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Vesicle-mediated secretion of human eosinophil granule-derived major basic protein (MBP)

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Abstract

Major basic protein (MBP), the predominant cationic protein of human eosinophil specific granules, is stored within crystalloid cores of these granules. Secretion of MBP contributes to the immunopathogenesis of varied diseases. Prior electron microscopy (EM) of eosinophils in sites of inflammation noted losses of granule cores in the absence of granule exocytosis and suggested that eosinophil granule proteins might be released through piecemeal degranulation (PMD), a secretory process mediated by transport vesicles. Because release of eosinophil granule-derived MBP through PMD has not been studied, we evaluated secretion of this cationic protein by human eosinophils. Intracellular localizations of MBP were studied within non-stimulated and eotaxin-stimulated human eosinophils by both immunofluorescence and a pre-embedding immunogold electron microscopy (EM) method which enables optimal epitope preservation and antigen access to membrane microdomains. In parallel, quantification of transport vesicles was assessed in eosinophils from a patient with hypereosinophilic syndrome (HES). Our data demonstrate vesicular trafficking of MBP within eotaxin-stimulated eosinophils. Vesicular compartments, previously implicated in transport from granules to the plasma membrane, including large vesiculotubular carriers termed eosinophil sombrero vesicles (EoSVs), were found to contain MBP. These secretory compartments were significantly increased in numbers within HES eosinophils. Moreover, in addition to granule-stored MBP, even unstimulated eosinophils contained appreciable amounts of MBP within secretory vesicles, as evidenced by immunogold EM and immunofluorescent co-localizations of MBP and CD63. These data suggest that eosinophil MBP, with its multiple extracellular activities, can be mobilized from granules via PMD into secretory vesicles and both granule- and secretory vesicle-stored pools of

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MBP are available for agonist-elicited secretion of MBP from human eosinophils. The recognition of PMD as a secretory process to release MBP is important to understand the pathological basis of allergic and other eosinophil-associated inflammatory diseases.

Keywords

CD63; eosinophils; human; inflammation; major basic protein, piecemeal degranulation; vesicular transport

Classical roles of eosinophils are based on their effector responses involving secretory processes that mobilize and release their preformed pools of granule-stored cationic proteins, including major basic protein (MBP), the most abundant eosinophil granule cationic protein. The roles of MBP in mediating cytotoxicity and allergic disorders such as asthma are long recognized. In addition to being cytotoxic to a variety of tissues, including heart, brain and bronchial epithelium, MBP increases smooth muscle reactivity by causing dysfunction of vagal muscarinic M2 receptors, which may contribute to the development of airway hyperreactivity, a cardinal feature of asthma (reviewed in (1-3)). MBP is also a potent helminthotoxin (4-6).

Within eosinophils, MBP is synthesized first as a precursor pro-MBP protein that is proteolytically processed within cytoplasmic granules into 14 kD MBP. MBP is then packaged and stored within the often electron dense crystalline cores of eosinophil secretory “specific” granules (7, 8). Early transmission electron microscopy (TEM) studies of lesional eosinophils in Crohn’s disease, eosinophilic gastroenteritis and hypereosinophilic syndrome (HES), noted that eosinophils lost their electron dense cores (9-11), and suggested that eosinophil granule core-derived MBP might be released by a mechanism not involving wholesale granule fusion at the plasma membrane. A candidate mechanism suggested by these early reports was that eosinophil granule proteins might be mobilized from within intracellular granules into vesicles that traffic to and release extracellularly at the cell surface, a process termed piecemeal degranulation (PMD) (12-14). Recent studies from our laboratory demonstrate the selective secretion by PMD of eosinophil granule-stored cytokines, such as IL-4 (15-17). Based principally to date on ultrastructural observations of eosinophil granules, PMD has been suggested to be involved in the release of products from activated eosinophils in a range of human diseases including allergic inflammation (18-24). In full support of these earlier studies, we now demonstrate increased numbers of large vesiculotubular carriers termed eosinophil sombrero vesicles (EoSvs), previously determined to function in cargo transport from granules to the plasma membrane (25), in the absence of granule-granule or granule-plasma membrane fusion events in eosinophils from a patient with HES. Recently, we detected the presence of MBP in transport vesicles within human eosinophils activated *in vitro* with the eosinophil-active chemokine, eotaxin (CCL-11) (25). These results, in concert with prior observations of the losses of MBP-containing cores within granules of eosinophils in sites of inflammation, provided inferential indications about mechanisms underlying MBP release. Although MBP is extensively detected in inflammatory sites (26-31), little is known about whether this eosinophil granule protein can be secreted by PMD.

With pre-embedding immunogold electron microscopy (EM) for precise epitope preservation and secondary antibody Fab fragments specifically conjugated with very small gold particles (1.4 nm) as a probe, we now demonstrate MBP in vesicles surrounding and extending from granules and beneath the plasma membrane within human eosinophils stimulated with eotaxin. Distinct vesicular compartments, including EoSVs (25), were also labeled with anti-MBP antibodies. Furthermore, our findings using immunogold EM and immunofluorescent co-localizations of MBP and CD63, a marker of eosinophil granule limiting membranes (32), indicate that secretory vesicles constitute substantial extragranular pools of MBP even within unstimulated human eosinophils. Our findings provide new insights into the intracellular mechanisms mediating secretion of eosinophil granule-derived proteins, corroborate previous evidence of PMD as a predominant mechanism of eosinophil secretion in eosinophilic patients (19-21, 24) and identify, for the first time, PMD as a secretory process to release the granule-derived cationic protein, MBP, from human eosinophils.

