RESEARCH PAPER

OPEN ACCESS OPEN ACCESS

Taylor & Francis

Taylor & Francis Group

Sensitivity and specificity evaluation of multiple neurodegenerative proteins for Creutzfeldt-Jakob disease diagnosis using a deep-learning approach

Sol Moe Lee ^[]^{a,b}, Jae Wook Hyeon^a, Soo-Jin Kim^b, Heebal Kim^b, Ran Noh^a, Seonghan Kim^a, Yeong Seon Lee^a, and Su Yeon Kim^a

^aDivision of Bacterial Disease Research, Center for Infectious Diseases Research, Korea National Institute of Health, Centers for Disease Control and Prevention, Cheongju-si, Chungcheongbuk-do, South Korea; ^bDepartment of Agricultural Biotechnology and Research Institute of Agriculture and Life Sciences, Seoul National University, Seoul, South Korea

ABSTRACT

The diagnosis of sporadic Creutzfeldt-Jakob disease (sCJD) can only be confirmed by abnormal protease-resistant prion protein accumulation in post-mortem brain tissue. The relationships between sCJD and cerebrospinal fluid (CSF) proteins such as 14–3-3, tau, and α -synuclein (a-syn) have been investigated for their potential value in pre-mortem diagnosis. Recently, deeplearning (DL) methods have attracted attention in neurodegenerative disease research. We established DL-aided pre-mortem diagnostic methods for CJD using multiple CSF biomarkers to improve their discriminatory sensitivity and specificity. Enzyme-linked immunosorbent assays were performed on phospho-tau (p-tau), total-tau (t-tau), a-syn, and β -amyloid (1–42), and western blot analysis was performed for 14-3-3 protein from CSF samples of 49 sCJD and 256 non-CJD Korean patients, respectively. The deep neural network structure comprised one input, five hidden, and one output layers, with 20, 40, 30, 20 and 12 hidden unit numbers per hidden layer, respectively. The best performing DL model demonstrated 90.38% accuracy, 83.33% sensitivity, and 92.5% specificity for the three-protein combination of t-tau, p-tau, and a-syn, and all other patients in a separate CSF set (n = 15) with other neuronal diseases were correctly predicted to not have CJD. Thus, DL-aided pre-mortem diagnosis may provide a suitable tool for discriminating CJD patients from non-CJD patients.

Introduction

Creutzfeldt-Jakob disease (CJD) is the most common human prion disease, which is characterized by the abnormal accumulation of misfolded prion protein (PrP^{Sc}), thus affecting the central nervous system [1,2]. The global incidence of CJD diagnosis is approximately 1 per million individuals per year. The most common form is sporadic CJD (sCJD), which accounts for about 85–95% of all known cases.

A definitive diagnosis of sCJD requires histological analysis of brain tissues obtained during autopsy [3], but it cannot always clinically distinguish CJD from other rapidly progressive neurodegenerative disorders in living patients, since these diseases share several clinical characteristics [4–6]. Therefore, there has been extensive research effort devoted to identifying cerebrospinal fluid (CSF) protein biomarkers for the premortem diagnosis of sCJD [7–10]. Indeed, diagnosis using CSF biomarkers has shown several benefits of reducing the diagnostic turn-around time with reproducible data without any risk of damage to the patient's brain. One of these candidate biomarkers for sCJD is the 14-3-3 protein, showing diagnostic sensitivities ranging from 61% to 96% and specificities ranfrom 67% to 95% [11–16]. Moreover, ging quantification of the CSF protein α -synuclein (a-syn) using an in-house enhanced chemiluminesence-based enzyme-linked immunosorbent assay (ELISA) was recently reported to be an excellent approach for sCJD diagnosis [10,17]. The total tau (t-tau) and phospho-tau (p-tau) levels, along with the p/t-tau ratio have also been suggested as clinically useful diagnostic markers [18,19]; however, these CSF biomarkers have not shown significantly high sensitivity or specificity in our experience with Korean patients with CJD, which is a major limitation of these biomarkers for this population.

An alternative diagnosis method is a real-time quaking-induced conversion (RT-QuIC) assay, which is a prion protein conversion assay that allows for the

ARTICLE HISTORY

Received 10 May 2019 Revised 26 June 2019 Accepted 29 June 2019

KEYWORDS

Deep learning; Tau; α-synuclein; 14-3-3; β-amyloid; prion diseases; Creutzfeldt-Jakob disease

CONTACT Su Yeon Kim 🔯 tenksy@korea.kr 🔄 Division of Bacterial Disease Research, Center for Infectious Diseases Research, Korea National Institute of Health, Centers for Disease Control and Prevention, Cheongju-si, Chungcheongbuk-do 28159, South Korea

Supplemental data for this article can be accessed here.

 $[\]ensuremath{\textcircled{\circ}}$ 2019 The Author(s). Published by Informa UK Limited, trading as Taylor & Francis Group.

This is an Open Access article distributed under the terms of the Creative Commons Attribution License (http://creativecommons.org/licenses/by/4.0/), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

detection of the abnormal amyloid form of PrP, and has been reported to exhibit high sensitivity and specificity for sCJD in the range of 76.5–97% and 99–100%, respectively [20–23].

