

STATE-OF-THE-ART REVIEW

Cancer Immunotherapy Beyond Checkpoint Blockade



JACC: CardioOncology State-of-the-Art Review

Nathan E. Welty, MD, PhD,^{a,b} Saar I. Gill, MD, PhD^{a,b}

ABSTRACT

Avoidance of immune destruction is recognized as one of the hallmarks of cancer development. Although first predicted as a potential antitumor treatment modality more than 50 years ago, the widespread clinical use of cancer immunotherapies has only recently become a reality. Cancer immunotherapy works by reactivation of a stalled pre-existing immune response or by eliciting a de novo immune response, and its toolkit comprises antibodies, vaccines, cytokines, and cell-based therapies. The treatment paradigm in some malignancies has completely changed over the past 10 to 15 years. Massive efforts in preclinical development have led to a surge of clinical trials testing innovative therapeutic approaches as monotherapy and, increasingly, in combination. Here we provide an overview of approved and emerging antitumor immune therapies, focusing on the rich landscape of therapeutic approaches beyond those that block the canonical PD-1/PD-L1 and CTLA-4 axes and placing them in the context of the latest understanding of tumor immunology. (J Am Coll Cardiol CardioOnc 2022;4:563-578) © 2022 The Authors. Published by Elsevier on behalf of the American College of Cardiology Foundation. This is an open access article under the CC BY-NC-ND license (<http://creativecommons.org/licenses/by-nc-nd/4.0/>).

A fundamental challenge for cancer treatments such as chemotherapy and targeted therapies is that cancers undergo natural selection. Genetic and epigenetic modifications accumulate in cancer subclones during treatment, and those that provide a selective advantage enable tumor growth or recurrence despite continued drug therapy. The power of immune therapy over other forms of treatment therefore rests on several fundamental features of the adaptive immune system: 1) antigen specificity: immune receptors are competitively tuned to produce exquisite specificity for particular antigens; 2) clonal selection: cells that obtain positive signaling

via these receptors expand and mature to become more adept at eliminating foreign antigens; and 3) memory: antigen-specific clones persist over the lifetime of the host, serving as a long-term reservoir to control foreign antigens. The combination of specificity, adaptation, and durability offered by immune-based therapies thus combats cancer clonal evolution through natural selection and offers the potential for long-term control without prohibitive off-target toxicities.

A basic principle of cancer immunotherapy is that oncogenesis requires escape from immune surveillance.¹ Because of immune ignorance (de novo

From the ^aCenter for Cellular Immunotherapies, University of Pennsylvania, Philadelphia, Pennsylvania, USA; and the ^bDivision of Hematology/Oncology, Department of Medicine, Perelman School of Medicine, University of Pennsylvania, Philadelphia, Pennsylvania, USA.

Kerry L. Reynolds, MD, served as the Guest Associate Editor for this paper. Paaladinesh Thavendiranathan, MD, MSc, served as the Guest Editor-in-Chief for this paper.

The authors attest they are in compliance with human studies committees and animal welfare regulations of the authors' institutions and Food and Drug Administration guidelines, including patient consent where appropriate. For more information, visit the [Author Center](#).

Manuscript received July 19, 2022; revised manuscript received November 2, 2022, accepted November 3, 2022.

**ABBREVIATIONS
AND ACRONYMS**

alloHCT = allogeneic hematopoietic stem cell transplantation
BITE = bispecific T cell engager
CAR = chimeric antigen receptor
CRS = cytokine-release syndrome
FDA = U.S. Food and Drug Administration
HLA = human leukocyte antigen
ICI = immune checkpoint inhibitor
IL = interleukin
mAb = monoclonal antibody
NK = natural killer
NSCLC = non-small cell lung cancer
TIL = tumor-infiltrating lymphocyte

absence of mutation-specific immune recognition), tolerance (acquired absence of mutation-specific immune recognition), or suppression (development of counter-regulatory mechanisms that prevent immune recognition), cancers that progress from precancerous lesions toward growth and metastasis are de facto insensitive to normal mechanisms of immunologic pressure against non-self-antigens. This process is known as cancer immunoediting.^{2,3} Successful cancer immunotherapies must overcome cancer immunoediting, either by provoking a de novo antitumor immune response or by reactivating an existing immune response that has become ineffective. In this review we provide an overview of approved and emerging antitumor immune therapies as they fit within these 2 broad mechanisms of action, discussing both cell-based and noncellular therapies, their mechanisms of action, and toxicities (**Central Illustration**). We emphasize the most recent

and clinically advanced approaches apart from immune checkpoint inhibitors (ICIs) targeting PD-1/PD-L1 or CTLA-4, which are amply covered in numerous other dedicated reviews.

**PROVOKING AN ANTITUMOR
IMMUNE RESPONSE**

TUMOR-SPECIFIC ANTIGENS. Cancer cells exhibit genetic instability and accumulate somatic mutations, gene fusions, chromosomal copy number variations, and other genomic, epigenetic, and post-transcriptional modifications.⁴ These events generate an intracellular pool of altered, potentially immunogenic peptides that are processed and presented on human leukocyte antigen (HLA) molecules to T cells, thereby stimulating immune responses⁵ (**Figure 1**). Peptides that are present only in malignant cells, known as neoantigens, can also act as therapeutic targets, demonstrated by data showing that neoantigen burden is a biomarker for response to immune checkpoint blockade.⁶⁻⁹ Neoantigens represent attractive targets because they are not expressed in normal tissues and therefore have higher potential immunogenicity and lower potential toxicity. However, they have also been challenging targets for several reasons. First, these mutations are generally private (ie, not shared between individuals), and drug development therefore requires

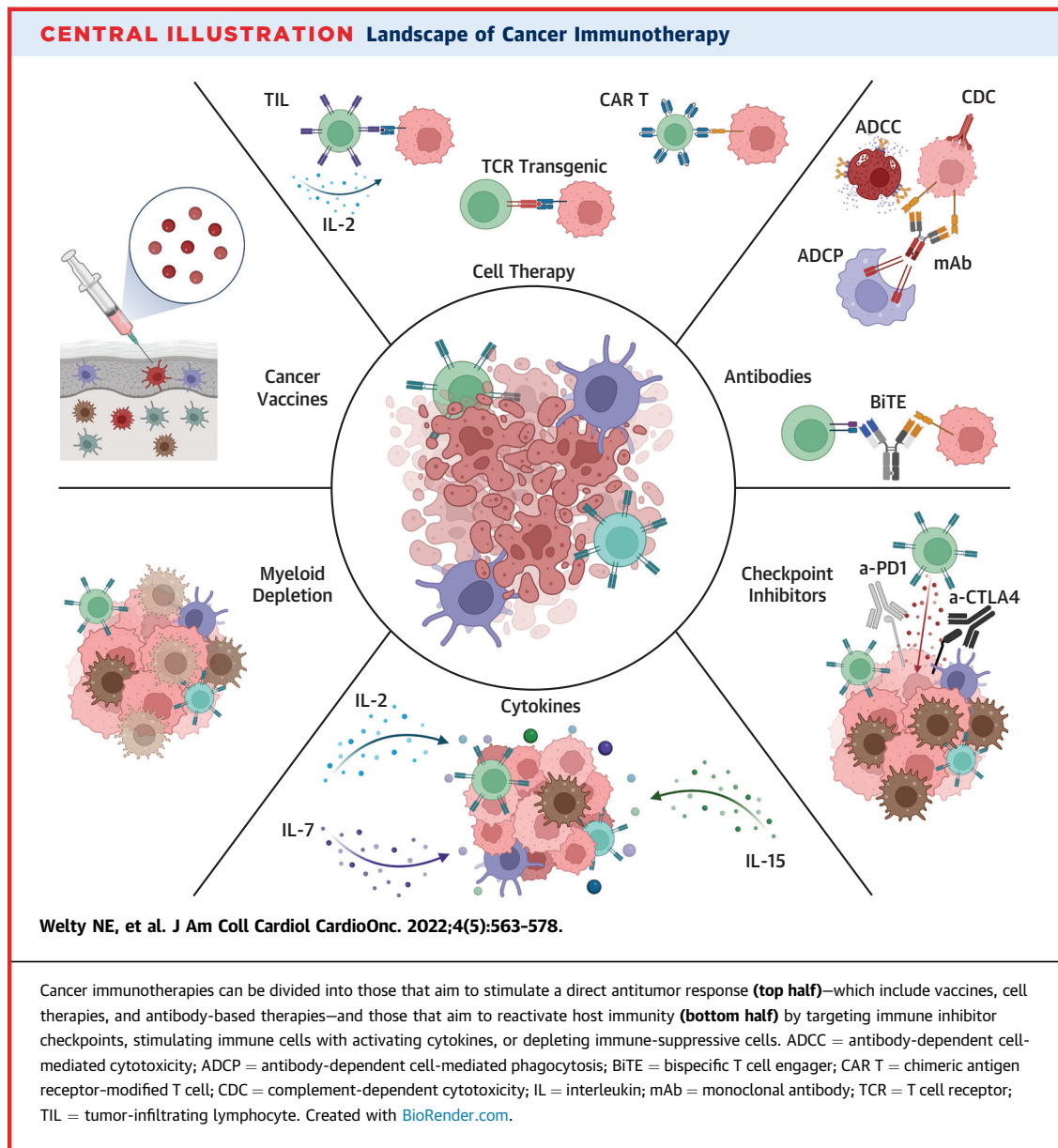
HIGHLIGHTS

- Cancer immunotherapies have undergone rapid commercialization over the past decade.
- Immunotherapy mechanisms, responses, and toxicities diverge from prior treatments.
- Future advances must build on these unique traits to address immunotherapy resistance.

intensive personalized medicine strategies such as whole-exome sequencing and bioinformatic analyses at the individual patient level.^{10,11} Second, neoantigen peptides are immunogenic only when presented by particular HLA molecules. HLA loci are among the most highly polymorphic regions of the human genome,^{12,13} presenting difficulties for drug development at the population level as well. Nonetheless, vaccine- and cell-based therapies targeting tumor-specific neoantigens are in early-phase development.¹¹