Material and Methods

Eosinophil isolation, stimulation and viability

Granulocytes were isolated from the blood of different healthy donors and from one patient with the HES (negative for Fip1-like 1/platelet-derived growth factor α -FIP1L1/PDGFR α -mutation). Eosinophils from the former donors were enriched and purified by negative selection using human eosinophil enrichment cocktail (StemSepTM, StemCell Technologies, Seattle WA) and the MACS bead procedure (Miltenyi Biotec, Auburn, CA), as described (33), with the exception that hypotonic red blood cell (RBC) lysis was omitted to avoid any potential for RBC lysis to affect eosinophil function. Experiments were approved by the Beth Israel Deaconess Medical Center Committee on Clinical Investigation, and informed consent was obtained from all subjects. Purified eosinophils (10^6 cells/mL) were stimulated with recombinant human eotaxin (100 ng/mL; R&D Systems, Minneapolis, MN) in RPMI-1640 medium plus 0.1% ovalbumin (OVA) (Sigma, St. Louis, MO) or medium alone at 37°C, for 1 h. Eosinophil viability and purity were greater than 99% as determined by ethidium bromide (Molecular Probes, Eugene, OR) incorporation and cytocentrifuged smears stained with HEMA 3 stain kit (Fisher Scientific, Houston, TX), respectively.

Antibody reagents

Anti-human mouse IgG₁ CD63 (clone H5C6) and irrelevant isotype control monoclonal antibodies (mAbs) (BD-Pharmingen, San Diego, CA) were used for flow cytometry (5 μ g/mL) and fluorescence (5 μ g/mL) and electron (2 μ g/mL) microscopy immunodetection studies. Secondary antibody (Ab) for CD63 immunofluorescence in eosinophils was an anti-mouse Alexa 594 pre-bound to the primary Ab using Zenon labeling as per the manufacturer's directions (Molecular Probes) and for CD63 flow cytometry of isolated granules was a goat anti-mouse fluorescein isothiocyanate (FITC)-conjugated Ab (1:100, Jackson ImmunoResearch). The secondary Ab for immunoEM was an affinity-purified goat anti-mouse Fab fragment conjugated to 1.4 nm gold (1:100; Nanogold®, Nanoprobes, Stony Brook, NY). Abs for MBP detection in eosinophils by immunofluorescence (5 μ g/ml for single staining, 2.5 μ g/ml for dual staining) and immunoEM (5 μ g/mL) were monoclonal

mouse anti-human MBP (clone AHE-2) and irrelevant isotype IgG1 control (BD-Pharmingen). Secondary Ab for MBP immunofluorescence in single-stain experiments was anti-mouse Alexa 488 (1:500, Molecular Probes), and for dual staining anti-MBP mAb was detected with anti-mouse Alexa 488 using Zenon labeling kits (Molecular Probes). Mouse anti-human MHC class I (HLA-ABC, clone G46-2.6, 14 µg/ml, BD Pharmingen) FITC-conjugated mAb was used with the respective FITC-conjugated IgG control mAb.

Subcellular fractionation

Eosinophils were resuspended in disrupting buffer as described (34), supplemented with 5 µg/ml dithiothreitol and subjected to nitrogen cavitation (Parr, Moline IL) under pressure of 600 psi (10 min). Post-nuclear supernatants, recovered after centrifugation (400 g, 10 min), were ultracentrifuged (100,000 g, 1 h at 4°C) in linear Accudenz™ or isotonic Optiprep (Axis-Shield PoC AS, Oslo, Norway) gradients (0-45% in disrupting buffer without protease inhibitors). Fractions (20 × 0.5 mL) were collected with a peristaltic pump. Eosinophil granules and EoSV vesicles were isolated, as previously described (16).

Preparation of cells and subcellular fractions for EM

Purified eosinophils and their isolated, purified subcellular fractions, were immediately fixed in a mixture of freshly prepared aldehydes (1% paraformaldehyde (PFO) and 1.25% glutaraldehyde) in 1 M sodium cacodylate buffer, pH 7.4 for 1 h, at room temperature (RT), washed in the same buffer and centrifuged at 1500 g for 1 min. Samples were then resuspended in molten 2% agar in 1M sodium cacodylate buffer, pH 7.4 and quickly recentrifuged. Resultant agar pellets were kept in the same buffer at 4°C for further processing. For immunoEM, cells or subcellular fractions were fixed in fresh 4% PFO in 0.02 M phosphate buffer saline (PBS), pH 7.4 (CD63 labeling) or 1% PFO and 1% glutaraldehyde in 1 M sodium cacodylate buffer, pH 7.4 (MBP labeling), for 30 min at RT, washed in PBS and embedded in molten 2% agar, as above. Pellets were immersed in 30% sucrose in PBS overnight at 4°C, embedded in OCT compound (Miles, Elkhart, IN), and stored in -180°C liquid nitrogen for subsequent use.

Conventional TEM

Agar pellets containing either intact eosinophils or their isolated eosinophil granules were post-fixed in 1% osmium tetroxide in Sym-Collidine buffer, pH 7.4, for 2 h at RT. After washing with sodium maleate buffer, pH 5.2, pellets were stained *en bloc* in 2% uranyl acetate in 0.05 M sodium maleate buffer, pH 6.0 for 2 h at RT and washed in the same buffer as before prior to dehydration in graded ethanols and infiltration and embedding with a propylene oxide-Epon sequence (Eponate 12 Resin; Ted Pella, Redding, CA). After polymerization at 60°C for 16 h, thin sections were cut using a diamond knife on an ultramicrotome (Leica, Bannockburn, IL). Sections were mounted on uncoated 200-mesh copper grids (Ted Pella) before staining with lead citrate and viewed with a transmission electron microscope (CM 10; Philips, Eindhoven, The Netherlands) at 60 KV. For EoSV quantification in granule fractions, electron micrographs from different subcellular fractionations (n=3) were randomly taken at 21,000 X and analyzed at the final magnification of 58,000 X. A total of 941 EoSVs were counted. To quantify the total

number of EoSVs within eosinophils from the HES donor, we randomly obtained 35 electron micrographs of cell sections showing the entire cell profile and nucleus at 12,000 X and analyzed these at a final magnification of 33,000 X. Data were compared using the Mann-Whitney *U*-test ($p < 0.05$).

