



Targeting Approaches of Nanomedicines in Acute Myeloid Leukemia

Xiao Huang¹ , Hai Lin², Feng Huang³, Yuning Xie¹,
Ka Hong Wong¹, Xiaoyu Chen¹, Dongyue Wu², Aiping Lu¹,
and Zhijun Yang¹

Abstract

Acute myeloid leukemia (AML) is a hematological malignancy, which is commonly associated with high incidence and mortality among adult patients. The standard induction regimen for AML has been substantially unchanged over the past 40 years, for which novel nanomedicines have represented a promising strategy in AML therapies. Despite developments of multiple nanoparticles formulated with drugs or genes, less there is not much information available about approaches in AML is available. This review presents an overview of nanomedicines currently being evaluated in AML. First, it briefly summarized conventional chemotherapies in use. Second, nanomedicines presently ongoing in clinical trials or preclinical researches were classified and described, with illustrative examples from recent literatures. Finally, limitations and potential safety issues concerns in clinical translation of AML treatment were discussed as well.

Keywords

nanomedicines, nanoparticles, cancer therapy, acute myeloid leukemia

Introduction

Acute myeloid leukemia (AML) represents as a hematologic malignancy, which is the most common leukemia among adults and more detrimental in older patients, with a higher incidence rate and almost 90% mortality rate for those older than 65 years,¹ as well as heterogeneous clinical presentations and subtypes.^{2,3} It was estimated that there were over 20 000 new cases and 10 670 deaths of patients with AML in the United States in 2018.⁴

Although progresses have been made in pathobiology and novel therapeutics targets, the standard induction therapies for AML have not been changed substantially for decades.⁴ Furthermore, with involvement of chemoresistance side effects, such as cardiotoxicity, myelosuppression, and infections, conventional therapeutics always lead to treatment failure or relapse; hence, the overall prognosis remains poor.^{5,6} Therefore, it is urgent to develop new therapeutic approaches.

Nano-based drug delivery systems have potentials to deliver drugs to specific area more efficiently and reduce side effects.⁷⁻⁹ Therefore, nanomedicines have been highlighted as a new strategy to optimize AML therapies.¹⁰⁻¹² Moreover, by conjugated with various ligands, drugs could actively target to AML cell-surface receptors, which may have a tremendous

improvement in treatment response or overall survival (OS) rates.^{13,14}

This review attempts to present an overview of nanomedicines for AML currently in experimental researches or clinical trials from published data, with a focus on targeting ligands. This article may not be exhaustive due to the rapid development of new drugs. Rather, we aim to illustrate some potential targeting ligands for nanoparticles (NPs) in AML, which could be utilized in therapeutic nanoplatfroms.

¹ School of Chinese Medicine, Hong Kong Baptist University, Hong Kong, China

² Department of Traditional Chinese Medicine, Beijing Friendship Hospital, Capital Medical University, Beijing, China

³ Institute of Acupuncture & Moxibustion, China Academy of Chinese Medical Sciences, Beijing, China

Received 07 July 2019; received revised 10 September 2019; accepted 23 September 2019

Corresponding Authors:

Zhijun Yang and Aiping Lu, School of Chinese Medicine, Hong Kong Baptist University, Hong Kong, China.

Emails: yzhijun@hkbu.edu.hk; aipinglu@hkbu.edu.hk



Current Treatments in AML

Standard Chemotherapy

The standard induction therapy is the combination of cytarabine and anthracycline (daunorubicin or idarubicin), which has been represented as “7+3 regimen” for more than 40 years.^{15,16} This regimen is administered as a continuous infusion of cytarabine for 7 days and anthracycline for the following 3 days. Consolidation is then conducted for the next phase after achieving complete remission (CR), which is called postremission therapy. Currently, cladribine combined with “7+3 regimen” is applied as a second standard regimen, which when received leads to an improvement in the CR rate and a 3-year OS.¹⁷ Fludarabine and etoposide are also used as an alternative regimen under poor heart functions.^{16,18} Recently, several new cytotoxic drugs have been utilized in clinic. Azacitidine and decitabine are 2 new DNA methyltransferase inhibitors recommended in the National Comprehensive Cancer Network guidelines.¹⁹

Target Therapy

FMS-like tyrosine kinase 3 (FLT3) mutation has been the foremost common change in patients with AML. Midostaurin was the first FLT3 inhibitor proved by the Food and Drug Administration (FDA) and commonly used in FLT3-mutated AML.²⁰ According to an international randomized phase III trial, combined with “7+3 regimen,” midostaurin could significantly improve median OS from 25.6 to 74.7 months in patients with FLT3-mutated AML; however, a large proportion of patients were relapsed within 2 years.²¹ Quizartinib and cabozantinib were investigated as the second generation FLT3 inhibitors, specifically targeting FLT3 wild-type and FLT3 internal tandem duplications (ITDs) mutation, which achieved higher CR, but still had inevitable drug resistance.^{22,23} Recently, a novel FLT3 inhibitor gilteritinib has been approved by FDA for relapsed or refractory AML harboring FLT3 mutation, with an achievement of 40% overall response rate and 8% CR.²⁴ Crenolanib is another FLT3 inhibitor enrolled within a phase III trial currently in patients with FLT3-ITD or FLT3-TKD mutated AML (NCT02298166).²⁵

Approximately 20% cases of AML are detected with IDH1 and IDH2 mutations.²⁶ The IDH1 inhibitor enasidenib and IDH2 inhibitor ivosidenib, proved by FDA, were reported achieving an effective overall response for 41.6%²⁷ and 40%,²⁸ respectively. However, acquired clinical resistances were subsequently detected after treatment.

B-cell lymphoma 2 (BCL-2) has been considered as an oncogene and overexpressed in patients with AML.²⁹ A BCL-2 inhibitor venetoclax, which is approved by FDA for chronic lymphocytic leukemia (CLL), has been evaluated in several clinical trials either as a single agent or combined treatment for AML (NCT02203773, NCT02993523, NCT02287233, and NCT03069352).

Immunotherapy

Gemtuzumab ozogamicin, an anti-CD33 monoclonal antibody conjugated with calicheamicin, was first approved by FDA for CD33-positive AML. CD33 is a membrane receptor and potential target, which is highly expressed on leukemic progenitor cells but less on normal hematopoietic stem cells.³⁰ Gemtuzumab ozogamicin was withdrawn in 2010 and ratified again in 2017 with dose and patient population modification. Currently, it is used in combined treatment with induction therapies or as a single agent in relapsed cases.³¹ However, toxicities in live cases still remain a concern because CD33 was also expressed on hepatocytes.^{32,33} Other novel anti-CD33 monoclonal antibodies, such as vadastuximab and AMG 330, are under clinical trials at present,³⁴⁻³⁶ which need further evaluations for dose effect or safety. Moreover, overexpression of CD123 has been observed in patients with resistance or relapsed AML, which is a potential target for the novel monoclonal antibody, SL-401. Currently, several phase I/II studies associated with SL-401 are ongoing.^{37,38}

There are some immune checkpoint inhibitors used to treat AML as well. It has been demonstrated that immune checkpoints, such as cytotoxic T-lymphocyte-associated protein 4 (CTLA4), programmed cell death 1 (PD-1), and programmed cell death-ligand 1 (PD-L1), play unique roles in maintaining malignancy survivals and synergistically inhibiting immune responses against tumors.³⁹ Inhibitors CTLA4 and PD-1/PD-L1 pathways have shown effects on promoting immune-mediated antileukemia responses and increasing survivals in murine AML models.^{40,41} Clinical trials of the PD-1 inhibitors nivolumab and pembrolizumab, PD-L1 inhibitors durvalumab and atezolizumab, as well as CTLA-4 inhibitor in patients with AML, which have shown well toleration and encouraging response to relapsed AML, are in progress at present.⁴²⁻⁴⁵

Chimeric antigen receptor (CAR) T-cell therapy is a novel antitumor immunotherapy that utilizes autologous lymphocytic T cells modified to express CARs, which could target on specific antigen of tumor. Chimeric antigen receptor T-cell therapy has received remarkable outcomes in patients with B-cell malignancies.⁴⁶ However, the approach has been restricted in AML treatment. Although several target antigens have been studied, there is no ideal molecule and no authority has approved CAR T-cell therapy for AML yet.^{47,48} In summary, AML is a complex hematological malignancy with high mortality; more studies and clinical trials are required for current chemotreatments to reduce toxicities and improve efficacies.

Approaches of Nanomedicines for AML

Nanoparticles have been highlighted in cancer therapies, with advantages of enhancing permeability, reducing adverse effects, getting around multidrug resistances (MDRs), improving bioavailability, and prolonging enhanced permeability and retention effects with drug circulations by loading and delivering anticancer agents to tumor site, which have been providing promising approaches to cancer therapies.^{49,50} Generally, NPs

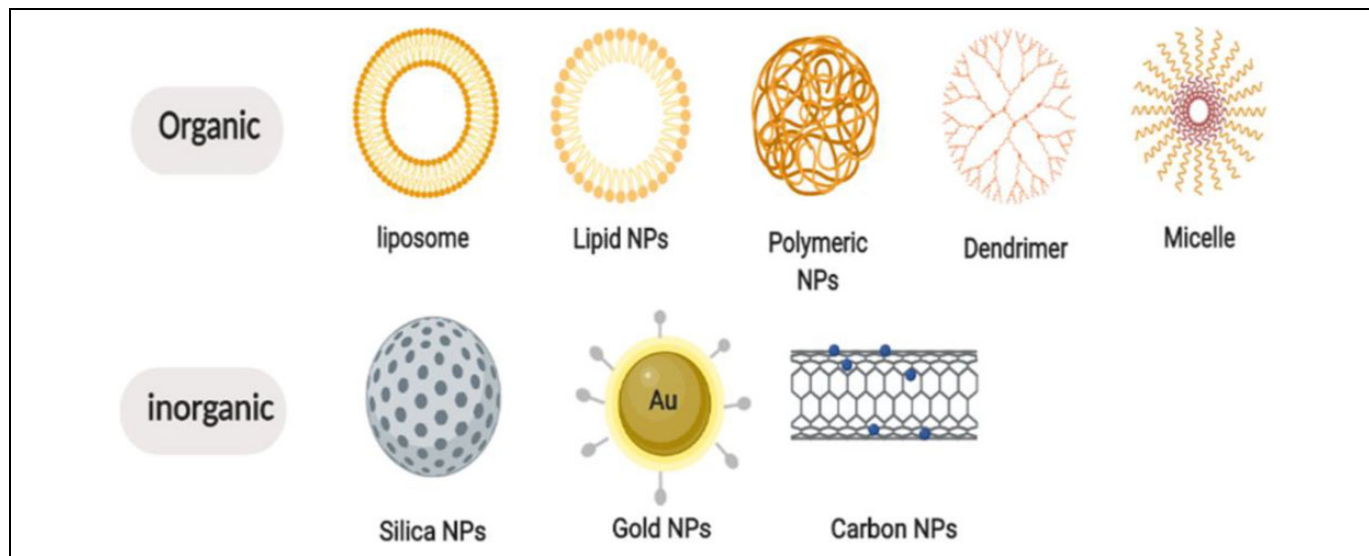


Figure 1. Different classes of nanoparticles.