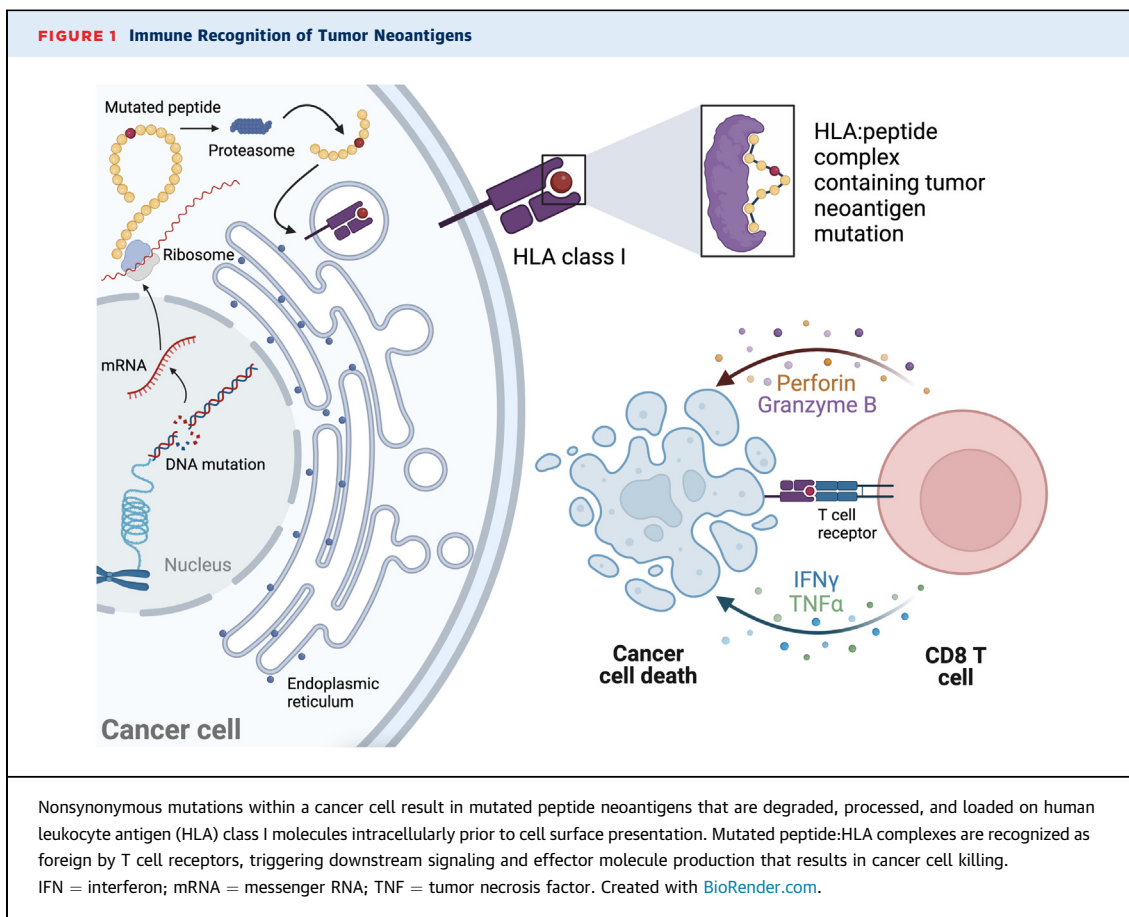
Cancer vaccines. Clinical trials targeting tumor-specific neoantigens largely use cancer vaccines to generate new immune responses for cancer prevention or treatment. In initial trials in melanoma, neoantigen-based cancer vaccines could expand existing antitumor T cells and elicit de novo T cell responses.¹⁴⁻¹⁶ However, results from phase 3 cancer vaccine trials, which include both neoantigen and non-neoantigen targets, have been disappointing.¹⁷ Failure of the elicited neoantigen immune responses to control cancer is multifactorial, and antigen target selection is only one of many barriers. However, an emerging strategy to improve neoantigen selection is to target mutated peptides in genes that are obligate for cancer development. These “driver” mutations are potentially favorable for a few reasons. Because they are oncogenic, these mutations are present in all cancer subclones and indispensable for continued cancer growth. Thus, further genetic aberrations that permit cancer cells to lose the mutation are unlikely. Additionally, these mutations are shared across individuals and tumor types, circumventing challenges of entirely personalized therapy.^{5,18-20}

Early-phase trials of neoantigen vaccination targeting driver mutations are currently recruiting (eg, [NCT03592888](#), [NCT04117087](#), [NCT05202561](#), [NCT04853017](#)). However, the HLA restriction of neoantigens remains a potential limitation.¹³



Additionally, HLA loss of heterozygosity in cancer cells is a known mechanism of immune escape,²¹ occurring, for instance, in up to 40% of patients with non-small cell lung cancer (NSCLC),²² and can prevent T cell recognition. Avoiding this phenomenon may require vaccination against multiple neopeptides presented on different HLAs, as is under way in a trial of vaccination against hundreds of recurrent mutations in patients with DNA mismatch repair-deficient cancers²³ (NCT04041310), which have a high neoantigen burden. This approach may be less

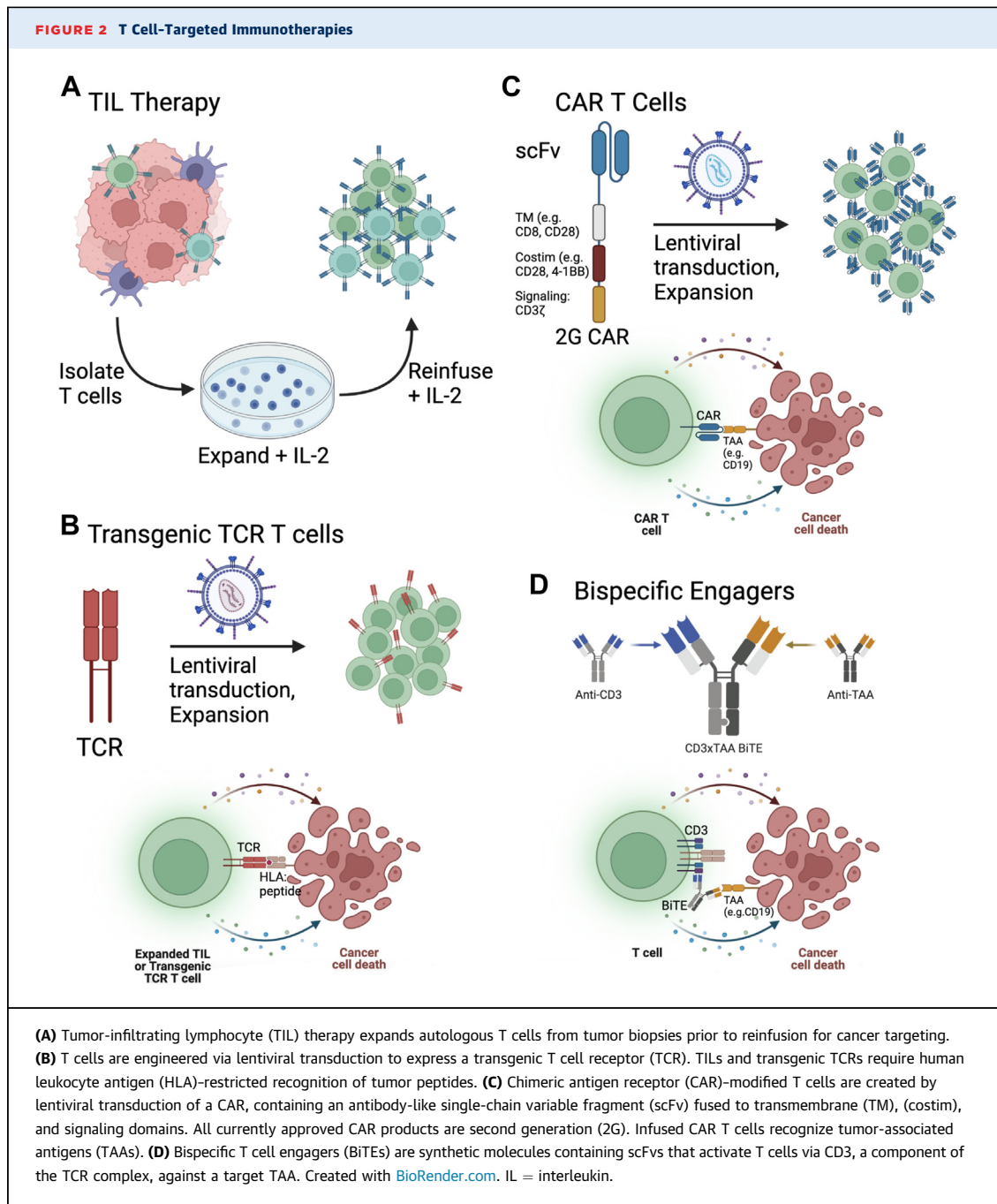
effective in cancers with a lower neoantigen burden, although several clinical trials of messenger RNA-based vaccines targeting multiple neoantigens are under way in melanoma, head and neck cancers, colorectal cancer, and others (eg, NCT03313778, NCT03897881, NCT03815058, NCT03289962). Ultimately the optimal strategy for anticancer vaccination against any antigen type remains unclear, despite trials of cell-based and acellular approaches using DNA, RNA, and peptides with various adjuvant strategies.^{11,17} Generally, vaccines work best in



combination with therapies that counteract immune suppression and in settings of low-volume disease, limiting use in patients with the most advanced cancers.¹⁷ Indeed, the only U.S. Food and Drug Administration (FDA)-approved vaccine for metastatic malignancy, sipuleucel-T, though well tolerated, shows only minimal clinical benefit²⁴ and is most effective in those patients with low-volume prostate cancer.²⁵ The only cardiovascular toxicity associated with sipuleucel-T is a low incidence of hypertension around the time of infusion²⁴; cardiotoxicities in phase 3 trials of other vaccines were uncommon and not clearly treatment related.²⁶⁻²⁸

Nonengineered cell therapies. One of the earliest cancer immunotherapies was autologous tumor-infiltrating lymphocytes (TILs) that were harvested from resected melanomas, expanded *in vitro* with interleukin (IL)-2, and then reinfused back into patients after lymphodepleting chemotherapy and in conjunction with additional *in vivo* IL-2 treatment²⁹ (Figure 2A). Later work demonstrated that antigens targeted by TIL therapy and associated with clinical

responses are in fact neoantigens.³⁰ Despite its early start, TIL therapy was overshadowed by the development of ICIs, which have an improved toxicity profile compared with systemic IL-2 and do not require the prolonged manufacturing process for personalized cellular therapy. Nonetheless, TIL therapies show a response rate of about 30% to 40%, even in patients with melanoma previously treated with checkpoint inhibitors,³¹⁻³³ suggesting nonoverlapping mechanisms of action. Responses are associated with high-dose IL-2, higher infused TIL number, and possibly with lack of prior anti-CTLA-4 treatment.³¹⁻³³ Responses can be very durable, and toxicities are typically within the first few weeks of treatment.³¹⁻³³ Although no phase 3 data have been reported for any TIL product, the first commercial TIL product for melanoma, lifileucel, has shown response rates of approximately 30% and received fast-track designation from the FDA.³³ TIL therapy is also an emerging treatment modality for other epithelial cancers, with early-phase clinical trials completed or ongoing in NSCLC (NCT04614103), cervical cancer (NCT03108495



), and others (NCT03449108).³⁴⁻³⁶ Cardiotoxicities observed with TIL therapy are common and include hypotension, arrhythmias, and edema (Table 1), with a major contribution from systemic IL-2 administration, although conditioning chemotherapy and TIL factors may also contribute.^{33,37,38}

Engineered cell therapies. While TILs are a non-engineered T cell therapy, advances in molecular

biology have enabled cell therapies that are augmented through genetic modification to enhance antitumor activity (Figure 2). Engineered cell therapies are usually generated by viral transduction with synthetic or naturally occurring molecules capable of modifying immune cell activity, although other approaches, such as clustered regularly interspaced short palindromic repeats-based gene editing, are

TABLE 1 Cardiovascular Toxicities Associated With Selected Immunotherapies

Agent(s)	Disease	Toxicity (Frequency)	Ref. #
Lifileucel (TIL) + chemotherapy + IL-2	Melanoma	Hypotension (36%), tachycardia (35%), edema (26%)	33
High-dose IL-2	RCC	Hypotension (96%), arrhythmia (14%), myocardial ischemia (2%), myocardial infarction (2%), cardiac arrest (2%), myocarditis (1%)	150
Tebentafusp	Uveal melanoma	CRS (89%), hypotension (38%), hypertension (6%)	68
Margetuximab	Breast cancer	Hypertension (5.3%), grade 3+ LV dysfunction (1.1%)	53
Obinutuzumab	NHL, CLL	Cardiac events (10.7%) ^a	54,174
Blinatumomab	B-ALL	CRS (14.2%), hypotension (12%), cardiac disorders (2.2% with blinatumomab vs 2.8% with chemotherapy)	175
CAR T (various products)	NHL (88%), myeloma (8%)	CRS (59%), elevated troponin (54%), decreased LVEF (28%), decompensated HF (3%-10%), arrhythmia (3%-4%), CV death (4%)	98-101
Allogeneic HCT	Various	Ischemic heart disease (5.4%), cardiomyopathy (12.3%), rhythm disorder (15.2%), hypertension (31.6%), cardiovascular death (5.2%)	114
PD-1 inhibitor ± CTLA-4 inhibitor (various products)	Various	Myocarditis (0.06%-0.15% for PD-1 vs 0.26%-0.27% for combination)	141,142
Pexidartinib	TGCT	Hypertension (15%)	162