ImmunoEM

Pre-embedding immunolabeling was done before standard EM processing (dehydration, infiltration, resin embedding and resin sectioning). Pre-embedding immunoEM optimizes antigen preservation and is more sensitive to detect small molecules than post-embedding labeling that is done after conventional EM processing. Moreover, to reach antigens at membrane microdomains such as small vesicles, we used labeling with very small (1.4 nm) gold particles (Nanogold). Immunonanogold was performed on cryostat 10 μm sections mounted on glass slides. All steps were done at RT as before (16), and modified as follows: cells or EoSVs isolated by subcellular fractionation were incubated in a mixture of PBS and bovine serum albumine (PBS-BSA buffer; 0.02 M PBS plus 1% BSA) containing 0.1% gelatin (20 min) followed by PBS-BSA plus 10% normal goat serum (NGS) and incubated with primary Ab (1 h). After blocking with PBS-BSA plus NGS (30 min), cells were incubated with secondary Ab (1 h), washed in PBS-BSA, post-fixed in 1% glutaraldehyde (10 min) and incubated with HQ silver enhancement solution (Nanoprobes) (10 min). Cells or fractions in cryostat sections were immersed in 5% sodium thiosulfate (5 min), post-fixed with 1% osmium tetroxide in distilled water (10 min), stained with 2% uranyl acetate in distilled water (5 min), embedded in Eponate and thin sectioned as described (35). Two controls were performed: (i) primary Ab was replaced by an irrelevant Ab and (ii) primary Ab was omitted. Specimens were examined as described for conventional TEM. Electron micrographs from different experiments ($n=4$) were randomly taken at magnifications of 12,000 to 40,000x to study the entire cell profile and vesicle features. A total of 195 electron micrographs were evaluated.

Immunofluorescence microscopy

Eosinophils (15×10^6 cells/mL) were gently mixed at 37°C with a low-gelling temperature 1.25% agarose (Pierce, Rockford, IL) in a 3:1 ratio (cells:agarose) and carefully spread onto slides (20 μL per slide). After agarose was solidified, a perfusion chamber (CoverWell™, Grace Bio-Labs, Bend, OR) was affixed over cells and medium added to maintain moisture. After several min to allow for cell attachment and spreading, chambers were removed and cells fixed in 2% PFO for 5 min at RT. Slides were then washed with $\text{Ca}^{++}/\text{Mg}^{++}$ free Hanks buffered salt solution (HBSS^{-/-}) alone, followed by a 5 min incubation in permeabilization solution (0.1% saponin, 5% milk, 1% NGS in HBSS^{-/-}). For MBP single-staining experiments, permeabilized cells were incubated with primary Ab for 1 h, washed and incubated with secondary Ab for 45 min at RT. For dual labeling with anti-MBP and anti-CD63, permeabilized cells were incubated with Zenon-labeled primary Abs for 45 min at RT. Cells were washed (2 \times 10 min) before drying and coverslipping. Analyses were performed by both phase contrast and fluorescence microscopy.

Deconvolution microscopy

Fluorescence images were acquired using a Retiga EXi cooled CCD camera (Burnaby, BC, Canada) coupled to a Provis AX-70 Olympus microscope (Olympus, Melville, NY) and an UPlanApo objective (100x 1.35). The microscope, Z-motor drive, shutters and camera were controlled by IPLab 3.6 for Mac (Scanalytics, VA). Acquired stacks were further processed for deconvolution with Volocity 2.6 (Improvision, Lexington, MA).

Flow cytometry

Non-permeabilized granules were incubated either with primary FITC-conjugated Ab (45 min) or primary (1 h) and then FITC-conjugated secondary (15 min) Abs on ice in the absence of granule fixation. After staining, granules were fixed in buffer containing 2% PFO without methanol (Electron Microscopy Sciences) for 5 min. Flow cytometric data, acquired using a FACScan with CellQuest software (BD Biosciences, Franklin Lakes, NJ), were analyzed by comparisons of mean fluorescent intensities (MFI) of unimodal histograms.

Results

Intracellular compartmentalization of MBP in unstimulated human eosinophils

With eosinophils isolated from normal donors and prepared for immunofluorescent staining on air-dried cytopins, punctate (luminal) granule staining was observed for MBP (Fig 1A), as previously reported with this anti-MBP mAb (34). In contrast, when eosinophils were kept moist in an agarose matrix that combines cell labeling with preservation of cell morphology, MBP staining was closely associated with granules and often exhibited circumferential, perigranular staining (Fig 1 B). To further evaluate the perigranular MBP staining, we utilized deconvolution microscopy of eosinophils stained for both MBP and CD63 (Fig 1C), a tetraspanin transmembrane protein that is localized at the cytoplasmic face of eosinophil granule membranes as evidenced by EM immunogold (Fig 1 D) and by flow cytometry showing anti-CD63 on the membranes of eosinophil granules isolated by subcellular fractionation (not shown) (25, 32). Deconvolution microscopy revealed CD63 granule membrane labeling, often associated with MBP labeling peripheral to the CD63 ring (Fig. 1C). Control cells assayed with an irrelevant Ab were negative. The two different MBP labeling patterns observed in the present study can be explained by the use of different cell preparation methodologies. Cell drying before labeling favors antibody access to the compact granule cores, but likely eliminates MBP labeling on other labile intracellular sites. On the other hand, these sites are detected only when the cells are kept moist, although in this case the antibodies cannot access granule cores.

