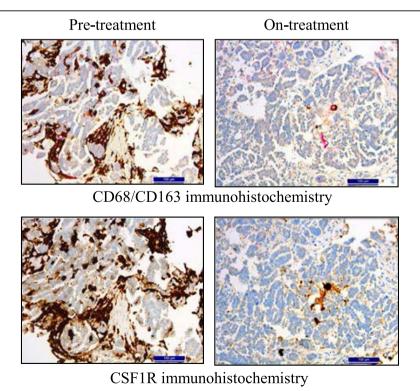


Fig. 2 Depletion of tumor-associated macrophages with emactuzumab in cancer patients. Immunohistochemistry of paired tumor biopsies from a representative ovarian cancer patient illustrating co-localization and reduction of CD68⁺CD163⁺ TAM (upper panel) and CSF1R⁺ cells (lower panel) after 4 weeks/two infusions of emactuzumab at the 1000 mg dose level. Permission for re-use granted by I. Klaman [18]

 Table 2
 Clinical trials with CSF1/CSF1R inhibitors in combination with anti-tumor therapies (excluding cancer-immunotherapy doublets)

Class	Target	Compound	Combination partner	Clinical phase	Clinical Sponsor phase	Indication	ClinicalTrials.gov Status/Results identifier	Status/Results	Reference
Small molecules		CSF1R (and Pexidartinib cKT, Flt3) (PLX3397, PLX108-01)	Androgen-deprivation therapy plus external radiotherapy	-	Plexxikon/Daiichi Sankyo Prostate cancer	Prostate cancer	NCT02472275	Ongoing	
			Paclitaxel	-	Plexxikon/Daiichi Sankyo Solid tumors	Solid tumors	NCT01525602	ORR: 4/23 (17%) CBR: 14/23 (61%)	[35]
			Eribulin	1/2	Plexxikon/Daiichi Sankyo	Breast cancer	NCT01596751	Ongoing	
			Temozolomide plus external radiotherapy	1/2	Plexxikon/Daiichi Sankyo GBM	GBM	NCT01790503	Not yet reported	1
			Vemurafenib	_	Plexxikon/Daiichi Sankyo	BRAF-mutated melanoma	NCT01826448	Terminated	1
			PLX9486 (KIT inhibitor)	1/2	Plexxikon/Daiichi Sankyo GIST	GIST	NCT02401815	Ongoing	ı
			Sirolimus	1/2	Plexxikon/Daiichi Sankyo	Plexxikon/Daiichi Sankyo Advanced sarcomas, MPNST	NCT02584647	Ongoing, not yet [73] reported	[73]
Monoclonal CSF1R	I CSF1R	Emactuzumab (RG7155) Paclitaxel	Paclitaxel	-	Roche	Ovarian and breast cancer	NCT01494688	Not yet reported	1
antibodies	CSF1R	Emactuzumab (RG7155) Paclitaxel	Paclitaxel plus bevacizumab	7	Roche	Ovarian cancer	NCT02923739	Ongoing	ı
	CSF1	MCS110	Carboplatin plus gemcitabine	7	Novartis	TNBC	NCT02435680	Ongoing	1
	CSF1	PD-0360324	Cyclophosphamide	7	Pfizer	Recurrent platinum-resistant epithelial ovarian, peritoneal, or fallopian tube cancer	NCT02948101	Ongoing	1

CBR clinical benefit rate, CSF1 colony-stimulating factor 1, CSF1R colony-stimulating factor 1 receptor, dt-GCT diffuse-type tenosynovial giant cell tumor, GBM glioblastoma, GIST gastrointestinal stromal tumor, MR objective response rate, TNBC triple-negative breast cancer

Table 3 Clinical trials with CSF1/CSF1R inhibitors in combination with cancer immunotherapy agents

Class	Target	Compound/Class	Combination partner	Clinical Sponsor phase	Sponsor	Indication	ClinicaTrials.gov Status/identifier	Status/ Reference Results
Small molecule	CSF1R (and cKT, Flt3)	CSF1R (and Pexidartinib cKIT, Flt3) (PLX3397, PLX108-01)	Pembrolizumab (anti-PD1 mAb)	1/2	Plexxikon/Daiichi Sankyo	Solid tumors, malignant melanoma GIST, NSCLC, ovarian carcinoma, TNBC, SCCHN, UBC, pancreatic cancer, gastric carcinoma, leiomyosarcoma, cholangio carcinoma, CRC (MSS)	NCT02452424	Ongoing [74]
			Durvalumab (anti-PDL1 mAb)	·	Astra Zeneca	Pancreatic carcinoma, CRC	NCT02777710	Ongoing -
	CSF1R	ARRY-382	Pembrolizumab (anti-PD1 mAb)		Array BioPharma	Solid tumors, melanoma, NSCLC	NCT02880371	Ongoing -
	CSF1R	BLZ945	PDR001 (anti-PD1 mAb)	1/2	Novartis	Solid tumors	NCT02829723	Ongoing -
Monoclonal antibody	CSF1R	Emactuzumab (RG7155)	Atezolizumab (anti-PDL1 mAb)	-	Roche	Solid tumors, TNBC, gastric cancer, soft tissue sarcoma, UBC, ovarian cancer, NSCLC, melanoma	NCT02323191	Ongoing -
			RG7876 (CD40 agonist mAb)	-	Roche	TNBC, gastric cancer, mesothelioma, CRC, melanoma, pancreatic cancer	NCT02760797	Ongoing -
	CSF1R	AMG820	Pembrolizumab (anti-PD1 mAb)	·	Amgen	Solid tumors	NCT02713529	Ongoing -
	CSF1R	Cabiralizumab (FPA008)	Nivolumab (anti-PD1 mAb)	-	FivePrime/BMS	Solid tumors, NSCLC, SCCHN, pancreatic cancer, ovarian cancer, RCC, GBM	NCT02526017	Ongoing [75]
	CSF1R	IMC-CS4 (LY3022855)	IMC-CS4 (LY3022855) Durvalumab (anti-PDL1 mAb) or Tremelimumab (anti-CTLA4 mAb)	-	eli Lilly	Solid tumors	NCT02718911	Ongoing [76]
	CSF1	MCS110	PDR001 (anti-PD1 mAb)	1/2	Novartis	Solid tumors, TNBC, pancreatic cancer, melanoma, endometrial cancer	NCT02807844	Ongoing -
	CSF1	PD-0360324	Avelumab (anti-PDL1 mAb)	-	Pfizer	Solid tumors	NCT02554812	Ongoing -

CRC colorectal cancer, CSF1 colony-stimulating factor 1, CSF1R colony-stimulating factor 1 receptor, CTLA4 cytotoxic T-lymphocyte-associated protein 4, GBM glioblastoma, GIST gastrointestinal stromal tumor, mAb monoclonal antibody, MSS microsatellite stable, NSCLC non-small cell lung cancer, PD1 programmed cell death protein 1, PDL1 programmed cell death ligand 1, RCC renal cell carcinoma, SCCHN squamous cell carcinoma of the head and neck, TNBC triple-negative breast cancer, UBC urothelial bladder carcinoma

CSF1R-signaling axis. Unresectable dt-GCT is rarely, if ever, a lethal disease but rather a debilitating chronic illness of high unmet medical need, frequently requiring several surgical procedures.