^aDefined as any preferred term in the System Organ Class cardiac disorders.
B-ALL = B cell acute lymphoblastic leukemia; CAR T = chimeric antigen receptor–modified T cell; CLL = chronic lymphocytic leukemia; CRS = cytokine-release syndrome (any grade, various grading systems); CV = cardiovascular; HCT = hematopoietic stem cell transplantation; HF = heart failure; IL = interleukin; LV = left ventricular; LVEF = left ventricular ejection fraction; NHL = non-Hodgkin lymphoma; RCC = renal cell carcinoma; TGCT = tenosynovial giant cell tumor; TIL = tumor-infiltrating lymphocyte.

also rapidly developing.³⁹⁻⁴² For instance, T cells are readily harvested from peripheral blood by apheresis and virally transduced to express a receptor targeting known neoantigens prior to expansion and reinfusion into patients, thereby stimulating an antitumor immune response (Figure 2B). Clinical trials using this method to target driver neoantigen mutations in solid tumors are ongoing (eg, NCT03745326, NCT03190941, NCT04146298). These T cells are engineered to express an HLA-restricted murine T cell receptor recognizing mutated Kristen rat sarcoma viral oncogene homologue peptides complexed with HLA-A*11:01.⁴³ Similarly, a recent case report described a clinical response in a patient treated with engineered autologous T cells transduced with an HLA-C*08:02-restricted receptor targeting a mutant Kristen rat sarcoma viral oncogene homologue G12D peptide.⁴⁴ Further discovery efforts to identify antigen recognition domains that specifically bind neoantigen:HLA complexes are ongoing.^{45,46} These may enable the development of neoantigen-targeting chimeric antigen receptor (CAR) T cells (Figure 2C), which typically use antibody-derived single-chain variable fragments to recognize their targets, or even noncellular immunotherapies, such as bispecific T cell engagers (BiTEs)⁴⁷ (Figure 2D). Important safety concerns with these agents include cross-recognition of alternative peptides presented by HLA molecules, as occurred with MAGE-A3-targeted transgenic T cell receptor T cells that caused cardiogenic shock and early mortality in 2 patients through recognition of a titin

peptide in beating cardiac tissue.^{48,49} Developing preclinical model systems that can predict such toxicities has been challenging.⁵⁰

TUMOR-ASSOCIATED ANTIGENS. As opposed to tumor-specific neoantigens, most immunotherapy targets are shared surface antigens that are highly expressed on tumor cells but are not necessarily cancer specific. In hematologic malignancies, these targets are typically lineage-associated maturation antigens that are expressed during development of both normal and malignant cells, for instance, CD19 and CD20 in B cell neoplasms and CD33 in myeloid neoplasms. In some malignancies, they may be oncofetal (also known as “cancer germline” or “cancer testis”) antigens that are typically expressed during fetal development and in cancer but not on normal adult tissues, antigens that are overexpressed on tumor cells, or normal antigens with aberrant post-transcriptional modifications, such as glycosylation variants, in the context of cancer.²⁰ Unlike neoantigens, these targets present higher potential for on-target, off-tumor toxicities because of expression on normal cells in addition to malignant ones. Despite these drawbacks, their more uniform expression both within tumors and across individuals, and the lack of HLA restriction, have been more amenable for drug development.

Monoclonal antibodies. Monoclonal antibodies (mAbs) directed at cell membrane antigens form the largest class of FDA-approved cancer immunotherapies. By “creating” antigens for immune recognition

from normally nonimmunogenic cell surface proteins, mAbs induce antitumor immunity through their Fc domains, which engage myeloid, natural killer (NK) cells, and other immune cells to cause antibody-dependent cellular cytotoxicity, antibody-dependent cell phagocytosis, and/or complement-dependent cytotoxicity of targeted cells⁵¹ (**Central Illustration**). Although in hematologic malignancies direct immune-mediated cytotoxicity is appreciated as the major mechanism of action of mAbs, in solid tumors attention has focused more on their disruption of cancer cell signaling pathways rather than their immune effects.⁵² However, the clinical importance of these immune effects in solid tumors is recently highlighted by the FDA approval of margetuximab-cmkb, a mAb targeting the same HER-2 epitope as trastuzumab, which was previously approved to treat HER-2-positive breast cancer, but with a redesigned Fc domain to enhance immune-mediated cytotoxicity. Margetuximab showed modest but significantly improved progression-free survival compared with trastuzumab (5.8 months vs 4.9 months) given with chemotherapy in patients with previously treated breast cancer. Despite a small increase in infusion reactions, rates of cardiotoxicity (**Table 1**) were similar to those seen with trastuzumab, which is known to be associated with decreased left ventricular ejection fraction.⁵³ However, other trials of engineered mAbs designed to enhanced immune activity have been less successful. For instance, the newer engineered anti-CD20 mAbs obinutuzumab and ofatumumab showed no improvement over standard rituximab for the treatment of non-Hodgkin lymphomas despite increased immune toxicities and infusion reactions. Overall cardiac events are also increased with these agents compared with rituximab, although these were not well defined in the registration trials, and the mechanism remains unclear (**Table 1**).^{54,55}

Naked mAbs have limited single-agent activity, and most regimens combine antibodies with traditional chemotherapy.⁵⁶⁻⁵⁹ This finding is somewhat counterintuitive given the immunosuppression caused by traditional chemotherapy. This synergy may relate to immunogenic cell death, depletion of immune-suppressive cells, or other immune stimulatory effects of chemotherapy.⁶⁰⁻⁶³ Beyond naked mAbs, numerous antibody-drug conjugates have received FDA approval. Antibody-drug conjugates exploit the tumor-targeting capacity of mAbs; upon cross-linking their antibody target antigen, these drugs deliver high intracellular doses of chemotherapy, radiation, or other cytotoxic payloads directly to cancer cells,⁵² expanding the therapeutic

window and avoiding prohibitive toxicities that would otherwise be associated with these agents. Although antibody-drug conjugates have much higher single-agent activity, their mechanism of action is due chiefly to the cytotoxicity of the drug conjugate. Antibody-drug conjugates and their toxicities, including cardiotoxicities, have been extensively reviewed in other publications.^{64,65}

Bispecific engagers. Although most antibody-based treatments target a single tumor-associated antigen, there is increasing research and development of antibody-like bispecific products that simultaneously bind 2 different targets to mediate antitumor immunity (**Figure 2D**). Amivantamab-vmjw is one such agent that simultaneously targets epidermal growth factor and mesenchymal-epithelial transition factor receptors on the surface of NSCLC via single-chain variable fragment domains, mediating antitumor activity via its Fc domain. This drug received FDA approval for the treatment of patients with NSCLC with epidermal growth factor receptor exon 20 insertion mutations, showing a response rate of 40% in a population that does not respond to drugs that treat most epidermal growth factor receptor-mutated lung cancers.⁶⁶ Peripheral edema and hypoalbuminemia, known class effects of mesenchymal-epithelial transition factor inhibition, occur with this agent, but cardiac toxicities are not seen.⁶⁶ Like mAbs, this Fc-mediated mechanism of action only indirectly leads to T cell activation, as T cells generally do not express Fc receptors. However, other bispecific products, known as BiTEs, use antibody fragments to directly activate T cells. BiTEs typically contain a domain recognizing CD3, a component of the T cell receptor, linked to a second domain targeting a cancer cell surface antigen.⁶⁷ Binding of the BiTE to both targets mimics the action of T cell receptor HLA-peptide binding and activates T cell effector functions against the target cell, “redirecting” T cells to kill tumor targets they would not otherwise recognize.

Tebentafusp-tebn is a bispecific molecule that redirects CD3⁺ T cells to kill tumor cells displaying a peptide from the melanoma antigen gp100 in the context of HLA-A*02:01. It is the only FDA-approved BiTE for solid tumors, with 1-year overall survival of 73% compared with 59% in the control group in treated patients with uveal melanoma.⁶⁸ However, cutaneous toxicities with this medication are also observed, likely due to on-target, off-tumor toxicity, as gp100 is not a neoantigen but is also expressed in normal melanocytes.^{68,69} The only BiTE to receive FDA approval to date for hematologic malignancies is blinatumomab, a CD3 × CD19-targeting molecule that

TABLE 2 Current FDA-Approved CAR T Cell Products

Product (Brand Name)	Target	Costimulation/Unique Features	FDA Indication	ORR (CRR)	Ref. #
Tisagenlecleucel (Kymriah)	CD19	4-1BB	B-ALL, LBCL	82% (60%), 52% (40%)	176,177
Axicabtagene ciloleucel (Yescarta)	CD19	CD28	LBCL, FL	82% (40%), 92% (74%)	178,179
Brexucabtagene autoleucel (Tecartus)	CD19	CD28/T cell enrichment of apheresis product	B-ALL, MCL	71% (56%), 93% (67%)	180,181
Lisocabtagene maraleucel (Breyanzi)	CD19	4-1BB/separate manufacture/infusion of CD4/CD8 T cells	LBCL	73% (53%)	182
Idecabtagene vicleucel (Abecma)	BCMA	4-1BB	MM	73% (33%)	183
Ciltacabtagene autoleucel (Carvykti)	BCMA	4-1BB/two VHHs targeting distinct BCMA epitopes	MM	97% (67%)	184

BCMA = B cell maturation antigen; CRR = complete response rate; FDA = U.S. Food and Drug Administration; FL = follicular lymphoma; LBCL = large B cell lymphoma; MCL = mantle cell lymphoma; MM = multiple myeloma; ORR = overall/objective response rate; VHH = heavy chain variable domain; other abbreviations as in [Table 1](#).

activates killing of B cells, for the treatment of B cell acute lymphoblastic leukemia, with a response rate of 69% in relapsed/refractory patients.⁷⁰⁻⁷² Additional agents targeting CD20, CD38, or BCMA in B cell and plasma cell malignancies and CD33 or CD123 in myeloid leukemias are in clinical trials.⁶⁷ Contrasting with treatments whose mechanism of action is based on Fc recognition, which typically have low immune toxicity,^{66,73} immune-related adverse events with BiTEs are more common. Both cytokine-release syndrome (CRS) and neurotoxicity are observed, likely because of direct T cell engagement.^{68,74,75} Although cardiotoxicities were observed with blinatumomab, rates were not significantly higher compared with the chemotherapy treatment group ([Table 1](#)). Mechanisms of resistance to BiTEs remain underexplored, although CD19 loss by tumor cells can occur in patients treated with blinatumomab.^{71,76}

Cell therapies. Nonengineered T cell products enriched for tumor-associated antigen specificity can be generated by exposing peripheral blood-derived T cells to autologous antigen presenting cells loaded with peptides selected from a list of known tumor-associated antigens (eg, PRAME, MAGEA4, NYESO1, survivin), followed by ex vivo manufacturing with activating cytokines. Treatment with these products has generated interesting antitumor responses in patients with hematologic malignancies, with few toxicities and no cardiotoxicities.^{77,78} However, the most clinically advanced approaches to cell therapy involve the use of immune effector cells that are genetically engineered to target tumor-associated antigens. Immune effector cells include T cells, NK cells, NK/T cells, and macrophages. Of these, T cells bearing CARs represent the most advanced technology to date. CAR T cells are typically manufactured from autologous T cells isolated from peripheral blood and lentivirally transduced to express a CAR, which redirects their specificity toward a tumor-associated antigen, prior to reinfusion after

lymphodepleting chemotherapy ([Figure 2C](#)). CAR T cells have FDA labels for the treatment of pediatric and adult B cell acute lymphoblastic leukemia, adults with B cell non-Hodgkin lymphomas, and multiple myeloma ([Table 2](#)) and are in clinical trials for other hematologic and solid tumors, with encouraging early data in T cell malignancies,⁷⁹ Hodgkin lymphoma,⁸⁰ acute myeloid leukemia,⁸¹ and some gastrointestinal,⁸² prostate,⁸³ and brain⁸⁴ cancers. Two recent publications of CAR-modified NK cells⁸⁵ and NK/T cells⁸⁶ have also shown promise. Overall, as of September 2022, 347 clinical trials of CAR-modified immune effector cells were registered as actively recruiting on ClinicalTrials.gov, including 152 in the United States, 165 in China, and 31 in Europe. Most use T cells, whereas 6 use NK cells, 2 use NK/T cells, and 1 uses macrophages. An excellent recent review of the therapeutic landscape for CAR-engineered cells identified 64 different targets for T, NK, or NK/T cells in more than 500 clinical trials encompassing more than 20,000 patients.⁸⁷ The potential of genetically engineered cell therapies to be a “one and done” treatment for cancer is underscored by reports of deep and durable responses in patients with multiply relapsed hematologic malignancies such as chronic lymphocytic leukemia,⁸⁸ results that may justify the high initial cost of these therapies.⁸⁹