Vesicular transport of MBP in activated eosinophils

To ascertain the intracellular localizations of MBP within activated eosinophils, we utilized immunogold EM protocols likely to preserve extragranular membranous organelles (i.e., vesicles) and detect antigenic epitopes (16). With pre-embedding immunogold EM for epitope preservation and a secondary Fab Ab fragment conjugated to very small gold particles to access possible vesicular domains positive for MBP, MBP was immunolocalized within eosinophils stimulated with an agonist, eotaxin, that elicits mobilization of eosinophil

granule proteins (16). MBP was detected within granule matrices (Fig. 2Ai arrows, and 2Aii) and within granules with apparently mobilized crystalline cores (Fig 2A, D). Moreover, MBP was present within vesicles attached to or surrounding the surface of emptying granules (Fig. 2Aii, 2B and 3C). Vesicles containing MBP were distributed around mobilized granules (Fig. 2B), in the cytoplasm and beneath the cell surface (Fig. 2C and 3A). MBP-positive vesicles fused with the plasma membrane (Fig. 2C and 3B). Of note, the MBP-positive vesicular system was associated with secretory pathways operating from eosinophil granules and not with a synthetic route from the *trans*-Golgi network, which was rarely labeled for MBP (not shown). Control cells in which the primary Ab was replaced by an irrelevant Ab were negative (not shown). Altogether, these findings indicate that in activated eosinophils granule-derived MBP can be segregated into secretory vesicles and released through PMD.

Intracellular transport of MBP is mediated by large vesiculotubular carriers

MBP-containing vesicles included both spherical small vesicles (Fig. 2C) and morphologically distinct, large vesiculotubular EoSV compartments (Fig. 3). EoSVs, recently associated with eosinophil secretory pathways, represent vesiculotubular pathways for transport of eosinophil granule products for their rapid extracellular release at the plasma membrane (16, 36). The lumina of EoSVs consistently labeled for MBP (Fig. 3). EM quantitative analyses revealed up to 39 gold particles/vesicle section. EoSVs were distributed in the cytoplasm, beneath plasma membranes (Fig. 3A) and at times fused with plasma membranes (Fig. 3B), and attached to secretory granules undergoing PMD in *in vitro* eotaxin-stimulated eosinophils (Fig. 3C).

Because EoSVs are prominently formed within *in vitro*-activated eosinophils (36, 37) and these vesicles were extensively labeled for MBP, we next investigated the distribution of EoSVs within otherwise unstimulated eosinophils from a patient with HES. This disorder is characterized by increased numbers of activated eosinophils in the blood and tissues (38-40) and was previously associated with losses of specific granule cores (11). Our EM quantitative analyses found that the total number of EoSVs in HES eosinophils was significantly higher compared to eosinophils from healthy donors [35.4 ± 11.4 per section for HES versus 21.3 ± 4.9 for normal eosinophils (mean \pm SEM), $n = 35$ cells, $p < 0.05$] (Fig. 4A). In the eosinophils of HES subjects, EoSVs were distributed in the cytoplasm and were clearly observed in contact with granules showing disassembled cores (Fig. 4B and 5). Granule-granule or granule-plasma membrane fusion events were not observed within HES eosinophils (Fig. 5).

To corroborate a role for EoSVs in transporting MBP, we utilized subcellular fractionation (16) to isolate EoSVs from unstimulated eosinophils. By immunogold EM, isolated EoSVs contained MBP (Fig. 3D). As observed in intact eosinophils (Fig. 3A-C), MBP was densely labeled within the tubular lumina of EoSVs (Fig. 3D). These results provide new documentation that even within unstimulated human eosinophils MBP is found within secretory vesicles of eosinophils and document that large vesiculotubular carriers, a secretory compartment increased in HES patients, can mediate transport of this cationic granule-derived protein.

Unstimulated eosinophils contain pools of MBP-loaded cytoplasmic vesicles

While activation of eosinophils leads to mobilization of granule-derived MBP into secretory vesicles, several findings, noted above, indicate that even unstimulated eosinophils contain extragranular, vesicular pools of MBP. For instance, these vesicular pools were imaged both by immunofluorescence as labeling circumferential to specific granules in wet preparations (agarose matrix) (Fig. 1B, C), and by immunogold EM as a collar of MBP-loaded vesicles (Fig. 6A, B). The presence of pools of MBP-positive vesicles in unstimulated eosinophils was confirmed by immunogold EM after vesicle isolation by subcellular fractionation (Fig. 3D).

We further evaluated associations between MBP-positive vesicles and specific granules in unstimulated eosinophils. Using the agarose matrix system, we analyzed free granules released from mechanically disrupted unstimulated eosinophils after double immunolabeling for CD63 and MBP. Clusters of free eosinophil granules evaluated at high magnification exhibited granules with the same MBP/CD63 pattern of labeling as observed in intact cells, i.e., with membrane CD63 labeling associated with MBP labeling peripheral to CD63 (Fig. 6C). This indicates that MBP-loaded vesicles are associated with and likely derived from eosinophil specific granules. TEM of subcellularly isolated granules (25), identified EoSVs, with their typical morphology and size (150-300nm), in granule-enriched fractions (Fig. 7A). These subcellular fractions were free of other organelles and exhibited granules with intact surrounding membranes. EM analyses were done using 48 electron micrographs randomly taken from denser granule and supragranule eosinophil subcellular fractionations (n=3) isolated from isotonic Optiprep gradients for optimal membrane preservation. A total of 941 EoSVs were counted. Our analyses indicated that part of this EoSV population (38%) was attached to the granule delimiting membrane in isolated granules (Fig. 7A, B), as seen in intact cells (Fig. 3C) and a larger fraction (62%) was not visibly attached to granules. Of note, granules isolated by subcellular fractionation did not express contaminating plasma membrane or endosomal MHC class I protein as assessed by flow cytometry after staining with anti-MHC I mAb (data not shown). Taken together, our findings identify cytoplasmic vesicles, derived from eosinophils granules, as intracellular sites of MBP localization in unstimulated eosinophils.