Initial clinical activity was seen in a dt-GCT patient who was treated with the BCR-ABL tyrosine kinasetargeting agent imatinib (Gleevec®) and achieved a complete response [20]. Thereafter, several phase I studies tested CSF1R-targeting compounds in selected dt-GCT patients as a proof-of-concept disease (Table 4). Substantial clinical activity was observed in the study by Cassier et al. (2015), in which 22 of 28 patients (79%) treated with the CSF1R-targeting mAb emactuzumab achieved a partial response, two patients (7%) had a complete response, three patients (11%) had stable disease, and no patient had disease progression [12]. Another study showed unconfirmed partial responses in four of four patients (100%) treated with the CSF1-targeting mAb MCS110 [21]. With the small-molecule inhibitor pexidartinib, 12 of 23 patients (52%) had a partial response, seven (30%) had stable disease, and one (4%) had progressive disease [22]. Responses in these studies were durable (>1 year for pexidartinib and >1.9 years for emactuzumab), and the median PFS had not been reached at the time of publication [12, 22]. A phase 3 study has started in patients with dt-GCT or giant cell tumor of the tendon sheath (GCT-TS) treated with pexidartinib versus placebo (ClinicalTrials.gov identifier NCT02371369). Although exciting clinical activity with CSF1R inhibition has been confirmed in dt-GCT, the safety profile of CSF1R-targeting compounds needs to be considered carefully in this non-life-threatening disease (see the next section for a detailed safety discussion).

In addition to tumor shrinkage as a measure of efficacy, functional and symptomatic improvement is another important aspect to assessing clinical benefit in patients with dt-GCT. Patient-reported outcome measures could add evidence for superior clinical benefit of CSF1R inhibitors over surgery as the current mainstay of therapy. Test instruments such as the Western Ontario and McMasters Universities Osteoarthritis Index (WOMAC) questionnaire, the Brief Pain Inventory (BPI), the worst pain numeric rating scale (NRS), and the Patient-Reported Outcomes Measurement Information System (PROMIS) physical functioning items were introduced into clinical studies to investigate whether tumor shrinkage correlated with clinical benefit for these patients [12, 23]. Preliminary results from 22 patients treated with pexidartinib showed trends toward improvement in both pain and joint stiffness over time [24]

Clinical safety and tolerability of CSF1R inhibitors

Preliminary safety results from phase 1 and 2 studies have been reported for CSF1R inhibitor monotherapy in a variety of settings, including healthy subjects and patients with rheumatoid arthritis, cHL, or advanced solid tumors. In some studies no dose-limiting toxicities (DLTs) were reported [12, 18, 25], whereas others have observed DLTs defining a maximum tolerated dose (MTD) [17, 19, 22]. Overall, the adverse event (AE) profile of CSF1R inhibitors has been characterized quite extensively for the different compounds. Frequently reported AEs for both small molecules and mAbs include fatigue, elevated liver enzymes, facial and peripheral edema, asthenia, pruritus, rash, nausea/vomiting, headache, dry skin, increased lacrimation, and decreased appetite [12, 17, 18, 22, 26–30]. Increases in creatine kinase, lactate dehydrogenase, aspartate aminotransferase (AST), and alanine transaminase (ALT) were seen across studies [12, 17, 19, 22, 25-27, 30, 31]. Most studies reported that, despite elevations of these enzymes, patients did not experience

Table 4 Clinical trials with CSF1R inhibitors for the treatment of dt-GCT

Class	Target	Compound	Clinical phase	Sponsor	Indication	ClinicalTrials.gov identifier	Status/ Results	Reference
Small molecules	CSF1R (and cKIT, Flt3)	Pexidartinib (PLX3397, PLX108-01)	1	Plexxikon	Solid tumors and extension for MEC, dt-GCT, GIST, ATC, metastatic solid tumors	NCT01004861	Ongoing ORR: 12/23 (52%) CBR: 19/23 (83%)	[22]
			3	Plexxikon/ Daiichi Sankyo	dt-GCT or GCT-TS	NCT02371369	Ongoing	-
Monoclonal antibodies	CSF1R	Emactuzumab (RG7155)	1	Roche	Solid tumors and dt-GCT	NCT01494688	ORR: 24/28 (86%) CBR: 27/28 (96%	[12]
	CSF1R	Cabiralizumab (FPA008)	1/2	FivePrime	dt-GCT	NCT02471716	Ongoing	-
	CSF1	MCS110	2	Novartis	dt-GCT or GCT-TS	NCT01643850	Ongoing ORR: 4/4 (100%)	[21]

ATC anaplastic thyroid cancer, CBR clinical benefit rate, CSF1 colony-stimulating factor 1, CSF1R colony-stimulating factor 1 receptor, dt-GCT diffuse-type tenosynovial giant cell tumor, GCT-TS giant cell tumor of the tendon sheath, GIST gastrointestinal stromal tumor, MEC mucoepidermoid carcinoma of the lung, ORR objective response rate

clinical signs of liver toxicity, and bilirubin levels remained within the normal range [22, 27, 30, 31]. Short-lived elevations of liver enzymes have also been observed in healthy volunteers [28]. The asymptomatic increases in liver enzymes with CSF1R-targeting treatment are most likely caused by a decrease in physiologic clearance through partial depletion of sessile macrophages of the liver (CSF1R⁺ Kupffer cells) [13, 32]. Therefore, liver enzyme elevations can be considered a class effect of CSF1R-targeting compounds. In general, it seems this is not associated with functional liver impairment or structural damage to hepatocytes.

However, there might be differences between CSF1R inhibition with mAbs and with small molecules. For example, although facial edema is reported for up to 64% of patients treated with the mAb emactuzumab [12], it seems to occur to a lesser extent for the small molecule pexidartinib (seen in 26% of patients [22]), and periorbital edema was not reported in a phase 2 study in 63 rheumatoid arthritis patients treated with the small molecule JNJ-40346527 [31]. Potentially immune-related AEs have been described for mAbs [12], whereas serious liver injuries have not been reported. In contrast, enrolment into a phase 3 study with pexidartinib (NCT02371369) was recently suspended because two of 121 patients experienced non-fatal serious liver toxicity [33]. Whether hepatotoxicity may be triggered by inhibition of other receptor kinases, for example, as suggested by hair color changes with pexidartinib treatment in up to 74% of patients (likely due to inhibition of KIT kinase), remains unclear [22]. Potent KIT inhibitors such as dasatinib and pazopanib also cause ALT and AST elevations in about 50% of tumor patients, and hepatocellular necrosis has been shown in patients treated with pazopanib [34]. Hence, it cannot be ruled out that inhibition of tyrosine kinases other than CSF1R contribute to an aggravation of liver toxicities, particularly with smallmolecule inhibitors.