Machine learning (ML) and deep learning (DL), which are subfields of AI, are powerful tools for extracting patterns and characteristics from big data using neural networks [24-26]. In contrast to traditional ML, DL uses multiple neural networks with many hidden layers. For the past decade, DL had remarkable successes in various fields such as computer vision [27,28], natural language processing [29,30], and audio signal processing [31]. Recently, it has gained attention for disease diagnosis owing to their good representational power [32], including research on CJD [33-35]. Specifically, Morabito et al. [35] reported that a DL-based analysis, which used electroencephalography signals, showed 89% accuracy in discriminating between CJD patients and patients with other forms of rapid progressive dementia, with 92% sensitivity and 89% specificity.

The aims of the present study were to establish a pre-mortem diagnostic method using a DL approach, which would (1) facilitate diagnostic decision making for CJD, (2) improve accuracy compared to conventional pre-mortem diagnostic methods using CSF protein markers, and (3) help to identify more CJD-related CSF biomarkers as landmarks. We then tested the ability of the established DL technique to discriminate CJD patients from non-CJD patients based on detection levels of the key CSF biomarkers 14–3-3, a-syn, amyloid beta (A β), t-tau, and p-tau.

Materials and methods

Subjects

We collected CSF samples from 49 patients with sCJD (5 definite and 44 probable) and 11 with possible sCJD, and 256 non-CJD patients (these cases were referred to as "suspected CJD" to the KNIH, but were not confirmed as CJD) ascertained through routine surveillance by the Korea National Institute of Health (KNIH) according to diagnostic criteria (Table 1). Sequencing of all patient samples revealed a methionine residue at codon 129 of the prion protein-coding gene PRNP (129M/M homo-zygotes). These samples, except for those of 11 patients with possible sCJD, were used as the training, validation, and test sets (test set_A) for ML and DL analysis. In total, 15 CSF samples from patients who were diagnosed

Table	 Patients' 	classification	and	clinical	features	of	the
definit	e, probable,	possible CJD	cases	and ot	her neuro	nal	dis-
eases	patients used	d in this study	/.				

	No. of	·
Groups	patients	Diagnostic criteria/Symptoms
Definite CJD	5	Diagnosed by neuropathological techniques, immunohistochemcal detection of PrP ^{Sc} of autopsied or biopsied brain tissues from patients with probable or possible sCJD
Probable CJD	44	Rapid progressive dementia of less than 2 years' duration and at least two of the following features such as myoclonus, visual or cerebellar disturbance, pyramidal, extrapyramidal dysfunction, akinetic mutism, with a EEG data, and/or a positive result of 14–3-3 assav
Possible CJD	11	Progressive dementia and at least two of the following features such as myoclonus, visual or cerebellar disturbance, pyramidal, extrapyramidal dysfunction, akinetic mutism, without a EEG data, and/or a positive result of 14–3-3 assay
Non-CJD	256	Cases referred to as "suspected CJD" to the KNIH, but were not fulfilling the criteria of probable or possible CJD
Neuronal	3	Hydrocephalus, unspecified
diseases (test	4	Normal Pressure Hydrocephalus
set_B)	1	Multi-system degeneration
	1	Alzheimer's Disease
	1	Nila cognitive disorder
	1	Spinal muscular atrophy, and related syndromes
	1	Ćerebral palsy
	2	Other disorders of brain

with other types of neuronal diseases were also collected and used as a separate test set (test set_B).

This study was approved by the Institutional Review Board (IRB) of the Korea Centers for Disease Control and Prevention (IRB No. 2017–03-09-C-A), and all the experiments were performed in accordance with relevant guidelines of the IRB. Written informed consent was obtained from the patients or their legal guardians.

Biochemical analysis

The 14–3-3 protein levels in the CSF samples from the patients were determined by western blot analysis as previously described [8]. Quantitative determination of CSF p-tau, t-tau, $A\beta$, and a-syn proteins was conducted with the INNOTEST* hTAU Ag (Fujirebio, Gent, Belgium), INNOTEST* PHOSPHO-TAU(181P) (Fujirebio, Gent, Belgium), INNOTEST* β -AMYLOID(1–42) (Fujirebio, Gent, Belgium), and SensoLyte Anti- α -Synuclein (Human, Mouse, Rat) ELISA Kit (AnaSpec, Inc., Fremont, CA), respectively, according to the manufacturers' instructions. The p-tau to t-tau ratio (p/t-tau ratio) was also calculated based on these results.

Data handling and scoring

To standardize the quantitative data of protein levels for the different candidate biomarkers, a positive band of 14–3-3 protein in the western blot was given a score of 2, whereas a negative or weakly positive finding was given a score of 1. When the a-syn or A β concentration was below the detection limit of the ELISA kits (7.813 ng/µl or 63 ng/µl, respectively), the case was given a score of 1.

To resolve the detrimental effect of imbalanced data on the performance of DL algorithms, we adopted an 'oversampling' approach [36–38]. Oversampling was performed after splitting the original dataset into the 'training with validation set' and 'test set_A.' This resulting dataset was then further split into 'training with validation set (n = 253)' and 'test set_A (n = 52)'. In the training with validation set, the results of the minor group (i.e. CJD patients; n = 37) were replicated five times (500%) to achieve a 1:1 ratio between the CJD and non-CJD groups (n = 216). Subsequently, the training with the validation set was split into separate training and validation sets with a 9:1 ratio, respectively.