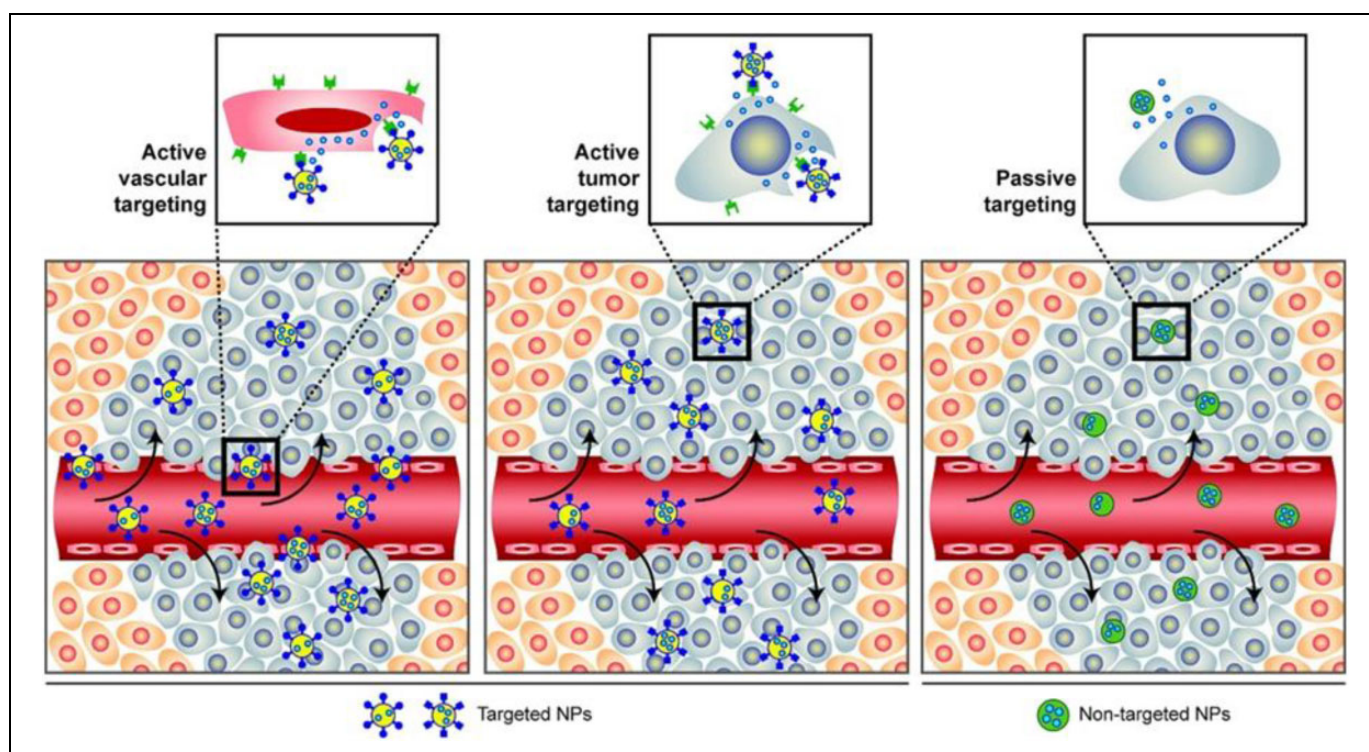


Figure 2. A schematic diagram of passive and active targeting. Nanoparticles incorporated with ligands could specifically active target on receptors of blood vessel or cells (left and middle), while the other way is passively targeting and accumulating through enhanced permeability and retention effect (right). Adapted with permission from Farokhzad and Langer.⁵³ Copyright (2009) American Chemical Society.

conjugate drugs are already used in AML therapies. Depending on formulation compositions and materials, NPs are mainly constituted with 2 types, organic NPs, such as liposomes, polymeric, micelle, dendrimer, carbon, and so on, and inorganic NPs, such as silica, metal, semiconductor, and so on,^{51,52} as shown in Figure 1. Generally, there are 2 kinds of classifications for NP-mediated drug deliveries. One is known as

passive targeting, which enables different types of NPs passing across capillary endothelium of tumor vessels and accumulating in tumor site. The other is called active targeting, which could recognize cell-surface receptor and target tumor phenotypes directly by utilizing specific ligands (antibodies, proteins, peptides etc.) conjugating with NPs (Figure 2).⁵³ Table 1 presents a summary of clinical trials ongoing in AML, among the 20

Table 1. Ongoing Clinical Trials Involving Nanomedicines in Acute Myeloid Leukemia.

Class	Compound	Clinical Phase	Trial Name	Status	Identifier
Liposomes	Annamycin	I/II	Study of Liposomal Annamycin for the Treatment of Subjects With AML	Recruiting	NCT00430443
		II	Bortezomib and Doxil for the Treatment of Patients With AML	Not yet recruiting	NCT01736943
	Doxorubicin	III	International Randomized Phase III Clinical Trial in Children With AML	Recruiting	NCT02724163
			(daunorubicin-cytarabine)	II	
	CPX-351		Liposome-encapsulated Daunorubicin-Cytarabine and Venetoclax in Treating Participants With Relapsed, Refractory or Untreated AML	Recruiting	NCT0362917
		I/II	Cytarabine and Daunorubicin in Combination With Ruxolitinib for the Treatment of Secondary AML Transformed From MDS	Recruiting	NCT03878199
		II	Investigator Initiated Trial of CPX-351 for Untreated AML	Recruiting	NCT03335267
		I	A Trial to Evaluate the Potential Impact of Renal Impairment on the Pharmacokinetics and Safety of CPX-351	Recruiting	NCT03555955
		II	Cytarabine, Idarubicin, Liposome-encapsulated Daunorubicin-Cytarabine or Decitabine in Treating Older Patients With AML	Recruiting	NCT03226418
		I	CPX-351 and Gemtuzumab Ozogamicin in Treating Patients With Relapsed AML	Not yet recruiting	NCT03904251
		I	CPX-351+GO in Subjects 55 Years Old, or Older, With AML	Not yet recruiting	NCT03878927
		IV	The Feasibility of Safely Managing Patients Receiving Induction With Liposomal Daunorubicin and Cytarabine (CPX-351) for AML in an Outpatient Environment	Not yet recruiting	NCT03988205
		I/II	Phase I/II Trial of CPX-351 + Palbociclib in Patients With AML	Not yet recruiting	NCT03844997
		II	CPX-351 in Treating Patients With Newly Diagnosed, High-Risk AML	Not recruiting	NCT02286726
	III	Phase III Study of CPX-351 Versus 7+3 in Patients 60-75 Years Old With Untreated High Risk (Secondary) AML	Not recruiting	NCT01696084	
BP1001	II	Clinical Trial of BP1001 (Liposomal Grb2 Antisense Oligonucleotide) in Combination With Decitabine in AML/High Risk MDS	Recruiting	NCT02781883	
	I/II	Clinical Trial of BP1001 (Liposomal Grb2 Antisense Oligonucleotide) in Combination With Dasatinib in Patients With Ph + CML Who Have Failed TKI, Ph+ AML, Ph+ MDS	Recruiting	NCT02923986	
	I	Recruiting Clinical Trial of BP1001 (L-Grb-2 Antisense Oligonucleotide) in CML, AML, ALL & MDS	Not recruiting	NCT01159028	
Vincristine	I	EphB4-HSA Fusion Protein and Cytarabine /or Liposomal Vincristine in Patients With Recurrent or Refractory Acute Leukemia	Recruiting	NCT03519984	
Polymeric NPs	AZD2811	I	A Phase I Study of Safety, Tolerability, and PK of AZD2811 in Patients With Advanced Solid Tumors		NCT02579226
		I/II	Safety, Tolerability, Pharmacokinetics, and Efficacy of AZD2811 Nanoparticles as Monotherapy or in Combination in Acute Myeloid Leukemia Patients		NCT03217838

Abbreviations: ALL, acute lymphoblastic leukemia; AML, acute myeloid leukemia; CML, chronic myeloid leukemia; MDS, myelodysplastic syndrome; NPs, nanoparticle; Ph+, Philadelphia chromosome positive; PK, pharmacokinetics.

clinical trials, of which 18 were identified using liposomal formulations and only 2 were based on polymeric NPs. No other type of NPs is under clinical investigation currently in AML.