In line with the overall favorable safety profile of CSF1R inhibitors, combination treatment studies have been initiated for both chemotherapies and targeted therapies or immunotherapies. For example, pexidartinib was tested together with paclitaxel in solid tumor patients, and no DLTs were reported [35]. As described in the next section, combinations with programmed cell death protein 1 (PD1) and programmed cell death-ligand 1 (PDL1) inhibitors are ongoing. As with most combination therapies, the promise of increased clinical activity is accompanied by the risk of additive toxicity and therefore requires careful assessment. However, the lack of significant overlapping toxicities of the single-agent AE profiles means these two classes of compounds are promising candidates for successful combination strategies.

Clinical combinations including CSF1R inhibitors

Rational combination therapies investigating CSF1R inhibition have been investigated in preclinical cancer models (recently reviewed by Ries et al. [13]). Various small-molecule kinase inhibitors and antibodies directed against CSF1 or CSF1R were combined with chemotherapies, irradiation, anti-angiogenic or cancer immunotherapies using immunocompetent and immunodeficient mouse models. Notably, the attack of tumor cells via chemotherapy or irradiation induced an upregulation of tumor-derived CSF1 secretion followed by enhanced TAM infiltration that provided additional growth and survival factors for the tumor. A similar mechanism was described for anti-angiogenic therapy that resulted in enhanced supply of vascular endothelial growth factor (VEGF) by TAM. Hence, the combination of tumortargeted or anti-angiogenic therapies and CSF1R inhibitors resulted in improved anti-tumoral activity. Another important link between TAM and cytotoxic CD8⁺ T cells was established using combinations with adoptive T-cell therapy or immune checkpoint inhibitors. In this context, TAM-derived suppressive cytokines such as IL-10 or the general T-cell suppressive functions of TAM provided the basis for increased tumor-inhibitory effects of CSF1R inhibitors combined with immunotherapies. On the basis of these results, multiple clinical trials combining various CSF1/CSF1R inhibitors with agents of diverse mechanisms of action were initiated. Examples include combinations with radiation and androgen-deprivation therapy in prostate cancer, radiation therapy and temozolomide in GBM, rapamycin in peripheral nerve sheet tumors, paclitaxel and eribulin in breast cancer, vemurafenib in melanoma, and KIT inhibitors in gastrointestinal stromal tumors (GISTs). Details and references are summarized in Table 2.

Clinical trials assessing the combination of CSF1R inhibitors with immune checkpoint inhibition clearly outnumber other ongoing combination efforts. This may be because immune checkpoint inhibitors have revolutionized therapeutic strategies in oncology, due to the durable clinical benefit experienced by a fraction of patients from enhancing systemic anti-tumor immunity. The cytotoxic T-lymphocyte-associated protein 4 (CTLA4) antagonist ipilimumab was the first checkpoint inhibitor to demonstrate clinical activity that led to a first approval in metastatic melanoma in 2011 [36]. Anti-PD1 and anti-PDL1 mAbs such as nivolumab, pembrolizumab, or atezolizumab have proven superior efficacy over standard-of-care therapy in a variety of indications such as melanoma, nonsmall-cell lung cancer (NSCLC), bladder cancer, and others [37-42]. Despite the sustained and deep responses observed in some patients, the majority of cancer patients do not respond to these agents. The underlying primary and secondary resistance mechanisms are not well understood; however, evidence is increasing that overcoming the immunosuppressive TME is key to improving the clinical activity of cancer immunotherapy. It is well documented that TAM and other myeloid cells contribute to an immunosuppressive TME. CSFR1 blockade has been shown to reduce T-cell-suppressive TAM infiltrates [13, 18]. Therefore, CSF1R inhibitors represent a promising combination partner for T-cell-enhancing immunotherapies. Based on the widespread use of PD1 and PDL1 inhibitors across tumor entities, a variety of clinical trials combining these agents with CSF1R inhibitors have been initiated. Most of these trials are currently in the dose-finding phase or are evaluating safety and preliminary efficacy in expansion cohorts (Table 3). Clinical results are eagerly awaited, and investigators are aiming to establish superiority of the combination regimen over checkpoint inhibition monotherapy, with the ultimate goal of replacing the current standard of care in various tumor types.

Apart from combining CSF1R inhibitors with PD1/PDL1 or CTLA4 antagonists, alternative strategies to further enhance anti-tumor efficacy of the host immune system are being investigated. Repolarization of the TME is being pursued by combining the CD40 agonist RO7009789 with the anti-CSF1R antibody emactuzumab in a phase 1 clinical trial (NCT02760797). Preclinically, Mok and coworkers reported that depletion of alternatively activated macrophages with pexidartinib improved the efficacy of adoptive cell transfer in a melanoma mouse model [43]. However, this combination has not been tested clinically. Other approaches include, for example, CSF1R inhibition within a triple combination with chemotherapy and anti-angiogenic treatment in platinum-resistant ovarian cancer (NCT02923739).

Local tissue macrophage and disease specificity impacting CSF1R-directed treatment strategies

CSF1R-targeting agents exhibit a rather benign safety profile; however, to date, only modest clinical activity as monotherapy has been reported outside of CSF1-driven dt-GCT. Currently, important patient data sets for solid and hematological malignancies, especially those from ongoing combination trials, are still pending. The complex phenotypic and functional properties of macrophages are heavily influenced by a continuum of survival, differentiation, recruitment, and polarization signals within their specific tissue environment. Therefore, a key question remaining to be answered in clinical studies is whether these agents provide benefit to all cancer patients by depleting CSF1R+ TAM, or whether certain patients and/or tumor types are more likely to respond to CSF1R inhibition. Evidence is increasing that the individual underlying tumor histology as well as organ site-specific features of CSF1R⁺ cells need to be considered.

