

RESEARCH

Open Access

Identification of a novel lipid metabolism-related gene signature within the tumour immune microenvironment for breast cancer



Xu Chang and Peng Xing* 

Abstract

Background: Systemic factors can strongly affect how tumour cells behave, grow, and communicate with other cells in breast cancer. Lipid metabolic reprogramming is a systemic process that tumour cells undergo; however, the formation and dynamics of lipids associated with the tumour immune microenvironment (TIME) remain unclear. The investigation of the sophisticated bidirectional crosstalk of tumour cells with cancer metabolism, gene expression, and TIME could have the potential to identify novel biomarkers for diagnosis, prognosis, and immunotherapy. This study aimed to construct a prognostic signature to detect the bicrosstalk between the lipid metabolic system and the TIME of breast cancer.

Methods: To detect the expression of LRGs and execute GO/KEGG analysis, the R program was chosen. Considering the clinical information and pathological features, a prognostic gene signature was constructed by LASSO Cox regression analysis. TMB, MSI, and immune infiltration analyses were performed, and consensus cluster analysis of LRGs was also performed.

Results: These 16 lipid metabolism-related genes (LRGs) were mainly involved in the process of lipid metabolism and fatty acid binding in breast cancer. Prognosis analysis identified the prognostic value of FABP7 (Fatty acid binding protein 7) and NDUFAB1 (NADH:ubiquinone oxidoreductase subunit AB1) in breast cancer patients. The prognostic gene signature constructed with FABP7 and NDUFAB1 was significantly related to immune cell infiltration and could predict the overall survival rate with above average correctness of breast cancer patients. FABP7 and NDUFAB1 were proven to have relevance in immune cell infiltration and tumour mutation burden (TMB). Consensus cluster analysis identified that the upregulated mRNAs were mostly related to the oncogenesis process, while the downregulated mRNAs were associated with immune-related signalling pathways.

Conclusion: A comprehensive analysis was performed to evaluate the lipid metabolic system and identified a signature constructed by two prognostic genes for immunotherapies in breast cancer. The results also revealed evidence of vulnerabilities in the interplay between the lipid metabolic system and the TIME in breast cancer. Further data with clinical studies and experiments are warranted.

* Correspondence: pxing@cmu.edu.cn

Department of Surgical Oncology, Breast Surgery, General Surgery, First Affiliated Hospital of China Medical University, No.77 PuHe Road, Shenyang North New Area, Shenyang 110122, China



© The Author(s). 2022 **Open Access** This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit <http://creativecommons.org/licenses/by/4.0/>. The Creative Commons Public Domain Dedication waiver (<http://creativecommons.org/publicdomain/zero/1.0/>) applies to the data made available in this article, unless otherwise stated in a credit line to the data.

Keywords: Breast cancer, Lipid metabolism, Tumour immune microenvironment, Immune-related analysis, Immunotherapy

Introduction

Breast cancer has become the most prevalent malignancy worldwide according to the latest statistical results [1]. This highly heterogeneous malignancy, which comprises different subtypes, is still a serious threat to the health of women, whereas the triple-negative breast cancer (TNBC) subtype has always been known to have the worst prognosis [2–4]. Multiple research studies have proven that how tumour cells grow, behave, and communicate with other cells are determined not only by the characteristics of cancer cells but also by their sophisticated surrounding environment [5, 6]. The tumour microenvironment (TME) has been proven to be a dynamic community containing tumour cells and tumour-related cells [7]. The TIME (the tumour immune microenvironment), which represents the immune part of the TME, plays crucial roles, and studies have illustrated the complicated bidirectional crosstalk between tumour cells and the TIME in breast cancer [8].

Reprogramming of energy metabolism that can actively contribute to cancer development has been recognized as one of the cancer hallmarks [9, 10]. Carcinogenic events can alter the regulation of metabolic pathways, which in turn enable the proliferation and survival of cancer cells in the microenvironment by providing selective advantages [11, 12]. Lipid metabolism, including fatty acid metabolism and fatty acid transport, which can be influenced by factors such as age, obesity, menopause, drugs, and diet, is also highly activated in breast cancer cells [13, 14] and can both promote and inhibit the oncogenesis and progression of cancer cells by reassigning nutrients in the microenvironment of breast cancer [15, 16]. FABP7 (Fatty acid binding protein 7) is a member of the FABP intracellular lipid chaperone family that regulates lipid metabolism by increasing fatty acid uptake, FAO, and lipolysis [17]. NDU-FAB1 (NADH:ubiquinone oxidoreductase subunit AB1), on the other hand, is a mitochondrial acyl carrier protein that participates in lipid metabolism by interacting with other mitochondrial proteins [18].

Immunotherapies such as immune checkpoint blockades and other immunotherapeutic strategies have furnished new hopes for breast cancer patients [19, 20]; however, the low response rate limits the application of tumour immunotherapy [21]. Hence, improved analysis of how the lipid metabolic system with the TIME modulates cancer development and evasion from tumour-suppressive surveillance may reveal clues for novel anticancer immunotherapeutic strategies directed at lipid metabolic targets.

Therefore, in this study, 16 lipid metabolism-related genes (LRGs) were selected to detect the bidirectional

interplay of the lipid metabolic system of tumour cells with the TIME and construct a prognostic signature to explore the dynamic lipid metabolic signature difference in breast cancer. The results could provide new evidence for identifying novel prognostic biomarkers of immunotherapies for breast cancer and contribute to revealing the heterogeneity of the TIME in breast cancer.

Materials and methods

Date and sample source

The RNA-sequencing (RNA-seq) data and clinical information of breast cancer patients were retrieved from The Cancer Genome Atlas (TCGA) database, which was released on June 1, 2021. R software (version 4.0.3) was used to develop all data analysis methods and the R package. All the obtained expression data of breast cancer patients are shown in Table S1 and were normalized to Fragments Per Kilobase per Million (FPKM) values for subsequent investigation. The patients with breast cancer were diagnosed mainly in 2008–2010, and the clinical information were uploaded to TCGA in 2016. The workflow of this study is shown in Fig. S1.

Identifying the different expressions of LRGs

Sixteen LRGs in total that participate in the lipid metabolic system in breast cancer were selected. The limma and reshape2 R packages were used to detect the difference in LRG expression in breast cancer and normal tissues [22, 23]. The STRING database was used to search the hub genes by the set with a minimum interaction score of 0.9 [24].

Functional enrichment analysis

To further identify the function of these LRGs in breast cancer, GO and KEGG databases were selected, and the data were analysed by functional enrichment analysis. The GO (Gene Ontology) database is a web-based tool for determining gene function for MF (molecular function), BP (biological pathways), and CC (cellular components) [25]. Gene set enrichment analysis detected gene pathway enrichments in KEGG (Kyoto Encyclopedia of Genes and Genomes), an open resource [26]. To better understand the carcinogenesis of these LRGs, the ClusterProfiler R package (version 3.14.0) was used to analyse the GO functions and KEGG pathways of these potential targets [27].

Construction of the lipid metabolism-related gene prognostic model

Cox regression analysis was utilised to determine the prognostic significance of the LRGs. The Kaplan–Meier

approach was used to calculate the prognostic value of these 16 LRGs, which were then examined using the log-rank test and univariate Cox proportional hazard regression, yielding *P* values and hazard ratios (HRs) with 95% confidence intervals (CIs). LRGs with substantial prognostic value, as demonstrated by Kaplan–Meier survival curves, were chosen. The chi-square test was selected to clarify the correlation between the prognostic LRGs and clinical TNM stage, and the Wilcoxon test was used to explore the correlation of the age factor with the prognostic LRGs. Based on the prognostic value of these LRGs, a prognostic model containing the two prognostic LRGs was developed by LASSO Cox regression analysis for breast cancer patients using 10-fold cross-validation to determine the optimal value of penalty parameter λ . According to the risk score, the patients with breast cancer were separated into two subgroups with low risk and high risk, and the overall survival (OS) possibility between these two groups was compared by the Kaplan–Meier method. Receiver operating characteristic (ROC) analysis was selected to predict the diagnostic accuracy of each gene. Considering the pathological characteristics, a predicted nomogram was developed to predict the 1-year, 3-year, and 5-year overall survival possibility through the forestplot package in R software [28].

Immune cell infiltration analysis of the prognostic LRGs

The correlation between the prognostic LRGs and immune cell infiltration was evaluated using the ssGESA R package for comprehensive analysis of tumour-infiltrating immune cells in breast cancer. In the analysis of the correlation between the immune checkpoints and the prognostic LRGs, the ggplot2 R package was selected. Spearman’s correlation analysis was chosen for the examination of tumour mutation

burden (TMB) and microsatellite instability (MSI), with a *P* value less than 0.05.

The cluster analysis of LRGs

The raw counts of RNA-sequencing data of patients with breast cancer and accompanying clinical information were received from the TCGA-BRCA cohort, with the collection and application methods complying with the recommendations and rules. The ConsensusClusterPlus R package (v1.54.0) was used for consistency analysis [29], and the pheatmap R package (v1.0.12) was used for clustering heatmaps. Genes with SD > 0.1 were kept in the gene expression heatmap. If the number of input genes was greater than 1000, the SD was sorted, and the top 25% of the genes were extracted. The Limma R package (version: 3.40.2) was used to study the differential expression of mRNAs. The adjusted *P* value was analysed to correct for false-positive results in TCGA or GTEx. The results with “Adjusted *P* < 0.05 and Log (Fold Change) > 1 or Log (Fold Change) < -1” were defined as the thresholds for the screening of differential expression of mRNAs. Fold-change numbers and corrected *P* values were used to create volcano graphs. Hierarchical clustering was used to look for mRNAs that were differentially expressed in tumour and normal tissues.