Passive Targeting Approaches in AML

Liposome-based nanomedicines. Liposomes are spherical vesicles composed of phospholipid bilayers, which could capture either

hydrophilic agents by aqueous core or lipophilic agents by lipid bilayer during transportation without structural changes.^{54,55}

For the purpose of increasing solubility, bioactivities, and distributions, liposomes have been considered the most prevalent category. At present, there are 16 liposomal drugs ratified by FDA or/and European Medicines Agency (EMA).^{56,57}

Liposomal nanomedicines approved in AML therapy. CPX-351 (Vyxeos) is a liposomal-encapsulated cytarabine and

daunorubicin with a fixed molar ratio (5:1), which was approved by FDA and EMA in 2017 for treating newly diagnosed therapy-related AML and/or AML with myelodysplasia-related changes.⁵⁸ CPX-351 was initially synthesized and evaluated in *in vitro* and *in vivo* studies with leukemia cell lines. The results indicated that the liposomal-encapsulated cytarabine and daunorubicin could display a best synergistic effect and minimum antagonism at the ratio of 5:1, with higher proportions of response rates, more durable remissions, and longer maintenances in bone marrows, compared to a free drug cocktail of cytarabine and daunorubicin with their maximum tolerated doses (MTD).⁵⁹⁻⁶¹ The therapeutic efficacy and frequency of CPX-351 were then examined using human leukemia xenograft model.⁶¹ Moreover, a later study tested in primary blood cells from patients with AML and normal bone marrow donors with CPX-351 and the same ratio of free drugs has shown that normal peripheral blood and bone marrow were more sensitive to free drugs, which illustrated that CPX-351 could preferentially accumulate in AML cells over normal cells.⁶² Subsequently, phase I study for CPX-351 was conducted in patients with refractory/relapsed AML in 2011, with a dose ranging from 32 U/m² of the initial response dose to 101 U/m² of MTD.⁶³ It showed that 9 of 43 patients received CR, accompanied with side effects such as hypertensive crisis, congestive heart failure, and prolonged cytopenias.⁶⁴ Further, a phase II trial explored efficiency and safety of CPX-351 in 126 older patients with AML and those with secondary AML (sAML) in 2014.⁶⁵ This trial demonstrated that there was a tendency of higher CR and CR with incomplete blood count recovery for CPX-351 compared to conventional “7+3” therapy (66.7% vs 51.2%, $P = .07$), especially in sAML subgroup, with a trend toward improving response rate (57.6% vs 31.6%, $P = .06$), prolonging event-free survival (EFS; hazard ratio [HR], 0.59; $P = .08$), and OS (HR, 0.46; $P = .01$). Although prolonged cytopenias and higher risk of infections were detected as well, there were lower rates in infection-related deaths (3.5% vs 7.3%) and 60-day mortality (4.7% vs 14.6%). With the potential clinical benefit of CPX-351, another phase II study followed up in 2015 comparing CPX-351 to “7+3” induction therapy in 125 patients with first relapsed AML.⁶⁶ Despite no improvement in 1-year EFS or OS, there was also a higher response rate (39.3% vs 27.6%), lower 60-day mortality rate (16.1% vs 24.1%), improved EFS (HR, 0.63; $P = .08$), and OS (HR, 0.55; $P = .02$) for European Prognostic Index–defined, poor-risk patients in CPX-351 group.

Based on these encouraging results, CPX-351 was advanced into phase III clinical studies for further ascertainment. A randomized phase III study comparing first-line CPX-351 (100 U/m²) with “7+3” regimen (daunorubicin, 60 mg/m²; cytarabine, 100 mg/m²) in 309 elderly patients (60-75 years) with high-risk sAML, indicated a significantly improved OS (9.56 months vs 5.95 months), composite response rates (47.7% vs 33.3%), and lower early mortality rates (5.9% and 13.7% vs 10.6% and 21.2%, through 30-day and 60-day, respectively), whereas a comparable frequency and severity of grade 3 to 5 adverse events.⁶⁷ These

encouraging results were presented at 2016 American Society of Clinical Oncology meeting and finally led to FDA approval in 2017. Lancet et al⁶⁸ further analyzed the data, consistent with these observations; CPX-351 indicated a significant improvement in survival over standard induction chemotherapy for high-risk patients with AML, older patients with sAML, and poor-risk subgroup of patients with AML.

Liposomal nanomedicines under clinical trials in AML therapy. As shown in Table 1, liposomal formulations of vincristine, doxorubicin, annamycin, daunorubicin, and BP1001 are being evaluated in clinical trials at phase I or II stages currently. Liposomal doxorubicin (Doxil) and non-PEGylated liposomal doxorubicin (Myocet) have already been approved for the treatment of AIDS-related Kaposi sarcoma, multiple myeloma (MM), ovarian cancer, and breast cancer.⁶⁹⁻⁷¹ Melillo et al⁷² have assessed Myocet combined with fludarabine, cytarabine, and granulocyte colony-stimulating factor (FLAG) in 35 elderly patients with AML, showing a median disease-free survival (DFS) at 12 months, 1-year, and 2-year DFS of 78.9% and 26.7%, and CR and partial remission of 63.8% and 8.5%, respectively, with a 20% resistance and 17% of severe cardiovascular toxicity. Another clinical study utilizing the same regimen was conducted in 18 children with refractory or relapsed AML, which achieved a CR rate of 18% (11/18), OR at 3 years of 38%, EFS at 3 years of 40%, and DFS at 3 years of 58% after hematopoietic stem cell transplantation, with well tolerant and remarkable low toxicity.⁷³ There is one phase II clinical trial ongoing currently (NCT03059615), which is designed to evaluate the safety and efficacy of bortezomib combined with liposomal doxorubicin in patients with relapsed MM, CLL, and non-Hodgkin lymphoma as well as elderly patients with relapsed/refractory AML who are not candidates for standard induction therapy.

Liposomal daunorubicin (DaunoXome) is a non-PEGylated liposomal-encapsulated anthracycline daunorubicin. A phase III study was conducted by the International Berlin-Frankfurt-Münster Study Group in 2013 among pediatric patients with relapsed AML. Patients were randomly assigned to regimens of FLAG and FLAG plus daunorubicin. Although OS and grade 3 to 4 toxicities were similar, FLAG plus daunorubicin regimen showed an improved day 28 BM status (80% vs 70%), higher CR rate (69% vs 59%) compared to FLAG regimen.⁷⁴ There is an international randomized phase III clinical trial that enrolled liposomal daunorubicin ongoing in children with AML (NCT02724163).

Antisense oligonucleotides, which refer to a class of small interfering RNA (siRNA), microRNA, or short hairpin RNA, have shown a great potential in cancer therapy and been approved for ALL treatment by FDA in 2017.⁷⁵ However, the clinical application is limited due to instability circulation, inefficiency delivery, and off-target adverse effects. BP1001 is a liposomal formulation of growth factor receptor-bound protein-2 antisense oligodeoxynucleotide (L-Grb-2 antisense oligonucleotide). Previously, a single-center, dose-escalation phase I/Ib clinical trial combined with low-dose cytarabine in

patients with refractory or relapsed AML, Philadelphia chromosome-positive (Ph+) chronic myeloid leukemia (CML), Ph+ ALL, or Ph+ myelodysplastic syndrome (MDS) demonstrated a well tolerated with an improved therapeutic activity.⁷⁶ There are 3 clinical trials underway. A phase I trial (NCT01159028) is to evaluate the highest safe dose for patients with AML, Ph+ CML, Ph+ ALL, and Ph+ MDS, in addition to the safety and toxicity in combination with low-dose Ara-C for patients with AML. Another phase I/II trial (NCT02923986) is to determine the efficacy with dose-limiting toxicity and maximal tolerated dose in combination with dasatinib in patients with Ph+ AML, Ph+ CML, or high-risk Ph+ MDS. The third phase II trial (NCT02781883) is to assess the efficacy in combination with decitabine in patients with AML or high-risk MDS.

Liposomal vincristine (Marqibo) was approved by FDA in 2012 for patients with Ph- ALL. There is also a phase I clinical trial ongoing (NCT03519984) that enrolls liposomal vincristine as part of a regimen containing EphB4-HSA fusion protein and cytarabine, for detecting patients with different types of acute leukemia, including AML.

Liposomal nanomedicines investigated in preclinical stage for AML therapy. Other liposomal nanomedicines are mostly being tested in vitro or in vivo. A liposome formulation of safinol was designed and evaluated with antitumor activities in human AML cell lines, patient samples, and murine xenograft models, as well as a longer survival time in U937-inoculated mice.⁷⁷ Subsequently, a liposomal coencapsulation of safinol/C2-ceramide was developed, which indicated effectivity in vitro and xenograft models, with a dose reduction of 33% compared to liposomal safinol or liposomal C2-ceramide alone.⁷⁸ Myhren et al⁷⁹ have reported a PEGylated liposome coencapsulating anthracycline daunorubicin (DNR) and emetine with folate modification, which enhanced loading ability than DNR alone. Leukemia stem cell (LSC) with overexpression of miR-126 has been considered as a potential therapeutic target for AML.⁸⁰ Dorrance et al⁸¹ designed a liposomal formulation containing antagomiR-126 with ligands of transferrin or antibody CD45 on surface. The formulation was tested on murine xenograft models and showed a significant improvement in survival rate with an interference with LSC. GTI-2040 is a 20-mer antisense oligonucleotide complementary to a coding region in the messenger RNA (mRNA) of the R2 small subunit component of human ribonucleotide reductase. Li et al⁸² designed and evaluated an immunoliposome-encapsulated GTI-2040, grafted with a target ligand of anti-CD33 to AML cells. It substantially downregulated the mRNA and protein expression of R2, reduced 15 times of IC₅₀ than Ara-C, and decreased tumor volumes in Kasumi-1 xenografted model. The delivery efficiency of liposomes can be affected by physicochemical properties, such as particle size, zeta potential, or drug-release kinetics. Liposomes have achieved a series of encouraging results and are in different stages of evaluation. However, it is still unable to predict biological interactions by

physicochemical properties accurately, which restrict converting preclinical effects to clinical benefits in AML.⁸³⁻⁸⁵

Polymer-based nanomedicines in AML therapy. Polymers containing polymeric particles, micelles, and dendrimers have been mainly investigated in preclinical studies to date. With advantages of biocompatible, tailored release, prolonging circulation, and functionalizing with specific peptide targeting ligands or antibodies, polymers have been the most intensively explored materials in drug delivery systems.⁸⁶⁻⁸⁸