Discussion

Release of granule-derived MBP by human eosinophils is a key event in the pathogenesis of allergic airway diseases, such as rhinitis and asthma, and other eosinophil-mediated diseases; but little is known about the cellular mechanisms of MBP release from eosinophils. Granule-stored products are released from eosinophils through different modes; i) classical exocytosis by which granules release their entire contents following granule fusion with the plasma membrane, including compound exocytosis, which also involves intracellular granule-granule fusion before extracellular release; ii) piecemeal degranulation (PMD), which is mediated through vesicular transport and secretion of granule-derived proteins; and iii) cytolysis, which involves the extracellular deposition of intact granules upon lysis of the cell (reviewed in (41-43)). PMD is likely the predominant mode of secretion from eosinophils in different human diseases (18, 20, 22, 23); and both PMD and cytolysis are

reported to occur in the airway lumen and tissues (21, 44, 45). Of note, one eosinophil granule-derived protein, eosinophil cationic protein (ECP), has been documented in subcellular fractionations studies to be localized in cytosolic vesicles isolated from eosinophils of allergic patients specifically during their seasonal allergen exposures (19). During eosinophil development, MBP is sequentially synthesized, processed and then packaged within the crystalline cores of eosinophil specific granules (7, 46). Despite observed extracellular MBP deposition, granule-mobilized MBP had not previously been detectable within eosinophil cytoplasmic vesicles in association with eosinophil-associated diseases. For instance, in human atopic dermatitis, while extracellular deposition of MBP was observed in parallel with ultrastructural changes within eosinophils indicative of PMD (granule core lucencies and vesicles budding from granules), for likely technical reasons, MBP was not demonstrable within eosinophil secretory vesicles (28). Prior MBP localization by immunoEM has been evaluated only by post-embedding techniques (28, 47) i.e., when labeling is performed after dehydration and resin embedding, steps that may affect epitope preservation in membrane microdomains. In addition, these techniques used larger sized gold particles (around 15 nm) conjugated to polyclonal Abs (28, 47) that might also compromise subcellular localization of MBP in vesicular compartments. In contrast, our findings used a pre-embedding technique combined with a smaller Fab secondary Ab fragment conjugated with very small gold particles to facilitate access to vesicles. These immunogoldEM techniques were complemented with additional experimental approaches.

To evaluate potential PMD-mediated release of granule MBP from human eosinophils, we studied eosinophils from normal and eotaxin-activated human eosinophils. Eotaxin stimulation of eosinophils *in vitro* elicits gross alterations in granule ultrastructure typical of PMD (25, 37), as well-recognized to occur within activated eosinophils *in vivo* (19, 21), including the mobilization of granule core and/or matrix contents. We found that both unstimulated and *in vitro*-eotaxin-stimulated eosinophils demonstrated vesicular trafficking of MBP. A prominent system of secretory vesicles from specific granules to plasma membrane was consistently labeled for MBP. This important process has likely been previously underestimated because of technical issues — inadequate preservation of vesicles and/or inability of Ab access to them. In fact, using different technical approaches for cell preparation before labeling we were able to detect two different MBP labeling patterns. Our present immunofluorescent findings showed that extragranular sites for MBP can be detected only in wet preparations of eosinophils (Fig 1). MBP detection within granules by immunofluorescence depended on cell drying which favors Ab access to the compacted crystalline core, but likely eliminates the vesicular pool. It is important to note that while our EM approach using pre-embedding labeling with mAbs strongly favored vesicle labeling, it did not enable optimal Ab access to the intact crystalline cores. Granules showing different degrees of core disarrangement, a process that seems fundamental for MBP release by PMD, were densely labeled for this cationic protein.

MBP was consistently detected in large tubular vesicles (EoSVs) (Fig 3), a vesicular system actively formed when eosinophils are stimulated with classical eosinophil agonists (16). Moreover, we demonstrate that the total numbers of EoSVs are significantly increased

within HES eosinophils (Fig. 4A), which are typically activated (38), compared to normal donors. These findings are important because they would explain the reason for the loss of electron dense cores (enriched in crystallized MBP) observed in tissue eosinophils from a range of disorders, such as Crohn's disease, eosinophilic gastroenteritis and HES (9-11). In fact, deposition of MBP can be demonstrated in affected tissues of patients with HES (48) and vesicular trafficking is likely involved in this secretory mechanism.

Increasing evidence has shown that large tubular carriers provide an additional mechanism to rapidly transport material between membranes in different secretory pathways and are also responsible for moving the bulk of the secretory traffic between distant compartments (49, 50). In eosinophils, EoSVs may be fundamental for the diversity of proteins that need to be rapidly transported from within these cells (36). Interestingly, MBP was observed preferentially within the lumen of EoSVs (Fig 3) in contrast to vesicle membrane-bound localization of eosinophil cytokines, such as IL-4 (16), a mechanism mediated by cognate, membrane-inserted receptors, as recently demonstrated by our group (17). The present finding showing preferential MBP labeling within vesicle lumina instead of at vesicle membranes provides more evidence for the occurrence of distinct cellular mechanisms involved in the mobilization of specific proteins from eosinophil granules. In fact, it is now apparent that eosinophils have a remarkable capacity to select proteins to be secreted from their cytoplasmic granules (25, 41), an event that underlies their multiple functions. An understanding of the intrinsic complexity of the eosinophil secretory pathway is beginning to emerge.

Rapid release of MBP may involve the presence of small storage/transient MBP sites (vesicular pools) in the cytoplasm as identified, for the first time, in the present work in unstimulated eosinophils (Fig 1). These extragranular sites appear to be relevant for the rapid release of small concentrations of MBP under cell activation without immediate disarrangement of the intricate crystalline cores within eosinophil specific granules. This is important because it may underlie eosinophil functions as an immunoregulatory cell. Indeed, the eosinophil role as a regulator of local immune and inflammatory responses has increasingly been appreciated (reviewed in Jacobsen, 2007 #418; Adamko, 2005 #126; Akuthota, 2008 #597}; and MBP, in addition to being a recognized molecule for defense against parasites, seems to be involved in the regulation of cytokine responses (51).