ML and DL analysis

ML analysis was performed using Waikato Environment for Knowledge Analysis 3.8.2 (WEKA) [39]. Evaluation of the J48 decision tree (confidence factor = 0.25, minimal number of objects = 2), naïve Bayes (the number of decimal places = 2), and the random forest classifier (the number of trees = 100) were performed using a default setting. Support vector machine (SVM) with sequential minimal optimization (SMO) evaluation was performed using default settings (C value = 1) with radial basis function kernel (gamma = 10). The Keras (http://keras.io) neural network library and the TensorFlow (https://www.tensor flow.org) software library were used for deep neural network (DNN) construction. Specifically, we used a multi-layer feed-forward artificial neural network with the standard back-propagation algorithm to perform binary classification [40,41]. For each DNN in the resulting ensemble, multiple hyper-parameters were adjusted, including the number of hidden layers, number of neurons in each layer, choice of activation function, choice of optimization method, and regularization techniques.

The best DNN structure consisted of one input, five hidden, and one output layers, with 20, 40, 30, 20 and 12 hidden unit numbers of each hidden layer, respectively. Ultimately, two DNNs were used to

discriminate between CJD and non-CJD patients. All of the layers were subjected to a kernel initializer with the 'random_normal' option for normalization of each layer. ReLu [42,43] was used as the activation function in each hidden layer, and softmax was used in the last layer. The loss function was binary crossentrophy [44], and Nesterov Adam (NAdam) [45] with default values (learning rate = 0.002, beta_1 = 0.9, and beta_2 = 0.999) was used as the optimizer of the loss function. Dropout [46] was used to overcome overfitting with a probability of 0.1 after each layer. Neural networks were constructed using the NN-SVG tool (alexlenail.me/NN-SVG/index.html). The best DNN structure was also used for two- or threevariable combination analysis from 6 variables. Therefore, 35 combinations were tested to find the best combination set for improving the discrimination accuracy and identifying potential 'landmark CSF markers'.

Statistical analysis

The performance of the ML and DL was evaluated according to the quantitative parameters of the true positive (TP), true negative (TN), false positive (FP), and false negative (FN) rates, which were then used to calculate the accuracy, sensitivity, and specificity according to the following equations:

Accuracy = (TP + TN)/(TP + TN + FP + FN) Sensitivity = TP/(TP + FN) Specificity = TN/(TN + FP)

The area under the receiver operating characteristic curve (AUC) was calculated to determine the best model.

10-fold cross-validation classification performance

Cross-validation estimation was performed using the oversampled dataset which used in ML and DL analysis to determine the predictive performance of the model and for tuning hyper-parameters as described previously [47,48]. In brief, the oversampled dataset excluding test set_A (n = 52) was split into 10 subsets. A repetition consisted of 10 iterations, and one fold was used as validation data for each iteration, while the other folds were used during training performance. After the initial training period, the performance of the network was analysed based on the validation data for tuning hyper-parameters. The training process was repeated 50 times to obtain a stable result. Each iteration was performed independently, so that each iteration had no prior knowledge about the chosen learning models in the other iterations.

Results

The concentrations of six CSF markers (14–3-3, t-tau, p-tau, p/t-tau ratio, $A\beta$, and a-syn) in 5 definite, 44 probable, 11 possible CJD patients, and 256 non-CJD patients, which were determined using ELISA and western blotting, are summarized in Supplementary Table 1. A correlation heat map among the six biomarkers and sCJD patients, except for patients with possible CJD and non-CJD patients, is presented in Figure 1. None of the biomarkers showed a particularly strong correlation. Figure 2 shows the discrimination plots of the patients in the dataset according to the six biomarkers, in which each pixel represents a given CSF protein level or 14–3-3 western blot result (positive or negative): orange pixels depict CJD patients and blue pixels represent non-CJD patients. No bivariate combination was found to effectively separate the two groups. The diagnostic performances of 14–3-3, t-tau, the p/t-tau ratio, and a-syn were assessed according to the reported diagnostic criteria (Table 2). The sensitivity and specificity of the 14–3-3 protein analysis were 67.35% and 67.58%, respectively, and the age in definite and probable CJD cases with 14–3-3 positivity (mean age 69.94 ± 10.73 years) was higher than those of cases with 14–3-3 negativity (mean age 61 ± 11.6 years, supplementary Table 2).

The highest diagnostic specificity in non-CJD patients was 81.6% using t-tau, and in neuronal diseases patients was 93.33% using a-syn level or t-tau combination test with p/t-tau ratio. The diagnostic sensitivity in possible CJD patients using the a-syn protein was lower than that of other proteins in these patients. The concentration of t-tau, p-tau and a-syn of the test set_B are described in Supplementary Table 1.



Figure 1. Heat map of the correlation of t-tau, p-tau, p/t-tau ratio, A β , a-syn and 14–3-3 levels with patients. The colour of each square depicts the correlation level, ranging from black (negative correlation) to red (intermediate correlation value) to white (positive correlation value).



Figure 2. Discrimination plot for discrimination between CJD (orange) and non-CJD (blue) patients based on values of six CSF biomarkers.

The performance of each classifier is described in Table 3. In the machine learning analysis using training with a validation set, the highest accuracy was observed in the analysis using J48 and the random forest classifier (78.85%). However, the machine analysis sensitivity was 41.67% or 33.33%. Although the highest specificity was observed in the analysis using the random forest classifier (92.5%), the related sensitivity was the lowest compared with analysis using the other classifiers. The best performing classifier was the DNN model, and it showed 86.54% discrimination accuracy and an AUC value of 0.90 (Figure 3).

