



The Role of Microglia in the Spread of Tau: Relevance for Tauopathies

Juan R. Perea^{1,2†}, María Llorens-Martín^{1,3†}, Jesús Ávila^{1,2} and Marta Bolós^{1,2*}

¹Department of Molecular Neuropathology, Centro de Biología Molecular "Severo Ochoa", CBMSO, CSIC, Madrid, Spain, ²Network Center for Biomedical Research on Neurodegenerative Diseases (CIBERNED), Madrid, Spain, ³Department of Molecular Biology, Faculty of Sciences, Universidad Autónoma de Madrid, Madrid, Spain

Tauopathies are neurodegenerative diseases which course with the accumulation of Tau, mainly in neurons. In addition, Tau accumulates in a hyperphosphorylated and aggregated form. This protein is released into the extracellular space and spreads following a stereotypical pattern, inducing the development of the disease through connected regions of the brain. Microglia—the macrophages of the brain—are involved in maintaining brain homeostasis. They perform a variety of functions related to the surveillance and clearance of pathological proteins, among other dead cells and debris, from the extracellular space that could compromise brain equilibrium. This review focuses on the role played by microglia in tauopathies, specifically in Alzheimer's disease (AD), and how the uncoupling of activation/phagocytosis functions can have fatal consequences leading to the development of the pathology.

OPEN ACCESS

Edited by:

Rocío Martínez De Pablos, Universidad de Sevilla, Spain

Reviewed by:

Nicolas Sergeant, Institut National de la Santé et de la Recherche Médicale (INSERM), France Fouad Janat, Independent Researcher, Westerly, RI, United States Victoria Campos-Peña, Instituto Nacional de Neurología y Neurocirugía (INNN), Mexico

***Correspondence:** Marta Bolós

mbolos@cbm.csic.es

[†]These authors have contributed equally to this work.

Received: 09 April 2018 **Accepted:** 01 June 2018 **Published:** 10 July 2018

Citation:

Perea JR, Llorens-Martín M, Ávila J and Bolós M (2018) The Role of Microglia in the Spread of Tau: Relevance for Tauopathies. Front. Cell. Neurosci. 12:172. doi: 10.3389/fncel.2018.00172 Keywords: microglia, tauopathies, Alzheimer's disease, tau protein, phagocytosis

INTRODUCTION

Microglia are the macrophages of the brain that are in charge of maintaining central nervous system (CNS) homeostasis. These cells are constantly surveying their microenvironment and, in response to small variations, they rapidly change their morphology, become activated, secrete proand anti-inflammatory molecules, and phagocytose foreign material and other potentially toxic elements that can alter the equilibrium of the brain (Ransohoff and El Khoury, 2016; Wolf et al., 2017).

Some neurodegenerative diseases known as tauopathies, such as Alzheimer's disease (AD), course with the abnormal intracellular accumulation of pathological Tau species (Ballatore et al., 2007; Goedert and Spillantini, 2011; Arendt et al., 2016). This toxic Tau is released outside the cells (Simón et al., 2012; Pooler et al., 2013), and in the extracellular space, it can interact with other cells (Gómez-Ramos et al., 2006), be internalized (Wu et al., 2013; Bolós et al., 2015), and trigger cell to cell Tau spread (Medina and Avila, 2014a; Calafate et al., 2015). Therefore, in order to protect healthy cells, microglia should activate mechanisms to remove this extracellular pathogenic protein and, by doing so, halt the progression of the disease. However, in some pathological situations, the uncoupling of the activation/phagocytosis functions of microglia can accelerate the progression of neurodegeneration (Sierra et al., 2013; Bolós et al., 2017a; Leyns and Holtzman, 2017).