Results

The expression of LRGs in breast cancer

The expression levels of 16 LRGs in breast cancer and normal breast tissues were first detected by data from TCGA-breast cancer. A total of two genes showed no significant change in breast cancer (Fig. 1). More specifically, the expression of FABP5, FABP7, FABP3, FABP6, NDU-FAB1, FABP2, FABP1, KLF5, LPN1, LPN3, and EP300 was upregulated compared with that in normal tissues,

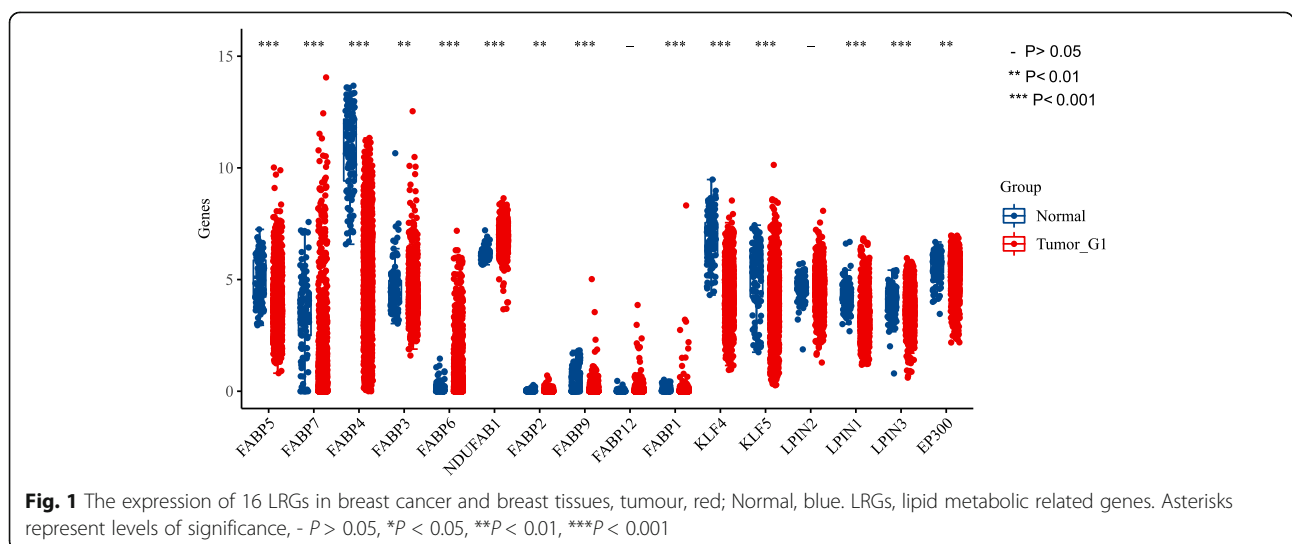


Fig. 1 The expression of 16 LRGs in breast cancer and breast tissues, tumour, red; Normal, blue. LRGs, lipid metabolic related genes. Asterisks represent levels of significance, - *P* > 0.05, **P* < 0.05, ***P* < 0.01, ****P* < 0.001

while the expression of FABP4, FABP9, and KLF4 was downregulated (all $P < 0.05$).

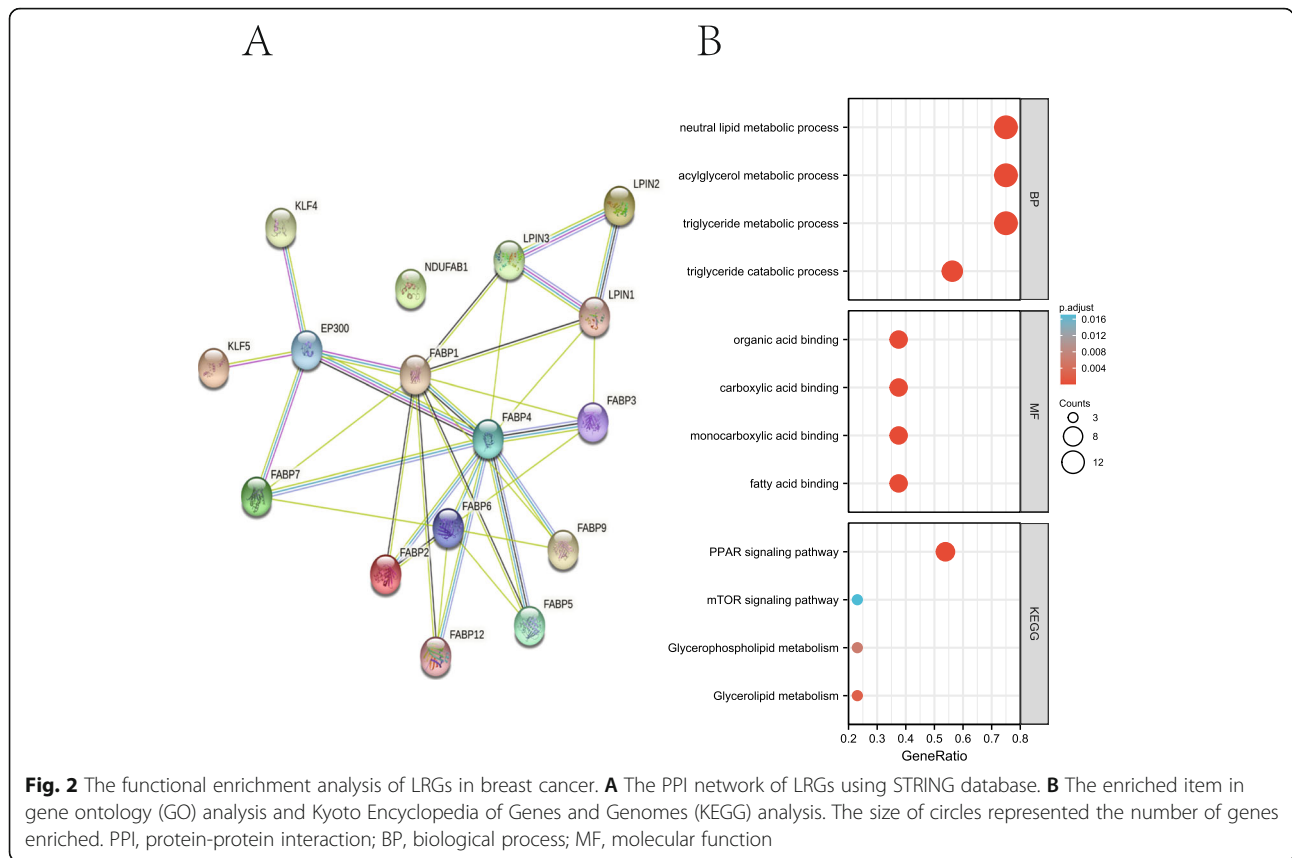
Functional enrichment analysis of LRGs

A protein–protein interaction (PPI) network was constructed to explore the correlation of these LRGs, which was set with a minimum interaction score of 0.9, as shown in Table 1. The analysis results revealed that EP300, FABP1, FABP2, FABP4, FABP7, KLF4, and KLF5 were hub genes (Fig. 2A). GO and KEGG databases were selected to screen the function of the LRGs. The results suggested that

these 16 LRGs were mainly involved in the positive regulation of the triglyceride metabolic process, acylglycerol metabolic process, neutral lipid metabolic process, triglyceride catabolic process, fatty acid binding, monocarboxylic acid binding, carboxylic acid binding, and organic acid binding in GO analysis. Moreover, the analysis results from the KEGG database revealed that 16 LRGs were primarily concerned with the PPAR signalling pathway, glycerolipid metabolism, glycerophospholipid metabolism, and mTOR signalling pathway (Fig. 2B, Table 2).

Table 1 The protein–protein interaction (PPI) network constructed by STRING database to explore the interactions of these LRGs

node1	node2	node1_external_id	node2_external_id	combined_score
EP300	KLF4	ENSP00000263253	ENSP00000363804	0.999
EP300	KLF5	ENSP00000263253	ENSP00000366915	0.991
LPIN3	LPIN1	ENSP00000362354	ENSP00000397908	0.937
FABP2	FABP1	ENSP00000274024	ENSP00000295834	0.926
FABP4	EP300	ENSP00000256104	ENSP00000263253	0.918
EP300	FABP7	ENSP00000263253	ENSP00000357429	0.907
EP300	FABP1	ENSP00000263253	ENSP00000295834	0.906
FABP4	FABP1	ENSP00000256104	ENSP00000295834	0.897
FABP2	FABP6	ENSP00000274024	ENSP00000377549	0.876
LPIN2	LPIN3	ENSP00000261596	ENSP00000362354	0.858
FABP1	FABP3	ENSP00000295834	ENSP00000362817	0.841
FABP4	FABP6	ENSP00000256104	ENSP00000377549	0.831
LPIN2	LPIN1	ENSP00000261596	ENSP00000397908	0.815
FABP3	FABP6	ENSP00000362817	ENSP00000377549	0.808
FABP1	FABP5	ENSP00000295834	ENSP00000297258	0.719
FABP4	LPIN3	ENSP00000256104	ENSP00000362354	0.695
FABP4	FABP2	ENSP00000256104	ENSP00000274024	0.695
FABP4	FABP5	ENSP00000256104	ENSP00000297258	0.673
FABP4	FABP3	ENSP00000256104	ENSP00000362817	0.667
FABP4	FABP7	ENSP00000256104	ENSP00000357429	0.659
FABP4	FABP12	ENSP00000256104	ENSP00000353650	0.657
FABP4	FABP9	ENSP00000256104	ENSP00000368362	0.655
FABP12	FABP6	ENSP00000353650	ENSP00000377549	0.654
FABP9	FABP6	ENSP00000368362	ENSP00000377549	0.628
FABP1	FABP7	ENSP00000295834	ENSP00000357429	0.607
FABP7	FABP6	ENSP00000357429	ENSP00000377549	0.588
FABP5	FABP6	ENSP00000297258	ENSP00000377549	0.584
FABP1	FABP9	ENSP00000295834	ENSP00000368362	0.532
FABP3	LPIN1	ENSP00000362817	ENSP00000397908	0.517
FABP4	LPIN1	ENSP00000256104	ENSP00000397908	0.505
FABP1	LPIN3	ENSP00000295834	ENSP00000362354	0.502
FABP1	LPIN1	ENSP00000295834	ENSP00000397908	0.448
FABP1	FABP12	ENSP00000295834	ENSP00000353650	0.43