Novel toxicities have emerged with the implementation of CAR T cell therapies. These include expected on-target, off-tumor toxicities from depletion of healthy cells that share the targeted antigen. For instance, anti-CD19-targeted CAR T cells cause aplasia of normal CD19⁺ B cells, which is therefore a pharmacodynamic marker of CAR T persistence.⁹⁰ CAR T cells also cause previously unanticipated toxicities, including CRS, immune cell-associated neurotoxicity syndrome, and prolonged bone marrow aplasia. CRS is a potentially life-threatening immune activation syndrome occurring in the first days to 2 weeks after infusion associated with elevated serum

inflammatory markers and cytokines. CRS can cause pyrexia, hypoxia, and vasodilatory shock.⁹¹ It occurs in 60% to 90% of patients because of inflammatory cytokine (IL-6 and IL-1) production by myeloid cells in response to T cell cytokines and inflammatory tumor cell death.⁹²⁻⁹⁴ Risk factors for severe CRS include higher disease burden and high infused CAR T dose,⁹¹ with severity graded per consensus guidelines from the American Society of Transplantation and Cell Therapy.⁹⁵ The mainstay of management is the IL-6 receptor inhibitor tocilizumab, which was empirically effective in initial registration trials, although steroids, IL-1 antagonists, and other agents are also used.^{91,92,96,97} Retrospective studies have identified cardiovascular toxicities in patients treated with CAR T cells (Table 1). The chief cardiotoxicity is heart failure, although arrhythmias are also observed, with the main risk factor being grade 2 or higher CRS.⁹⁸ Other risk factors include delayed tocilizumab administration and elevated baseline creatinine.^{98,99} Left ventricular ejection fraction recovery is seen in most patients with the resolution of CRS, although increased cardiovascular mortality is observed.⁹⁸⁻¹⁰¹ Like CRS, immune cell-associated neurotoxicity syndrome was not originally predicted as a toxicity of CAR T therapy, but it occurs in up to 40% to 50% of patients and can have serious sequelae.⁹¹ The pathophysiology of immune cell-associated neurotoxicity syndrome is less well understood but involves endothelial damage and blood-brain barrier permeabilization due to IL-1 and tumor necrosis factor- α but not IL-6 production.⁹²⁻⁹⁴ Immune cell-associated neurotoxicity syndrome management includes steroids and supportive care, although the efficacy of steroids has not been well established. Tocilizumab is not effective.^{91,92,96} Finally, prolonged bone marrow suppression is emerging as a frequent complication of CAR T cell therapy even when the targeted antigen is not expressed on bone marrow stem cells and is seen in up to 40% of patients. The etiology remains unclear; however, risk factors include degree and duration of CRS.¹⁰²⁻¹⁰⁵

UNKNOWN OR POORLY CHARACTERIZED ANTIGENS.

Arguably the cellular immunotherapy modality in widest routine clinical use is allogeneic hematopoietic stem cell transplantation (alloHCT), also known as bone marrow transplantation, for the treatment of hematopoietic cancers. Minor histocompatibility mismatches between the donor and host hematopoietic systems, among other potential immunogenic antigens, leads to a graft-vs-leukemia effect, whereby donor lymphocytes attack and eliminate the tumor-bearing host hematopoietic system.¹⁰⁶⁻¹⁰⁸ The

corollary of this desired graft-vs-leukemia effect is graft-vs-host disease; maintaining graft-vs-leukemia effects without exacerbating graft-vs-host disease is the key challenge of alloHCT.^{107,108} Sources for alloHCT donor grafts include bone marrow, peripheral blood (following administration of recombinant granulocyte colony-stimulating factor to induce CD34⁺ hematopoietic stem cell egress into peripheral blood), or umbilical cord blood, and donor source can affect the incidence of graft-vs-host disease.^{109,110} The etiology of cardiovascular complications after alloHCT is multifactorial given intensive pre-transplantation chemotherapy and/or radiation that predisposes to cardiac dysfunction, with anthracycline dose representing a strong predictor.^{111,112} Nonetheless, graft-vs-host disease also predisposes cardiovascular complications, which include pericardial effusions or tamponade,¹¹³ arrhythmias,^{114,115} and other morbidities related to long-term cardiac risk factor modification, suggesting immune-mediated mechanisms as well. The increasing use of haploidentical (half-matched) alloHCT grafts has also led to the development of CRS-like reactions, with significant impacts on mortality.¹¹⁶ Further cardiac complications of alloHCT are more fully discussed in other recent publications.^{117,118}

(RE)ACTIVATING EXISTING ANTITUMOR IMMUNITY

In contrast to tumor target-specific strategies to generate a de novo antitumor immune response, many cancer immunotherapies aim to reverse general pre-existing immune suppression (Central Illustration). The most well-established examples of this approach are the ICIs, mAbs directed against PD-1, PD-L1, and CTLA-4, as well as other emerging targets. These antibodies block immune inhibitory signals from cancer, stromal, and immune-suppressive myeloid elements within the tumor microenvironment to T cells, thus reactivating dysfunctional anti-tumor T cell responses. Since their first approvals in melanoma, ICIs have garnered numerous additional indications in a variety of tumor types, including lung cancer, genitourinary cancers, gastrointestinal malignancies, Hodgkin lymphoma, and all solid tumors with DNA mismatch repair deficiency or high tumor mutational burden regardless of tissue origin.^{119,120} As PD-1/PD-L1 and CTLA-4 checkpoint inhibitors, their mechanism of action, and toxicities have been covered in depth in prior publications,¹²¹ we focus on novel immune checkpoints and combination therapies as well as noncheckpoint strategies to activate existing anti-tumor immune responses.

NOVEL ICIs AND COMBINATIONS. Although the PD-1 and CTLA-4 pathways are the most well studied, they are not the only immune-suppressive checkpoints exerted on T cells within the tumor microenvironment. Both PD-1 and CTLA-4 are early T cell activation markers that are up-regulated as a counter-regulatory mechanism against overactivation. Their prolonged expression during long-term T cell stimulation is eventually associated with an “exhausted” and dysfunctional T cell state.¹²² LAG-3 is a similar T cell inhibitory receptor, and high LAG-3 expression is associated with T cell exhaustion.¹²³ Recently, in a phase 3 first-line study for unresectable or metastatic melanoma, treatment with the LAG-3 inhibitor relatlimab in combination with nivolumab, a PD-1 inhibitor, significantly improved progression-free survival compared with PD-1 inhibition alone (10.1 months vs 4.6 months)¹²⁴ and has now received FDA approval for this indication. A key advantage of this combination over the previously approved combination of PD-1/CTLA-4 inhibitors is the lower rate of grade 3 or 4 immune-related toxicities, which were >50% with combined PD-1/CTLA-4 blockade and 18.9% for PD-1/LAG-3 inhibition.¹²⁴⁻¹²⁶ Myocarditis was uncommon, occurring in 1.7% of patients with PD-1/LAG-3 inhibition, consistent with other ICI studies. Numerous other products, both mono- and bispecific, targeting LAG-3 or other noncanonical immune checkpoints are in preclinical and early-phase development.¹²³ The durability of responses with these novel agents compared with existing ICIs, which can allow treatment discontinuation in some patients,^{127,128} remains unclear.

ICIs have also been combined with other drug classes, which has steadily expanded the population of patients with cancer eligible for these therapies. The most illustrative example is NSCLC, for which the PD-1 inhibitor pembrolizumab has single-agent FDA approval for PD-L1-positive patients.¹²⁹ Subsequently, even patients with negative PD-L1 staining became eligible for pembrolizumab plus chemotherapy, which showed a 12-month overall survival benefit compared with chemotherapy alone (69.2% vs 49.4%).¹³⁰ ICI and chemotherapy combinations have also been approved in gastroesophageal and biliary tract cancers.^{131,132} ICIs have also been found to synergize with angiogenesis inhibitors, and these combinations are approved in several solid tumors, including hepatocellular carcinoma,¹³³ endometrial cancer,¹³⁴ and renal cell carcinoma.¹³⁵⁻¹³⁷ The exact mechanism for this synergy is unclear, particularly as many antiangiogenic agents are multikinase inhibitors that affect several different pathways.^{138,139} Last, newly engineered versions of existing ICIs may

improve responses in presently immune-refractory tumor types, as was shown recently in a phase 1 trial of botensilimab, an Fc-engineered CTLA-4 inhibitor, which when combined with a PD-1 inhibitor achieved a 24% response rate (73% disease control) with high durability in patients with heavily pretreated microsatellite-stable colorectal cancer who are resistant to traditional ICIs.¹⁴⁰

Cardiovascular toxicities are rare with ICI monotherapy, with myocarditis occurring in <1% of patients, though more commonly with dual ICI treatment (**Table 1**). These rates may be underestimated, as increased recognition has led to increases in reporting over time.¹⁴¹⁻¹⁴³ Despite its rarity, this is a feared complication that carries a high mortality rate.^{141,144} Additionally, emerging evidence shows that ICI combination with anti-VEGF or multikinase targeted agents is associated with major adverse cardiovascular events.^{141,145} As these agents alone lead to a known increase in cardiovascular risk factors such as hypertension, thromboembolism, and cardiomyopathy,¹⁴⁶⁻¹⁴⁸ the contribution of immune toxicity to major adverse cardiovascular event risk remains unclear, particularly given that these are mostly nonmyocarditis events,¹⁴¹ although recent retrospective data have suggested that ICIs are also associated with major adverse cardiovascular events, particularly in predisposed patients.¹⁴⁹

CYTOKINES. Cytokines are intercellular communication molecules that are secreted by or act upon immune cells in an autocrine, paracrine, or endocrine manner. The relevance of cytokines for immunoncology pertains to their direct antitumor effect or their ability to stimulate effector immune cells (**Central Illustration**). Recombinant human IL-2 (aldesleukin) has an FDA label for the treatment of metastatic renal cell carcinoma and for metastatic melanoma despite the low fraction of responding patients (15% and 16% overall responders, respectively).^{150,151} As administration of high doses of IL-2 is an integral component of TIL therapy (as discussed earlier), the high incidence of severe toxicities due to capillary leak syndrome when administered at the requisite dose remains relevant (**Table 1**).^{33,152} More recent developments in synthetic biology have resulted in synthetic cytokines with favorable toxicity profiles that can be administered as single agents and are currently in clinical trials.¹⁵³ Synthetic cytokines can also be used in cell therapies by creating immune cells that secrete proinflammatory cytokines,¹⁵⁴ synthetic cytokines that act as partial agonists to expand T cells without inducing terminal differentiation,¹⁵⁵ or entirely synthetic cytokine and cytokine receptor

TABLE 3 Selected Cytokines Investigated in Immunotherapy Clinical Trials

Cytokine	Desired Immunologic Activity	Ref. #
IL-2	Stimulation of CD4 and CD8 T cell proliferation and survival	33,152,153
IL-7	Stimulation of CD4 and CD8 T cell proliferation and survival	185-187
IL-10	Increase expansion and cytotoxicity of tumor-infiltrating CD8 T cells	188
IL-12	NK, NK/T, and CD8 T cell proliferation, T cell differentiation to produce IFN γ , enhancement of dendritic cell antigen presentation	189
IL-15	Stimulation of NK and CD8 T cell proliferation	157,190
IL-18	Augmentation of antibody-dependent cellular cytotoxicity in NK cells	191
IL-21	CD8 T cell proliferation, survival, and resistance to regulatory T cell suppression	192,193

IL = interleukin; NK = natural killer.