We also demonstrated that MBP-positive vesicles, including a large number of EoSVs, are tightly associated with specific granules. This finding supports our previous EM tomographic studies showing that EoSVs can arise from specific granules (16). Moreover, our present results shed more light on the understanding of the eosinophil secretory pathway. MBP-loaded vesicles maintain an effective interaction with the secretory granules and seem to be, at least in part, structurally linked to them. This interaction may be important for vesicular replenishment of MBP from granules. As noted here, the MBP-positive vesicular system was associated with a secretory pathway transporting MBP from eosinophil specific granules and not with a synthetic route from the *trans*-Golgi network, which was rarely labeled for MBP.

In summary, our studies have identified vesicular trafficking in activated human eosinophils that directs transport of MBP from secretory granules to the cell surface. We also demonstrated a pool of secretory vesicles as an additional intermediate storage site for MBP in unstimulated eosinophils. The recognition of PMD as a secretory process to release MBP is important to understand the pathological basis of allergic and other eosinophil-associated inflammatory diseases.

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Abbreviations:

Ab	antibody
FITC	fluorescein isothiocyanate
HES	hypereosinophilic syndrome
HBSS	Hanks buffered salt solution
mAb	monoclonal antibody
MBP	major basic protein
NGS	normal goat serum
PBS	phosphate buffered saline
PMD	piecemeal degranulation
TEM	transmission electron microscopy
RT	room temperature

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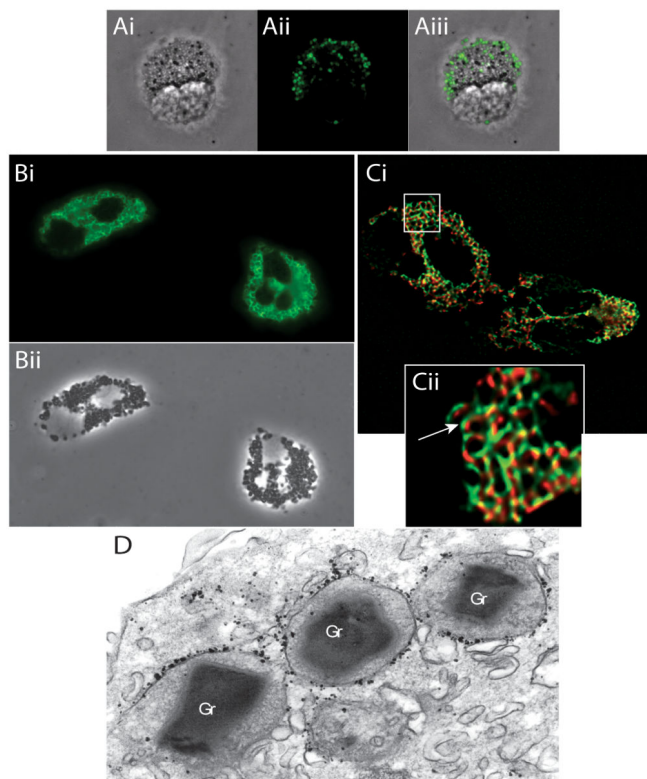


Figure 1. Major basic protein (MBP) and CD63 colocalize within human unstimulated eosinophils

(A) When eosinophils were allowed to dry in cytospin preparations, MBP immunodetection (green fluorescence) was observed within specific granules. A single eosinophil is shown in phase contrast (Ai), fluorescence microscopy (Aii) and an overlay of these two images (Aiii). (B) Fluorescence (Bi) and phase contrast (Bii) microscopy of an identical field of eosinophils kept in a wet agarose preparation show green MBP labeling often circumferentially peripheral to specific granules. (Ci) MBP (green) and CD63 (red) colocalizations, evaluated by deconvolution microscopy, reveal both fluorescent sites marginal to cytoplasmic granules, with MBP labeling often peripheral to membrane-bound CD63 (arrow). (Cii) is a higher magnification of the boxed area in (Ci). (D) Immunogold electron microscopy demonstrates CD63 labeling primarily on the cytoplasmic surface of eosinophil granule limiting membranes. Fluorescence images were acquired using a Retiga EXi cooled CCD camera coupled to a Provis AX-70 Olympus microscope and an UPlanApo objective (100 \times 1.35). Acquired stacks were further processed for deconvolution with the software Volocity 2.6. Fluorochromes used were Alexa 488 (MBP) and Alexa 594 (CD63). The electron micrograph was obtained on a CM 10 (Philips) transmission electron microscope at 60 Kv. Bar, 6 μ m (A, B); 5 μ m (Ci); 2 μ m (Cii); 400 nm (D). Gr, specific granules.