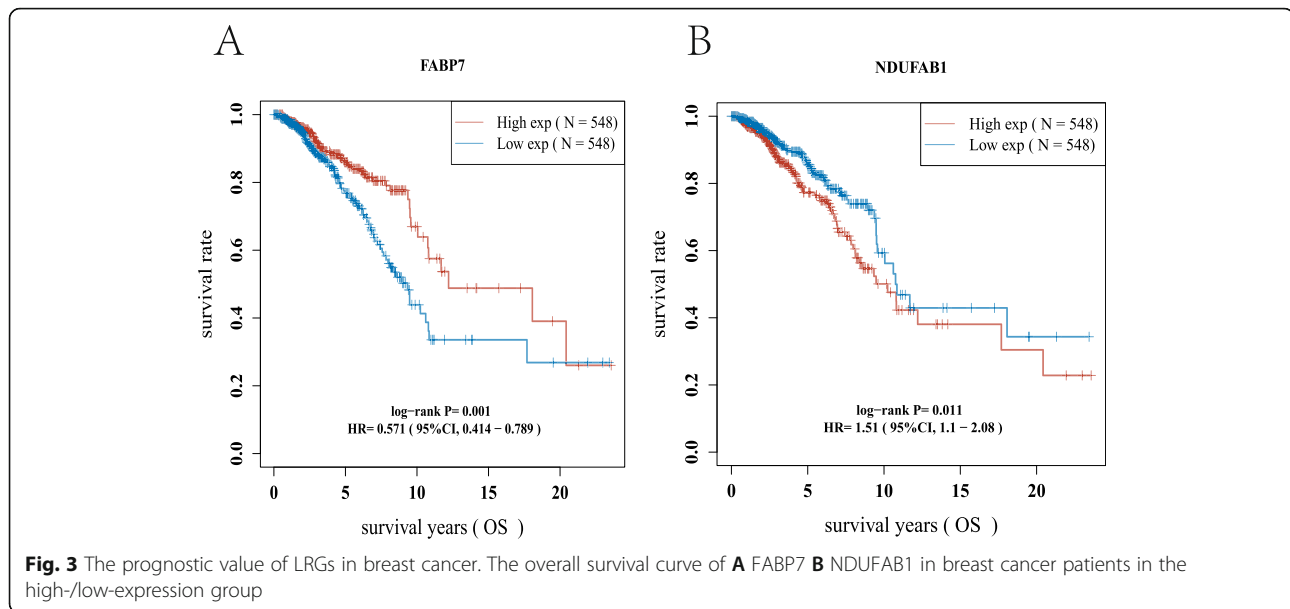
Construction of the prognostic gene model

To clarify the prognostic value of these LRGs, a prognostic gene model was built using univariate Cox regression analysis. The findings revealed that two LRG genes were identified as having prognostic significance, and the Kaplan–Meier survival curves are displayed in Fig. 3. The findings

of the prognostic analysis revealed that individuals with breast cancer who had FABP7 downregulation had a low chance of survival (Fig. 3A, $P = 0.001$) and upregulation of NDUFAB1 (Fig. 3B, $P = 0.011$). Based on the prognostic value of FABP7 and NDUFAB1 in breast cancer, a prognostic gene signature containing these two LRGs (FABP7

Table 2 Pathway enrichment analysis items of 16 LRGs from KEGG

ONTOLOGY	ID	Description	GeneRatio	BgRatio	P value	P.adjust	qvalue
BP	GO:0006641	triglyceride metabolic process	10/11	105/18670	2.23e-22	9.62e-20	5.50e-20
BP	GO:0006639	acylglycerol metabolic process	10/11	129/18670	1.90e-21	2.96e-19	1.69e-19
BP	GO:0006638	neutral lipid metabolic process	10/11	130/18670	2.06e-21	2.96e-19	1.69e-19
BP	GO:0019433	triglyceride catabolic process	8/11	35/18670	1.06e-20	1.14e-18	6.52e-19
BP	GO:0046461	neutral lipid catabolic process	8/11	43/18670	6.51e-20	4.68e-18	2.67e-18
MF	GO:0005504	fatty acid binding	5/11	34/17697	8.82e-12	3.53e-10	1.30e-10
MF	GO:0033293	monocarboxylic acid binding	5/11	64/17697	2.40e-10	4.79e-09	1.77e-09
MF	GO:0005324	long-chain fatty acid transporter activity	3/11	11/17697	2.94e-08	3.92e-07	1.44e-07
MF	GO:0031406	carboxylic acid binding	5/11	193/17697	6.42e-08	5.18e-07	1.91e-07
MF	GO:0036041	long-chain fatty acid binding	3/11	14/17697	6.48e-08	5.18e-07	1.91e-07
KEGG	hsa03320	PPAR signaling pathway	6/9	78/8076	5.48e-11	1.70e-09	1.50e-09
KEGG	hsa04975	Fat digestion and absorption	2/9	43/8076	9.74e-04	0.015	0.013
KEGG	hsa00561	Glycerolipid metabolism	2/9	61/8076	0.002	0.020	0.018
KEGG	hsa00564	Glycerophospholipid metabolism	2/9	98/8076	0.005	0.038	0.034
KEGG	hsa04150	mTOR signaling pathway	2/9	155/8076	0.012	0.075	0.066



and NDUFAB1) was constructed by LASSO Cox regression analysis (Fig. 4A, B), and the final results were calculated with the formula risk score = $(-0.1013) * FABP7 + (0.3367) * NDUFAB1$. Based on the risk score, the two groups were separated into high-risk and low-risk groups. The distribution of risk scores, survival status, and expression of these two genes were all present. As the risk score increased, the risk of death of breast cancer patients increased, while the survival time decreased (Fig. 4C). The Kaplan–Meier survival curves showed that breast cancer patients with high-risk scores (median time = 9.5 years) had a worse overall survival (OS) rate than those with low-risk scores (median time = 10.8 years) ($P = 0.0053$), and the 1-year, 3-year, and 5-year ROC curves had AUCs of 0.596, 0.591, and 0.608, respectively (Fig. 4D, E).

The construction of the predictive nomogram

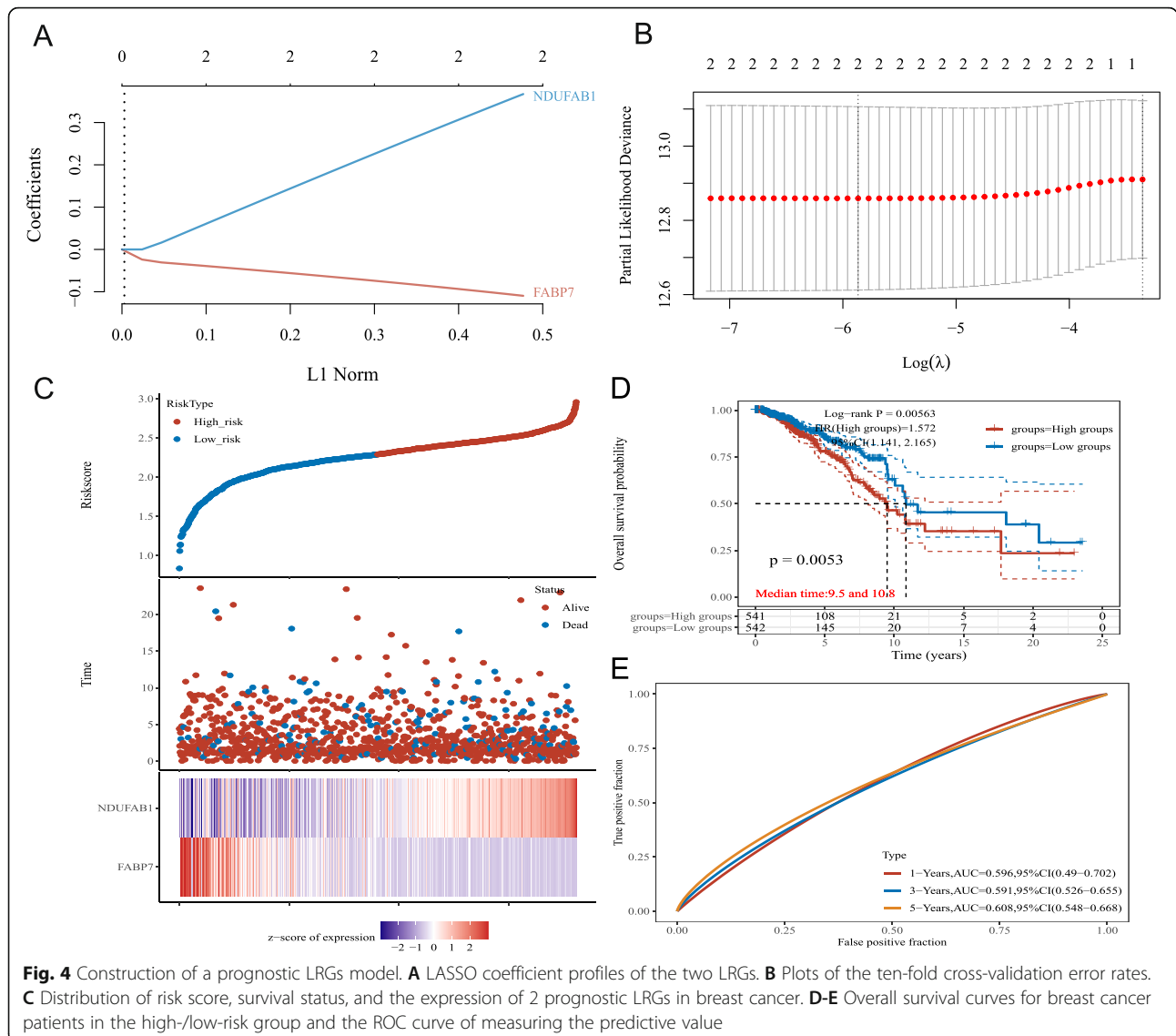
Allowing for the correlation between pathological features and these two prognostic LRGs (FABP7 and NDUFAB1), a predictive nomogram was subsequently built to predict the survival probability. By analysing the results, univariate analyses identified the expression of FABP7 and NDUFAB1, and the stage of pT, pN, and pM were the factors that could affect the prognosis of breast cancer patients. More interestingly, the univariate and multivariate analyses illustrated that age was the factor affecting prognosis, and the results are shown in Fig. 5A–B. After analysing the expression of FABP7 and NDUFAB1 with clinical characteristics, the results demonstrated that the expression of FABP7, as well as NDUFAB1, was significantly correlated with T stage and age, and the results are shown in Tables 3 and 4. Furthermore, the 3- and 5- year overall survival (OS) rates in the complete cohort could be predicted reasonably well by assessing the predictive nomogram data, as shown in Fig. 5C, D.

The correlation between prognostic LRGs and immune cell infiltration in breast cancer

In this study, correlation analysis for the expression of prognostic LRGs (FABP7 and NDUFAB1) and immune cell infiltration in breast cancer was also performed using the ssGSEA R package. The findings revealed a strong relationship between the expression of prognostic LRGs (FABP7 and NDUFAB1) and the quantity of immunologically infiltrating cells, such as CD8+ T cells, macrophages, neutrophils, cytotoxic cells, eosinophils, NK cells, and Treg cells (Fig. 6 B). A B, all $P < 0.05$. This evidence suggested a significant correlation between the prognostic LRGs and tumour immune infiltration. Moreover, the study detected the correlation between the immune checkpoints (TIGIT, PDCD1, CD274, LAG3, CTLA4) and the prognostic LRGs by the ggplot2 R package, and the results revealed a significant correlation between the immune checkpoints and the two prognostic LRGs (Fig. 7 A B, all $P < 0.05$).