circuits, such as the IL-2/IL-2 receptor beta chain system, to selectively expand T cells.¹⁵⁶ By engineering synthetic cytokine receptors to be expressed in adoptively transferred CAR T cells that respond only to engineered cytokines, it is possible to achieve a high level of biological specificity, avoiding both on- and off-target effects of the native cytokine on normal immune cells. Although most efforts to stimulate antitumor immunity with cytokines have focused on native or engineered versions of IL-2, other cytokines with immunologic activity are also under investigation in clinical trials, including IL-7, IL-12, IL-15, and others (Table 3). Although cardiotoxicities associated with systemic IL-2 are significant (Table 1), early-phase trials of these other cytokines have shown lower severity of cardiovascular side effects, for instance, less severe hypotension and capillary leak syndrome in phase 1 trial of systemic IL-15.¹⁵⁷

TARGETING IMMUNOSUPPRESSIVE CELLS. The tumor microenvironment poses a key hurdle against antitumor immunity. The tumor microenvironment consists of hematopoietic and stromal cells that are recruited by tumor cells and provide negative regulatory signals to T cells to prevent effective antitumor immunity.¹⁵⁸ As myeloid cells, either in the form of tumor-associated macrophages or myeloid-derived suppressor cells, form a significant fraction of these immune-suppressive bystander cells, depletion of myeloid cells or blockade of their recruitment is an emerging objective in cancer immunotherapy (Central Illustration).¹⁵⁹ Some of the activity of traditional chemotherapeutic agents has been attributed to their ability to deplete immune-suppressive myeloid cells.^{62,160,161} More targeted strategies have included the use of pexidartinib, a small-molecule inhibitor of the macrophage colony-stimulating factor receptor, which showed a response rate of 39% compared with 0% in a

placebo-controlled trial and received FDA approval for the treatment of tenosynovial giant cell tumors.¹⁶² Tenosynovial giant cell tumors overexpress macrophage colony-stimulating factor, leading to increased recruitment of myeloid-derived suppressor cells into the tumor microenvironment. This pathway is blocked by pexidartinib, the only systemic therapy to show robust activity in tenosynovial giant cell tumors.¹⁶³ Other strategies to block immune-suppressive myeloid cells include targeting other chemokine and receptor pathways involved in myeloid recruitment, such as the C-C chemokine receptor 2 and 5 pathways,¹⁶⁴ or use of a CD33 \times CD3 BiTE to redirect T cells for myeloid depletion that has early-phase activity in solid tumors.¹⁶⁵ Use of CAR T cells to deplete components of the immune-suppressive microenvironment rather than tumor cells themselves has also been proposed and tested in preclinical models,¹⁶⁶ although this therapeutic strategy has not yet reached clinical phase development. In addition to myeloid depletion, reactivating myeloid cell intrinsic antitumor immune functions through inhibition of the “don’t eat me” signal CD47¹⁶⁷ or through stimulatory engagement with CD40 agonists^{168,169} is also under investigation. Pexidartinib is associated with hypertension (Table 1); cardiovascular toxicities of other myeloid depletion strategies remain unknown.

CONCLUSIONS

Although predicted as a potential antitumor treatment modality more than 50 years ago,¹⁷⁰ clinically tractable cancer immunotherapies have become a reality only in the past decade. This achievement is based on many decades of research advances elucidating basic mechanisms of both immunity and tumorigenesis. With the identification of new targets and novel combinations, use of immunotherapy in

oncology is likely to grow in the coming decades because of its distinct advantages over more traditional anticancer drugs in terms of both depth and durability of potential responses. Rapid expansion of the therapeutic arsenal has presented challenges to clinicians, regulators, and the health care system, as mechanisms of action, response patterns, toxicities, and costs of these therapies diverge significantly from prior generations of cancer treatments. However, a few hallmarks of immune-based therapies have emerged from their widespread clinical adoption, including marked and durable survival benefits despite a low initial responding fraction of patients^{171,172} as well as loss of immunotherapy sensitivity in some treated patients.¹²¹ These response patterns highlight future directions for research and development, which must continue to

address issues of immune priming as well as both primary and secondary immune resistance^{121,173} to achieve durable cancer control and cures.

FUNDING SUPPORT AND AUTHOR DISCLOSURES

This work was supported by the Emerson Collective and a Conquer Cancer Foundation Young Investigator Award. Dr Gill is named as an inventor on multiple CAR T-related patents. Dr Welty has reported that he has no relationships relevant to the contents of this paper to disclose.

ADDRESS FOR CORRESPONDENCE: Dr Saar I. Gill, Smilow Center for Translational Research, Room 8-101, 3400 Civic Center Boulevard, Philadelphia, Pennsylvania 19104, USA. E-mail: saar.gill@penntmedicine.upenn.edu.

REFERENCES

- Hanahan D. Hallmarks of cancer: new dimensions. *Cancer Discov*. 2022;12:31-46.
- O'Donnell JS, Teng MWL, Smyth MJ. Cancer immunoeediting and resistance to T cell-based immunotherapy. *Nat Rev Clin Oncol*. 2019;16:151-167.
- Schreiber RD, Old LJ, Smyth MJ. Cancer immunoeediting: integrating immunity's roles in cancer suppression and promotion. *Science*. 2011;331:1565-1570.
- Negrini S, Gorgoulis VG, Halazonetis TD. Genomic instability—an evolving hallmark of cancer. *Nat Rev Mol Cell Biol*. 2010;11:220-228.
- McGranahan N, Swanton C. Neoantigen quality, not quantity. *Sci Transl Med*. 2019;11:eaax7918.
- Snyder A, Makarov V, Merghoub T, et al. Genetic basis for clinical response to CTLA-4 blockade in melanoma. *N Engl J Med*. 2014;371:2189-2199.
- Lee CH, Yelensky R, Jooss K, Chan TA. Update on tumor neoantigens and their utility: why it is good to be different. *Trends Immunol*. 2018;39:536-548.
- Van Allen EM, Miao D, Schilling B, et al. Genomic correlates of response to CTLA-4 blockade in metastatic melanoma. *Science*. 2015;350:207-211.
- Rizvi NA, Hellmann MD, Snyder A, et al. Cancer immunology. Mutational landscape determines sensitivity to PD-1 blockade in non-small cell lung cancer. *Science*. 2015;348:124-128.
- Wells DK, van Buuren MM, Dang KK, et al. Key parameters of tumor epitope immunogenicity revealed through a consortium approach improve neoantigen prediction. *Cell*. 2020;183:818-834.e13.
- Blass E, Ott PA. Advances in the development of personalized neoantigen-based therapeutic cancer vaccines. *Nat Rev Clin Oncol*. 2021;18:215-229.
- Trowsdale J, Knight JC. Major histocompatibility complex genomics and human disease. *Annu Rev Genomics Hum Genet*. 2013;14:301-323.
- Maiers M, Gragert L, Klitz W. High-resolution HLA alleles and haplotypes in the United States population. *Hum Immunol*. 2007;68:779-788.
- Carreno BM, Magrini V, Becker-Hapak M, et al. Cancer immunotherapy. A dendritic cell vaccine increases the breadth and diversity of melanoma neoantigen-specific T cells. *Science*. 2015;348:803-808.
- Ott PA, Hu Z, Keskin DB, et al. An immunogenic personal neoantigen vaccine for patients with melanoma. *Nature*. 2017;547:217-221.
- Sahin U, Derhovanessian E, Miller M, et al. Personalized RNA mutanome vaccines mobilize poly-specific therapeutic immunity against cancer. *Nature*. 2017;547:222-226.
- Saxena M, van der Burg SH, Melief CJM, Bhardwaj N. Therapeutic cancer vaccines. *Nat Rev Cancer*. 2021;21:360-378.
- Bear AS, Blanchard T, Cesare J, et al. Biochemical and functional characterization of mutant KRAS epitopes validates this oncoprotein for immunological targeting. *Nat Commun*. 2021;12:4365.
- Chaft JE, Litvak A, Arcila ME, et al. Phase II study of the GI-4000 KRAS vaccine after curative therapy in patients with stage I-III lung adenocarcinoma harboring a KRAS G12C, G12D, or G12V mutation. *Clin Lung Cancer*. 2014;15:405-410.
- Leko V, Rosenberg SA. Identifying and targeting human tumor antigens for t cell-based immunotherapy of solid tumors. *Cancer Cell*. 2020;38:454-472.
- Tran E, Robbins PF, Lu YC, et al. T-cell transfer therapy targeting mutant KRAS in cancer. *N Engl J Med*. 2016;375:2255-2262.
- McGranahan N, Rosenthal R, Hiley CT, et al. Allele-specific HLA loss and immune escape in lung cancer evolution. *Cell*. 2017;171:1259-1271.e11.
- Leoni G, D'Alise AM, Cotugno G, et al. A genetic vaccine encoding shared cancer neoantigens to treat tumors with microsatellite instability. *Cancer Res*. 2020;80:3972-3982.
- Kantoff PW, Higano CS, Shore ND, et al. Sipuleucel-T immunotherapy for castration-resistant prostate cancer. *N Engl J Med*. 2010;363:411-422.
- Schellhammer PF, Chodak G, Whitmore JB, Sims R, Frohlich MW, Kantoff PW. Lower baseline prostate-specific antigen is associated with a greater overall survival benefit from sipuleucel-T in the Immunotherapy for Prostate Adenocarcinoma Treatment (IMPACT) trial. *Urology*. 2013;81:1297-1302.
- Vansteenkiste JF, Cho BC, Vanakesa T, et al. Efficacy of the MAGE-A3 cancer immunotherapeutic as adjuvant therapy in patients with resected MAGE-A3-positive non-small-cell lung cancer (MAGRIT): a randomised, double-blind, placebo-controlled, phase 3 trial. *Lancet Oncol*. 2016;17:822-835.
- Butts C, Socinski MA, Mitchell PL, et al. Tecemotide (L-BLP25) versus placebo after chemoradiotherapy for stage III non-small-cell lung cancer (START): a randomised, double-blind, phase 3 trial. *Lancet Oncol*. 2014;15:59-68.
- Middleton G, Silcocks P, Cox T, et al. Gemcitabine and capecitabine with or without telomerase peptide vaccine GV1001 in patients with locally advanced or metastatic pancreatic cancer (TeloVac): an open-label, randomised, phase 3 trial. *Lancet Oncol*. 2014;15:829-840.
- Rosenberg SA, Packard BS, Aebersold PM, et al. Use of tumor-infiltrating lymphocytes and interleukin-2 in the immunotherapy of patients with metastatic melanoma. A preliminary report. *N Engl J Med*. 1988;319:1676-1680.