To improve the discrimination and identify potential 'landmark CSF markers' for discriminating between CJD and non-CJD patients, the structure of the bestperforming model described above was used for analysis with two- or three-variable combinations. As shown in Table 4, AUC is not necessarily equivalent to the accuracy measured at a threshold because an AUC score considers the sensitivity and specificity of the various threshold changes in each classification model. The best accuracy and AUC values were obtained for the three-marker combination of t-tau, p-tau, and a-syn. The scores for the test set_A were 90.38% (47/ 52) accuracy, 83.33% (10/12) sensitivity, and 92.5% (37/ 40) specificity. Using the established discrimination model, 10-fold cross-validation was performed. The validation was repeated 50 times, and the AUC and accuracy scores were calculated to obtain a stable result (Supplementary Table 3). The model was then reevaluated using test set_B, which only consisted of CSF samples from patients with neuronal diseases, and all the samples were predicted correctly to not have CID.

		Sensitivity % (TP)	/all)	Specifi		
Markers	Criteria	Definite and probable	Possible	Non-CJD	Neuronal diseases	References
14–3-3	14–3-3 positive ^a	67.35	54.54	67.58	N/A	[52]
		(33/49)	(6/11)	(173/256)		
t-tau	> 1,000 pg/mL	59.18	72.73	77.34	86.67	[49]
		(29/49)	(8/11)	(198/256)	(13/15)	
t-tau	> 1,300 pg/mL	53.06	72.73	80.86	86.67	[50]
		(26/49)	(8/11)	(207/256)	(13/15)	
t-tau with p/t-tau ratio	t-tau > 1,000 pg/ml with	55.1	72.73	79.3	93.33	[49]
·	p/t tau ratio < 0.04	(27/49)	(8/11)	(203/256)	(14/15)	
Αβ	< 445 pg/mL	69.39	81.82	28.91	N/A	[51]
		(34/49)	(9/11)	(74/256)		
a-syn	> 820 pg/mL	55.1	27.27	78.91	93.33	[10]
,	13	(27/49)	(3/11)	(202/256)	(14/15)	
a-syn	> 680 pg/mL	55.1	27.27	74.61	86.67	[17]
,	15	(27/49)	(3/11)	(191/256)	(13/15)	

Table 2. Diagnostic performance of various cerebrospinal fluid markers for Creutzfeldt-Jakob disease according to defined criteria.

 $^{\rm a} {\rm Weak}$ positive result considered as negative N/A: not analysed

Tab	le 3. Eval	luation of	classifier p	performances (test set A	, 12 of	CJD cases a	and 40 of	f non-CJD	patients).	
-----	------------	------------	--------------	----------------	------------	---------	-------------	-----------	-----------	------------	--

		· _ ·		
Classifier	Accuracy	Sensitivity	Specificity	AUC
SVM	76.92% (40/52)	66.67% (8/12)	80% (32/40)	0.73
Decision tree (J48)	78.85% (41/52)	41.67% (5/12)	90% (36/40)	0.8
Naïve Bayes	76.92% (40/52)	66.67% (8/12)	80% (32/40)	0.72
Random Forest	78.85% (41/52)	33.33% (4/12)	92.5% (37/40)	0.82
DNN	86.54%(45/52)	83.33%(10/12)	87.5% (35/40)	0.90

Accuracy: TP+TN/No. of cases in test set_A

Sensitivity: TP/No. of CJD cases in test set_A

Specificity: TN/No. of non-CJD cases in test set_A



Figure 3. Deep neural network showing the best performance for CJD discrimination. Our network structure consisted of one input, five hidden, and one output layer. The five hidden layers consisted of 20, 40, 30, 20, and 12 hidden unit numbers of each layer, respectively. The last two output units were used to distinguish between CJD and non-CJD patients.

Table 4. Analysis of two- and three-protein combinations for discrimination between CJD and non-CJD patients. All values were calculated using the oversampled validation set and test set_A.