TMB and MSI analysis of LRGs

Tumour mutation burden (TMB), as well as microsatellite instability (MSI) analysis, could be utilized to anticipate the immunotherapeutic efficacy of breast cancer treatment. To clarify whether these two prognostic LRGs could serve as biomarkers for immunotherapy, the correlation between the two prognostic LRGs and TMB as well as MSI in breast cancer was analysed. The results indicated a favourable relationship between TMB and FABP7 (Fig. 8A, $P = 2.26e-06$) and NDUFAB1 (Fig. 8B, $P = 0.005$). The findings demonstrated that prognostic LRGs were strongly linked to tumour immune cell infiltration and could serve as biomarkers of immunotherapies for breast cancer.



Tumour immune infiltration analysis of the prognostic gene model

To further screen the correlation of the gene prognostic model containing the prognostic LRGs (FABP7 and NDUFAB1) with the tumour immune microenvironment (TIME) in breast cancer, the ssGSEA method was selected to perform the immune infiltration analysis of this prognostic signature. The analysis results illustrated a negative correlation between the prognostic model containing FABP7 and NDUFAB1 (Fig. 9). The above results revealed a significant correlation of the gene signature with the TIME in breast cancer.

Consensus clustering analysis of LRGs in breast cancer

To explore the different functions of these 16 LRGs, a consensus clustering analysis algorithm was performed. The delta area curve of consensus clustering (Fig. 10A)

indicated the relative change in the area under the cumulative distribution function (CDF) curve for each category number *k* compared with the *k*–1 consistency analysis (Fig. 10B), and the number of clusters was reduced to two in the end (Fig. 10C, D).

Functional enrichment analysis of the consensus clusters

Volcano plots were constructed using fold-change values and adjusted *P* values. The red point in the plot represents the overexpressed mRNAs, and the blue point indicates the downregulated mRNAs with statistical significance. The results revealed that 846 mRNAs were upregulated and 255 mRNAs were downregulated (Fig. 11A). Analysis of mRNAs that were differentially expressed between tumour and normal tissues was conducted using hierarchical clustering (Fig. 11B).

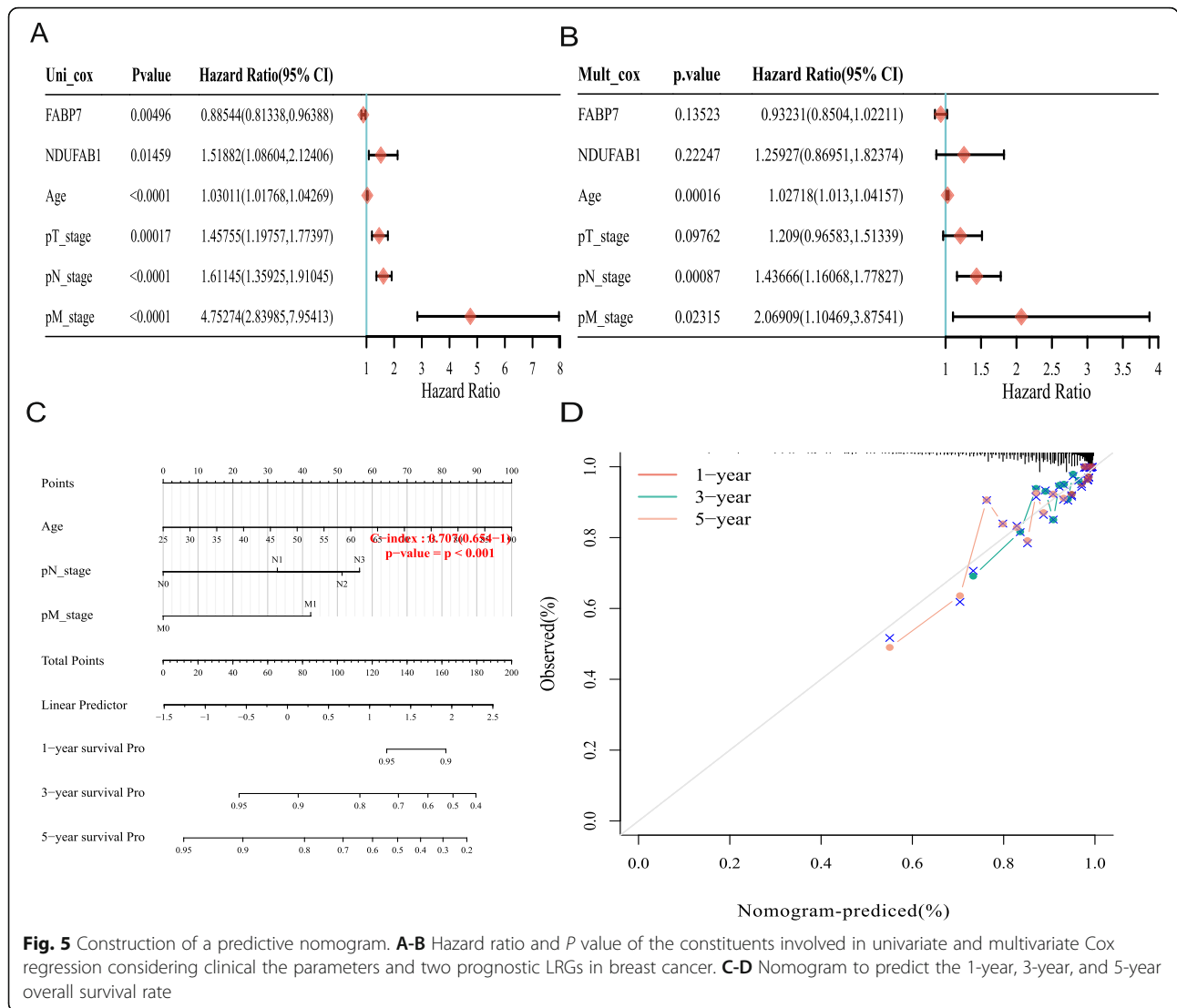


Fig. 5 Construction of a predictive nomogram. **A-B** Hazard ratio and *P* value of the constituents involved in univariate and multivariate Cox regression considering clinical the parameters and two prognostic LRGs in breast cancer. **C-D** Nomogram to predict the 1-year, 3-year, and 5-year overall survival rate

The data were evaluated using functional enrichment analysis via GO/KEGG techniques to further establish the underlying function of putative targets of these two clusters in breast cancer, and the findings with $P < 0.05$ or $FDR < 0.05$ were judged to be enriched in a meaningful pathway. The analysis results suggested that the up consensus cluster mRNAs were related to the PI3K – Akt signalling pathway, MAPK signalling pathway, insulin secretion, positive regulation of protein kinase B signalling, regulation of hormone secretion, regulation of insulin secretion, and reproductive system development (Fig. 11C). Meanwhile, the down cluster was closely related to the p53 signalling pathway, cell cycle, chemokine signalling pathway, lymphocyte chemotaxis, and IL – 17 signalling pathway (Fig. 11D). More interestingly, both consensus clusters were involved in the oestrogen signalling pathway.

The above results showed the different functions and signature pathways of the consensus clusters of LRGs in breast cancer, which may suggest that the process of the lipid metabolic system comprises lipid formation and dynamics and is related to different biological functions in breast cancer.

Discussion

Breast cancers, including different subtypes, have always threatened the health of women worldwide, and this is especially true for triple-negative breast cancer (TNBC) [30]. Numerous studies of the TME have redefined tumours from simple gatherings of tumour cells to a complicated community composed of multiple surrounding cells [31]. The tumour immune microenvironment (TIME), which represents the TME’s immunological components, plays decisive roles in breast cancer and prospective immunotherapeutic targets [32]. Studies

Table 3 The correlation between the clinical characteristics and the expression of FABP7

Characteristic	Low expression of FABP7	High expression of FABP7	P value	Method
n	541	542		
T stage, n (%)			0.014	Chisq.test
T1	125 (11.6%)	152 (14.1%)		
T2	325 (30.1%)	304 (28.1%)		
T3	64 (5.9%)	75 (6.9%)		
T4	25 (2.3%)	10 (0.9%)		
N stage, n (%)			0.694	Chisq.test
N0	247 (23.2%)	267 (25.1%)		
N1	177 (16.6%)	181 (17%)		
N2	62 (5.8%)	54 (5.1%)		
N3	40 (3.8%)	36 (3.4%)		
M stage, n (%)			0.258	Chisq.test
M0	448 (48.6%)	454 (49.2%)		
M1	13 (1.4%)	7 (0.8%)		
Age, median (IQR)	61 (50, 70)	55 (47, 64)	< 0.001	Wilcoxon

have shown that breast cancer cells can contain naturally processed and distinct exceptional mutations that can be identified by patients' immune systems [33]. The major objective of immunotherapy is to promote the switch from a protumour to an antitumour impact to maximize the efficacy of antitumour immunity, and accordingly, immunotherapies such as immune checkpoint blockades and others have offered new clinical strategies for breast cancer patients.