30. Robbins PF, Lu YC, El-Gamil M, et al. Mining exomic sequencing data to identify mutated antigens recognized by adoptively transferred tumor-reactive T cells. *Nat Med*. 2013;19:747-752.
31. Dafni U, Michielin O, Lluemas SM, et al. Efficacy of adoptive therapy with tumor-infiltrating lymphocytes and recombinant interleukin-2 in advanced cutaneous melanoma: a systematic review and meta-analysis. *Ann Oncol*. 2019;30:1902-1913.
32. Forget MA, Haymaker C, Hess KR, et al. Prospective analysis of adoptive TIL therapy in patients with metastatic melanoma: response, impact of anti-CTLA4, and biomarkers to predict clinical outcome. *Clin Cancer Res*. 2018;24:4416-4428.
33. Sarnaik AA, Hamid O, Khushalani NI, et al. Lifileucel, a tumor-infiltrating lymphocyte therapy, in metastatic melanoma. *J Clin Oncol*. 2021;39:2656-2666.
34. Creelan BC, Wang C, Teer JK, et al. Tumor-infiltrating lymphocyte treatment for anti-PD-1 resistant metastatic lung cancer: a phase 1 trial. *Nat Med*. 2021;27:1410-1418.
35. Jiang SS, Tang Y, Zhang YJ, et al. A phase I clinical trial utilizing autologous tumor-infiltrating lymphocytes in patients with primary hepatocellular carcinoma. *Oncotarget*. 2015;6:41339-41349.
36. Stevanovic S, Helman SR, Wunderlich JR, et al. A phase II study of tumor-infiltrating lymphocyte therapy for human papillomavirus-associated epithelial cancers. *Clin Cancer Res*. 2019;25:1486-1493.
37. Chandran SS, Somerville RPT, Yang JC, et al. Treatment of metastatic uveal melanoma with adoptive transfer of tumour-infiltrating lymphocytes: a single-centre, two-stage, single-arm, phase 2 study. *Lancet Oncol*. 2017;18:792-802.
38. Fradley MG, Damrongwatanasuk R, Chandrasekhar S, Alomar M, Kip KE, Sarnaik AA. Cardiovascular toxicity and mortality associated with adoptive cell therapy and tumor-infiltrating lymphocytes for advanced stage melanoma. *J Immunother*. 2021;44:86-89.
39. Kitada T, DiAndreth B, Teague B, Weiss R. Programming gene and engineered-cell therapies with synthetic biology. *Science*. 2018;359:eaad1067.
40. Stadtmauer EA, Fraietta JA, Davis MM, et al. CRISPR-engineered T cells in patients with refractory cancer. *Science*. 2020;367:eaba7365.
41. Finck AV, Blanchard T, Roselle CP, Golinelli G, June CH. Engineered cellular immunotherapies in cancer and beyond. *Nat Med*. 2022;28:678-689.
42. Weber EW, Maus MV, Mackall CL. The emerging landscape of immune cell therapies. *Cell*. 2020;181:46-62.
43. Wang QJ, Yu Z, Griffith K, Hanada K, Restifo NP, Yang JC. Identification of T-cell receptors targeting KRAS-mutated human tumors. *Cancer Immunol Res*. 2016;4:204-214.
44. Leidner R, Sanjuan Silva N, Huang H, et al. Neoantigen T-cell receptor gene therapy in pancreatic cancer. *N Engl J Med*. 2022;386:2112-2119.
45. Miller MS, Douglass J, Hwang MS, et al. An engineered antibody fragment targeting mutant beta-catenin via major histocompatibility complex I neoantigen presentation. *J Biol Chem*. 2019;294:19322-19334.
46. Skora AD, Douglass J, Hwang MS, et al. Generation of MANAbodies specific to HLA-restricted epitopes encoded by somatically mutated genes. *Proc Natl Acad Sci U S A*. 2015;112:9967-9972.
47. Douglass J, Hsueh EH, Mog BJ, et al. Bispecific antibodies targeting mutant RAS neoantigens. *Sci Immunol*. 2021;6:eabd5515.
48. Linette GP, Stadtmauer EA, Maus MV, et al. Cardiovascular toxicity and titin cross-reactivity of affinity-enhanced T cells in myeloma and melanoma. *Blood*. 2013;122:863-871.
49. Cameron BJ, Gerry AB, Dukes J, et al. Identification of a Titin-derived HLA-A1-presented peptide as a cross-reactive target for engineered MAGE A3-directed T cells. *Sci Transl Med*. 2013;5:197ra103.
50. Donnadieu E, Luu M, Alb M, et al. Time to evolve: predicting engineered T cell-associated toxicity with next-generation models. *J Immunother Cancer*. 2022;10:e003486.
51. Clynes RA, Towers TL, Presta LG, Ravetch JV. Inhibitory Fc receptors modulate in vivo cytotoxicity against tumor targets. *Nat Med*. 2000;6:443-446.
52. Tsoo LC, Force J, Hartman ZC. Mechanisms of therapeutic antitumor monoclonal antibodies. *Cancer Res*. 2021;81:4641-4651.
53. Rugo HS, Im SA, Cardoso F, et al. Efficacy of margetuximab vs trastuzumab in patients with pretreated ERBB2-positive advanced breast cancer: a phase 3 randomized clinical trial. *JAMA Oncol*. 2021;7:573-584.
54. Vitolo U, Trnny M, Belada D, et al. Obinutuzumab or rituximab plus cyclophosphamide, doxorubicin, vincristine, and prednisone in previously untreated diffuse large B-Cell lymphoma. *J Clin Oncol*. 2017;35:3529-3537.
55. Maloney DG, Ogura M, Fukuhara N, et al. A phase 3 randomized study (HOMER) of ofatumumab vs rituximab in iNHL relapsed after rituximab-containing therapy. *Blood Adv*. 2020;4:3886-3893.
56. Coiffier B, Haioun C, Ketterer N, et al. Rituximab (anti-CD20 monoclonal antibody) for the treatment of patients with relapsing or refractory aggressive lymphoma: a multicenter phase II study. *Blood*. 1998;92:1927-1932.
57. Coiffier B, Lepage E, Briere J, et al. CHOP chemotherapy plus rituximab compared with CHOP alone in elderly patients with diffuse large-B-cell lymphoma. *N Engl J Med*. 2002;346:235-242.
58. Baselga J, Tripathy D, Mendelsohn J, et al. Phase II study of weekly intravenous recombinant humanized anti-p185HER2 monoclonal antibody in patients with HER2/neu-overexpressing metastatic breast cancer. *J Clin Oncol*. 1996;14:737-744.
59. Pegram MD, Lipton A, Hayes DF, et al. Phase II study of receptor-enhanced chemosensitivity using recombinant humanized anti-p185HER2/neu monoclonal antibody plus cisplatin in patients with HER2/neu-overexpressing metastatic breast cancer refractory to chemotherapy treatment. *J Clin Oncol*. 1998;16:2659-2671.
60. Galluzzi L, Buque A, Kepp O, Zitvogel L, Kroemer G. Immunogenic cell death in cancer and infectious disease. *Nat Rev Immunol*. 2017;17:97-111.
61. Ghiringhelli F, Menard C, Puig PE, et al. Metronomic cyclophosphamide regimen selectively depletes CD4⁺CD25⁺ regulatory T cells and restores T and NK effector functions in end stage cancer patients. *Cancer Immunol Immunother*. 2007;56:641-648.
62. Alizadeh D, Trad M, Hanke NT, et al. Doxorubicin eliminates myeloid-derived suppressor cells and enhances the efficacy of adoptive T-cell transfer in breast cancer. *Cancer Res*. 2014;74:104-118.
63. Srivastava S, Furlan SN, Jaeger-Ruckstuhl CA, et al. Immunogenic chemotherapy enhances recruitment of CAR-T cells to lung tumors and improves antitumor efficacy when combined with checkpoint blockade. *Cancer Cell*. 2021;39:193-208.e10.
64. Chau CH, Steeg PS, Figg WD. Antibody-drug conjugates for cancer. *Lancet*. 2019;394:793-804.
65. Drago JZ, Modi S, Chandralapaty S. Unlocking the potential of antibody-drug conjugates for cancer therapy. *Nat Rev Clin Oncol*. 2021;18:327-344.
66. Park K, Haura EB, Leigh NB, et al. Amivantamab in EGFR exon 20 insertion-mutated non-small-cell lung cancer progressing on platinum chemotherapy: initial results from the CHRYSALIS phase I study. *J Clin Oncol*. 2021;39:3391-3402.
67. Duell J, Lammers PE, Djuretic I, et al. Bispecific antibodies in the treatment of hematologic malignancies. *Clin Pharmacol Ther*. 2019;106:781-791.
68. Nathan P, Hassel JC, Rutkowski P, et al. Overall survival benefit with tebentafusp in metastatic uveal melanoma. *N Engl J Med*. 2021;385:1196-1206.
69. Pham JP, Star P, Ardolino L, Smith A, Joshua AM. A review of the cutaneous toxicities of tebentafusp-Featuring two cases involving superficial bullous reactions. *Australas J Dermatol*. 2022;63:e279-e282.
70. Gokbuget N, Dombret H, Bonifacio M, et al. Blinatumomab for minimal residual disease in adults with B-cell precursor acute lymphoblastic leukemia. *Blood*. 2018;131:1522-1531.
71. Topp MS, Gokbuget N, Zugmaier G, et al. Phase II trial of the anti-CD19 bispecific T cell-engager blinatumomab shows hematologic and molecular remissions in patients with relapsed or refractory B-precursor acute lymphoblastic leukemia. *J Clin Oncol*. 2014;32:4134-4140.
72. Zugmaier G, Gokbuget N, Klinger M, et al. Long-term survival and T-cell kinetics in relapsed/refractory ALL patients who achieved MRD response after blinatumomab treatment. *Blood*. 2015;126:2578-2584.
73. Hainsworth JD, Litchy S, Barton JH, et al. Single-agent rituximab as first-line and maintenance treatment for patients with chronic