Combinations	Accuracy in validation set	AUC in validation set	Accuracy in test set_A	AUC in test set_A
t-tau and p-tau	70.73%	0.83	69.23%	0.71
t-tau and p/t-tau ratio	65.85%	0.77	73.08%	0.76
t-tau and Åβ	80.49%	0.91	75%	0.76
t-tau and a-syn	80.49%	0.91	82.69%	0.86
t-tau and 14–3-3	63.41%	0.80	69.23%	0.79
p-tau and p/t-tau ratio	65.85%	0.83	61.54%	0.58
p-tau and Aβ	51.22%	0.51	46.15%	0.55
p-tau and a-syn	58.54%	0.72	65.38%	0.84
p-tau and 14–3-3	65.85%	0.71	65.38%	0.60
p/t-tau ratio and Aβ	48.78%	0.46	48.08%	0.52
p/t-tau ratio and a-syn	53.66%	0.62	59.62%	0.84
p/t-tau ratio and 14–3-3	68.29%	0.77	69.23%	0.75
Åβ and a-syn	65.85%	0.76	69.23%	0.74
Aβ and 14–3-3	56.1%	0.76	38.46%	0.42
a-syn and 14–3-3	60.98%	0.69	75%	0.90
t-tau, p-tau and p/t-tau ratio	85.21%	0.85	78.85%	0.85
t-tau, p-tau and Åβ	78.05%	0.88	75%	0.74
t-tau, p-tau and a-syn	87.8%	0.95	90.38%	0.88
t-tau, p-tau and 14-3-3	78.05%	0.91	71.15%	0.71
t-tau, p/t-tau ratio and Aβ	75.61%	0.85	80.77%	0.77
t-tau, p/t-tau ratio and a-syn	75.61%	0.90	75%	0.84
t-tau, p/t-tau ratio and 14–3-3	70.73%	0.82	76.92%	0.78
t-tau, Åβ and a-syn	78.05%	0.85	86.54%	0.86
t-tau, Aβ and 14–3-3	87.8%	0.95	78.85%	0.75
t-tau, a-syn and 14–3-3	87.8%	0.98	78.85%	0.86
p-tau, p/t-tau ratio and Aβ	46.34%	0.54	51.92%	0.60
p-tau, p/t-tau ratio and a-syn	68.29%	0.72	71.15%	0.77
p-tau, p/t-tau ratio and 14–3-3	70.73%	0.84	71.15%	0.65
p-tau, Aβ and a-syn	73.17%	0.88	84.62%	0.91
p-tau, Aβ and 14–3-3	58.54%	0.75	40.38%	0.44
p-tau, a-syn and 14–3-3	65.85%	0.70	71.15%	0.74
p/t-tau ratio, Aβ and a-syn	65.85%	0.83	69.23%	0.72
p/t-tau ratio, Aβ and 14–3-3	56.1%	0.55	26.92%	0.40
p/t-tau ratio, a-syn and 14–3-3	60.98%	0.62	71.15%	0.82
Aβ, a-syn and 14–3-3	63.41%	0.76	67.31%	0.75

Accuracy: TP+TN/No. of cases in test set_A

Sensitivity: TP/No. of CJD cases in test set_A

Specificity: TN/No. of non-CJD cases in test set_A

Discussion

At present, sCJD can only be diagnosed with certainty after a patient is deceased, based on histological examination of the brain tissue. Analysis of protein biomarkers related to CJD has proven to be a potentially useful alternative pre-mortem diagnosis method. The 14-3-3 protein has been reported to be a biomarker for rapid progressive neurodegenerative disorders, including CJD. In our data, analysis for CSF 14-3-3 revealed a dependency on age; the sensitivity was higher in definite and probable sCJD cases than in possible sCJD cases. However, the sensitivity and specificity of CSF 14-3-3 in sCJD cases were found to be lower than those reported previously [17,52] but were similar to those indicated in some studies in Japan [22], China [53] and USA [54,55]. The lower sensitivity and specificity in our results compared to those of other studies described above is assumed to be due to the following reasons. First, the 129 MM2 type, for which the 14-3-3 protein is reported to have relatively low sensitivity, was not excluded while calculating the sensitivity and specificity of 14-3-3 protein analysis. Second, experimental differences and/or laboratory errors could

have been the contributing factors. Furthermore, 14–3-3 protein concentration could have changed in patients from the date of collection of the CSF sample to date of discrimination analysis.

The specificities of tau and a-syn analysis in neuronal disease patients, which were used as test set_B, were higher (86.67%–93.33%) than these values in the non-CJD group (74.61%–80.86%). The possibility of misdiagnosis due to similarities of symptoms could not be excluded in some cases in the non-CJD group. However, the protein concentration patterns of the non-CJD group significantly differed from those of the sCJD cases.

Here, we performed ML and DL-based analysis using combination of CSF markers as a pre-mortem diagnostic method. Our initial ML and DL model had an imbalanced learning problem owing to the lower number of CSF samples from CJD patients than that from non-CJD patients, which would result in a dominant influence of the major group on the analysis. We chose to use an oversampling method rather than an under-sampling method to resolve this imbalance and avoid losing information on the larger group (non-CJD) after drastic reduction to balance the ratio between the numbers of sCJD and non-CJD cases. In particular, an ensemble of DNNs using ELISA data of t-tau, p-tau, and a-syn outperformed (90.38% accuracy, 83.33% sensitivity and 92.5% specificity) the diagnostic performance of diagnostic performance of any single CSF marker described in Table 2 and other combination with DL analysis.

RT-QuIC analysis has been used to detect PrP^{Sc} in CSF samples directly, showing high diagnostic specificity [20,21,56]. However, the sensitivity of this method might rely on the specific PrP^{Sc} concentration in each CSF sample. Thus, cases in which the PrP^{Sc} concentration in a CSF sample is below the detection limit of the RT-QuIC assay, the DL-aided discrimination method could be an alternative pre-mortem CJD diagnosis method.

Despite the good performances of the models developed in this study, the discrimination between CJD and non-CJD patients was not completely accurate. We consider the following limitations of the proposed method that may have contributed to this result. First, noise among samples might have been derived from misdiagnosis for cases in which the symptoms of CJD are misinterpreted, since they are similar to those of other neurodegenerative disorders [57,58]. Second, additional combinations with other proteins associated with neurodegenerative disorders such as S-100 should be tested. Third, the hyper-parameters combination might not have been optimized since there is currently no established optimization method. In addition, the relatively small sample size might have limited the ability to construct a robust algorithm to effectively discriminate between CJD and non-CJD patients. Typically, deep learning analyses require an extremely large dataset; therefore, performing deep learning analysis with a relatively small dataset is a significant challenge. However, there are several methods to overcome the limitation of small sample size; we used the oversampling method [36,38] and 10-fold cross validation [47,48].