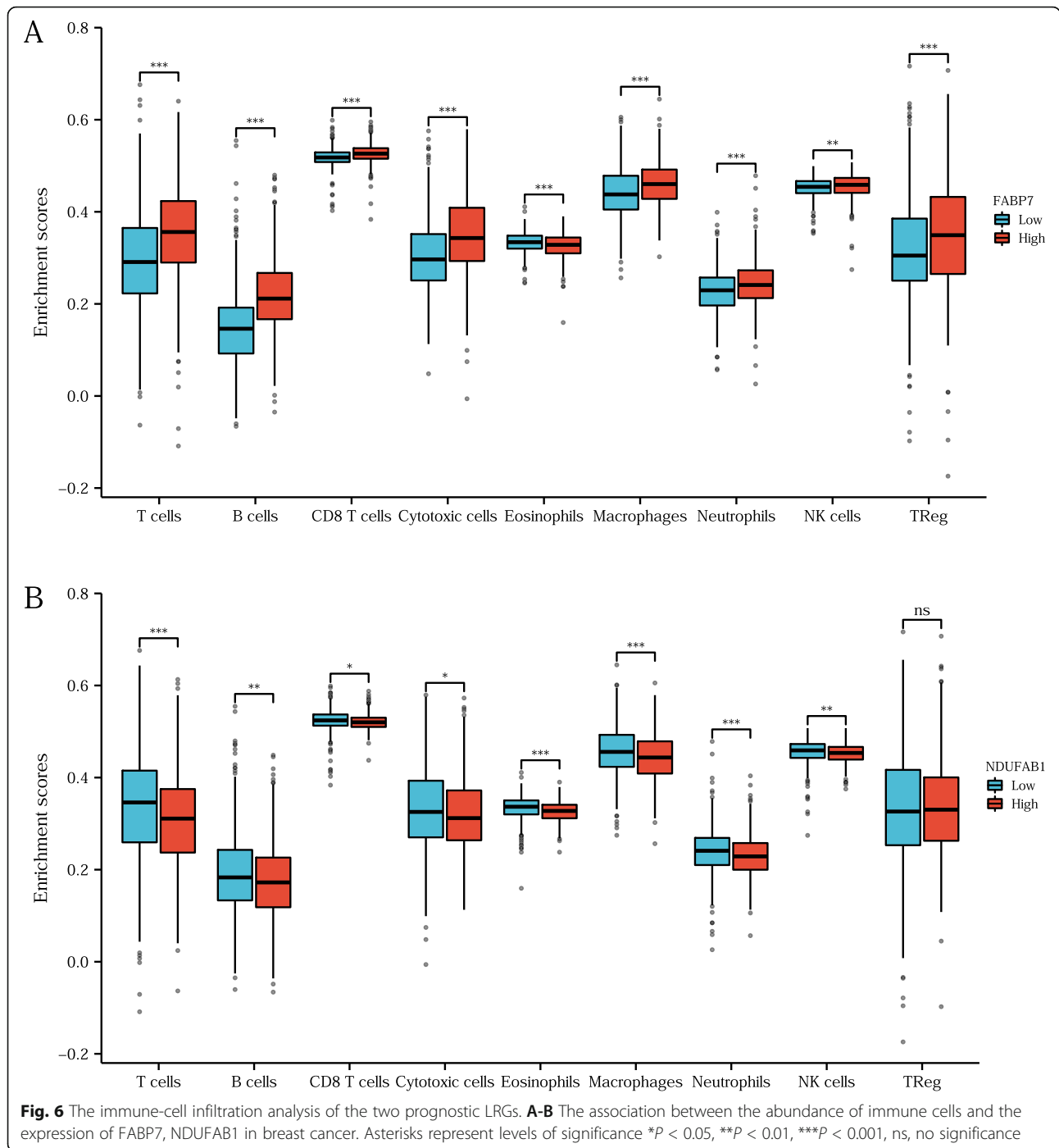
Cancer cells have been known to be able to take advantage of the altered metabolic community to maintain their survival and proliferation [34–36]. The altered lipid

metabolic process of cancer cells can further reprogram and impact other cells in the tumour microenvironment (TME), which contributes to the regulation of oncogenesis, aggravation, metastasis, and recurrence of breast cancer [37, 38].

In this study, the expression and prognostic value of these 16 lipid metabolism-related genes (LRGs) in breast cancer were first evaluated, and the results showed that the expression of 11 LRGs was increased compared with that in normal tissues, while the expression of 3 LRGs was decreased. Kaplan–Meier survival analysis results identified the prognostic value of

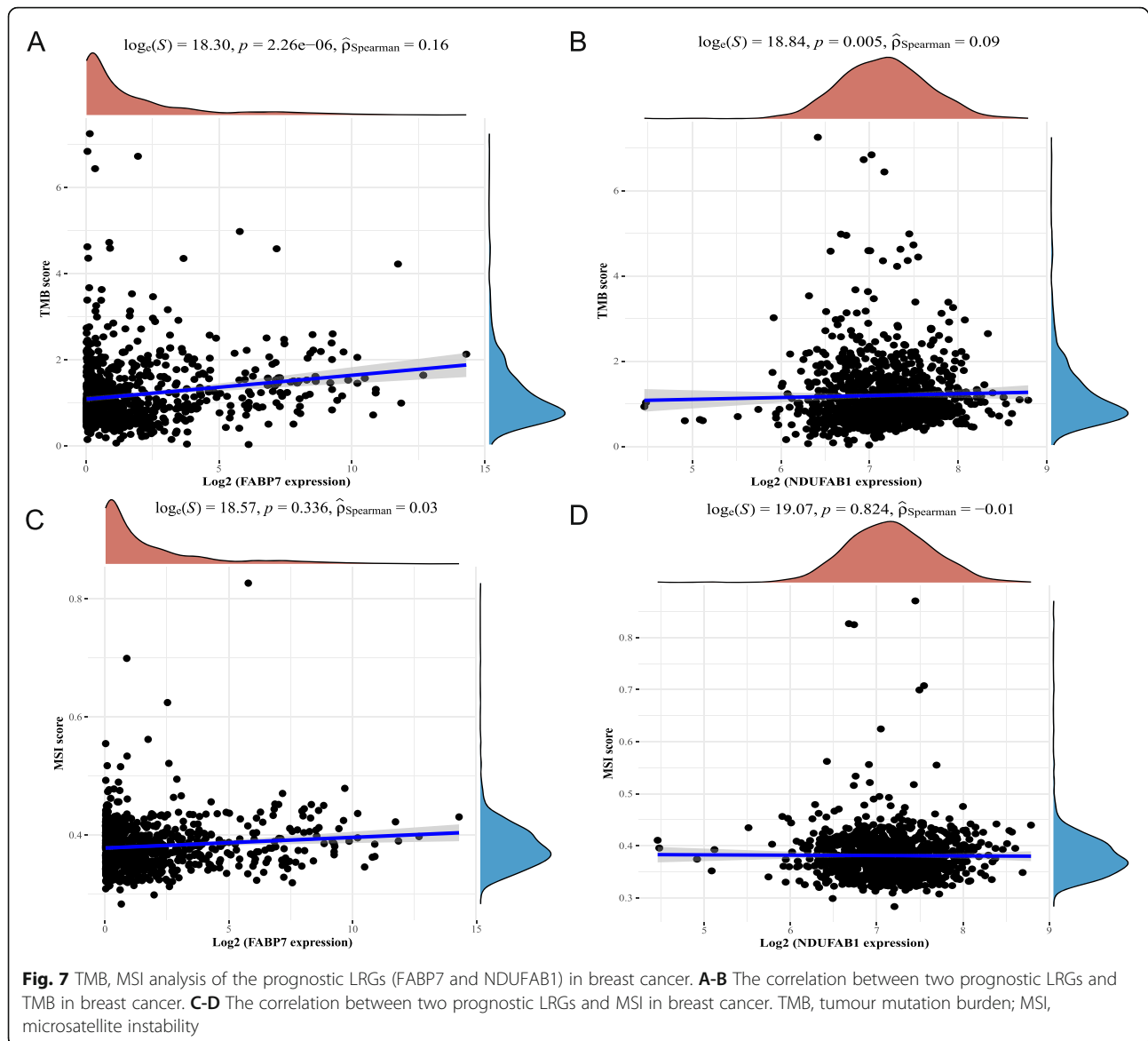
Table 4 The correlation between the clinical characteristics and the expression of NDUFB1

Characteristic	Low expression of NDUFB1	High expression of NDUFB1	P value	Method
n	541	542		
T stage, n (%)			0.014	Chisq.test
T1	159 (14.7%)	118 (10.9%)		
T2	296 (27.4%)	333 (30.8%)		
T3	71 (6.6%)	68 (6.3%)		
T4	13 (1.2%)	22 (2%)		
N stage, n (%)			0.908	Chisq.test
N0	262 (24.6%)	252 (23.7%)		
N1	175 (16.4%)	183 (17.2%)		
N2	58 (5.5%)	58 (5.5%)		
N3	40 (3.8%)	36 (3.4%)		
M stage, n (%)			0.041	Chisq.test
M0	457 (49.6%)	445 (48.3%)		
M1	5 (0.5%)	15 (1.6%)		
Age, median (IQR)	56 (47, 65)	59 (50, 69)	0.002	Wilcoxon



FABP7 and NDUFAB1 in breast cancer patients. GO/KEGG functional enrichment analysis was subsequently performed, and the results illustrated that these 16 LRGs were involved in the triglyceride metabolic process, fatty acid binding, PPAR signalling pathway, and mTOR signalling pathway. FABP7, which regulates lipid metabolism, has been found to be up-regulated in triple negative breast cancer (TNBC) and to affect TNBC cell death as a metabolic mediator in

nutrient-depleted conditions [39]. It's also a regulator of lipid metabolism reprogramming in HER2+ breast cancer cells, allowing metastatic cells to adapt and survive in the brain microenvironment [17]. In the meantime, NDUFAB1 has been identified as an important endogenous regulator of mitochondrial bioenergetics and a contributor to lipoic acid production [18]. The above results identified that the selected LRGs were related to the process of lipid metabolic