In the healthy lung, for instance, resident alveolar macrophages that develop from fetal monocytes are mainly regulated by the presence of local GM-CSF [44, 45]. Downstream of GM-CSF signaling, lung-specific transcription factor peroxisome proliferator-activated receptor (PPAR)-y and nuclear repressor Bach2 are responsible for surfactant clearance in macrophages that are participating in host defense [46–48]. This supports the hypothesis that alveolar macrophages in healthy steady state are predominantly of the immunostimulatory M1 macrophage subtype. These cells produce T_b1 cytokines and promote T-cell stimulation while expressing low levels of CSF1R and proving largely resistant to CSF1R inhibitors [13]. Two recent publications report high levels of CSF1R and low human leukocyte antigen-antigen D-related (HLA-DR) as well as increased levels of macrophage markers CD68 and CD163 and decreased levels of inducible nitric oxide synthase (iNOS) on myeloid cells derived from tumors of lung cancer patients [10, 49]. Thus, tumor cells may induce a reprogramming of steady-state alveolar macrophages from the M1, CSF1Rlow phenotype towards the M2, CSF1R^{high} phenotype, making this tumor type a good candidate for CSF1R-targeting therapies. However, the literature documents contradictory prognostic relevance of TAM in NSCLC [9]. This may in part be due to the different detection methods and markers used, but it is more likely that the contradictory reports indicate differences in the local TAM phenotypes in lung cancer subtypes. Depending on the predominant TAM phenotype, CSF1R inhibition may either a) reprogram an M2 TAMdominated, immunosuppressive TME through depletion of CSF1R-dependent TAM or b) boost an ongoing antitumor response by increasing the M1/M2 TAM ratio in an M1 TAM-dominated TME. For both strategies, the choice of an appropriate combination partner will be key to reprogramming a tumor-promoting TME or boosting a pre-existing anti-tumor immune response. Recently, in vitro-differentiated CD206-expressing human macrophages were shown to be rescued from emactuzumab-induced depletion in the presence of IL-4 [50], demonstrating the importance of the local cytokine micromilieu. As CD206 expression is high on alveolar macrophages, an elevated concentration of IL-4 in lung cancer patients may result in resistance to CSF1R inhibitors. Patient data on local IL-4 concentrations and the effect of CSF1R-targeting agents are not available yet.

The physiological steady state of intestinal macrophages in colon tissue is quite different from those in the lung. Intestinal macrophages also originate from monocytes [51] but exhibit a reduced inflammatory activation state that allows the healthy co-existence with commensal bacteria achieved by locally produced autocrine IL-10 as well as regulatory T-cell-derived IL-10 production [52]. This M2 macrophage phenotype is able

to protect colon cancer cells from tumor necrosis factorrelated apoptosis-inducing ligand (TRAIL)-induced cell death [53] and is dependent on CSF1R-related signaling, which was demonstrated for different species (i.e., mice, monkeys) and in human colorectal cancer (CRC) patients, where intestinal macrophages were reduced significantly with CSF1R-targeting therapy [13, 18, 54, 55]. This predominant immunosuppressed environment, together with the observed CSF1R dependency of intestinal macrophages, supports using CSF1R inhibition in CRC patients. However, again, available data on the prognostic relevance of macrophages in CRC patients are contradictory [56]. In contrast to stromal macrophages, peritumoral TAM showed an anti-tumor M1 macrophage phenotype in CRC [57]. The pro- or anti-tumor effect of TAM may therefore also depend on their localization within the TME. However, the degree of CSF1R-signaling dependency of macrophages at these different locations is still unclear. In addition to the location within the TME, the genetic stability of tumor cells may influence the general immune status of the TME. In a CRC subset with high microsatellite instability (MSI-high), TAM infiltration correlated with higher immunity compared with microsatellite stable (MSS) tumors [58, 59]. Therefore, the MSI-high subset of patients may represent a promising target population in which to include CSF1R inhibition to boost pre-existing tumor immunity. Despite the lower number of TAM (and immune infiltrates in general) in MSS CRC tumors, CSF1R-targeted therapies may still also be beneficial in this subset of CRC patients. In MSS CRC patients, a higher expression of genes involved with epithelial to mesenchymal transition (EMT) has been reported, an event that is associated with the invasiveness and spreading of tumor cells [60]. In addition to their immunoregulatory capacities, M2 macrophages can be key contributors to the priming of the pre-metastatic niche [61] by, for example, inducing cytokine-mediated EMT [57, 62] and matrix remodeling [63]. Whether CSF1R blockade in MSS CRC patients is able to control the invasiveness and metastasis remains to be investigated.

In contrast to monocyte-derived alveolar and intestinal macrophages, microglia of the central nervous system (CNS) are resident yolk sac-derived macrophages. They are scavengers for non-functional synapses [52, 64, 65], and CSF1R-mediated signaling is required both during early CNS development [66] and for survival in adults [67]. Inhibition of CSF1R signaling via small molecules BLZ9445 [68] or PLX3397 [69] leads to profound depletion of microglia in the CNS without overt behavioral abnormalities or reduced performance in cognitive function in mice. Although of different origin than, for example, intestinal macrophages, the observed function of microglia seem to be

similar to those of the M2 macrophage subtype described for intestinal macrophages in the healthy gut. In contrast to the contradictory reports for lung and CRC, the presence of TAM in human gliomas seems to be exclusively associated with tumor growth, grade, and poor prognosis [70, 71]. Targeting of microglia using BLZ9445 resulted in improved survival and regression of tumors in a mouse proneural GBM model. In this tumor model, CSF1R blockade did not result in depletion of microglia as in healthy control mice. Here, the local TME under anti-CSF1R treatment was dominated by tumor cell-derived GM-CSF and IFN-y. Interestingly, this local cytokine milieu did not result in unresponsiveness of TAM to anti-CSF1R monotherapy treatment but rather in reprogramming of microglia from M2 into M1 TAM. A phase 2 clinical study assessing pexidartinib in GBM patients is currently ongoing (NCT01790503). In the aforementioned GBM in vivo model, long-term anti-CSF1R treatment led to acquired resistance driven by elevated macrophage-derived insulin-like growth factor 1 (IGF-1) and high IGF-1 receptor (IGF-1R) levels on tumor cells, resulting in enhanced glioma cell survival and invasion [72]. Whether high IL-4 levels together with CD206 expression on TAM may also play a role in this acquired resistance to CSF1R inhibition in GBM is currently unknown. Potential translational approaches to prevent or resolve resistance to CSF1R inhibition may consist of treatment schedules other than continuous administration of CSF1R inhibitors. Alternative schedules could, for example, pursue an initial anti-CSF1R-mediated debulking of M2 TAM followed by other treatment modalities to maintain or induce tumor immunity.

With several CSF1R-targeting therapies currently under evaluation in the clinic, we are still only beginning to understand which covariates impact on macrophage phenotypes and the respective role of CSF1R-mediated signaling in cancer. The aforementioned examples illustrate that the origin and the presence of a predominant macrophage phenotype in healthy tissue cannot alone predict pro- or antitumor effects of TAM during tumorigenesis. Furthermore, it is difficult to identify tumor types that might or might not benefit from CSF1R-targeted therapies without taking into account further sub-classification of tumors and their respective impact on the local TME. Both primary and disseminated tumor cells may induce a profound functional reprogramming of resident tissue macrophages by changing the local cytokine milieu. Only once the impact of organspecific CSF1R blockade is better understood will a more precise selection of anti-CSF1R-containing treatment regimens and prediction of clinical benefit for patients be possible.