Polymeric NPs could conjugate hydrophobic drugs by encapsulating to solid cores or water-soluble drugs by covalently attaching, which could control drug release, prolong circulation time, or reduce toxicity.⁸⁹⁻⁹² Polylactide (PLA) and poly(lactide-co-glycolide) (PLGA) are commonly used polymers at present. AZD2811 is a polymeric NP loaded with aurora kinase B inhibitor. It has been assessed in AML xenografts model and shown an improved efficacy in inhibiting tumor growth and inducing apoptosis compared to free aurora kinase B inhibitor (AZD1152). Moreover, this formulation also demonstrated a transient cellular reduction in the bone marrow, which may have a potential agent for targeting residual disease.⁹³ There are 2 clinical trials ongoing for evaluating the safety, tolerability, and pharmacokinetics of AZD2811 (NCT02579226, NCT03217838). Poly(lactide-co-glycolide) is a FDA-approved polymer widely used as a nanocarrier. Simon et al⁹⁴ developed a PLGA polymeric NPs-encapsulated all-trans-retinoic acid (ATRA), which prolonged the drug release and induced differentiation as well as inhibited proliferation in AML cells. A PEG-PLGA polymeric micelle loaded with edelfosine and conjugated with transferrin was designed by Sun and Sun.⁹⁵ This formulation prolonged blood circulation, leading to a continuous drug release and biological activity maintenance, which resulted in a higher cytotoxic effect and apoptosis in K562 cells. Zhu et al⁹⁶ designed a PLGA and Pluronic85 copolymeric NP encapsulated with doxorubicin and grafted with transferrin, which was further evaluated in AML cell lines and relevant animal models. This formulation led to a reduction of tumor volume in vivo and an enhancement of cytotoxicity in doxorubicin-resistant cells. A polymeric NPs-loaded specific CD-44 siRNA was designed and performed in vitro.⁹⁷ It was demonstrated to inhibit stem cells-progenitor cells interactions and sensitize chemotherapies by silencing and decreasing CD44 surface levels in AML cell lines, which induced apoptosis and decreased adherence of primary AML cells to bone marrow mesenchymal stem cell. Chandran et al⁹⁸ developed a PLGA polymer-protein core-shell formulation, loaded with everolimus, sorafenib, and inhibitors of mTOR, MAPK, and STAT5, additionally conjugated with anti-CD33 antibody. The result showed that it could cause synergistic lethality against leukemic cells by simultaneous inhibition, without affecting normal blood cells.⁹⁸ Recently, another methoxy PEG-PLGA polymeric NP with encapsulated idarubicin was synthesized. The study demonstrated that, compared with free idarubicin, it could decrease cell proliferation and induce apoptosis more remarkable in vivo, and improve the OS more significantly in

murine models.⁹⁹ PCX is a polymeric NP loaded with AMD3100, a FDA-approved CXCR4 inhibitor and carrying siRNA simultaneously. This PCX/siRNA nanomedicine exerted a higher cytotoxic effect on AML cell lines compared to other CXCR4 inhibitors. In addition, it could deliver siRNAs against the transcription factor RUNX1, which was typically required in AML subtypes.¹⁰⁰

Micelles are promising copolymers nanomaterial composed of hydrophobic core and hydrophilic corona, which are good candidates for encapsulating hydrophobic anticancer agents and ensuring solubility.¹⁰¹ L-PLA micelle formulation of paclitaxel (Genexol-PM), docetaxel-loaded polymeric micelle (NanoxelM), and paclitaxel micellar (Paclical) have been approved and used in clinic for the treatment of various types of cancers, such as breast cancer, ovarian cancer, and lung cancer, however, not yet in AML.¹⁰²⁻¹⁰⁴ Dextran is a polymeric micelle loaded with doxorubicin and grafted with folic and retinoic acid. The cytotoxicity of dextran was higher in KG-1 cells than free drug, which reduced approximately half of IC₅₀.¹⁰⁵ SP1049C is another micellar formulation of doxorubicin based on pluronic, which has completed phase II trials in patients with advanced esophagus and gastroesophageal junction adenocarcinoma. Alakhova et al¹⁰⁶ compared antitumor activities of SP1049C with doxorubicin in P388 murine leukemia ascitic tumor model. The tumor formation frequency and aggression were much more reduced in SP1049C group than that in control group. Further evaluations are needed for clinical application in AML.

Dendrimers are spherical polymers composed of a central core, multibranches, and an outer layer with functional groups, which could conjugate with charged polar molecules through electrostatic interaction by outer layers and encapsulate uncharged molecules by hydrophobic inner.¹⁰⁷ These properties of dendrimers enable covalent attach to hydrophobic anticancer agents and increase bioavailability. There is no clinical trial and few in vivo studies for AML associated with dendrimer formulations. The cytotoxicity and apoptosis of a dendrimer NPs encapsulated with cytarabine was assessed in 1301 and HL-60 cell lines previously, which showed an enhancement compared to free drug.¹⁰⁸

Metallic nanomedicines in AML therapy. Metal NPs, such as gold or silver, which are inorganic and nontoxic nanomaterials, have been considered to be useful candidates in cancer therapy for attaching and delivering drugs by use of surface plasmon resonances and photophysical properties.¹⁰⁹ There is no metallic nanomedicine under clinical trials to date, although quite a few preclinical studies have reported for AML indications. A gold NP loaded with tyrosine kinase inhibitors was designed and showed an increased efficacy compared with free drug.¹¹⁰ Another study newly synthesized a gold NP with adsorbed high-density lipoprotein loaded with BMS309403 (BMS), an AML-promoting factor fatty acid-binding protein 4 inhibitor. The result showed this formulation could induce cell differentiation and reduce progression of AML.¹¹¹

Table 2. Active Targeting Nanoparticles Investigated in AML.

Nanoparticle	Ligand	Therapeutic Agents	Ref
Liposome	Anti-CD33 mAb	GTI-2040	16
Polymer	Anti-CD33 mAb	Ara-C	17
PLGA polymer	Anti-CD33 mAb	mTOR, MAPK, and STAT5 inhibitor	18
Liposome	Anti-CD123 mAb	Daunorubicin	20
Niosome	Anti-CD123 mAb	Daunorubicin	21
Silica	Anti-B220 mAb	Daunorubicin	23
Liposome	Anti-CD45 mAb	AntagomiR-126	24,25
Polymer	Anti-CD44 mAb	siRNA	8
Polymer	Transferrin	Doxorubicin	27
Micelle	Transferrin	Edelfosine	28

Abbreviations: AML, acute myeloid leukemia; PLGA, poly(lactide-co-glycolide); siRNA, small interfering RNA.

Lipid-based nanomedicines in AML therapy. Lipid NPs are designed to encapsulate lipophilic drugs.¹¹² Currently, there is no lipid-based nanomedicines approved in clinical trials for AML, as some sporadic experimental reported. It is known that sparingly water solubility has confined clinical applications of etoposide. Khajavinia et al¹¹³ synthesized a lipid NP loaded with etoposide and conjugated with transferrin. The result indicated an enhanced cellular uptake and higher cytotoxicity in etoposide lipid-based NPs compared to that in free etoposide. An ATRA-loaded lipid-based NP was obtained and showed a significantly suppression in AML cell lines compared with ATRA in solution.¹¹⁴ Previously, fingolimod, a sphingosine analog that could activate protein phosphatase 2A in leukemia, has been demonstrated to be a potential treatment option for AML.¹¹⁵ A lipid-based NP loaded with fingolimod was therefore designed. Results showed it could induce a higher apoptosis and enhance the oral bioavailability compared with bare fingolimod solution.¹¹⁶

Other types of nanomedicines in AML therapy. Chandran et al¹¹⁷ reported a silico-based nanomedicine loaded with vorinostat, which showed a selective and superior anticancer activity against patient with primary AML cells and AML cell lines. It demonstrated a lower IC₅₀, enhanced histone deacetylase inhibition, apoptosis, and oxidative injury compared to free vorinostat, without toxicity to healthy bone marrow. Carbon-based materials, especially new discovery of grapheme, are another new kind of organic NPs that possess attachment sites on surface of ligands and deliver drugs into cytoplasm of cancer cells by carbon nanotubes. Currently, to our knowledge, there is no carbon base nanomedicine detected in AML.

Active Targeting Approaches in AML

In order to improve delivery efficiency and reduce toxicity to normal cells, there is another type of drug delivery defined as active targeting, with surface of NPs decorated with different ligands.⁵³ Ligands typically include peptides, antibodies, folic acid, retinoic acid, vitamins, and transferrins.¹¹⁸⁻¹²³ So far, investigations for AML are still in laboratory stage, and summary is presented in Table 2.

Cell-penetrating peptides (CPPs) are composed of 5 to 30 amino acids and able to carry a variety of cargoes, including NPs and antisense oligonucleotides. Cell-penetrating peptides could cross across cell membranes and deliver drugs into cells, with the advantages of highly selective for tumor site and multivalent conjugation to anticancer agents.¹²⁴ The accurate cellular uptake mechanisms of CPPs remain uncertain, accompanied with 2 major theories. One is energy-independent endocytic process and directly through the lipid bilayer, the other is energy-dependent endocytic process.¹²⁵ There is no report published on nanomedicines equipped with CPPs in AML to date; however, relevant experimental results may represent potential strategies for further investigations. A specific peptide (CP-EPS8-NLS) derived from the nuclear localization signal of epidermal growth factor receptor pathway substrate no.8 (EPS8) was synthesized and analyzed in AML cells as well as related xenograft models, which showed potential cytotoxicity effects.¹²⁶ Agarwal et al¹²⁷ evaluated the specificity and efficacy of a CPP OP449 in antagonizing SET oncoprotein in AML cells and animal models. It has been reported the CPP inhibitor of mucin 1-C-terminal subunit (MUC1-C) oncoprotein could arrest tumor cell growth, induce late apoptosis, and increase the reactive oxygen species, which resulted to induce a terminally differentiated myeloid phenotype in AML cell lines.¹²⁸ Lastly, a research showed CPP inhibitor GO-203 could depress MUC1-C aberrantly expressed in AML and downregulate the FLT3, which conferred a poor prognosis in AML.¹²⁹ Li et al¹³⁰ indicated that a TLR2-binding peptide motif (Pep2) could target and penetrate into AML cells in a dependent manner, thus inducing apoptosis in AML cell lines as well as patient samples and depressing progression in TLR2^{high} AML mice. These results indicated several optional CPPs for further investigations in AML.