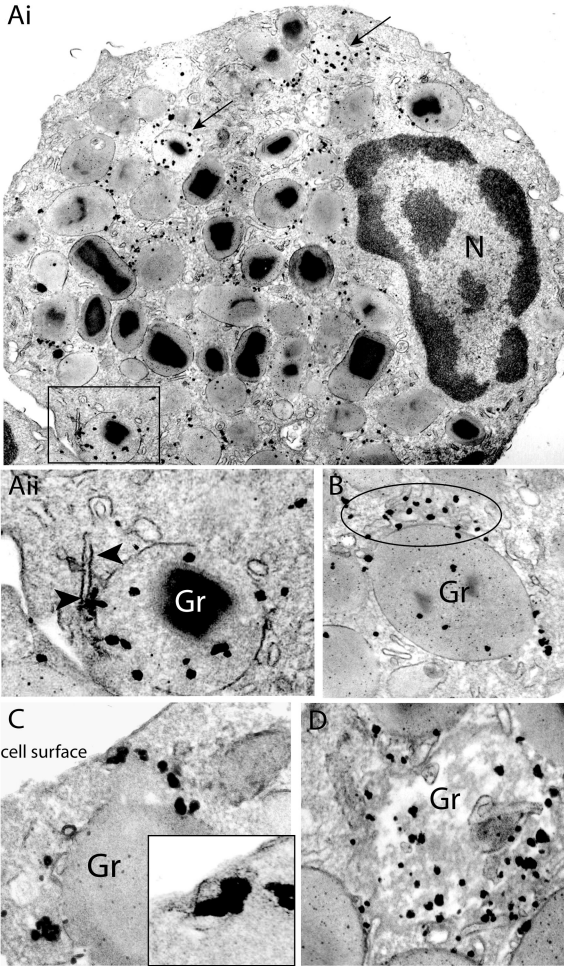


Figure 2. Major basic protein (MBP) is localized to specific granules and cytoplasmic vesicles in human eotaxin-stimulated eosinophils
 Immunonanogold electron microscopy revealed MBP-positive vesicles surrounding (A, B) and arising (C) from granules undergoing depletion of their contents. (Aii) is the boxed area of (Ai) and shows a tubular vesicle (arrowheads) at the surface of a specific granule. Note that MBP is present in the granule and in the attached vesicle. In (B), a pool of vesicles around a secretory granule is labeled for MBP. Observe that (Aii) and (B) show specific granules in progressive stages of content emptying, characterized by reduced electron density. In (C), a linear array of small, MBP-positive vesicles bud from a granule surface into the cell surface. The inset shows a vesicle fused with the plasma membrane at higher magnification. In (D), a granule, which demonstrates its disarranged core and matrix, is densely labeled for MBP. A similar image is seen at low magnification in Ai (arrows). Bar, 800 nm (Ai); 300 nm (Aii, C); 500 nm (B, D). Gr, specific granules.

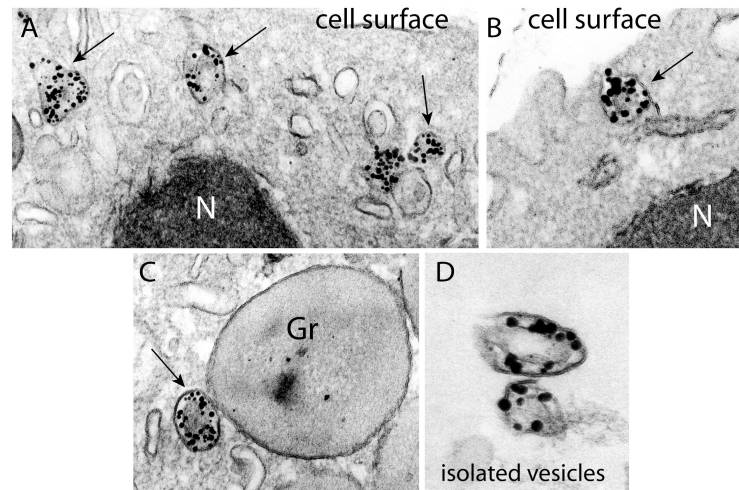


Figure 3. Large tubular carriers actively transport MBP

(A-C) Eosinophil sombrero vesicles - EoSVs - (arrows) are observed in the cytoplasm of activated eosinophils by transmission electron microscopy (TEM) after immunogold labeling for MBP. Vesicles are seen beneath the plasma membrane in the cytoplasm (A), fused with the plasma membrane (B) and attached to an enlarged partially empty granule (C), typically indicative of PMD. In (D), EoSVs, isolated by subcellular fractionation from unstimulated eosinophils, are densely labeled for MBP. Note that MBP is preferentially localized within the vesicle lumen. Eosinophils were stimulated by eotaxin as described in Material and Methods. Bar, 400 nm (A); 230 nm (B); 250 nm (C); 200 nm (D). Gr, specific granules; N, nucleus.

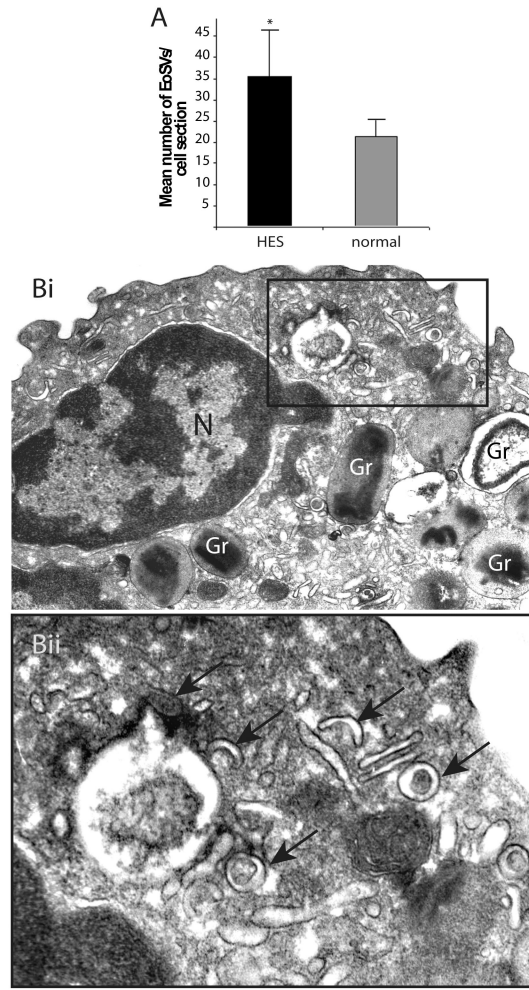


Figure 4. Eosinophil Sombrero Vesicles (EoSvs) are significantly increased within HES eosinophils

(A) Significant increases in EoSv numbers are observed within eosinophils from a hypereosinophilic patient compared to normal donors (* $p < 0.05$). (Bi and Bii) The boxed area shows EoSvs (arrows), with typical morphology, close or associated with an emptying specific granule. A total of 958 EoSvs were counted in 35 cell sections showing the entire cell profile ad nucleus. Bar, 600 nm (Bi); 250 nm (Bii). Gr, specific granules; N, nucleus; HES, hypereosinophilic syndrome.