Conclusions

CSF1R inhibitors represent an exciting new class of immune-modulatory drugs. Scientific understanding of macrophage and CSF1R biology is evolving rapidly, and more data from clinical trials investigating CSF1R-directed therapies will become available shortly. Whereas clinical tolerability seems to have been established for this group of agents, it is increasingly clear that the organ site and tumor-type specifics of TAM will need to be considered for the selection of both the right patients population and the appropriate combination partner to achieve a meaningful clinical benefit for cancer patients.

Abbreviations

AE: Adverse event; ALT: Alanine aminotransferase; AST: Aspartate aminotransferase; ATC: Anaplastic thyroid cancer; BPI: Brief Pain Inventory; CBR: Clinical benefit rate; cHL: Classical Hodgkin lymphoma; CNS: Central nervous system; CRC: Colorectal cancer; CSF1: Colony-stimulating factor 1; CSF1R: Colony-stimulating factor 1 receptor; CTLA4: Cytotoxic T-lymphocyteassociated protein 4; DC: Dendritic cells; DLT: Dose-limiting toxicity; dt-GCT: Diffuse-type tenosynovial giant cell tumors; EMT: Epithelial to mesenchymal transition; FLT3: fms-like tyrosine kinase 3; GBM: Glioblastoma; GCT-TS: Giant cell tumor of the tendon sheath; GIST: Gastrointestinal stromal tumor; GM-CSF: Granulocyte-macrophage colony-stimulating factor; HLA-DR: Human leukocyte antigen-antigen D related; IDO: Indolamin-2,3-Dioxygenase; IFNγ: Interferon γ; IGF-1: Insulin-like growth factor 1; IHC: Immunohistochemistry; IL: Interleukin; iNOS: Inducible nitric oxide synthase; mAb: Monoclonal antibody; MCSF: Macrophage colony-stimulating factor; MDSC: Myeloid derived suppressor cells; MEC: Mucoepidermoid carcinoma of the lung; MPNST: Malignant peripheral nerve sheath tumor; MSI: Microsatellite instability; MSS: Microsatellite stable; MTD: Maximum tolerated dose; NRS: Numeric rating scale; NSCLC: Non-small cell lung cancer; ORR: Objective response rate; PD1: Programmed cell death protein 1; PDGFR: Platelet-derived growth factor receptor; PDL1: Programmed cell death-ligand 1; PFS: Progression-free survival; PPAR-y: Peroxisome proliferator-activated receptor-y; PROMIS: Patient-Reported Outcomes Measurement Information System; PVNS: Pigmented villonodular synovitis; RCC: Renal cell carcinoma; SCCHN: Squamous cell carcinoma of the head and neck; TAM: Tumor-associated macrophages; Th cells: T helper cells; TME: Tumor microenvironment; TNBC: Triple-negative breast cancer; TNFα: Tumor necrosis factor α; TRAIL: Tumor necrosis factor related apoptosis inducing ligand; UBC: Urothelial bladder carcinoma; VEGF: Vascular Endothelial Growth Factor; WOMAC: Western Ontario and McMasters Universities Osteoarthritis Index

Acknowledgements

The authors thank Dr. Irina Klaman, pRED Roche Innovation Center Munich, for providing the immunohistochemistry analysis in Fig. 2 (Gomez-Roca et al. J Clin Oncol 2015;33). The authors thank Dr. William Pao, Dr. Friedrich Graf Finckenstein, and Dr. Monika Baehner for critical review of the manuscript.

Funding

Not applicable.

Availability of data and materials

Not applicable.

Authors' contributions

MW, CHR, MAC, WJ, AJ, and DR drafted and critically revised the manuscript. All authors read and approved the final manuscript.

Competing interests

M.W., C.H.R., M.A.C., W.J., A.J., and D.R. are employees of Roche Diagnostics GmbH. C.H.R., M.A.C., and D.R. are inventors of granted and pending patent applications relating to RG7155. M.W., C.H.R., M.A.C., W.J., and D.R. hold stock and stock options in F. Hoffman La Roche. The authors declare that they have no competing interests.

Consent for publication

Not applicable.

Ethics approval and consent to participate

Not applicable.

Publisher's Note

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

Received: 10 February 2017 Accepted: 5 June 2017 Published online: 18 July 2017

References

- Lamagna C, Aurrand-Lions M, Imhof BA. Dual role of macrophages in tumor growth and angiogenesis. J Leukoc Biol. 2006;80:705–13.
- Mills CD. M1 and M2 Macrophages: Oracles of Health and Disease. Crit Rev Immunol. 2012;32:463–88.
- 3. Noy R, Pollard JW. Tumor-associated macrophages: from mechanisms to therapy. Immunity. 2014;41:49–61.
- Ostuni R, Kratochvill F, Murray PJ, Natoli G. Macrophages and cancer: from mechanisms to therapeutic implications. Trends Immunol. 2015;36:229–39.
- Pedersen MB, Danielsen AV, Hamilton-Dutoit SJ, Bendix K, Norgaard P, Moller MB, Steiniche T, d'Amore F. High intratumoral macrophage content is an adverse prognostic feature in anaplastic large cell lymphoma. Histopathology. 2014;65:490–500.
- Lenzo JC, Turner AL, Cook AD, Vlahos R, Anderson GP, Reynolds EC, Hamilton JA. Control of macrophage lineage populations by CSF-1 receptor and GM-CSF in homeostasis and inflammation. Immunol Cell Biol. 2012;90:429–40.
- Stanley ER, Chitu V: CSF-1 receptor signaling in myeloid cells. Cold Spring Harb Perspect Biol 2014, 6.
- Achkova D, Maher J. Role of the colony-stimulating factor (CSF)/CSF-1 receptor axis in cancer. Biochem Soc Trans. 2016;44:333–41.
- Zhang QW, Liu L, Gong CY, Shi HS, Zeng YH, Wang XZ, Zhao YW, Wei YQ. Prognostic significance of tumor-associated macrophages in solid tumor: a meta-analysis of the literature. PLoS One. 2012;7:e50946.
- Holmgaard RB, Zamarin D, Lesokhin A, Merghoub T, Wolchok JD. Targeting myeloid-derived suppressor cells with colony stimulating factor-1 receptor blockade can reverse immune resistance to immunotherapy in indoleamine 2,3-dioxygenase-expressing tumors. EBioMedicine. 2016;6:50–8.
- Bronte V, Brandau S, Chen SH, Colombo MP, Frey AB, Greten TF, Mandruzzato S, Murray PJ, Ochoa A, Ostrand-Rosenberg S, et al. Recommendations for myeloid-derived suppressor cell nomenclature and characterization standards. Nat Commun. 2016;7:12150.
- Cassier PA, Italiano A, Gomez-Roca CA, Le Tourneau C, Toulmonde M, Cannarile MA, Ries C, Brillouet A, Muller C, Jegg AM, et al. CSF1R inhibition with emactuzumab in locally advanced diffuse-type tenosynovial giant cell tumours of the soft tissue: a dose-escalation and dose-expansion phase 1 study. Lancet Oncol. 2015;16:949–56.
- Ries CH, Cannarile MA, Hoves S, Benz J, Wartha K, Runza V, Rey-Giraud F, Pradel LP, Feuerhake F, Klaman I, et al. Targeting Tumor-Associated Macrophages with Anti-CSF-1R Antibody Reveals a Strategy for Cancer Therapy. Cancer Cell. 2014;25:846–59.
- Butowski N, Colman H, De Groot JF, Omuro AM, Nayak L, Wen PY, Cloughesy TF, Marimuthu A, Haidar S, Perry A, et al. Orally administered colony stimulating factor 1 receptor inhibitor PLX3397 in recurrent glioblastoma: an Ivy Foundation Early Phase Clinical Trials Consortium phase Il study. Neuro Oncol. 2016;18:557–64.
- Moskowitz CH, Younes A, de Vos S, Bociek RG, Gordon LI, Witzig TE, Gascoyne RD, West B, Nolop K, Steidl C: CSF1R Inhibition by PLX3397 in Patients with Relapsed or Refractory Hodgkin Lymphoma: Results From a Phase 2 Single Agent Clinical Trial. Blood 2012, 120.
- von Tresckow B, Morschhauser F, Ribrag V, Topp MS, Chien C, Seetharam S, Aquino R, Kotoulek S, Khan I, de Boer C, Engert A: A Phase 1 Study Of JNJ-40346527, a Colony Stimulating Factor-1 Receptor (CSF-1R) Inhibitor, In Patients With Relapsed Or Refractory Hodgkin Lymphoma. *Blood* 2013, 122.
- Bendell JC, Tolcher AW, Jones SF, Beeram M, Infante JR, Larsen P, Rasor K, Garrus JE, Li JF, Cable PL, et al.: A phase 1 study of ARRY-382, an oral inhibitor of colony-stimulating factor-1 receptor (CSF1R), in patients with advanced or metastatic cancers. Molecular Cancer Therapeutics 2013, 12.