Although the DL-aided discrimination model warrants further improvement in performance and classification accuracy using stacked sample sizes via further ELISA data collection and/or extended analysis with other biomarkers for use as a pre-mortem diagnostic method, the DL-based model has several advantages for clinical application. First, it can easily handle large amounts of medical data. Second, CJD and non-CJD patients can be discriminated rapidly (within one or two days). Third, the results can be obtained consistently and reproducibly without requiring a specialist to conduct the laboratory tests. Overall, our findings could help to facilitate clinical decision-making.

Acknowledgments

This study was supported by a research grant (2017-NI52004-00) of the Korea Centers for Disease Control and Prevention. The biospecimens and data which used as test set_B (n=15) in this study were provided by the Gyeongsang National University Hospital and Pusan National University Hospital, members of the Korea Biobank Network.

Data availability statement

All data generated or analysed during this study are included in https://github.com/varamos/DNN_CJD and its supplementary files 1–10.

Disclosure statement

No potential conflict of interest was reported by the authors.

Funding

This work was supported by the Korea Centers for Disease Control and Prevention [2017-NI52004-00].

ORCID

Sol Moe Lee D http://orcid.org/0000-0002-6935-462X

References

- Prusiner SB. Molecular biology of prion diseases. Science. 1991 Jun 14;252(5012):1515–1522. PubMed PMID: 1675487.
- Prusiner SB. Novel proteinaceous infectious particles cause scrapie. Science. 1982 Apr 9;216(4542):136–144. PubMed PMID: 6801762.
- [3] Belay ED, Holman RC, Schonberger LB. Creutzfeldt-Jakob disease surveillance and diagnosis. Clin Infect Dis. 2005 Sep 15;41(6):834–836. PubMed PMID: 16107982.
- [4] Hainfellner JA, Wanschitz J, Jellinger K, et al. Coexistence of Alzheimer-type neuropathology in Creutzfeldt-Jakob disease. Acta Neuropathol. 1998 Aug;96(2):116–122. PubMed PMID: 9705125.
- [5] Woerman AL, Stohr J, Aoyagi A, et al. Propagation of prions causing synucleinopathies in cultured cells. Proc Natl Acad Sci U S A. 2015 Sep 1;112(35):E4949–58. PubMed PMID: 26286986; PubMed Central PMCID: PMC4568231.
- [6] Prusiner SB, Woerman AL, Mordes DA, et al. Evidence for alpha-synuclein prions causing multiple system atrophy in humans with parkinsonism. Proc Natl Acad Sci U S A. 2015 Sep 22;112(38):E5308–17. PubMed PMID: 26324905; PubMed Central PMCID: PMC4586853.
- [7] Skinningsrud A, Stenset V, Gundersen AS, et al. Cerebrospinal fluid markers in Creutzfeldt-Jakob disease. Cerebrospinal Fluid Res. 2008 Aug 27;5:14. PubMed PMID: 18727840; PubMed Central PMCID: PMC2531166.

- [8] Hyeon JW, Kim SY, Lee J, et al. Alternative application of Tau protein in Creutzfeldt-Jakob disease diagnosis: improvement for weakly positive 14-3-3 protein in the laboratory. Sci Rep. 2015 Oct 28;5:15283. PubMed PMID: 26507666; PubMed Central PMCID: PMC4623667.
- [9] Koscova S, Slivarichova DZ, Tomeckova I, et al. Cerebrospinal fluid biomarkers in the diagnosis of Creutzfeldt-Jakob disease in slovak patients: over 10-year period review. Mol Neurobiol. 2017 Oct;54 (8):5919–5927. 10.1007/s12035-016-0128-4. PubMed PMID: 27665282.
- [10] Llorens F, Kruse N, Karch A, et al. Validation of alpha-Synuclein as a csf biomarker for sporadic Creutzfeldt-Jakob Disease. Mol Neurobiol. 2018 Mar;55(3):2249–2257. 10.1007/s12035-017-0479-5. PubMed PMID: 28321768; PubMed Central PMCID: PMC5840235.
- [11] Soomro S, Mohan C. Biomarkers for sporadic Creutzfeldt-Jakob disease. Ann Clin Transl Neurol. 2016 Jun;3(6):465–472. PubMed PMID: 27547775; PubMed Central PMCID: PMC4892001.
- [12] Peoc'h K, Schroder HC, Laplanche J, et al. Determination of 14-3-3 protein levels in cerebrospinal fluid from Creutzfeldt-Jakob patients by a highly sensitive capture assay. Neurosci Lett. 2001 Apr 6;301 (3):167–170. PubMed PMID: 11257424.
- [13] Bahl JM, Heegaard NH, Falkenhorst G, et al. The diagnostic efficiency of biomarkers in sporadic Creutzfeldt-Jakob disease compared to Alzheimer's disease. Neurobiol Aging. 2009 Nov;30(11):1834–1841. PubMed PMID: 18339451.
- [14] Chohan G, Pennington C, Mackenzie JM, et al. The role of cerebrospinal fluid 14-3-3 and other proteins in the diagnosis of sporadic Creutzfeldt-Jakob disease in the UK: a 10-year review. J Neurol Neurosurg Psychiatry. 2010 Nov;81(11):1243–1248. PubMed PMID: 20855493.
- [15] Pennington C, Chohan G, Mackenzie J, et al. The role of cerebrospinal fluid proteins as early diagnostic markers for sporadic Creutzfeldt-Jakob disease. Neurosci Lett. 2009 May 8;455(1):56–59. PubMed PMID: 19429106.
- [16] Stoeck K, Sanchez-Juan P, Gawinecka J, et al. Cerebrospinal fluid biomarker supported diagnosis of Creutzfeldt-Jakob disease and rapid dementias: a longitudinal multicentre study over 10 years. Brain. 2012 Oct;135(Pt 10):3051–3061. PubMed PMID: 23012332; PubMed Central PMCID: PMC3470713.
- [17] Llorens F, Kruse N, Schmitz M, et al. Evaluation of alpha-synuclein as a novel cerebrospinal fluid biomarker in different forms of prion diseases. Alzheimers Dement. 2017 Jun;13(6):710–719. PubMed PMID: 27870938.
- [18] Skillback T, Rosen C, Asztely F, et al. Diagnostic performance of cerebrospinal fluid total tau and phosphorylated tau in Creutzfeldt-Jakob disease: results from the Swedish mortality registry. JAMA Neurol. 2014 Apr;71(4):476–483. PubMed PMID: 24566866.
- [19] Wang GR, Gao C, Shi Q, et al. Elevated levels of tau protein in cerebrospinal fluid of patients with probable Creutzfeldt-Jakob disease. Am J Med Sci. 2010 Oct;340 (4):291–295. PubMed PMID: 20881758.