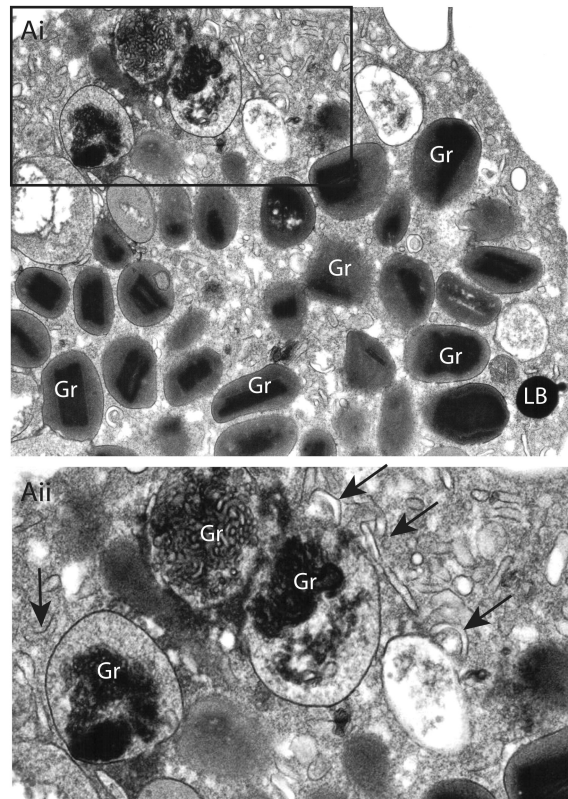


Figure 5. Ultrastructure of an eosinophil from a hypereosinophilic patient

(Ai) Specific granules (Gr) with typical crystalloid cores are observed together with enlarged granules with disassembled cores (boxed area in higher magnification in Aii). Arrows indicate Eosinophil Sonbrero Vesicles (EoSvs). Bar, 800 nm (Ai); 300 nm (Aii). Gr, specific granules; LB, lipid body.

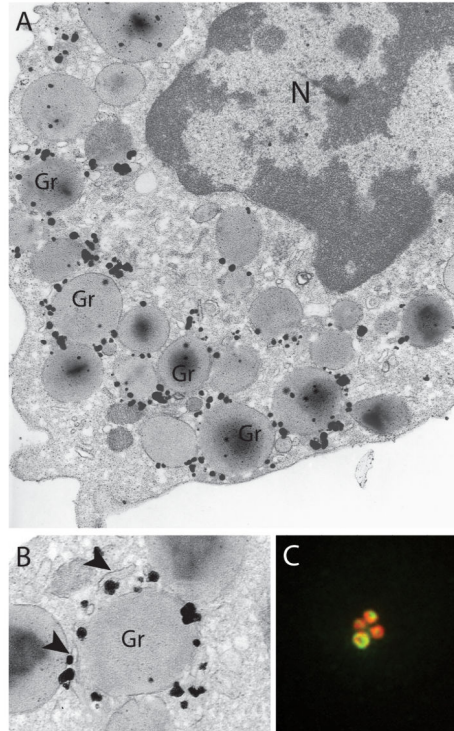


Figure 6. A major pool of MBP-loaded vesicles is tightly associated with secretory granules in unstimulated eosinophils

(A) Immunonanogold electron microscopy reveals MBP labeling within and around specific granules in an unstimulated eosinophil. (B) In higher magnification, labeling around granules is imaged as a consistent pool of MBP-positive vesicles. Labeling is observed in both round small and large vesiculotubular (arrowheads) carriers. Note that the vesicles are closely associated with the granule. (C) A cluster of free granules released from a disrupted eosinophil after double immunolabeling for CD63 and MBP show fluorescence for MBP (green) peripheral to CD63 (red) in some granules. Fluorescence image was acquired using a Retiga EXi cooled CCD camera coupled to a Provis AX-70 Olympus microscope and an UPlanApo objective (100 \times 1.35). Fluorochromes used were Alexa 488 (MBP) and Alexa 594 (CD63). Bar, 600 nm (A); 250 nm (B); 2 μ m (C). Gr, specific granules; N, nucleus.

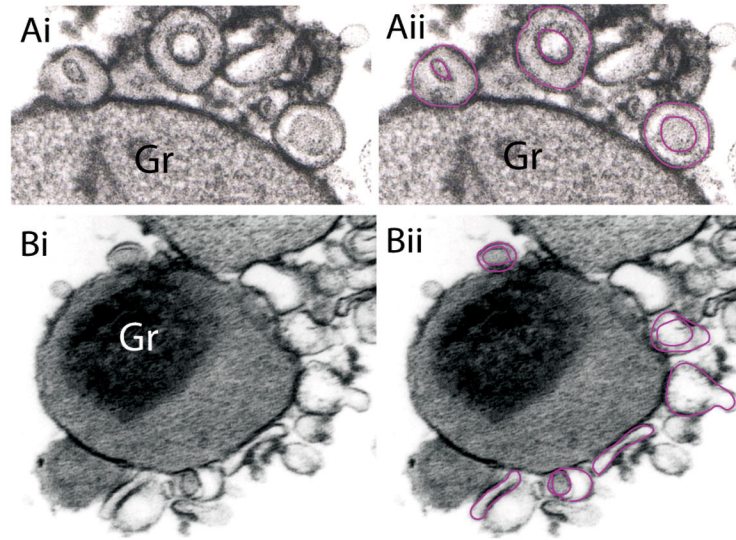


Figure 7. Ultrastructure of isolated eosinophil granules

(Ai, Bi) Transmission electron microscopy of a granule-enriched fraction isolated by subcellular fractionation shows typical EoSVs, the lumens of which are highlighted in pink in (Aii and Bii). EoSVs are imaged as open, curved, tubular-shaped structures. EoSVs are seen around and in close association with isolated granules. Granules were isolated and fixed for optimal membrane preservation as described in Material and Methods. Bar, 250 nm (A, B). Gr, specific granules.