- Gomez-Roca CA, Cassier PA, Italiano A, Cannarile M, Ries C, Brillouet A, Mueller C, Jegg AM, Meneses-Lorente G, Baehner M, et al.: Phase I study of RG7155, a novel anti-CSF1R antibody, in patients with advanced/metastatic solid tumors. *Journal of Clinical Oncology* 2015, 33.
- Papadopoulos K, Gluck L, Martin LP, Olszanski AJ, Ngarmchamnanrith G, Rasmussen E, Amore B, Nagorsen D, Hill JS, Stephenson J: First-in-human study of AMG 820, a monoclonal anti-CSF-1R (c-fms) antibody, in patients (pts) with advanced solid tumors. *Cancer Research* 2016, 76.
- Blay JY, El Sayadi H, Thiesse P, Garret J, Ray-Coquard I. Complete response to imatinib in relapsing pigmented villonodular synovitis/tenosynovial giant cell tumor (PVNS/TGCT). Ann Oncol. 2008;19:821–2.
- 21. Cheng EY ea: MCS110, an anti-CSF-1 antibody, for the treatment of pigmented villonodular synovitis (PVNS). In ISOLS/MSTS 2015 Annual Meeting, Abstract #11105, 2015.
- Tap WD, Wainberg ZA, Anthony SP, Ibrahim PN, Zhang C, Healey JH, Chmielowski B, Staddon AP, Cohn AL, Shapiro GI, et al. Structure-Guided Blockade of CSF1R Kinase in Tenosynovial Giant-Cell Tumor. N Engl J Med. 2015;373:428–37.
- Gelhorn H, Tong S, Maclaine G, Healey JH, Bukata SV, Keedy VL, Anthony SP, Wagner AJ, Von Hoff DD, Singh AS, et al.: Patient-reported outcome instruments meaningful and relevant for tenosynovial giant cell tumor (TGCT): A qualitative study. *Journal of Clinical Oncology* 2015, 33.
- Gelhorn HL, Tong S, McQuarrie K, Vernon C, Hanlon J, Maclaine G, Lenderking W, Ye X, Speck RM, Lackman RD, et al. Patient-reported Symptoms of Tenosynovial Giant Cell Tumors. Clin Ther. 2016;38:778–93.
- von Tresckow B, Morschhauser F, Ribrag V, Topp MS, Chien C, Seetharam S, Aquino R, Kotoulek S, de Boer CJ, Engert A. An Open-Label, Multicenter, Phase I/II Study of JNJ-40346527, a CSF-1R Inhibitor, in Patients with Relapsed or Refractory Hodgkin Lymphoma. Clin Cancer Res. 2015;21:1843–50.
- Butowski NA, Colman H, De Groot JF, Omuro AMP, Nayak L, Cloughesy TF, Marlmuthu A, Perry A, Phillips JJ, West B, et al.: A phase 2 study of orally administered PLX3397 in patients with recurrent glioblastoma. *Journal of Clinical Oncology* 2014, 32.
- Hambleton J, Zhou L, Rogers S, van Marle S, van Iersel T, Zanghi J, Masteller E, Baker K, Wong B. A Phase 1 Study of FPA008, an Anti-Colony Stimulating Factor 1 Receptor (anti-CSF1R) Antibody in Healthy Volunteers and Subjects with Rheumatoid Arthritis (RA): Preliminary Results. Arthritis Rheum. 2014;66:S657.
- Sadis S, Mukherjee A, Olson S, Dokmanovich M, Maher R, Cai C-H, Le V. Safety, Pharmacokinetics, And Pharmacodynamics Of PD-0360324, AHuman Monoclonal Antibody To Monocyte/Macrophage Colony Stimulating Factor, In Healthy Volunteers [abstract]. Arthritis Rheum. 2009;60 Suppl 10:408.
- Tap WD, Anthony SP, Chmielowski B, Staddon AP, Cohn AL, Shapiro G, Puzanov I, Kwak EL, Wagner AJ, Peterfy C, et al.: A pilot study of PLX3397, a selective colony-stimulating factor 1 receptor (CSF1R) kinase inhibitor, in pigmented villonodular synovitis (PVNS). *Journal of Clinical Oncology* 2014, 32.
- Zhou L, Sikorski R, Rogers S, Costin S, Korkosz M, Jaraczewska-Baumann M, Eva P, Rojkovich B, Bartalos J, Masteller E, et al.: A Phase 1 Study of FPA008, an Anti-Colony Stimulating Factor 1 Receptor (anti-CSF1R) Antibody in Patients (pts) with Rheumatoid Arthritis (RA): Preliminary Results. Arthritis & Rheumatology 2015, 67.
- Genovese MC, Hsia E, Belkowski SM, Chien C, Masterson T, Thurmond RL, Manthey CL, Yan XY, Ge TT, Franks C, Greenspan A. Results from a Phase IIA Parallel Group Study of JNJ-40346527, an Oral CSF-1R Inhibitor, in Patients with Active Rheumatoid Arthritis despite Disease-modifying Antirheumatic Drug Therapy. J Rheumatol. 2015;42:1752–60.
- Radi ZA, Koza-Taylor PH, Bell RR, Obert LA, Runnels HA, Beebe JS, Lawton MP, Sadis S. Increased Serum Enzyme Levels Associated with Kupffer Cell Reduction with No Signs of Hepatic or Skeletal Muscle Injury. Am J Pathol. 2011;179:240–7.
- ENLIVEN Phase 3 Study of Pexidartinib in Tenosynovial Giant Cell Tumor (TGCT) Will Continue to Completion Following Enrollment Discontinuation [media release]. Tokyo: Daiichi Sankyo Company Ltd; 21 October 2016. [http://www.daiichisankyo.com/media_investors/media_relations/press_ releases/detail/006538.html]
- Shah RR, Morganroth J, Shah DR. Hepatotoxicity of tyrosine kinase inhibitors: clinical and regulatory perspectives. Drug Saf. 2013;36:491–503.
- Rugo HS, Sharma N, Reebel L, Rodal MB, Peck A, West BL, Marimuthu A, Karlin DA, Dowlati A, Le MH, et al.: Phase lb Study of Plx3397, a Csf1r Inhibitor, and Paclitaxel in Patients with Advanced Solid Tumors. *Annals of Oncology* 2014, 25.