- lymphocytic leukemia or small lymphocytic lymphoma: a phase II trial of the Minnie Pearl Cancer Research Network. *J Clin Oncol*. 2003;21:1746-1751.
74. Schuster SJ, Bartlett NL, Assouline S, et al. Mosunetuzumab induces complete remissions in poor prognosis non-hodgkin lymphoma patients, including those who are resistant to or relapsing after chimeric antigen receptor T-cell (CAR-T) therapies, and is active in treatment through multiple lines. *Blood*. 2019;134:6.
75. Topp MS, Gokbuget N, Stein AS, et al. Safety and activity of blinatumomab for adult patients with relapsed or refractory B-precursor acute lymphoblastic leukaemia: a multicentre, single-arm, phase 2 study. *Lancet Oncol*. 2015;16:57-66.
76. Jabbour E, Dull J, Yilmaz M, et al. Outcome of patients with relapsed/refractory acute lymphoblastic leukemia after blinatumomab failure: no change in the level of CD19 expression. *Am J Hematol*. 2018;93:371-374.
77. Lulla PD, Tzannou I, Vasileiou S, et al. The safety and clinical effects of administering a multiantigen-targeted T cell therapy to patients with multiple myeloma. *Sci Transl Med*. 2020;12:eaz3339.
78. Vasileiou S, Lulla PD, Tzannou I, et al. T-cell therapy for lymphoma using nonengineered multiantigen-targeted T cells is safe and produces durable clinical effects. *J Clin Oncol*. 2021;39:1415-1425.
79. Pan J, Tan Y, Wang G, et al. Donor-derived CD7 chimeric antigen receptor T cells for T-cell acute lymphoblastic leukemia: first-in-human, phase I trial. *J Clin Oncol*. 2021;39:3340-3351.
80. Ramos CA, Grover NS, Beaven AW, et al. Anti-CD30 CAR-T cell therapy in relapsed and refractory Hodgkin lymphoma. *J Clin Oncol*. 2020;38:3794-3804.
81. Zhang H, Gan WT, Hao WG, Wang PF, Li ZY, Chang LJ. Successful anti-CLL1 CAR T-cell therapy in secondary acute myeloid leukemia. *Front Oncol*. 2020;10:685.
82. Qi C, Gong J, Li J, et al. Claudin18.2-specific CAR T cells in gastrointestinal cancers: phase 1 trial interim results. *Nat Med*. 2022;28:1189-1198.
83. Narayan V, Barber-Rotenberg JS, Jung IY, et al. PSMA-targeting TGFbeta-insensitive armored CAR T cells in metastatic castration-resistant prostate cancer: a phase 1 trial. *Nat Med*. 2022;28:724-734.
84. Majzner RG, Ramakrishna S, Yeom KW, et al. GD2-CAR T cell therapy for H3K27M-mutated diffuse midline gliomas. *Nature*. 2022;603:934-941.
85. Liu E, Marin D, Banerjee P, et al. Use of CAR-transduced natural killer cells in CD19-positive lymphoid tumors. *N Engl J Med*. 2020;382:545-553.
86. Heczey A, Courtney AN, Montalbano A, et al. Anti-GD2 CAR-NKT cells in patients with relapsed or refractory neuroblastoma: an interim analysis. *Nat Med*. 2020;26:1686-1690.
87. MacKay M, Afshinnekoo E, Rub J, et al. The therapeutic landscape for cells engineered with chimeric antigen receptors. *Nat Biotechnol*. 2020;38:233-244.
88. Melenhorst JJ, Chen GM, Wang M, et al. Decade-long leukaemia remissions with persistence of CD4⁺ CAR T cells. *Nature*. 2022;602:503-509.
89. Hay AE, Cheung MC. CAR T-cells: costs, comparisons, and commentary. *J Med Econ*. 2019;22:613-615.
90. Maude SL, Frey N, Shaw PA, et al. Chimeric antigen receptor T cells for sustained remissions in leukemia. *N Engl J Med*. 2014;371:1507-1517.
91. Frey N, Porter D. Cytokine release syndrome with chimeric antigen receptor T cell therapy. *Biol Blood Marrow Transplant*. 2019;25:e123-e127.
92. Morris EC, Neelapu SS, Giavridis T, Sadelain M. Cytokine release syndrome and associated neurotoxicity in cancer immunotherapy. *Nat Rev Immunol*. 2022;22:85-96.
93. Norelli M, Camisa B, Barbiera G, et al. Monocyte-derived IL-1 and IL-6 are differentially required for cytokine-release syndrome and neurotoxicity due to CAR T cells. *Nat Med*. 2018;24:739-748.
94. Giavridis T, van der Stegen SJC, Eyquem J, Hamieh M, Piersigilli A, Sadelain M. CAR T cell-induced cytokine release syndrome is mediated by macrophages and abated by IL-1 blockade. *Nat Med*. 2018;24:731-738.
95. Lee DW, Santomasso BD, Locke FL, et al. ASTCT consensus grading for cytokine release syndrome and neurologic toxicity associated with immune effector cells. *Biol Blood Marrow Transplant*. 2019;25:625-638.
96. Neelapu SS, Tummala S, Kebriaei P, et al. Chimeric antigen receptor T-cell therapy—assessment and management of toxicities. *Nat Rev Clin Oncol*. 2018;15:47-62.
97. Teachey DT, Bishop MR, Maloney DG, Grupp SA. Toxicity management after chimeric antigen receptor T cell therapy: one size does not fit "ALL". *Nat Rev Clin Oncol*. 2018;15:218.
98. Alvi RM, Frigault MJ, Fradley MG, et al. Cardiovascular events among adults treated with chimeric antigen receptor T-cells (CAR-T). *J Am Coll Cardiol*. 2019;74:3099-3108.
99. Lefebvre B, Kang Y, Smith AM, Frey NV, Carver JR, Scherrer-Crosbie M. Cardiovascular effects of CAR T cell therapy: a retrospective study. *J Am Coll Cardiol CardioOnc*. 2020;2:193-203.
100. Shalabi H, Sachdev V, Kulshreshtha A, et al. Impact of cytokine release syndrome on cardiac function following CD19 CAR-T cell therapy in children and young adults with hematological malignancies. *J Immunother Cancer*. 2020;8:e001159.
101. Ganatra S, Redd R, Hayek SS, et al. Chimeric antigen receptor T-cell therapy-associated cardiomyopathy in patients with refractory or relapsed non-Hodgkin lymphoma. *Circulation*. 2020;142:1687-1690.
102. Wudhikarn K, Palomba ML, Pennisi M, et al. Infection during the first year in patients treated with CD19 CAR T cells for diffuse large B cell lymphoma. *Blood Cancer J*. 2020;10:79.
103. Nahas GR, Komanduri KV, Pereira D, et al. Incidence and risk factors associated with a syndrome of persistent cytopenias after CAR-T cell therapy (PCTT). *Leuk Lymphoma*. 2020;61:940-943.
104. Fried S, Avigdor A, Biorlari B, et al. Early and late hematologic toxicity following CD19 CAR-T cells. *Bone Marrow Transplant*. 2019;54:1643-1650.
105. Wang L, Hong R, Zhou L, et al. New-onset severe cytopenia after CAR-T cell therapy: analysis of 76 patients with relapsed or refractory acute lymphoblastic leukemia. *Front Oncol*. 2021;11:702644.
106. Warren EH, Fujii N, Akatsuka Y, et al. Therapy of relapsed leukemia after allogeneic hematopoietic cell transplantation with T cells specific for minor histocompatibility antigens. *Blood*. 2010;115:3869-3878.
107. Kolb HJ. Graft-versus-leukemia effects of transplantation and donor lymphocytes. *Blood*. 2008;112:4371-4383.
108. Negrin RS. Graft-versus-host disease versus graft-versus-leukemia. *Hematology Am Soc Hematol Educ Program*. 2015;2015:225-230.
109. Rocha V, Wagner JE Jr, Sobocinski KA, et al. Graft-versus-host disease in children who have received a cord-blood or bone marrow transplant from an HLA-identical sibling. Eurocord and International Bone Marrow Transplant Registry Working Committee on Alternative Donor and Stem Cell Sources. *N Engl J Med*. 2000;342:1846-1854.
110. Schrezenmeier H, Passweg JR, Marsh JC, et al. Worse outcome and more chronic GVHD with peripheral blood progenitor cells than bone marrow in HLA-matched sibling donor transplants for young patients with severe acquired aplastic anemia. *Blood*. 2007;110:1397-1400.
111. Sakata-Yanagimoto M, Kanda Y, Nakagawa M, et al. Predictors for severe cardiac complications after hematopoietic stem cell transplantation. *Bone Marrow Transplant*. 2004;33:1043-1047.
112. Armenian SH, Yang D, Teh JB, et al. Prediction of cardiovascular disease among hematopoietic cell transplantation survivors. *Blood Adv*. 2018;2:1756-1764.
113. Pfeiffer TM, Rotz SJ, Ryan TD, et al. Pericardial effusion requiring surgical intervention after stem cell transplantation: a case series. *Bone Marrow Transplant*. 2017;52:630-633.
114. Chow EJ, Mueller BA, Baker KS, et al. Cardiovascular hospitalizations and mortality among recipients of hematopoietic stem cell transplantation. *Ann Intern Med*. 2011;155:21-32.
115. Tonorez ES, Stillwell EE, Calloway JJ, et al. Arrhythmias in the setting of hematopoietic cell transplants. *Bone Marrow Transplant*. 2015;50:1212-1216.
116. Imus PH, Blackford AL, Bettinotti M, et al. Severe cytokine release syndrome after haplo-identical peripheral blood stem cell transplantation. *Biol Blood Marrow Transplant*. 2019;25:2431-2437.
117. Rotz SJ, Ryan TD, Hayek SS. Cardiovascular disease and its management in children and adults

undergoing hematopoietic stem cell transplantation. *J Thromb Thrombolysis*. 2021;51:854-869.

118. Tuzovic M, Mead M, Young PA, Schiller G, Yang EH. Cardiac complications in the adult bone marrow transplant patient. *Curr Oncol Rep*. 2019;21:28.

119. Vaddepally RK, Kharel P, Pandey R, Garje R, Chandra AB. Review of indications of FDA-approved immune checkpoint inhibitors per NCCN guidelines with the level of evidence. *Cancers (Basel)*. 2020;12:738.

120. Marabelle A, Fakhri M, Lopez J, et al. Association of tumour mutational burden with outcomes in patients with advanced solid tumours treated with pembrolizumab: prospective biomarker analysis of the multicohort, open-label, phase 2 KEYNOTE-158 study. *Lancet Oncol*. 2020;21:1353-1365.

121. Morad G, Helmkamp BA, Sharma P, Wargo JA. Hallmarks of response, resistance, and toxicity to immune checkpoint blockade. *Cell*. 2021;184:5309-5337.

122. McLane LM, Abdel-Hakeem MS, Wherry EJ. CD8 T cell exhaustion during chronic viral infection and cancer. *Annu Rev Immunol*. 2019;37:457-495.

123. Andrews LP, Yano H, Vignali DAA. Inhibitory receptors and ligands beyond PD-1, PD-L1 and CTLA-4: breakthroughs or backups. *Nat Immunol*. 2019;20:1425-1434.

124. Tawbi HA, Schadendorf D, Lipson EJ, et al. Relatlimab and nivolumab versus nivolumab in untreated advanced melanoma. *N Engl J Med*. 2022;386:24-34.

125. Larkin J, Chiarion-Sileni V, Gonzalez R, et al. Combined nivolumab and ipilimumab or monotherapy in untreated melanoma. *N Engl J Med*. 2015;373:23-34.

126. Postow MA, Chesney J, Pavlick AC, et al. Nivolumab and ipilimumab versus ipilimumab in untreated melanoma. *N Engl J Med*. 2015;372:2006-2017.

127. Hamid O, Robert C, Daud A, et al. Five-year survival outcomes for patients with advanced melanoma treated with pembrolizumab in KEYNOTE-001. *Ann Oncol*. 2019;30:582-588.

128. Betof Warner A, Palmer JS, Shoushtari AN, et al. Long-term outcomes and responses to retreatment in patients with melanoma treated with PD-1 blockade. *J Clin Oncol*. 2020;38:1655-1663.

129. Mok TSK, Wu YL, Kudaba I, et al. Pembrolizumab versus chemotherapy for previously untreated, PD-L1-expressing, locally advanced or metastatic non-small-cell lung cancer (KEYNOTE-042): a randomised, open-label, controlled, phase 3 trial. *Lancet*. 2019;393:1819-1830.

130. Gandhi L, Rodriguez-Abreu D, Gadgeel S, et al. Pembrolizumab plus chemotherapy in metastatic non-small-cell lung cancer. *N Engl J Med*. 2018;378:2078-2092.

131. Janjigian YY, Shitara K, Moehler M, et al. First-line nivolumab plus chemotherapy versus

chemotherapy alone for advanced gastric, gastro-oesophageal junction, and oesophageal adenocarcinoma (CheckMate 649): a randomised, open-label, phase 3 trial. *Lancet*. 2021;398:27-40.

132. Oh DY, He AR, Qin S, et al. A phase 3 randomised, double-blind, placebo-controlled study of durvalumab in combination with gemcitabine plus cisplatin (GemCis) in patients (pts) with advanced biliary tract cancer (BTC): TOPAZ-1. *J Clin Oncol*. 2022;40, 378-378.

133. Finn RS, Qin S, Ikeda M, et al. Atezolizumab plus bevacizumab in unresectable hepatocellular carcinoma. *N Engl J Med*. 2020;382:1894-1905.

134. Makker V, Colombo N, Casado Herraez A, et al. Lenvatinib plus pembrolizumab for advanced endometrial cancer. *N Engl J Med*. 2022;386:437-448.

135. Motzer R, Alekseev B, Rha SY, et al. Lenvatinib plus pembrolizumab or everolimus for advanced renal cell carcinoma. *N Engl J Med*. 2021;384:1289-1300.

136. Rini BI, Plimack ER, Stus V, et al. Pembrolizumab plus axitinib versus sunitinib for advanced renal-cell carcinoma. *N Engl J Med*. 2019;380:1116-1127.

137. Choueiri TK, Powles T, Buratto M, et al. Nivolumab plus cabozantinib versus sunitinib for advanced renal-cell carcinoma. *N Engl J Med*. 2021;384:829-841.

138. Hegde PS, Wallin JJ, Mancao C. Predictive markers of anti-VEGF and emerging role of angiogenesis inhibitors as immunotherapeutics. *Semin Cancer Biol*. 2018;52:117-124.

139. Song Y, Fu Y, Xie Q, Zhu B, Wang J, Zhang B. Anti-angiogenic agents in combination with immune checkpoint inhibitors: a promising strategy for cancer treatment. *Front Immunol*. 2020;11:1956.

140. Bullock A, Grossman J, Fakhri M, et al. Botensilimab, a novel innate/adaptive immune activator, plus balstilimab (anti-PD-1) for metastatic heavily pretreated microsatellite stable colorectal cancer. *Ann Oncol*. 2022;33:5376.

141. Naqash AR, Moey MYY, Cherie Tan XW, et al. Major adverse cardiac events with immune checkpoint inhibitors: a pooled analysis of trials sponsored by the National Cancer Institute-Cancer Therapy Evaluation Program. *J Clin Oncol*. 2022;40:3439-3452.