- [20] Park JH, Choi YG, Lee YJ, et al. Real-time quaking-induced conversion analysis for the diagnosis of sporadic Creutzfeldt-Jakob disease in Korea. J Clin Neurol. 2016 Jan;12(1):101–106. PubMed PMID: 26541494; PubMed Central PMCID: PMC4712274.
- [21] Franceschini A, Baiardi S, Hughson AG, et al. High diagnostic value of second generation CSF RT-QuIC across the wide spectrum of CJD prions. Sci Rep. 2017 Sep 6;7 (1):10655. 10.1038/s41598-017-10922-w. PubMed PMID: 28878311; PubMed Central PMCID: PMC5587608.
- [22] Atarashi R, Satoh K, Sano K, et al. Ultrasensitive human prion detection in cerebrospinal fluid by real-time quaking-induced conversion. Nat Med. 2011 Feb;17(2):175–178. PubMed PMID: 21278748.
- [23] McGuire LI, Peden AH, Orru CD, et al. Real time quaking-induced conversion analysis of cerebrospinal fluid in sporadic Creutzfeldt-Jakob disease. Ann Neurol. 2012 Aug;72(2):278–285. PubMed PMID: 22926858; PubMed Central PMCID: PMC3458796.
- [24] Min S, Lee B, Yoon S. Deep learning in bioinformatics. Brief Bioinform. 2017 Sep 1;18(5):851–869. PubMed PMID: 27473064.
- [25] Ravi D, Wong C, Deligianni F, et al. Deep learning for health informatics. IEEE J Biomed Health Inform. 2017;21(1):4–21.
- [26] LeCun Y, Bengio Y, Hinton G. Deep learning. nature. 2015 May 28;521(7553):436. PubMed PMID: 26017442.
- [27] He K, Zhang X, Ren S, et al., editors. Deep residual learning for image recognition. Proceedings of the IEEE conference on computer vision and pattern recognition; Las Vegas (NV); 2016.
- [28] Choi Y, Choi M, Kim M, et al. Stargan: unified generative adversarial networks for multi-domain image-to-image translation. arXiv preprint. 2017;1711.
- [29] Bahdanau D, Cho K, Bengio Y Neural machine translation by jointly learning to align and translate. arXiv preprint arXiv:14090473SOR. 2014.
- [30] Gu X, Cho K, Ha J, et al. DialogWAE: multimodal response generation with conditional Wasserstein auto-encoder. arXiv preprint arXiv:180512352. 2018.
- [31] Van Den Oord A, Dieleman S, Zen H, et al., editors. WaveNet: A generative model for raw audio. SSW; 2016.
- [32] Libbrecht MW, Noble WS. Machine learning applications in genetics and genomics. Nat Rev Genet. 2015 Jun;16(6):321–332. PubMed PMID: 25948244; PubMed Central PMCID: PMC5204302.
- [33] Hung CI, Wang PS, Soong BW, et al. Blind source separation of concurrent disease-related patterns from EEG in Creutzfeldt-Jakob disease for assisting early diagnosis. Ann Biomed Eng. 2007 Dec;35(12):2168–2179. PubMed PMID: 17891454.
- [34] Wang PS, Wu YT, Hung CI, et al. Early detection of periodic sharp wave complexes on EEG by independent component analysis in patients with Creutzfeldt-Jakob disease. J Clin Neurophysiol. 2008 Feb;25(1):25–31. PubMed PMID: 18303557.
- [35] Morabito FC, Campolo M, Mammone N, et al. Deep learning representation from electroencephalography of early-stage Creutzfeldt-Jakob disease and features for differentiation from rapidly progressive dementia. Int J Neural Syst. 2017 Mar;27(2):1650039. PubMed PMID: 27440465.