Nanomedicines conjugated with antibody can specifically target on cell-surface receptor and delivery drugs into cells.¹³¹ GTI-2040 is an antisense oligonucleotide targeting the small subunit R2 of ribonucleotide reductase, which is underevaluation in clinical trials for AML. It has been known that CD33 is a membrane receptor expressed by AML progenitors but absent in normal bone marrow stem cells. GTI-2040-loaded immunoliposomes grafted with an anti-CD33 ligand has been synthesized and observed in AML cells as well as related animal models.⁸² The result indicated that it significantly downregulated expression of R2 and suppressed cell viabilities in AML cell lines. Moreover, after combining with Ara-C, there was a strengthened inhibition in tumor growth and a prolonged survival time for this immunoliposomal nanomedicine in AML xenograft model. A CD33-targeted pH-sensitive polymeric liposome encapsulated with Ara-C was designed and verified in AML cells, which obviously restrained cell viabilities and successfully internalized into CD33-positive AML cells. On the contrary, limited cellular internalization was found in control liposomes with an isotype antibody.¹³² As mentioned above, there was also an anti-CD33 incorporated multi-inhibitor-loaded PLGA polymer NPs developed.⁹⁸ CD123, which is overexpressed on AML cells, has been identified as

a potential target for treatment.¹³³ An anti-CD123 PEGylated liposomal encapsulated with daunorubicin (CD123-ILP) was synthesized and assessed in several AML cell lines with different densities of anti-CD123 antibody. CD123-ILP highly slowed down the growth of CD123⁺ AML cells and showed a stable release in vitro.¹³⁴ Another anti-CD123 niosome formulation loaded with daunorubicin was formed as well. The result demonstrated improved uptake efficiency than free antibody drug in AML cells with ligand density dependent. Moreover, it resulted in a higher cytotoxicity and prolonged survival time in vitro and in vivo treatment of AML.¹³⁵ B220, also known as CD45R, is an isoform of CD45. It has been proved that antigen B220 is specifically expressed on LSCs.¹³⁶ A liposomal formulation containing antagomiR-126 with ligands of transferrin or antibody CD45 has been discussed in previous section.^{80,81} Recently, an anti-B220-ligand mesoporous silica NP has been designed to deliver daunorubicin. This drug selectively reduced the growth of B220-positive AML LSCs and decreased progression in mice model.¹³⁷

Transferrin, a single-chain iron-transporting glycoprotein as well as a membrane receptor, is overexpressed on a majority of cancer cells and plays role in mediating intracellular uptake and regulating cancer growth.¹³⁸ It has reported a copolymeric NP conjugated with transferrin was designed to enhance the antileukemia efficacy of doxorubicin in AML cell lines.⁹⁶ There was also another polymeric NPs incorporating transferrin and edelfosine, which indicated a higher cytotoxic effect in AML cells.⁹⁵

Challenges in Clinical Translation of Nanomedicines for AML Treatment

It has been proved that nanomedicines offer abundant benefits, such as improving solubility, biocompatibility, bioavailability, distribution, and stability, as well as reducing toxicity and MDR, which have shown superior efficacy than conventional therapeutics. However, it remains difficult to facilitate the application in clinic. Despite a few nanomedicines approved and used in solid tumors,¹³⁹⁻¹⁴² CPX-351 is the only liposomal formulation approved by FDA to date. There are some risks and limitations concerning therapeutic effects and safeties due to the intrinsic physicochemical properties of NPs. Sizes are concerned with circulation time and half-life in blood or organs. Previous studies have demonstrated that NPs <4 nm may penetrate vacuoles and interfere cellular processed, such as metabolism, detoxification, transcription, and gene expression, including normal cells.^{143,144} Particles with sizes ranging from 10 to 250 nm could stay in the liver, spleen, kidneys, or other organs for several months.^{145,146} Nanoparticles ranging from 15 to 20 nm could penetrate and infiltrate through the blood-brain barrier and blood-retinal barrier, implicating that toxicities should be inevitably taken into a consideration.¹⁴⁷ Pharmacokinetics, distributions, and toxicities can also be affected by NPs. Imputable to technology limitations, it is still difficult to evaluated efficacies in loading or release drug exactly. After internalized into cells, NPs often face

degradation due to endosomes.¹⁴⁸ In addition, there are off-target risks due to some of the NP–antibody ligands that are also target on normal cells.^{149,150} Additionally, NPs may active immune responses and be cleared by immune cells.¹⁵¹ Therefore, assessing the toxicity and side effects should be taken into account before approval and utilize in clinic. Besides the biological hurdles, there are also technical challenges. It is not an easy approach to fulfill the producing demand in a manufactory rather than in a laboratory, which requires special production equipment and high manufacturing costs. Moreover, each subtype of NPs offers unique features or characteristics, which means general rules could not be laid down and massive efforts have to be made for case-by-case evaluation. In vivo tests are essential for clinical translation. However, the development and progress of tumors in conventional xenograft models are not spontaneous or humanized during nanomedicines evaluation in animal models, which are different from live cases and deviated from realistic situations. Consequently, further evaluations are needed for nanomedicines transferring from bench to bedside in AML.

Conclusions

Acute myeloid leukemia has been considered an intricate hematological malignancy with poor responses and high mortalities. Chemotherapies, targeting therapies, and immunotherapies are the conventional treatment strategies for AML. This review summarized current development and approaches of nanomedicines in AML, including related technologic rationale, efficacies, safety profiles, and limitations, which may provide a novel and highlighted therapeutic option for AML treatment in future. Previous studies have demonstrated a superior for that NPs can be superior in improving bioavailability, reducing adverse effects, and circumventing drug resistances. Liposomal and polymeric NPs have shown especially promising prospects with advantages of strengthened loading ability and drug-release controlling. Chemical formulations and specific ligands such as antibodies or CPPs are also in underdevelopment. On the contrary, due to physical properties, drugs decorated with metal or silica NPs are less biocompatible and biodegradable, which restrict their utilization in AML. However, according to encouraging results in clinical trials, we could inference that NPs may finally become a promising and clinically acceptable option in the treatment of AML.

Author's Note

Feng Huang is also affiliated with Beijing Hospital of Traditional Chinese Medicine, Capital Medical University, Beijing, China.


Declaration of Conflicting Interests

The author(s) declared no potential conflicts of interest with respect to the research, authorship, and/or publication of this article.

Funding

The author(s) disclosed receipt of the following financial support for the research, authorship, and/or publication of this article: This study is supported by grants from Hong Kong Baptist University Special Development Fund (SDF18-0319-P03).

ORCID iD

Xiao Huang  <https://orcid.org/0000-0002-6144-2860>

References

1. Howlader N, Noone AM, Krapcho M, et al. (eds). *SEER Cancer Statistics Review, 1975–2016*. National Cancer Institute. Bethesda, MD. http://seer.cancer.gov/csr/1975_2014/. Accessed May 15, 2019.
2. Harris NL, Jaffe ES, Diebold J, et al. The World Health Organization Classification of neoplastic diseases of the haematopoietic and lymphoid tissues: report of the Clinical Advisory Committee Meeting, Airlie House, Virginia, November 1997. *Histopathology*. 2000;36(1):69-86.
3. Arber DA, Orazi A, Hasserjian R, et al. The 2016 revision to the World Health Organization classification of myeloid neoplasms and acute leukemia. *Blood*. 2016;127(20):2391-2405.
4. Vakiti A, Mewawalla P. Cancer, acute myeloid leukemia (AML, erythroid leukemia, myelodysplasia-related leukemia, BCR-ABL chronic leukemia). In: *StatPearls*. Treasure Island, FL: StatPearls Publishing; 2019.
5. Swords R, Santini V. In elderly patients with AML, which patients should be considered fit or unfit for standard induction therapy? *Hematology Am Soc Hematol Educ Program*. 2012; 2012:74-75.
6. Yanada M, Naoe T. Acute myeloid leukemia in older adults. *Int J Hematol*. 2012;96(2):186-193.
7. Jain S, Hirst DG, O'Sullivan JM. Gold nanoparticles as novel agents for cancer therapy. *Brit J Radiol*. 2012;85(1010):101-113.
8. Zhou C, Yang Z, Teng L. Nanomedicine based on nucleic acids: pharmacokinetic and pharmacodynamic perspectives. *Curr Pharm Biotechnol*. 2014;15(9):829-838.
9. Chen Y, Liu X, Yuan H, et al. Therapeutic remodeling of the tumor microenvironment enhances nanoparticle delivery. *Adv Sci*. 2019;6(5):1802070.
10. Briot T, Roger E, Thepot S, Lagarce F. Advances in treatment formulations for acute myeloid leukemia. *Drug Discov Today*. 2018;23(12):1936-1949.
11. Guo J, Luan X, Cong Z, et al. The potential for clinical translation of antibody-targeted nanoparticles in the treatment of acute myeloid leukaemia. *J Control Release*. 2018;286:154-166.
12. Yang Z, Yu B, Zhu J, et al. A microfluidic method to synthesize transferrin-lipid nanoparticles loaded with siRNA LOR-1284 for therapy of acute myeloid leukemia. *Nanoscale*. 2014;6(16): 9742-9751.
13. Kurrikoff K, Aphkhasava D, Langel U. The future of peptides in cancer treatment. *Curr Opin Pharmacol*. 2019;47:27-32.
14. Vadevoo SMP, Gurung S, Khan F, et al. Peptide-based targeted therapeutics and apoptosis imaging probes for cancer therapy. *Arch Pharm Res*. 2019;42(2):150-158.