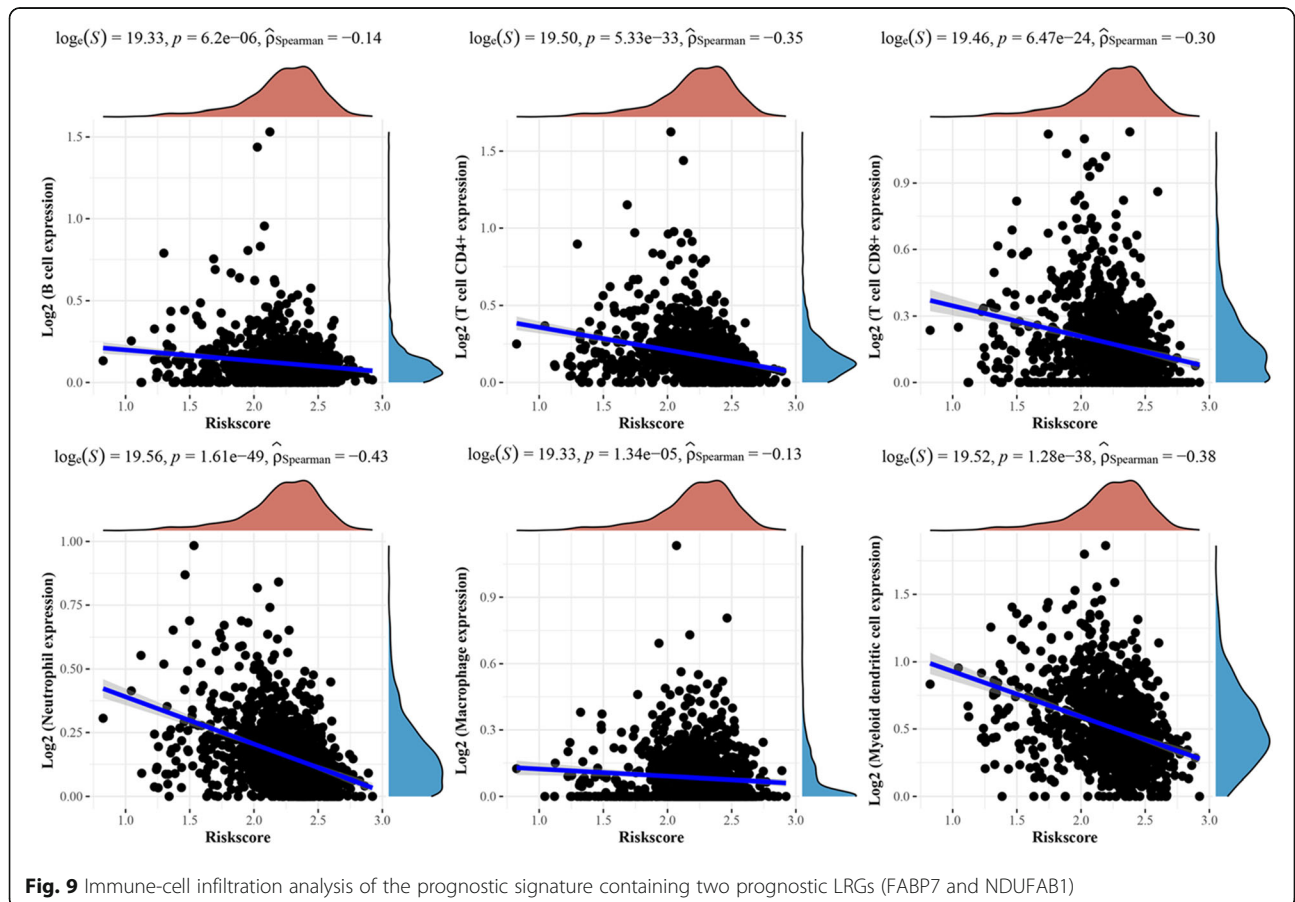
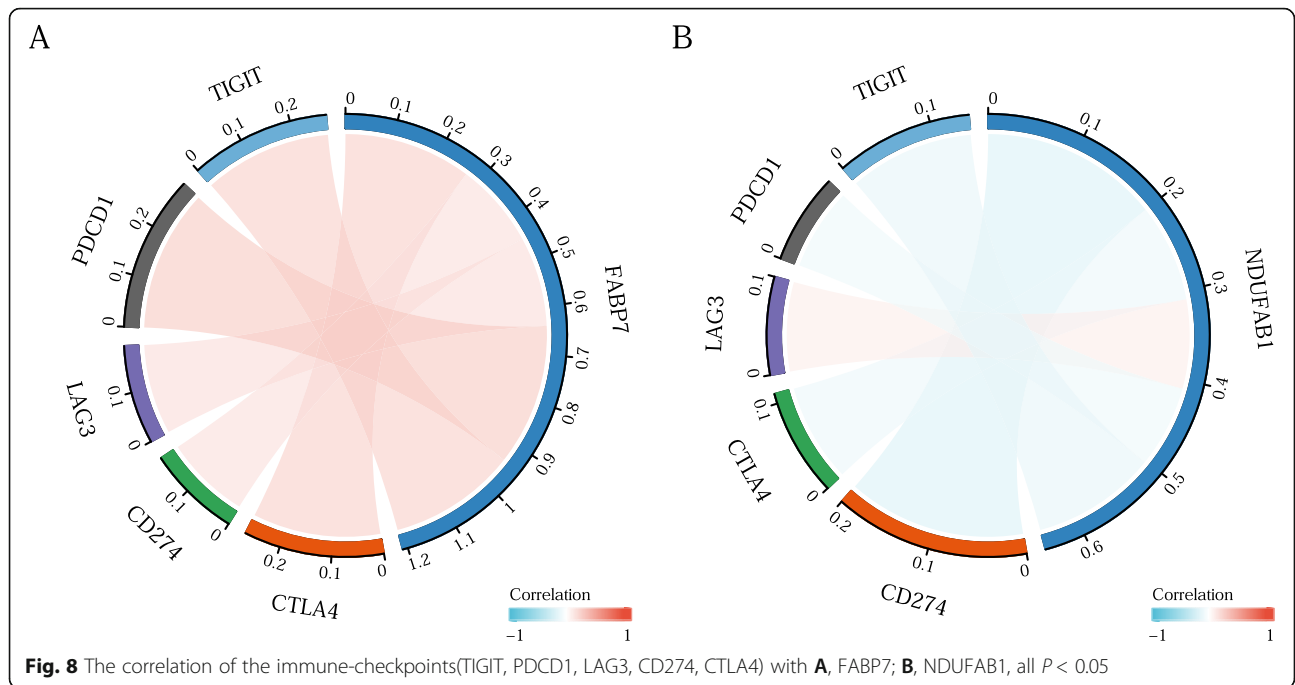


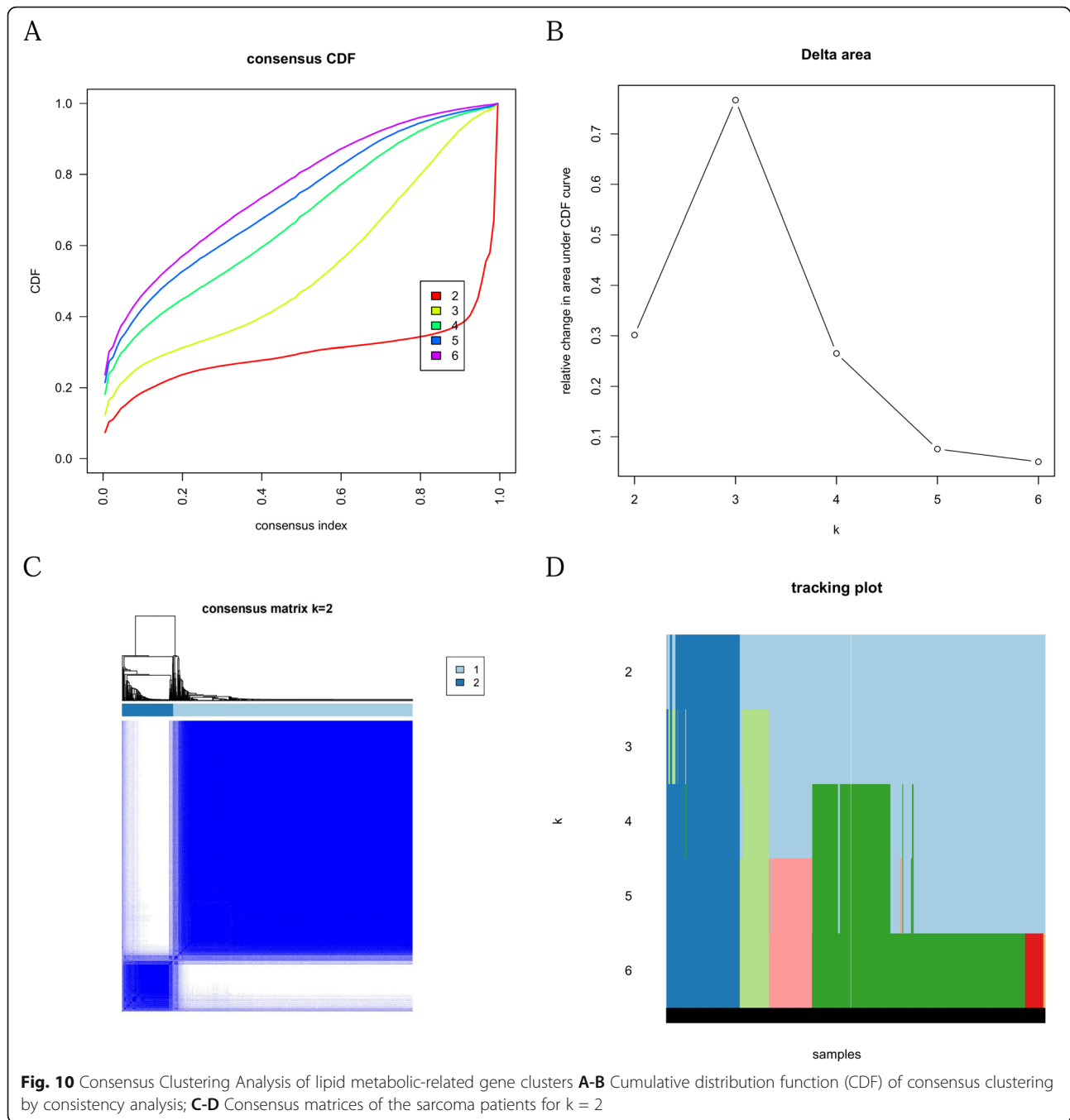
reprogramming, oncogenesis, and inflammation in breast cancer [40–42], of which the expression of FABP7 and NDUFAB1 could affect the prognosis of patients with breast cancer. Furthermore, as lipid metabolism regulators, FABP7 and NDUFAB1 might have the potential to elucidate the mechanism of breast cancer cell invasion and metastasis.

Based on the prognostic value of FABP7 and NDUFAB1, LASSO Cox regression analysis was used to build a prognostic gene model that revealed that breast cancer patients with high risk scores had a lower overall survival rate than those with low risk scores. The correlation of the clinical characteristics with these two prognostic LRGs (FABP7, NDUFAB1) in breast cancer patients was performed using the chi-square test, and the results revealed that both the expression of FABP7 and NDUFAB1 were significantly

related to the T stage of breast cancer patients. Moreover, when compared to an ideal model in the full cohort, the predictive nomogram showed that the 3-year and 5-year overall survival rates could be predicted pretty well. This was the first study to develop a lipid metabolism-related prognostic gene signature in breast cancer patients with clinical data, thus providing us with new alternatives for prognostic prediction in breast cancer.