- [36] He H, Garcia EA. Learning from Imbalanced Data. IEEE Trans Knowledge Data Eng. 2009 Sep;21(9):1263–1284.
 PubMed PMID: WOS:000268062400003; English.
- [37] Van Hulse J, Khoshgoftaar TM, Napolitano A Experimental perspectives on learning from imbalanced data. Proceedings of the 24th international conference on Machine learning; Corvalis (OR); 2007. p. 935–942.
- [38] Kim K-M, Jang H-Y, Zhang B-T. Oversampling-based ensemble learning methods for imbalanced data. KIISE Trans Comput Pract. 2014;20(10):549–554.
- [39] Hall M, Frank E, Holmes G, et al. The WEKA data mining software: an update. ACM SIGKDD Explorations Newsl. 2009;11(1):10–18.
- [40] Witten IH, Frank E, Hall MA, et al. Data mining: practical machine learning tools and techniques. Morgan Kaufmann; 2016.
- [41] Lin E, Kuo P-H, Liu Y-L, et al. A deep learning approach for predicting antidepressant response in major depression using clinical and genetic biomarkers. Front Psychiatry. 2018 Jul 6;9:290. PubMed PMID: WOS:000437691100001; English.
- [42] Nair V, Hinton GE, editors. Rectified linear units improve restricted boltzmann machines. Proceedings of the 27th international conference on machine learning (ICML-10); Haifa (Israel); 2010.
- [43] Clevert D-A, Unterthiner T, Hochreiter S Fast and accurate deep network learning by exponential linear units (elus). arXiv preprint arXiv:151107289. 2015.
- [44] De Boer P-T, Kroese DP, Mannor S, et al. A tutorial on the cross-entropy method. Ann Oper Res. 2005;134(1):19–67.
- [45] Dozat T Incorporating nesterov momentum into adam. 2016.
- [46] Srivastava N, Hinton G, Krizhevsky A, et al. Dropout: a simple way to prevent neural networks from overfitting. J Mach Learn Res. 2014 Jun;15(1):1929–1958. PubMed PMID: WOS:000344638300002; English.
- [47] James G, Witten D, Hastie T, et al. An introduction to statistical learning. Vol. 112. New York (NY): Springer; 2013.
- [48] Chen CL, Mahjoubfar A, Tai LC, et al. Deep learning in label-free cell classification. Sci Rep. 2016 Mar 15;6:21471. PubMed PMID: 26975219; PubMed Central PMCID: PMC4791545.
- [49] Satoh K, Shirabe S, Eguchi H, et al. 14-3-3 protein, total tau and phosphorylated tau in cerebrospinal fluid of patients with Creutzfeldt-Jakob disease and neurodegenerative disease in Japan. Cell Mol Neurobiol. 2006 Feb;26(1):45-52. PubMed PMID: 16633900.

- [50] Aksamit AJ Jr., Preissner CM, Homburger HA. Quantitation of 14-3-3 and neuron-specific enolase proteins in CSF in Creutzfeldt-Jakob disease. Neurology. 2001 Aug 28;57(4):728-730. PubMed PMID: 11524493.
- [51] Kapaki E, Kilidireas K, Paraskevas G, et al. Highly increased CSF tau protein and decreased β-amyloid (1-42) in sporadic CJD: a discrimination from Alzheimer's disease? J Neurol Neurosurg. 2001;71 (3):401-403.
- [52] Muayqil T, Gronseth G, Camicioli R. Evidencebased guideline: diagnostic accuracy of CSF 14-3-3 protein in sporadic Creutzfeldt-Jakob disease: report of the guideline development subcommittee of the american academy of neurology. Neurology. 2012 Oct 2;79(14):1499–1506. PubMed PMID: 22993290; PubMed Central PMCID: PMC3525296.
- [53] Zhao W, Zhang J-T, Xing X-W, et al. Chinese specific characteristics of sporadic Creutzfeldt-Jakob disease: a retrospective analysis of 57 cases. PloS One. 2013;8(3):e58442. PubMed PMID: 23516482; PubMed Central PMCID: PMC3597718.
- [54] Geschwind MD, Martindale J, Miller D, et al. Challenging the clinical utility of the 14-3-3 protein for the diagnosis of sporadic Creutzfeldt-Jakob disease. Arch Neurol. 2003 Jun;60(6):813–816. PubMed PMID: 12810484.
- [55] Forner SA, Takada LT, Bettcher BM, et al. Comparing CSF biomarkers and brain MRI in the diagnosis of sporadic Creutzfeldt-Jakob disease. Neurol Clin Pract. 2015 Apr;5(2):116–125. PubMed PMID: 26137420; PubMed Central PMCID: PMC4404282.
- [56] Atarashi R, Sano K, Satoh K, et al. Real-time quaking-induced conversion: a highly sensitive assay for prion detection. Prion. 2011 Jul-Sep;5(3):150–153. PubMed PMID: 21778820; PubMed Central PMCID: PMC3226039.
- [57] Lim JS, Kwon HM, Jang JW, et al. Characteristics of Korean patients with suspected Creutzfeldt-Jakob disease with 14-3-3 protein in cerebrospinal fluid: preliminary study of the Korean Creutzfeldt-Jakob disease active surveillance program. Prion. 2015;9(2):136–143. PubMed PMID: 25996401; PubMed Central PMCID: PMC4601202.
- [58] Jang J-W, Park YH, Lim JS, et al. Neurologists' awareness and preparedness on prion diseases in Korea. Dementia Neurocognitive Disord. 2013;12 (1):9.