- Hodi FS, O'Day SJ, McDermott DF, Weber RW, Sosman JA, Haanen JB, Gonzalez R, Robert C, Schadendorf D, Hassel JC, et al. Improved survival with ipilimumab in patients with metastatic melanoma. N Engl J Med. 2010;363:711–23.
- Ansell SM, Lesokhin AM, Borrello I, Halwani A, Scott EC, Gutierrez M, Schuster SJ, Millenson MM, Cattry D, Freeman GJ, et al. PD-1 blockade with nivolumab in relapsed or refractory Hodgkin's lymphoma. N Engl J Med. 2015;377:311–9
- Ferris RL, Blumenschein Jr G, Fayette J, Guigay J, Colevas AD, Licitra L, Harrington K, Kasper S, Vokes EE, Even C, et al. Nivolumab for Recurrent Squamous-Cell Carcinoma of the Head and Neck. N Engl J Med. 2016; 375:1856–67.
- Herbst RS, Baas P, Kim DW, Felip E, Perez-Gracia JL, Han JY, Molina J, Kim JH, Arvis CD, Ahn MJ, et al. Pembrolizumab versus docetaxel for previously treated, PD-L1-positive, advanced non-small-cell lung cancer (KEYNOTE-010): a randomised controlled trial. Lancet. 2016;387:1540–50.
- Motzer RJ, Escudier B, McDermott DF, George S, Hammers HJ, Srinivas S, Tykodi SS, Sosman JA, Procopio G, Plimack ER, et al. Nivolumab versus Everolimus in Advanced Renal-Cell Carcinoma. N Engl J Med. 2015;373:1803–13.
- Robert C, Long GV, Brady B, Dutriaux C, Maio M, Mortier L, Hassel JC, Rutkowski P, McNeil C, Kalinka-Warzocha E, et al. Nivolumab in previously untreated melanoma without BRAF mutation. N Engl J Med. 2015;372:320–30.
- Rosenberg JE, Hoffman-Censits J, Powles T, van der Heijden MS, Balar AV, Necchi A, Dawson N, O'Donnell PH, Balmanoukian A, Loriot Y, et al. Atezolizumab in patients with locally advanced and metastatic urothelial carcinoma who have progressed following treatment with platinum-based chemotherapy: a single-arm, multicentre, phase 2 trial. Lancet. 2016;387:1909–20.
- Mok S, Koya RC, Tsui C, Xu J, Robert L, Wu L, Graeber TG, West BL, Bollag G, Ribas A. Inhibition of CSF-1 receptor improves the antitumor efficacy of adoptive cell transfer immunotherapy. Cancer Res. 2014;74:153–61.
- Guilliams M, De Kleer I, Henri S, Post S, Vanhoutte L, De Prijck S, Deswarte K, Malissen B, Hammad H, Lambrecht BN. Alveolar macrophages develop from fetal monocytes that differentiate into long-lived cells in the first week of life via GM-CSF. J Exp Med. 2013;210:1977–92.
- 45. Hamilton JA. Colony-stimulating factors in inflammation and autoimmunity. Nat Rev Immunol. 2008;8:533–44.
- 46. Kuroki Y, Takahashi M, Nishitani C. Pulmonary collectins in innate immunity of the lung. Cell Microbiol. 2007;9:1871–9.
- 47. Nakamura A, Ebina-Shibuya R, Itoh-Nakadai A, Muto A, Shima H, Saigusa D, Aoki J, Ebina M, Nukiwa T, Igarashi K. Transcription repressor Bach2 is required for pulmonary surfactant homeostasis and alveolar macrophage function. J Exp Med. 2013;210:2191–204.
- Schneider C, Nobs SP, Kurrer M, Rehrauer H, Thiele C, Kopf M. Induction of the nuclear receptor PPAR-gamma by the cytokine GM-CSF is critical for the differentiation of fetal monocytes into alveolar macrophages. Nat Immunol. 2014;15:1026–37.
- Almatroodi SA, McDonald CF, Darby IA, Pouniotis DS. Characterization of M1/M2 Tumour-Associated Macrophages (TAMs) and Th1/Th2 Cytokine Profiles in Patients with NSCLC. Cancer Microenviron. 2016;9:1–11.
- Pradel LP, Ooi CH, Romagnoli S, Cannarile MA, Sade H, Ruttinger D, Ries CH. Macrophage Susceptibility to Emactuzumab (RG7155) Treatment. Mol Cancer Ther. 2016;15:3077–86.
- Bain CC, Bravo-Blas A, Scott CL, Gomez Perdiguero E, Geissmann F, Henri S, Malissen B, Osborne LC, Artis D, Mowat AM. Constant replenishment from circulating monocytes maintains the macrophage pool in the intestine of adult mice. Nat Immunol. 2014;15:929–37.
- Amit I, Winter DR, Jung S. The role of the local environment and epigenetics in shaping macrophage identity and their effect on tissue homeostasis. Nat Immunol. 2016;17:18–25.
- Kaler P, Augenlicht L, Klampfer L. Activating mutations in beta-catenin in colon cancer cells alter their interaction with macrophages; the role of snail. PLoS One. 2012;7:e45462.
- Kruse J, von Bernstorff W, Evert K, Albers N, Hadlich S, Hagemann S, Gunther C, van Rooijen N, Heidecke CD, Partecke LI. Macrophages promote tumour growth and liver metastasis in an orthotopic syngeneic mouse model of colon cancer. Int J Colorectal Dis. 2013;28:1337–49.
- MacDonald KP, Palmer JS, Cronau S, Seppanen E, Olver S, Raffelt NC, Kuns R, Pettit AR, Clouston A, Wainwright B, et al. An antibody against the colonystimulating factor 1 receptor depletes the resident subset of monocytes and tissue- and tumor-associated macrophages but does not inhibit inflammation. Blood. 2010;116:3955–63.