142. Johnson DB, Balko JM, Compton ML, et al. Fulminant myocarditis with combination immune checkpoint blockade. *N Engl J Med*. 2016;375:1749-1755.

143. Moslehi JJ, Salem JE, Sosman JA, Lebrun-Vignes B, Johnson DB. Increased reporting of fatal immune checkpoint inhibitor-associated myocarditis. *Lancet*. 2018;391:933.

144. Martins F, Sofiya L, Sykiotis GP, et al. Adverse effects of immune-checkpoint inhibitors: epidemiology, management and surveillance. *Nat Rev Clin Oncol*. 2019;16:563-580.

145. Chitturi KR, Xu J, Araujo-Gutierrez R, et al. Immune checkpoint inhibitor-related adverse

cardiovascular events in patients with lung cancer. *J Am Coll Cardiol CardioOnc*. 2019;1:182-192.

146. Nazer B, Humphreys BD, Moslehi J. Effects of novel angiogenesis inhibitors for the treatment of cancer on the cardiovascular system: focus on hypertension. *Circulation*. 2011;124:1687-1691.

147. Waliyan S, Sainani KL, Park LS, Zhang CA, Srinivas S, Witteles RM. Increase in blood pressure associated with tyrosine kinase inhibitors targeting vascular endothelial growth factor. *J Am Coll Cardiol CardioOnc*. 2019;1:24-36.

148. Moslehi JJ. Cardiovascular toxic effects of targeted cancer therapies. *N Engl J Med*. 2016;375:1457-1467.

149. Laenens D, Yu Y, Santens B, et al. Incidence of cardiovascular events in patients treated with immune checkpoint inhibitors. *J Clin Oncol*. 2022;40:3430-3438.

150. Fyfe G, Fisher RI, Rosenberg SA, Szoln M, Parkinson DR, Louie AC. Results of treatment of 255 patients with metastatic renal cell carcinoma who received high-dose recombinant interleukin-2 therapy. *J Clin Oncol*. 1995;13:688-696.

151. Atkins MB, Lotze MT, Dutcher JP, et al. High-dose recombinant interleukin 2 therapy for patients with metastatic melanoma: analysis of 270 patients treated between 1985 and 1993. *J Clin Oncol*. 1999;17:2105-2116.

152. Klapper JA, Downey SG, Smith FO, et al. High-dose interleukin-2 for the treatment of metastatic renal cell carcinoma: a retrospective analysis of response and survival in patients treated in the surgery branch at the National Cancer Institute between 1986 and 2006. *Cancer*. 2008;113:293-301.

153. Vaishampayan UN, Tomczak P, Muzaffar J, et al. Nemvaleukin alfa monotherapy and in combination with pembrolizumab in patients (pts) with advanced solid tumors: ARTISTRY-1. *J Clin Oncol*. 2022;40:2500.

154. Yeku OO, Purdon TJ, Koneru M, Spriggs D, Brentjens RJ. Armored CAR T cells enhance anti-tumor efficacy and overcome the tumor microenvironment. *Sci Rep*. 2017;7:10541.

155. Mo F, Yu Z, Li P, et al. An engineered IL-2 partial agonist promotes CD8⁺ T cell stemness. *Nature*. 2021;597:544-548.

156. Zhang Q, Hresko ME, Picton LK, et al. A human orthogonal IL-2 and IL-2Rbeta system enhances CAR T cell expansion and antitumor activity in a murine model of leukemia. *Sci Transl Med*. 2021;13:eabg6986.

157. Conlon KC, Lugli E, Welles HC, et al. Redistribution, hyperproliferation, activation of natural killer cells and CD8 T cells, and cytokine production during first-in-human clinical trial of recombinant human interleukin-15 in patients with cancer. *J Clin Oncol*. 2015;33:74-82.

158. Binnewies M, Roberts EW, Kersten K, et al. Understanding the tumor immune microenvironment (TIME) for effective therapy. *Nat Med*. 2018;24:541-550.

159. De Cicco P, Ercolano G, Ianaro A. The new era of cancer immunotherapy: targeting myeloid-

derived suppressor cells to overcome immune evasion. *Front Immunol.* 2020;11:1680.

160. Kanterman J, Sade-Feldman M, Biton M, et al. Adverse immunoregulatory effects of 5FU and CPT11 chemotherapy on myeloid-derived suppressor cells and colorectal cancer outcomes. *Cancer Res.* 2014;74:6022-6035.

161. Vincent J, Mignot G, Chalmin F, et al. 5-Fluorouracil selectively kills tumor-associated myeloid-derived suppressor cells resulting in enhanced T cell-dependent antitumor immunity. *Cancer Res.* 2010;70:3052-3061.

162. Tap WD, Gelderblom H, Palmerini E, et al. Pexidartinib versus placebo for advanced tenosynovial giant cell tumour (ENLIVEN): a randomised phase 3 trial. *Lancet.* 2019;394:478-487.

163. Benner B, Good L, Quiroga D, et al. Pexidartinib, a novel small molecule CSF-1R inhibitor in use for tenosynovial giant cell tumor: a systematic review of pre-clinical and clinical development. *Drug Des Devel Ther.* 2020;14:1693-1704.

164. Connolly KA, Belt BA, Figueroa NM, et al. Increasing the efficacy of radiotherapy by modulating the CCR2/CCR5 chemokine axes. *Oncotarget.* 2016;7:86522-86535.

165. Cheng P, Chen X, Dalton R, et al. Immuno-depletion of MDSC by AMV564, a novel bivalent, bispecific CD33/CD3 T cell engager, ex vivo in MDS and melanoma. *Mol Ther.* 2022;30:2315-2326.

166. Ruella M, Klichinsky M, Kenderian SS, et al. Overcoming the immunosuppressive tumor microenvironment of hodgkin lymphoma using chimeric antigen receptor T cells. *Cancer Discov.* 2017;7:1154-1167.

167. Sallman DA, Asch AS, Al Malki MM, et al. The first-in-class anti-CD47 antibody magrolimab (SF9) in combination with azacitidine is effective in MDS and AML patients: ongoing phase 1b results. *Blood.* 2019;134, 569-569.

168. O'Hara MH, O'Reilly EM, Varadhachary G, et al. CD40 agonistic monoclonal antibody APX005M (sotigalimab) and chemotherapy, with or without nivolumab, for the treatment of metastatic pancreatic adenocarcinoma: an open-label, multicentre, phase 1b study. *Lancet Oncol.* 2021;22:118-131.

169. Padron LJ, Maurer DM, O'Hara MH, et al. Sotigalimab and/or nivolumab with chemotherapy in first-line metastatic pancreatic cancer: clinical and immunologic analyses from the randomized phase 2 PRINCE trial. *Nat Med.* 2022;28:1167-1177.

170. Burnet FM. Immunological aspects of malignant disease. *Lancet.* 1967;1:1171-1174.

171. de Miguel M, Calvo E. Clinical challenges of immune checkpoint inhibitors. *Cancer Cell.* 2020;38:326-333.

172. Champiat S, Ferrara R, Massard C, et al. Hyperprogressive disease: recognizing a novel pattern to improve patient management. *Nat Rev Clin Oncol.* 2018;15:748-762.

173. Vonderheide RH. The immune revolution: a case for priming, not checkpoint. *Cancer Cell.* 2018;33:563-569.

174. Amitai I, Gafter-Gvili A, Shargian-Alon L, Raanani P, Gurion R. Obinutuzumab-related adverse events: a systematic review and meta-analysis. *Hematol Oncol.* 2021;39:215-221.

175. Kantarjian H, Stein A, Gokbuget N, et al. Blinatumomab versus chemotherapy for advanced acute lymphoblastic leukemia. *N Engl J Med.* 2017;376:836-847.

176. Maude SL, Laetsch TW, Buechner J, et al. Tisagenlecleucel in children and young adults with B-cell lymphoblastic leukemia. *N Engl J Med.* 2018;378:439-448.

177. Schuster SJ, Bishop MR, Tam CS, et al. Tisagenlecleucel in adult relapsed or refractory diffuse large B-cell lymphoma. *N Engl J Med.* 2019;380:45-56.

178. Neelapu SS, Locke FL, Bartlett NL, et al. Axicabtagene ciloleucel CAR T-cell therapy in refractory large B-cell lymphoma. *N Engl J Med.* 2017;377:2531-2544.

179. Jacobson CA, Chavez JC, Sehgal AR, et al. Axicabtagene ciloleucel in relapsed or refractory indolent non-Hodgkin lymphoma (ZUMA-5): a single-arm, multicentre, phase 2 trial. *Lancet Oncol.* 2022;23:91-103.

180. Shah BD, Ghobadi A, Oluwole OO, et al. KTE-X19 for relapsed or refractory adult B-cell acute lymphoblastic leukaemia: phase 2 results of the single-arm, open-label, multicentre ZUMA-3 study. *Lancet.* 2021;398:491-502.

181. Wang M, Munoz J, Goy A, et al. KTE-X19 CAR T-cell therapy in relapsed or refractory mantle-cell lymphoma. *N Engl J Med.* 2020;382:1331-1342.

182. Abramson JS, Palomba ML, Gordon LI, et al. Lisocabtagene maraleucel for patients with relapsed or refractory large B-cell lymphomas (TRANSCEND NHL 001): a multicentre seamless design study. *Lancet.* 2020;396:839-852.

183. Munshi NC, Anderson LD Jr, Shah N, et al. Idecabtagene vicleucel in relapsed and refractory multiple myeloma. *N Engl J Med.* 2021;384:705-716.

184. Berdeja JG, Madduri D, Usmani SZ, et al. Ciltacabtagene autoleucel, a B-cell maturation antigen-directed chimeric antigen receptor T-cell therapy in patients with relapsed or refractory multiple myeloma (CARTITUDE-1): a phase 1b/2 open-label study. *Lancet.* 2021;398:314-324.

185. Mackall CL, Fry TJ, Gress RE. Harnessing the biology of IL-7 for therapeutic application. *Nat Rev Immunol.* 2011;11:330-342.

186. Kim MY, Jayasinghe R, Devenport JM, et al. A long-acting interleukin-7, rIL-7-hyFc, enhances CAR T cell expansion, persistence, and anti-tumor activity. *Nat Commun.* 2022;13:3296.

187. Lee SW, Choi D, Heo M, et al. hIL-7-hyFc, A long-acting IL-7, increased absolute lymphocyte count in healthy subjects. *Clin Transl Sci.* 2020;13:1161-1169.

188. Naing A, Papadopoulos KP, Autio KA, et al. Safety, antitumor activity, and immune activation of pegylated recombinant human interleukin-10 (AM0010) in patients with advanced solid tumors. *J Clin Oncol.* 2016;34:3562-3569.

189. Strauss J, Heery CR, Kim JW, et al. First-in-human phase I trial of a tumor-targeted cytokine (NHS-IL12) in subjects with metastatic solid tumors. *Clin Cancer Res.* 2019;25:99-109.

190. Cooley S, He F, Bachanova V, et al. First-in-human trial of rIL-15 and haploidentical natural killer cell therapy for advanced acute myeloid leukemia. *Blood Adv.* 2019;3:1970-1980.

191. Robertson MJ, Stamatkin CW, Pelloso D, Weisenbach J, Prasad NK, Safa AR. A dose-escalation study of recombinant human interleukin-18 in combination with ofatumumab after autologous peripheral blood stem cell transplantation for lymphoma. *J Immunother.* 2018;41:151-157.

192. Hashmi MH, Van Veldhuizen PJ. Interleukin-21: updated review of phase I and II clinical trials in metastatic renal cell carcinoma, metastatic melanoma and relapsed/refractory indolent non-Hodgkin's lymphoma. *Expert Opin Biol Ther.* 2010;10:807-817.

193. Thompson JA, Curti BD, Redman BG, et al. Phase I study of recombinant interleukin-21 in patients with metastatic melanoma and renal cell carcinoma. *J Clin Oncol.* 2008;26:2034-2039.

KEY WORDS cancer, immune therapy, immunotherapy, innovation, treatment