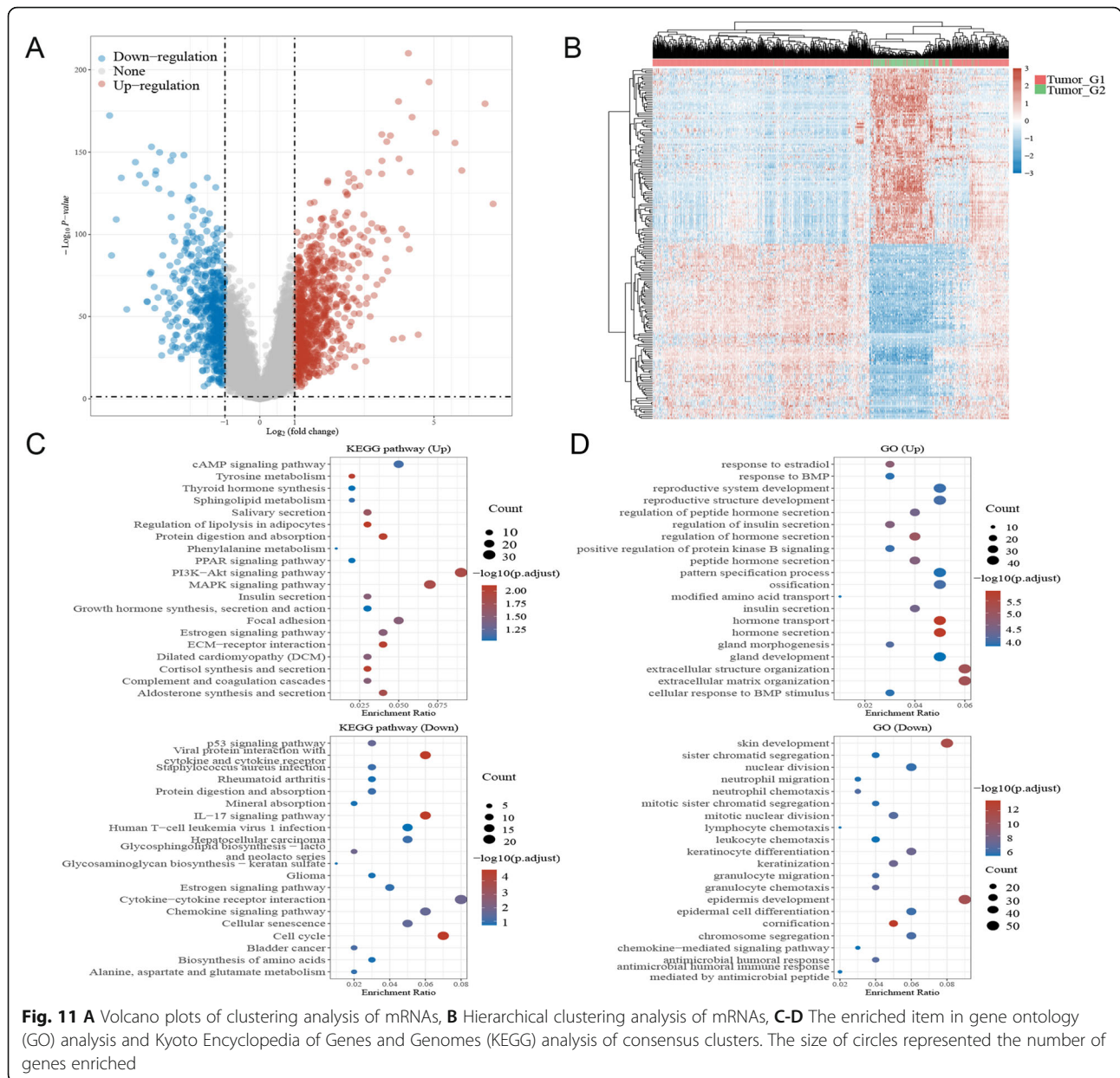
Tumour mutation burden (TMB) [43], as well as microsatellite instability (MSI) [44] analysis, can be used to predict the response to immunotherapies and have proven to be helpful biomarkers in breast cancer for identifying individuals who will benefit from immunotherapies. Using tumour immune cell infiltration-related analysis, the results illustrated that FABP7 and NDUFAB1 were significantly correlated with tumour





immune cell infiltration. More interestingly, by immune cell infiltration analysis of the prognostic signature constructed by FABP7 and NADUFAB1, a more significant correlation with tumour immune cells in breast cancer was demonstrated. The above results illustrated that FABP7 and NDUFAB1 could serve as predictive biomarkers in immunotherapies for breast cancer, which also suggested a significant correlation of FABP7 and NDUFAB1 with the tumour immune microenvironment (TIME).

Another important finding of this study revealed the difference in the function of these 16 LRGs by consistency analysis. To further confirm the underlying function of potential targets of these two consensus clusters in breast cancer, the data were analysed by functional enrichment using the GO and KEGG databases. The analysis results suggested that the up cluster was significantly related to the functions of oncogenesis and metabolic reprogramming processes of breast cancer, such as the PI3K–Akt signalling pathway, MAPK



signalling pathway, insulin secretion, and regulation of hormone secretion [45–47]. Meanwhile, the down consensus cluster was closely related to immune-related functions and signalling pathways, such as the IL–17 signalling pathway and chemokine signalling pathway [48, 49]. More interestingly, both consensus clusters were involved in the oestrogen signalling pathway, which is an important signalling pathway for breast cancer [50, 51]. Studies have proven that disturbances in the lipid metabolic system can modulate the menopausal status of women [52] which could also lead to the unbalanced distribution of nutrients between tumour cells and immune cells in the tumour microenvironment (TME) [53]. The above results showed the different functions

and signature pathways of the consensus clusters of LRGs in breast cancer. The results suggested that the lipid metabolic system might induce the reassignment of nutrients in the tumour microenvironment by the oestrogen signalling pathway, which could also modulate the menopausal status of patients with breast cancer.

Study strengths and limitations

This current study has certain strengths. This study directly detect the correlation between the lipid metabolism-related genes (LRGs) and the tumour immune cell infiltration in breast cancer, and the functional landscape of these LRGs was also illustrated using consensus analysis. These findings would have crucial

implications for breast cancer patients' clinical treatment, endocrine therapy, chemotherapy, and immunotherapy. There are also some limitations to this research. All studies were carried out with the TCGA-BRCA cohort, and more data with in vivo, in vitro research and clinical studies could be used to corroborate the results.

Conclusions

In conclusion, a thorough investigation of lipid metabolism-related genes was conducted for patients with breast cancer, and a prognostic signature encompassing two biomarkers (FABP7 and NDUFB1) was discovered for the application of immunotherapy. The results also provided new perspectives for deciphering the bidirectional interplay between the lipid metabolic system and the tumour immune microenvironment (TIME) in breast cancer. More data and investigations are required to corroborate these findings.

Abbreviations

LRGs: Lipid metabolic-related genes; FABP7: Fatty acid binding protein 7; NDUFB1: NADH:ubiquinone oxidoreductase subunit AB1; TME: The tumour microenvironment; TIME: The tumour immune microenvironment; TNBC: Triple negative breast cancer; HER2: Human epidermal growth factor receptor 2; GO: Gene Ontology; MF: Molecular function; BP: Biological pathways; CC: Cellular components; KEGG: Kyoto Encyclopedia of Genes and Genomes; TMB: Tumour mutation burden; MSI: Microsatellite instability; NK cell: Natural killer cell

Supplementary Information

The online version contains supplementary material available at <https://doi.org/10.1186/s12944-022-01651-9>.

Additional file 1: Table S1. Characteristics of patients with breast cancer in the TCGA cohort.

Additional file 2.

Acknowledgements

The authors thank the teachers and students for their help.

Authors' contributions

All authors participated in the analysis of the data, drafted or revised the article, gave final approval to the version to be published, and agreed to take responsibility for all aspects of the work. The author(s) read and approved the final manuscript.

Funding

This work was supported by the National Natural Science Foundation of China (81702593).

Availability of data and materials

The datasets used and/or analysed in this study are available from the corresponding author upon reasonable request.

Declarations

Ethics approval and consent to participate

Not applicable.

Consent for publication

Not applicable.

Competing interests

The authors report no conflicts of interest in this work.

Received: 18 October 2021 Accepted: 15 December 2021

Published online: 13 May 2022

References

- Sung H, Ferlay J, Siegel RL, Laversanne M, Soerjomataram I, Jemal A, et al. Global cancer statistics 2020: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. *CA Cancer J Clin.* 2021; (3):209–49. <https://doi.org/10.3322/caac.21660>.
- Yeo SK, Guan JL. Breast Cancer: multiple subtypes within a tumor? *Trends Cancer.* 2017;3(11):753–60. <https://doi.org/10.1016/j.trecan.2017.09.001>.
- Anastasiadi Z, Lianos GD, Ignatiadou E, Harissis HV, Mitsis M. Breast cancer in young women: an overview. *Updat Surg.* 2017;69(3):313–7. <https://doi.org/10.1007/s13304-017-0424-1>.
- Lyons TG. Targeted therapies for triple-negative breast Cancer. *Curr Treat Options in Oncol.* 2019;20(11):82. <https://doi.org/10.1007/s11864-019-0682-x>.
- Xiao Z, Dai Z, Locasale JW. Metabolic landscape of the tumor microenvironment at single cell resolution. *Nat Commun.* 2019;10(1):3763. <https://doi.org/10.1038/s41467-019-11738-0>.
- Wu K, Lin K, Li X, Yuan X, Xu P, Ni P, et al. Redefining tumor-associated macrophage subpopulations and functions in the tumor microenvironment. *Front Immunol.* 2020;11:1731. <https://doi.org/10.3389/fimmu.2020.01731>.
- Hessmann E, Buchholz SM, Demir IE, Singh SK, Gress TM, Ellenrieder V, et al. Microenvironmental determinants of pancreatic Cancer. *Physiol Rev.* 2020; 100(4):1707–51. <https://doi.org/10.1152/physrev.00042.2019>.
- Lei X, Lei Y, Li JK, Du WX, Li RG, Yang J, et al. Immune cells within the tumor microenvironment: biological functions and roles in cancer immunotherapy. *Cancer Lett.* 2020;470:126–33. <https://doi.org/10.1016/j.canlet.2019.11.009>.
- Chen F, Chen J, Yang L, Liu J, Zhang X, Zhang Y, et al. Extracellular vesicle-packaged HIF-1 α -stabilizing lncRNA from tumour-associated macrophages regulates aerobic glycolysis of breast cancer cells. *Nat Cell Biol.* 2019;21(4): 498–510. <https://doi.org/10.1038/s41556-019-0299-0>.
- Wang YP, Lei QY. Perspectives of reprogramming breast Cancer metabolism. *Adv Exp Med Biol.* 2017;1026:217–32. https://doi.org/10.1007/978-981-10-6020-5_10.
- Boroughs LK, DeBerardinis RJ. Metabolic pathways promoting cancer cell survival and growth. *Nat Cell Biol.* 2015;17(4):351–9. <https://doi.org/10.1038/ncb3124>.
- Fearon KC, Glass DJ, Guttridge DC. Cancer cachexia: mediators, signaling, and metabolic pathways. *Cell Metab.* 2012;16(2):153–66. <https://doi.org/10.1016/j.cmet.2012.06.011>.
- Sevinsky CJ, Khan F, Kokabee L, Darehshouri A, Maddipati KR, Conklyn DS. NDRG1 regulates neutral lipid metabolism in breast cancer cells. *Breast Cancer Res.* 2018;20(1):55. <https://doi.org/10.1186/s13058-018-0980-4>.
- Zhang ZG, Zhang HS, Sun HL, Liu HY, Liu MY, Zhou Z. KDM5B promotes breast cancer cell proliferation and migration via AMPK-mediated lipid metabolism reprogramming. *Exp Cell Res.* 2019;379(2):182–90. <https://doi.org/10.1016/j.yexcr.2019.04.006>.
- Keckesova Z, Donaher JL, De Cock J, Freinkman E, Lingrell S, Bachovchin DA, et al. LACTB is a tumour suppressor that modulates lipid metabolism and cell state. *Nature.* 2017;543(7647):681–6. <https://doi.org/10.1038/nature21408>.
- Madden E, Logue SE, Healy SJ, Manie S, Samali A. The role of the unfolded protein response in cancer progression: from oncogenesis to chemoresistance. *Biol Cell.* 2019;111(1):1–17. <https://doi.org/10.1111/boc.201800050>.
- Cordero A, Kanojia D, Miska J, Panek WK, Xiao A, Han Y, et al. FABP7 is a key metabolic regulator in HER2+ breast cancer brain metastasis. *Oncogene.* 2019;38(37):6445–60. <https://doi.org/10.1038/s41388-019-0893-4>.
- Hou T, Zhang R, Jian C, Ding W, Wang Y, Ling S, et al. NDUFB1 confers cardio-protection by enhancing mitochondrial bioenergetics through coordination of respiratory complex and supercomplex assembly. *Cell Res.* 2019;29(9):754–66. <https://doi.org/10.1038/s41422-019-0208-x>.
- Kwa MJ, Adams S. Checkpoint inhibitors in triple-negative breast cancer (TNBC): where to go from here. *Cancer.* 2018;124(10):2086–103. <https://doi.org/10.1002/cncr.31272>.