15. Ferrara F, Vitagliano O. Induction therapy in acute myeloid leukemia: is it time to put aside standard 3 + 7? *Hematol Oncol*. 2019. doi:10.1002/hon.2615.
16. Lin M, Chen B. Advances in the drug therapies of acute myeloid leukemia (except acute wpromyelocytic leukemia). *Drug Des Develop Ther*. 2018;12:1009-1017.
17. Holowiecki J, Grosicki S, Giebel S, et al. Cladribine, but not fludarabine, added to daunorubicin and cytarabine during induction prolongs survival of patients with acute myeloid leukemia: a multicenter, randomized phase III study. *J Clin Oncol*. 2012; 30(20):2441-2448.
18. Short NJ, Rytting ME, Cortes JE. Acute myeloid leukaemia. *Lancet (Lond Engl)*. 2018;392(10147):593-606.
19. Fenaux P, Mufti GJ, Hellstrom-Lindberg E, et al. Azacitidine prolongs overall survival compared with conventional care regimens in elderly patients with low bone marrow blast count acute myeloid leukemia. *J Clin Oncol*. 2010;28(4):562-569.
20. Kayser S, Levis MJ. Advances in targeted therapy for acute myeloid leukaemia. *Br J Haematol*. 2018;180(4):484-500.
21. Stone RM, Larson RA, Dohner H. Midostaurin in FLT3-mutated acute myeloid leukemia. *N Engl J Med*. 2017;377(19):1903.
22. Ko YC, Hu CY, Liu ZH, et al. Cytarabine-resistant FLT3-ITD leukemia cells are associated with TP53 mutation and multiple pathway alterations-possible therapeutic efficacy of cabozantinib. *Int J Mole Sci*. 2019;20(5):E1230.
23. Zhou F, Ge Z, Chen B. Quizartinib (AC220): a promising option for acute myeloid leukemia. *Drug Des Devel Ther*. 2019;13: 1117-1125.
24. Lee LY, Hernandez D, Rajkhowa T, et al. Preclinical studies of gilteritinib, a next-generation FLT3 inhibitor. *Blood*. 2017; 129(2):257-260.
25. Bohl SR, Bullinger L, Rucker FG. New targeted agents in acute myeloid leukemia: new hope on the rise. *Int J Mol Sci*. 2019; 20(8):E1983.
26. DiNardo CD, Ravandi F, Agresta S, et al. Characteristics, clinical outcome, and prognostic significance of IDH mutations in AML. *Am J Hematol*. 2015;90(8):732-736.
27. DiNardo CD. Ivosidenib in IDH1-mutated acute myeloid leukemia. *N Engl J Med*. 2018;379(12):1186.
28. Abou Dalle I, DiNardo CD. The role of enasidenib in the treatment of mutant IDH2 acute myeloid leukemia. *Ther Adv Hematol*. 2018;9(7):163-173.
29. Adams JM, Cory S. Bcl-2-regulated apoptosis: mechanism and therapeutic potential. *Curr Opin Immunol*. 2007;19(5):488-496.
30. Norsworthy KJ, Ko CW, Lee JE, et al. FDA approval summary: Mylotarg for treatment of patients with relapsed or refractory CD33-positive acute myeloid leukemia. *Oncologist*. 2018;23(9): 1103-1108.
31. FDA approves gemtuzumab ozogamicin for CD33-positive AML. 2017; <https://www.fda.gov/drugs/resources-information-approve-d-drugs/fda-approves-gemtuzumab-ozogamicin-cd33-positive-aml>. Accessed May 20, 2018.
32. Gao Z, McAlister VC, Williams GM. Repopulation of liver endothelium by bone-marrow-derived cells. *Lancet (London, England)*. 2001;357(9260):932-933.
33. Thol F, Schlenk RF. Gemtuzumab ozogamicin in acute myeloid leukemia revisited. *Expert Opin Biol Ther*. 2014;14(8): 1185-1195.
34. Fathi AT, Erba HP, Lancet JE, et al. A phase 1 trial of vadastuximab talirine combined with hypomethylating agents in patients with CD33-positive AML. *Blood*. 2018;132(11):1125-1133.
35. Walter RB. Investigational CD33-targeted therapeutics for acute myeloid leukemia. *Expert Opin Investig Drugs*. 2018;27(4): 339-348.
36. Aigner M, Feulner J, Schaffer S, et al. T lymphocytes can be effectively recruited for ex vivo and in vivo lysis of AML blasts by a novel CD33/CD3-bispecific BiTE antibody construct. *Leukemia*. 2013;27(5):1107-1115.
37. Pemmaraju N, Lane AA, Sweet KL, et al. Results from phase 2 trial ongoing expansion stage of SL-401 in patients with blastic plasmacytoid dendritic cell neoplasm (BPDCN). *Blood*. 2016; 128:342-342.
38. Sweet KL, Pemmaraju N, Lane AA, et al. Lead-in stage results of a pivotal trial of SL-401, an interleukin-3 receptor (IL-3R) targeting biologic, in patients with blastic plasmacytoid dendritic cell neoplasm (BPDCN) or acute myeloid leukemia (AML). *Blood*. 2015;126(23):3795-3795.
39. Sharma P, Allison JP. Immune checkpoint targeting in cancer therapy: toward combination strategies with curative potential. *Cell*. 2015;161(2):205-214.
40. Fevery S, Billiau AD, Sprangers B, et al. CTLA-4 blockade in murine bone marrow chimeras induces a host-derived antileukemic effect without graft-versus-host disease. *Leukemia*. 2007; 21(7):1451-1459.
41. Zhang L, Gajewski TF, Kline J. PD-1/PD-L1 interactions inhibit antitumor immune responses in a murine acute myeloid leukemia model. *Blood*. 2009;114(8):1545-1552.
42. Zeidner JF, Vincent BG, Ivanova A, et al. Phase II study of high dose cytarabine followed by pembrolizumab in relapsed/refractory acute myeloid leukemia (AML). *Blood*. 2017;130: 1349-1349.
43. Daver N, Garcia-Manero G, Basu S, et al. Efficacy, safety, and biomarkers of response to azacitidine and nivolumab in relapsed/refractory acute myeloid leukemia: a non-randomized, open-label, phase 2 study. *Cancer Discov*. 2019;9(3):370-383.
44. Giannopoulos K. Targeting immune signaling checkpoints in acute myeloid leukemia. *J Clin Med*. 2019;8(2):E236.
45. ASH 2018: Azacitidine with nivolumab plus ipilimumab vs azacitidine plus nivolumab in relapsed or refractory AML. 2018; <https://www.ascopost.com/News/59523>. Accessed May 12, 2019.
46. Jetani H, Garcia-Cadenas I, Nerretter T, et al. CAR T-cells targeting FLT3 have potent activity against FLT3(-)ITD(+) AML and act synergistically with the FLT3-inhibitor crenolanib. *Leukemia*. 2018;32(5):1168-1179.
47. Sauer T, Rooney CM. Current challenges for CAR T-cell therapy of acute myeloid leukemia. *Transfusion*. 2019;59(4): 1171-1173.
48. Cummins KD, Gill S. Will CAR T cell therapy have a role in AML? Promises and pitfalls. *Semin Hematol*. 2019;56(2): 155-163.

49. Sutradhar KB, Amin ML. Nanotechnology in cancer drug delivery and selective targeting. *ISRN Nanotechnol.* 2014;2014. doi: 10.1155/2014/939378.
50. Dawar S, Singh N, Kanwar RK, et al. Multifunctional and multi-targeted nanoparticles for drug delivery to overcome barriers of drug resistance in human cancers. *Drug Discov Today.* 2013; 18(23-24):1292-1300.
51. Fu HB, Yao JN. Size effects on the optical properties of organic nanoparticles. *J Am Chem Soc.* 2001;123(7):1434-1439.
52. Scholes GD. Controlling the optical properties of inorganic nanoparticles. *Adv Funct Mater.* 2008;18(8):1157-1172.
53. Farokhzad OC, Langer R. Impact of nanotechnology on drug delivery. *ACS Nano.* 2009;3(1):16-20.
54. Yang X, Yang S, Chai H, et al. A novel isoquinoline derivative anticancer agent and its targeted delivery to tumor cells using transferrin-conjugated liposomes. *PLoS One.* 2015;10(8): e0136649.
55. Wang X, Huang X, Yang Z, et al. Targeted delivery of tumor suppressor microRNA-1 by transferrin-conjugated lipopolyplex nanoparticles to patient-derived glioblastoma stem cells. *Curr Pharm Biotechnol.* 2014;15(9):839-846.
56. Bobo D, Robinson KJ, Islam J, Thurecht KJ, Corrie SR. Nanoparticle-based medicines: a review of FDA-approved materials and clinical trials to date. *Pharm Res.* 2016;33(10): 2373-2387.
57. D'Mello SR, Cruz CN, Chen ML, Kapoor M, Lee SL, Tyner KM. The evolving landscape of drug products containing nanomaterials in the United States. *Nat Nanotechnol.* 2017;12(6):523-529.
58. Krauss AC, Gao X, Li L, et al. FDA approval summary: (daunorubicin and cytarabine) liposome for injection for the treatment of adults with high-risk acute myeloid leukemia. *Clin Cancer Res.* 2019;25(9):2685-2690.
59. Tardi P, Johnstone S, Harasym N, et al. In vivo maintenance of synergistic cytarabine: daunorubicin ratios greatly enhances therapeutic efficacy. *Leuk Res.* 2009;33(1):129-139.
60. Lim WS, Tardi PG, Dos Santos N, et al. Leukemia-selective uptake and cytotoxicity of CPX-351, a synergistic fixed-ratio cytarabine: daunorubicin formulation, in bone marrow xenografts. *Leuk Res.* 2010;34(9):1214-1223.
61. Lim WS, Tardi PG, Xie X, et al. Schedule- and dose-dependency of CPX-351, a synergistic fixed ratio cytarabine: daunorubicin formulation, in consolidation treatment against human leukemia xenografts. *Leuk Lymphoma.* 2010;51(8):1536-1542.
62. Kim HP, Gerhard B, Harasym TO, Mayer LD, Hogge DE. Liposomal encapsulation of a synergistic molar ratio of cytarabine and daunorubicin enhances selective toxicity for acute myeloid leukemia progenitors as compared to analogous normal hematopoietic cells. *Exp Hematol.* 2011;39(7):741-750.
63. Feldman EJ, Lancet JE, Kolitz JE, et al. First-in-man study of CPX-351: a liposomal carrier containing cytarabine and daunorubicin in a fixed 5:1 molar ratio for the treatment of relapsed and refractory acute myeloid leukemia. *J Clin Oncol.* 2011;29(8): 979-985.
64. Feldman EJ, Kolitz JE, Trang JM, et al. Pharmacokinetics of CPX-351; a nano-scale liposomal fixed molar ratio formulation of cytarabine:daunorubicin, in patients with advanced leukemia. *Leuk Res.* 2012;36(10):1283-1289.
65. Lancet JE, Cortes JE, Hogge DE, et al. Phase 2 trial of CPX-351, a fixed 5:1 molar ratio of cytarabine/daunorubicin, vs cytarabine/daunorubicin in older adults with untreated AML. *Blood.* 2014; 123(21):3239-3246.
66. Cortes JE, Goldberg SL, Feldman EJ, et al. Phase II, multicenter, randomized trial of CPX-351 (cytarabine: daunorubicin) liposome injection versus intensive salvage therapy in adults with first relapse AML. *Cancer.* 2015;121(2):234-242.
67. Lancet JE, Uy GL, Cortes JE, et al. Final results of a phase III randomized trial of CPX-351 versus 7+3 in older patients with newly diagnosed high risk (secondary) AML. *Am Soc Clin Oncol.* 2016. doi:10.1200/jco.2016.34.15_suppl.7000.
68. Lancet JE, Uy GL, Cortes JE, et al. CPX-351 (cytarabine and daunorubicin) liposome for injection versus conventional cytarabine plus daunorubicin in older patients with newly diagnosed secondary acute myeloid leukemia. *J Clin Oncol.* 2018;36(26): 2684-2692.
69. Gabizon AA, Patil Y, La-Beck NM. New insights and evolving role of pegylated liposomal doxorubicin in cancer therapy. *Drug Resist Update.* 2016;29:90-106.
70. Franco YL, Vaidya TR, Ait-Oudhia S. Anticancer and cardioprotective effects of liposomal doxorubicin in the treatment of breast cancer. *Breast Cancer (Dove Medical Press).* 2018;10: 131-141.
71. Huang YF, Kuo MT, Liu YS, Cheng YM, Wu PY, Chou CY. A dose escalation study of trientine plus carboplatin and pegylated liposomal doxorubicin in women with a first relapse of epithelial ovarian, tubal, and peritoneal cancer within 12 months after platinum-based chemotherapy. *Front Oncol.* 2019;9:437.
72. Melillo L, Valente D, D'Arena G, et al. Combination treatment of flag with non-pegylated liposomal doxorubicin (MYOCET(TM)) in elderly patients with acute myeloid leukemia: a single center experience. *Int J Immunopathol Pharmacol.* 2011;24(3):703-709.
73. Quarello P, Berger M, Rivetti E, et al. FLAG-liposomal doxorubicin (Myocet) regimen for refractory or relapsed acute leukemia pediatric patients. *J Pediatr Hematol Oncol.* 2012;34(3):208-216.
74. Kaspers GJ, Zimmermann M, Reinhardt D, et al. Improved outcome in pediatric relapsed acute myeloid leukemia: results of a randomized trial on liposomal daunorubicin by the international BFM study group. *J Clin Oncol.* 2013;31(5):599-607.
75. Chi X, Gatti P, Papoian T. Safety of antisense oligonucleotide and siRNA-based therapeutics. *Drug Discov Today.* 2017;22(5): 823-833.
76. Ohanian M, Tari Ashizawa A, Garcia-Manero G, et al. Liposomal Grb2 antisense oligodeoxynucleotide (BP1001) in patients with refractory or relapsed haematological malignancies: a single-centre, open-label, dose-escalation, phase 1/1b trial. *Lancet Haematol.* 2018;5(4):e136-e146.
77. Tan KB, Ling LU, Bunte RM, Chng WJ, Chiu GN. In vivo efficacy of a novel liposomal formulation of safinol in the treatment of acute myeloid leukemia. *J Control Release.* 2012;160(2): 290-298.
78. Tan KB, Ling LU, Bunte RM, Chng WJ, Chiu GN. Liposomal codelivery of a synergistic combination of bioactive lipids in the