- Owusu BY, Vaid M, Kaler P, Klampfer L. Prognostic and Predictive Significance of Stromal Fibroblasts and Macrophages in Colon Cancer. Biomark Cancer. 2015;7:29–37.
- 57. Erreni M, Mantovani A, Allavena P. Tumor-associated Macrophages (TAM) and Inflammation in Colorectal Cancer. Cancer Microenviron. 2011;4:141–54.
- Kim CG, Ahn JB, Jung M, Beom SH, Kim C, Kim JH, Heo SJ, Park HS, Kim JH, Kim NK, et al. Effects of microsatellite instability on recurrence patterns and outcomes in colorectal cancers. Br J Cancer. 2016;115:25–33.
- Xiao Y, Freeman GJ. The microsatellite instable subset of colorectal cancer is a particularly good candidate for checkpoint blockade immunotherapy. Cancer Discov. 2015;5:16–8.
- Mlecnik B, Bindea G, Angell HK, Maby P, Angelova M, Tougeron D, Church SE, Lafontaine L, Fischer M, Fredriksen T, et al. Integrative Analyses of Colorectal Cancer Show Immunoscore Is a Stronger Predictor of Patient Survival Than Microsatellite Instability. Immunity. 2016;44:698–711.
- Qian BZ, Li J, Zhang H, Kitamura T, Zhang J, Campion LR, Kaiser EA, Snyder LA, Pollard JW. CCL2 recruits inflammatory monocytes to facilitate breasttumour metastasis. Nature. 2011;475:222–5.
- 62. Techasen A, Loilome W, Namwat N, Dokduang H, Jongthawin J, Yongvanit P. Cytokines released from activated human macrophages induce epithelial mesenchymal transition markers of cholangiocarcinoma cells. Asian Pac J Cancer Prev. 2012;13(Suppl):115–8.
- Afik R, Zigmond E, Vugman M, Klepfish M, Shimshoni E, Pasmanik-Chor M, Shenoy A, Bassat E, Halpern Z, Geiger T, et al. Tumor macrophages are pivotal constructors of tumor collagenous matrix. J Exp Med. 2016;213:2315–31.
- Fourgeaud L, Traves PG, Tufail Y, Leal-Bailey H, Lew ED, Burrola PG, Callaway P, Zagorska A, Rothlin CV, Nimmerjahn A, Lemke G. TAM receptors regulate multiple features of microglial physiology. Nature. 2016;532:240–4.
- 65. Ransohoff RM, Cardona AE. The myeloid cells of the central nervous system parenchyma. Nature. 2010;468:253–62.
- Erblich B, Zhu LY, Etgen AM, Dobrenis K, Pollard JW: Absence of Colony Stimulation Factor-1 Receptor Results in Loss of Microglia, Disrupted Brain Development and Olfactory Deficits. *Plos One* 2011, 6.
- Ginhoux F, Greter M, Leboeuf M, Nandi S, See P, Gokhan S, Mehler MF, Conway SJ, Ng LG, Stanley ER, et al. Fate Mapping Analysis Reveals That Adult Microglia Derive from Primitive Macrophages. Science. 2010;330:841–5.
- Pyonteck SM, Akkari L, Schuhmacher AJ, Bowman RL, Sevenich L, Quail DF, Olson OC, Quick ML, Huse JT, Teijeiro V, et al. CSF-1R inhibition alters macrophage polarization and blocks glioma progression. Nat Med. 2013;19:1264–72.
- Elmore MR, Najafi AR, Koike MA, Dagher NN, Spangenberg EE, Rice RA, Kitazawa M, Matusow B, Nguyen H, West BL, Green KN. Colony-stimulating factor 1 receptor signaling is necessary for microglia viability, unmasking a microglia progenitor cell in the adult brain. Neuron. 2014;82:380–97.
- Hussain SF, Yang D, Suki D, Aldape K, Grimm E, Heimberger AB. The role of human glioma-infiltrating microglia/macrophages in mediating antitumor immune responses. Neuro Oncol. 2006;8:261–79.
- Komohara Y, Ohnishi K, Kuratsu J, Takeya M. Possible involvement of the M2 anti-inflammatory macrophage phenotype in growth of human gliomas. J Pathol. 2008;216:15–24.
- Quail DF, Bowman RL, Akkari L, Quick ML, Schuhmacher AJ, Huse JT, Holland EC, Sutton JC, Joyce JA. The tumor microenvironment underlies acquired resistance to CSF-1R inhibition in gliomas. Science. 2016;352:aad3018.
- Gulam Abbas Manji PP, Shing Mirn Lee, Nanette Matos, Edward Bentlyewski, Brian Andrew Van Tine, Khanh Tu Do, Suzanne George, Gary K. Schwartz: Phase 1/2 study of combination therapy with pexidartinib and sirolimus to target tumor-associated macrophages in malignant peripheral nerve sheath tumors. In J Clin Oncol 34, 2016 (suppl; abstr TPS11070); 2016.
- 74. Hu-Lieskovan S, Patnaik A, Eisenberg P, Sachdev J, Weise A, Kaufman DR, Aromin I, West BL, Tong S, Ribas A. Phase 1/2a study of double immune suppression blockade by combining a CSF1R inhibitor (pexidartinib/ PLX3397) with an anti PD-1 antibody (pembrolizumab) to treat advanced melanoma and other solid tumors. Ann Oncol. 2015;26:7–7.
- Brahmer J, Rasco D, Chen MJ, Masteller E, Qazi I, Rogers S, Sankar N, Sikorski R, Hambleton J, Hodi FS: A phase 1a/1b study of FPA008 in combination with nivolumab in patients with selected advanced cancers. Cancer Immunology Research 2016, 4.
- Bauer T. Phase Ia/Ib trial investigating the CSF-1R inhibitor LY3022855 in combination with durvalumab (MEDI4736) or tremelimumab in patients with advanced solid malignancies. J ImmunoTher Cancer. 2016;4 Suppl 1:P137.

Submit your next manuscript to BioMed Central and we will help you at every step:

- We accept pre-submission inquiries
- Our selector tool helps you to find the most relevant journal
- We provide round the clock customer support
- Convenient online submission
- Thorough peer review
- Inclusion in PubMed and all major indexing services
- Maximum visibility for your research

Submit your manuscript at www.biomedcentral.com/submit