20. Yu LY, Tang J, Zhang CM, Zeng WJ, Yan H, Li MP, et al. New Immunotherapy Strategies in Breast Cancer. *Int J Environ Res Public Health*. 2017;14(1):14(1). <https://doi.org/10.3390/ijerph14010068>.
21. Emens LA. Breast Cancer immunotherapy: facts and hopes. *Clin Cancer Res*. 2018;24(3):511–20. <https://doi.org/10.1158/1078-0432.CCR-16-3001>.
22. Ritchie ME, Phipson B, Wu D, Hu Y, Law CW, Shi W, Smyth GK: limma powers differential expression analyses for RNA-sequencing and microarray studies. *Nucleic Acids Res*. 2015;43(7):e47. <https://doi.org/10.1093/nar/gkv007>.
23. Symsock A, Gaunaud I, Thaper A, Springer B, Bennett C, Clemens S, et al. Usability assessment of the rehabilitation lower-limb orthopedic assistive device by service members and veterans with lower limb loss. *Mil Med*. 2021;186(3–4):379–86. <https://doi.org/10.1093/milmed/usaa428>.
24. Szklarczyk D, Gable AL, Nastou KC, Lyon D, Kirsch R, Pyysalo S, et al. The STRING database in 2021: customizable protein-protein networks, and functional characterization of user-uploaded gene/measurement sets. *Nucleic Acids Res*. 2021;49(D1):D605–d612. <https://doi.org/10.1093/nar/gkaa1074>.
25. The Gene Ontology Resource. 20 years and still GOing strong. *Nucleic Acids Res*. 2019;47(D1):D330–d338. <https://doi.org/10.1093/nar/gky1055>.
26. Zhang YF, Huang Y, Ni YH, Xu ZM. Systematic elucidation of the mechanism of geraniol via network pharmacology. *Drug Des Devel Ther*. 2019;13:1069–75. <https://doi.org/10.2147/DDDT.S189088>.
27. Yu G, Wang LG, Han Y, He QY: clusterProfiler: an R package for comparing biological themes among gene clusters. *Omic*. 2012;16(5):284–7. <https://doi.org/10.1089/omi.2011.0118>.
28. Garge NR, Bobashev G, Eggleston B. Random forest methodology for model-based recursive partitioning: the mobForest package for R. *BMC Bioinformatics*. 2013;14(1):125. <https://doi.org/10.1186/1471-2105-14-125>.
29. Tao C, Huang K, Shi J, Hu Q, Li K, Zhu X. Genomics and prognosis analysis of epithelial-mesenchymal transition in glioma. *Front Oncol*. 2020;10:183. <https://doi.org/10.3389/fonc.2020.00183>.
30. Pascual J, Turner NC. Targeting the PI3-kinase pathway in triple-negative breast cancer. *Ann Oncol*. 2019;30(7):1051–60. <https://doi.org/10.1093/annonc/mdz133>.
31. Petitprez F, Meylan M, de Reyniès A, Sautès-Fridman C, Fridman WH. The tumor microenvironment in the response to immune checkpoint blockade therapies. *Front Immunol*. 2020;11:784. <https://doi.org/10.3389/fimmu.2020.00784>.
32. Baci D, Bosi A, Gallazzi M, Rizzi M, Noonan DM, Poggi A, et al. The ovarian cancer Tumor Immune Microenvironment (TIME) as target for therapy: a focus on innate immunity cells as therapeutic effectors. *Int J Mol Sci*. 2020; 21(9). <https://doi.org/10.3390/ijms21093125>.
33. Criscitiello C, Curigliano G. Immunotherapy of breast Cancer. *Prog Tumor Res*. 2015;42:30–43. <https://doi.org/10.1159/000437183>.
34. Sun L, Suo C, Li ST, Zhang H, Gao P. Metabolic reprogramming for cancer cells and their microenvironment: beyond the Warburg effect. *Biochim Biophys Acta Rev Cancer*. 2018;1870(1):51–66. <https://doi.org/10.1016/j.bbcan.2018.06.005>.
35. DeBerardinis RJ, Chandel NS. Fundamentals of cancer metabolism. *Sci Adv*. 2016;2(5):e1600200. <https://doi.org/10.1126/sciadv.1600200>.
36. Dias AS, Almeida CR, Helguero LA, Duarte IF. Metabolic crosstalk in the breast cancer microenvironment. *Eur J Cancer*. 2019;121:154–71. <https://doi.org/10.1016/j.ejca.2019.09.002>.
37. Yi M, Li J, Chen S, Cai J, Ban Y, Peng Q, et al. Emerging role of lipid metabolism alterations in Cancer stem cells. *J Exp Clin Cancer Res*. 2018; 37(1):118. <https://doi.org/10.1186/s13046-018-0784-5>.
38. Wang T, Fahrman JF, Lee H, Li YJ, Tripathi SC, Yue C, et al. JAK/STAT3-Regulated Fatty Acid β -Oxidation Is Critical for Breast Cancer Stem Cell Self-Renewal and Chemoresistance. *Cell Metab*. 2018;27(1): 136–150.e135.
39. Kwong SC, Jamil AHA, Rhodes A, Taib NA, Chung I. Metabolic role of fatty acid binding protein 7 in mediating triple-negative breast cancer cell death via PPAR- α signaling. *J Lipid Res*. 2019;60(11):1807–17. <https://doi.org/10.1194/jlr.M092379>.
40. Sultan G, Zubair S, Tayubi IA, Dahms HU, Madar IH. Towards the early detection of ductal carcinoma (a common type of breast cancer) using biomarkers linked to the PPAR(γ) signaling pathway. *Bioinformatics*. 2019; 15(11):799–805. <https://doi.org/10.6026/97320630015799>.
41. Miricescu D, Totan A, Stanescu S II, Badoiu SC, Stefani C, Greabu M. PI3K/AKT/mTOR signaling pathway in breast cancer: from molecular landscape to clinical aspects. *Int J Mol Sci*. 2020;22(1):22(1). <https://doi.org/10.3390/ijms22010173>.
42. Hao J, Zhang Y, Yan X, Yan F, Sun Y, Zeng J, et al. Circulating adipose fatty acid binding protein is a new link underlying obesity-associated breast/mammary tumor development. *Cell Metab*. 2018;28(5):689–705.e685.
43. Barroso-Sousa R, Jain E, Cohen O, Kim D, Buendia-Buendia J, Winer E, et al. Prevalence and mutational determinants of high tumor mutation burden in breast cancer. *Ann Oncol*. 2020;31(3):387–94. <https://doi.org/10.1016/j.annonc.2019.11.010>.
44. Dudley JC, Lin MT, Le DT, Eshleman JR. Microsatellite instability as a biomarker for PD-1 blockade. *Clin Cancer Res*. 2016;22(4):813–20. <https://doi.org/10.1158/1078-0432.CCR-15-1678>.
45. Ellis H, Ma CX. PI3K inhibitors in breast Cancer therapy. *Curr Oncol Rep*. 2019;21(12):110. <https://doi.org/10.1007/s11912-019-0846-7>.
46. Peng WX, Huang JG, Yang L, Gong AH, Mo YY. Linc-RoR promotes MAPK/ERK signaling and confers estrogen-independent growth of breast cancer. *Mol Cancer*. 2017;16(1):161. <https://doi.org/10.1186/s12943-017-0727-3>.
47. Cui P, Chen Y, Waili N, Li Y, Ma C, Li Y. Associations of serum C-peptide and insulin-like growth factor binding proteins-3 with breast cancer deaths. *PLoS One*. 2020;15(11):e0242310. <https://doi.org/10.1371/journal.pone.0242310>.
48. Bastid J, Dejou C, Docquier A, Bonnefoy N. The emerging role of the IL-17B/IL-17RB pathway in Cancer. *Front Immunol*. 2020;11:718. <https://doi.org/10.3389/fimmu.2020.00718>.
49. Liu Q, Li A, Tian Y, Wu JD, Liu Y, Li T, et al. The CXCL8-CXCR1/2 pathways in cancer. *Cytokine Growth Factor Rev*. 2016;31:61–71. <https://doi.org/10.1016/j.cytogfr.2016.08.002>.
50. Kulkoyluoglu-Cotul E, Arca A, Madak-Erdogan Z. Crosstalk between estrogen signaling and breast Cancer metabolism. *Trends Endocrinol Metab*. 2019; 30(1):25–38. <https://doi.org/10.1016/j.tem.2018.10.006>.
51. Saha T, Makar S, Swetha R, Gutti G, Singh SK. Estrogen signaling: an emanating therapeutic target for breast cancer treatment. *Eur J Med Chem*. 2019;177:116–43. <https://doi.org/10.1016/j.ejmech.2019.05.023>.
52. Ko SH, Kim HS. Menopause-associated lipid metabolic disorders and foods beneficial for postmenopausal women. *Nutrients*. 2020;12(1):12(1). <https://doi.org/10.3390/nu12010202>.
53. Leone RD, Powell JD. Metabolism of immune cells in cancer. *Nat Rev Cancer*. 2020;20(9):516–31. <https://doi.org/10.1038/s41568-020-0273-y>.

Publisher's Note

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

Ready to submit your research? Choose BMC and benefit from:

- fast, convenient online submission
- thorough peer review by experienced researchers in your field
- rapid publication on acceptance
- support for research data, including large and complex data types
- gold Open Access which fosters wider collaboration and increased citations
- maximum visibility for your research: over 100M website views per year

At BMC, research is always in progress.

Learn more biomedcentral.com/submissions