- treatment of acute myeloid leukemia. *Nanomedicine (Lond)*. 2014;9(11):1665-1679.
79. Myhren L, Nilssen IM, Nicolas V, Doskeland SO, Barratt G, Herfindal L. Efficacy of multi-functional liposomes containing daunorubicin and emetine for treatment of acute myeloid leukemia. *Eur J Pharm Biopharm*. 2014;88(1):186-193.
 80. Pollyea DA, Jordan CT. Therapeutic targeting of acute myeloid leukemia stem cells. *Blood*. 2017;129(12):1627-1635.
 81. Dorrance AM, Neviani P, Ferencak GJ, et al. Targeting leukemia stem cells in vivo with antagomiR-126 nanoparticles in acute myeloid leukemia. *Leukemia*. 2015;29(11):2143-2153.
 82. Li H, Xu S, Quan J, et al. CD33-targeted lipid nanoparticles (aCD33LNs) for therapeutic delivery of GTI-2040 to acute myelogenous leukemia. *Mol Pharm*. 2015;12(6):2010-2018.
 83. Drummond DC, Noble CO, Hayes ME, Park JW, Kirpotin DB. Pharmacokinetics and in vivo drug release rates in liposomal nanocarrier development. *J Pharm Sci*. 2008;97(11):4696-4740.
 84. Sarin H, Kanevsky AS, Wu H, et al. Physiologic upper limit of pore size in the blood-tumor barrier of malignant solid tumors. *J Transl Med*. 2009;7:51.
 85. Zamboni WC, Torchilin V, Patri AK, et al. Best practices in cancer nanotechnology: perspective from NCI nanotechnology alliance. *Clin Cancer Res*. 2012;18(12):3229-3241.
 86. Wang Y, Wu H, Wang Z, et al. Optimized synthesis of biodegradable elastomer pegylated poly(glycerol sebacate) and their biomedical application. *Polymers*. 2019;11(6):E965.
 87. Hao F, Li Y, Zhu J, et al. Polyethylenimine-based formulations for delivery of oligonucleotides. *Curr Med Chem*. 2019;26(13):2264-2284.
 88. Pan S, Xing H, Fu X, et al. The effect of photothermal therapy on osteosarcoma with polyacrylic acid-coated gold nanorods. *Dose-Response*. 2018;16(3).doi:10.1177/1559325818789841.
 89. Sha L, Chen Z, Chen Z, Zhang A, Yang Z. Polylactic acid based nanocomposites: promising safe and biodegradable materials in biomedical field. *Int J Polym Sci*. 2016;2016:1-11.
 90. Xie J, Teng L, Yang Z, Zhou C, Liu Y, Yung BC, Lee RJ. A polyethylenimine-linoleic acid conjugate for antisense oligonucleotide delivery. *Biomed Res Int*. 2013;2013. doi:10.1155/2013/710502.
 91. Sun J, Shen J, Chen S, et al. Nanofiller reinforced biodegradable PLA/PHA composites: current status and future trends. *Polymers*. 2018;10(5):E505.
 92. Kang C, Sun Y, Zhu J, et al. Delivery of nanoparticles for treatment of brain tumor. *Curr Drug Metab*. 2016;17(8):745-754.
 93. Floc'h N, Ashton S, Taylor P, et al. Optimizing therapeutic effect of aurora B inhibition in acute myeloid leukemia with AZD2811 nanoparticles. *Mol Cancer Ther*. 2017;16(6):1031-1040.
 94. Simon AM, Jagadeeshan S, Abraham E, et al. Poly (D, L-lactico-glycolide) nanoparticles for the improved therapeutic efficacy of all-trans-retinoic acid: a study of acute myeloid leukemia (AML) cell differentiation in vitro. *Med Chem*. 2012;8(5):805-810.
 95. Sun Y, Sun ZL. Transferrin-conjugated polymeric nanomedicine to enhance the anticancer efficacy of edelfosine in acute myeloid leukemia. *Biomed Pharmacother*. 2016;83:51-57.
 96. Zhu B, Zhang H, Yu L. Novel transferrin modified and doxorubicin loaded pluronic 85/lipid-polymeric nanoparticles for the treatment of leukemia: in vitro and in vivo therapeutic effect evaluation. *Biomed Pharmacother*. 2017;86:547-554.
 97. Gul-Uludag H, Valencia-Serna J, Kucharski C, et al. Polymeric nanoparticle-mediated silencing of CD44 receptor in CD34⁺ acute myeloid leukemia cells. *Leuk Res*. 2014;38(11):1299-1308.
 98. Chandran P, Gupta N, Retnakumari AP, et al. Simultaneous inhibition of aberrant cancer kinome using rationally designed polymer-protein core-shell nanomedicine. *Nanomedicine*. 2013;9(8):1317-1327.
 99. Liang B, Li N, Zhang S, et al. Idarubicin-loaded methoxy poly (ethylene glycol)-b-poly(L-lactide-co-glycolide) nanoparticles for enhancing cellular uptake and promoting antileukemia activity. *Int J Nanomed*. 2019;14:543-556.
 100. Wang Y, Xie Y, Williams J, et al. Use of polymeric CXCR4 inhibitors as siRNA delivery vehicles for the treatment of acute myeloid leukemia. *Cancer Gene Ther*. 2019.
 101. Gui R, Wan A, Liu X, Jin H. Intracellular fluorescent thermometry and photothermal-triggered drug release developed from gold nanoclusters and doxorubicin dual-loaded liposomes. *Chem Commun*. 2014;50(13):1546-1548.
 102. Ventola CL. Progress in nanomedicine: approved and investigational nanodrugs. *P T*. 2017;42(12):742-755.
 103. Do VQ, Park KH, Park JM, Lee MY. Comparative in vitro toxicity study of docetaxel and nanoxel, a docetaxel-loaded micellar formulation using cultured and blood cells. *Toxicol Res*. 2019;35(2):201-207.
 104. Bernabeu E, Cagel M, Lagomarsino E, Moreton M, Chiappetta DA. Paclitaxel: what has been done and the challenges remain ahead. *Int J Pharm*. 2017;526(1-2):474-495.
 105. Varshosaz J, Hassanzadeh F, Sadeghi Aliabadi H, Nayebsadrian M, Banitalebi M, Rostami M. Synthesis and characterization of folate-targeted dextran/retinoic acid micelles for doxorubicin delivery in acute leukemia. *Biomed Res Int*. 2014;2014:525684.
 106. Alakhova DY, Zhao Y, Li S, Kabanov AV. Effect of doxorubicin/pluronic SP1049C on tumorigenicity, aggressiveness, DNA methylation and stem cell markers in murine leukemia. *PLoS One*. 2013;8(8): e72238.
 107. Lim J, Simanek EE. Triazine dendrimers as drug delivery systems: from synthesis to therapy. *Adv Drug Deliv Rev*. 2012;64(9):826-835.
 108. Szulc A, Pulaski L, Appelhans D, Voit B, Klajnert-Maculewicz B. Sugar-modified poly(propylene imine) dendrimers as drug delivery agents for cytarabine to overcome drug resistance. *Int J Pharm*. 2016;513(1-2):572-583.
 109. Sharma H, Mishra PK, Talegaonkar S, Vaidya B. Metal nanoparticles: a theranostic nanotool against cancer. *Drug Discov Today*. 2015;20(9):1143-1151.
 110. Petrushev B, Boca S, Simon T, et al. Gold nanoparticles enhance the effect of tyrosine kinase inhibitors in acute myeloid leukemia therapy. *Int J Nanomed*. 2016;11:641-660.
 111. Shen N, Yan F, Pang J, et al. HDL-AuNPs-BMS nanoparticle conjugates as molecularly targeted therapy for leukemia. *ACS Appl Mater Interfaces*. 2018;10(17):14454-14462.

112. Huynh NT, Passirani C, Saulnier P, Benoit JP. Lipid nanocapsules: a new platform for nanomedicine. *Int J Pharm.* 2009; 379(2):201-209.
113. Khajavinia A, Varshosaz J, Dehkordi AJ. Targeting etoposide to acute myelogenous leukaemia cells using nanostructured lipid carriers coated with transferrin. *Nanotechnology.* 2012;23(40): 405101.
114. Silva EL, Lima FA, Carneiro G, et al. Improved in vitro antileukemic activity of all-trans retinoic acid loaded in cholesteryl butyrate solid lipid nanoparticles. *J Nanosci Nanotechnol.* 2016; 16(2):1291-1300.
115. Enjeti AK, D'Crus A, Melville K, Verrills NM, Rowlings P. A systematic evaluation of the safety and toxicity of fingolimod for its potential use in the treatment of acute myeloid leukaemia. *Anti-Cancer Drugs.* 2016;27(6):560-568.
116. Estella-Hermoso de Mendoza A, Castello-Cros R, Imbuluzqueta E et al. Lipid nanosystems enhance the bioavailability and the therapeutic efficacy of FTY720 in acute myeloid leukemia. *J Biomed Nanotechnol.* 2015;11(4):691-701.
117. Chandran P, Kavalakatt A, Malarvizhi GL, et al. Epigenetics targeted protein-vorinostat nanomedicine inducing apoptosis in heterogeneous population of primary acute myeloid leukemia cells including refractory and relapsed cases. *Nanomedicine.* 2014;10(4):721-732.
118. Narmani A, Rezvani M, Farhood B, et al. Folic acid functionalized nanoparticles as pharmaceutical carriers in drug delivery systems. *Drug Dev Res.* 2019;80(4):404-424.
119. Kalmouni M, Al-Hosani S, Magzoub M. Cancer targeting peptides. *Cell Mol Life Sci.* 2019;76(11):2171-2183.
120. Carter T, Mulholland P, Chester K. Antibody-targeted nanoparticles for cancer treatment. *Immunotherapy.* 2016;8(8):941-958.
121. Ngo B, Van Riper JM, Cantley LC, Yun J. Targeting cancer vulnerabilities with high-dose vitamin C. *Nat Rev Cancer.* 2019;19(5):271-282.
122. Choudhury H, Pandey M, Chin PX, et al. Transferrin receptors-targeting nanocarriers for efficient targeted delivery and transcytosis of drugs into the brain tumors: a review of recent advancements and emerging trends. *Drug Deliv Transl Res.* 2018;8(5):1545-1563.
123. Tortorella S, Karagiannis TC. Transferrin receptor-mediated endocytosis: a useful target for cancer therapy. *J Membr Biol.* 2014;247(4):291-307.
124. Ye J, Liu E, Yu Z, et al. CPP-assisted intracellular drug delivery, what is next? *Int J Mol Sci.* 2016;17(11).
125. Regberg J, Srimanee A, Langel U. Applications of cell-penetrating peptides for tumor targeting and future cancer therapies. *Pharmaceuticals (Basel).* 2012;5(9):991-1007.
126. Chen Y, Xie X, Wu A, et al. A synthetic cell-penetrating peptide derived from nuclear localization signal of EPS8 exerts anticancer activity against acute myeloid leukemia. *J Exp Clin Cancer Res.* 2018;37(1):12.
127. Agarwal A, MacKenzie RJ, Pippa R, et al. Antagonism of SET using OP449 enhances the efficacy of tyrosine kinase inhibitors and overcomes drug resistance in myeloid leukemia. *Clin Cancer Res.* 2014;20(8):2092-2103.
128. Yin L, Wu Z, Avigan D, et al. MUC1-C oncoprotein suppresses reactive oxygen species-induced terminal differentiation of acute myelogenous leukemia cells. *Blood.* 2011;117(18): 4863-4870.
129. Liu S, Yin L, Stroopinsky D, et al. MUC1-C oncoprotein promotes FLT3 receptor activation in acute myeloid leukemia cells. *Blood.* 2014;123(5):734-742.
130. Li K, Lv XX, Hua F, et al. Targeting acute myeloid leukemia with a proapoptotic peptide conjugated to a Toll-like receptor 2-mediated cell-penetrating peptide. *Int J Cancer.* 2014;134(3): 692-702.
131. Hughes B. Antibody-drug conjugates for cancer: poised to deliver? *Nat Rev Drug Discov.* 2010;9:665.
132. Simard P, Leroux JC. PH-sensitive immunoliposomes specific to the CD33 cell surface antigen of leukemic cells. *Int J Pharm.* 2009;381(2):86-96.
133. Al-Hussaini M, Rettig MP, Ritchey JK, et al. Targeting CD123 in acute myeloid leukemia using a T-cell-directed dual-affinity retargeting platform. *Blood.* 2016;127(1):122-131.
134. Wang Y, Liu F, Wang Q, et al. A novel immunoliposome mediated by CD123 antibody targeting to acute myeloid leukemia cells. *Int J Pharm.* 2017;529(1-2):531-542.
135. Liu FR, Jin H, Wang Y, et al. Anti-CD123 antibody-modified niosomes for targeted delivery of daunorubicin against acute myeloid leukemia. *Drug Deliv.* 2017;24(1):882-890.
136. Deshpande AJ, Buske C. Lymphoid progenitors as candidate cancer stem cells in AML: new perspectives. *Cell Cycle (Georgetown, Tex).* 2007;6(5):543-545.
137. Mandal T, Beck M, Kirsten N, Linden M, Buske C. Targeting murine leukemic stem cells by antibody functionalized mesoporous silica nanoparticles. *Sci Rep.* 2018;8(1):989.
138. Daniels TR, Delgado T, Rodriguez JA, Helguera G, Penichet ML. The transferrin receptor part I: biology and targeting with cytotoxic antibodies for the treatment of cancer. *Clin Immunol.* 2006;121(2):144-158.
139. Al-Hatamleh MAI, Ahmad S, Boer JC, et al. A perspective review on the role of nanomedicine in the modulation of TNF-TNFR2 axis in breast cancer immunotherapy. *J Oncol.* 2019. doi:10.1155/2019/6313242.
140. Gudbergsson JM, Jonsson K, Simonsen JB, Johnsen KB. Systematic review of targeted extracellular vesicles for drug delivery—considerations on methodological and biological heterogeneity. *J Control Release.* 2019;306:108-120.
141. Liu JF, Jang B, Issadore D, Tsourkas A. Use of magnetic fields and nanoparticles to trigger drug release and improve tumor targeting. *Wiley Interdiscip Rev Nanomed Nanobiotechnol.* 2019;11(6):e1571. doi:10.1002/wnan.1571.
142. Xiong Y, Wang Y, Tiruthani K. Tumor immune microenvironment and nano-immunotherapeutics in colorectal cancer. *Nanomedicine.* 2019;21:102034. doi:10.1016/j.nano.2019. 102034.
143. Liu Y, Meyer Zaika W, Franzka S, Schmid G, Tsoli M, Kuhn H. Gold-cluster degradation by the transition of B-DNA into A-DNA and the formation of nanowires. *Angew Chem Int Ed Engl.* 2003;42(25):2853-2857.

144. Semmler Behnke M, Kreyling WG, Lipka J, et al. Biodistribution of 1.4- and 18-nm gold particles in rats. *Small*. 2008;4(12):2108-2111.
145. De Jong WH, Hagens WI, Krystek P, Burger MC, Sips AJ, Geertsma RE. Particle size-dependent organ distribution of gold nanoparticles after intravenous administration. *Biomaterials*. 2008;29(12):1912-1919.
146. Zhang G, Yang Z, Lu W, et al. Influence of anchoring ligands and particle size on the colloidal stability and in vivo biodistribution of polyethylene glycol-coated gold nanoparticles in tumor-xenografted mice. *Biomaterials*. 2009;30(10):1928-1936.
147. Kim JH, Kim JH, Kim KW, Kim MH, Yu YS. Intravenously administered gold nanoparticles pass through the blood-retinal barrier depending on the particle size, and induce no retinal toxicity. *Nanotechnology*. 2009;20(50):505101.
148. Sahay G, Alakhova DY, Kabanov AV. Endocytosis of nanomedicines. *J Control Release*. 2010;145(3):182-195.
149. Konopleva M, Pollyea DA, Potluri J, et al. Efficacy and biological correlates of response in a phase II study of venetoclax monotherapy in patients with acute myelogenous leukemia. *Cancer Discov*. 2016;6(10):1106-1117.
150. Truong NP, Whittaker MR, Mak CW, Davis TP. The importance of nanoparticle shape in cancer drug delivery. *Expert Opin Drug Deliv*. 2015;12(1):129-142.
151. Butcher NJ, Mortimer GM, Minchin RF. Drug delivery: unraveling the stealth effect. *Nat Nanotechnol*. 2016;11(4):310.