PHYSIOLOGICAL AND PATHOLOGICAL TAU SPECIES

First discovered by Weingarten et al. (1975), Tau is a microtubule-associated protein present in the CNS. The human Tau gene, *Mapt*, is situated on chromosome 17q21 and comprises 16 exons (Pittman et al., 2006). Some of these exons in the CNS, namely 1, 4, 5, 7, 9, 11, 12 and 13, are

1

constitutive, while the others, 2, 3 and 10, are subjected to alternative splicing (Andreadis, 2005). The latter process generates six isoforms of Tau in the adult human brain that differ in the number of N-terminal insertions, tubulin binding domains and microtubule-binding repeats (Goedert and Spillantini, 2011). Alternative splicing of exon 10 is highly regulated through a complex mechanism. The deregulation of this process contributes to neurodegeneration (Liu and Gong, 2008).

Tau is largely found in axons (Aronov et al., 2001). It binds to microtubules and regulates the dynamics of these structures, thus allowing the reorganization of the cytoskeleton and axonal transport. However, Tau is also localized to the somatodendritic compartment, where it plays key roles at synapses (Drubin and Kirschner, 1986; Ittner and Götz, 2011). The expression of Tau protein, as well as its function, are complexly regulated by alternative splicing and several post-transcriptional modifications, including phosphorylation, glycosylation, acetylation and truncation, among others (Morris et al., 2015; Avila et al., 2016).

Tauopathies are characterized by the accumulation of aggregated and/or hyperphosphorylated Tau (Ballatore et al., 2007; Holtzman et al., 2011; Medina et al., 2016). Tauopathies are a group of neurological disorders that include corticobasal degeneration (CBD), frontotemporal dementia (FTD) and Parkinsonism linked to chromosome 17 (FTDP-17), progressive supranuclear palsy (PSP), Pick's disease (PiD), chronic traumatic encephalopathy (CTE), argyrophilic grain disease (AGD) and AD, the latter being the most common tauopathy (Spillantini and Goedert, 2013; Arendt et al., 2016). In tauopathies, Tau which is a highly soluble hydrophilic protein, detaches from microtubules and accumulates, forming intracellular hyperphosphorylated aggregates or inclusions, such as the neurofibrillary tangles (NFTs) found in AD brains (Brion et al., 1985; Wood et al., 1986; Wischik et al., 1988; Zhang et al., 2009). These structures damage cell function and produce neuronal cell death and neurodegeneration.

SPREAD OF TAU

Current hypotheses postulate that Tau is released from its intracellular localization into the extracellular space, from where it spreads following a stereotypic pattern, from the transentorhinal cortex to the hippocampus and, finally, to the neocortex (de Calignon et al., 2012; Holmes and Diamond, 2014; Hyman, 2014; Brettschneider et al., 2015; Hu et al., 2016). The spread of Tau through connected areas of the brain contributes to cognitive decline in age-associated tauopathy. It has been proposed that Tau is released by dying neurons, and, in addition, that it can be secreted physiologically (Pooler et al., 2013; Yamada et al., 2014; Kanmert et al., 2015; Pérez et al., 2016). Indeed, its physiological release was postulated as a mechanism to prevent the toxicity caused by elevated intracellular Tau (Karch et al., 2012; Simón et al., 2012; Pooler et al., 2013).

Extracellular Tau can interact with neighboring cells, such as neurons or glia (Gómez-Ramos et al., 2006; Bolós et al., 2015; Luo et al., 2015), and cause severe damage (Bolós et al., 2017b). The spread of Tau from adjacent cells has been proposed to be one of the mechanisms underlying the progression of tauopathies (Medina and Avila, 2014b; Mirbaha et al., 2015). Tau propagation between synaptically connected neurons have been reported by many groups (de Calignon et al., 2012; Yamada et al., 2014; Brettschneider et al., 2015); however, other spread mechanisms may occur. In this regard, we and others have recently demonstrated that extracellular soluble Tau, which encompasses small oligomers of Tau, interacts with microglia and is internalized by these cells (Luo et al., 2015; Sanchez-Mejias et al., 2016; Bolós et al., 2017a). Therefore, the dysfunction of microglia can also contribute to the spread of Tau (Maphis et al., 2015). Remarkably, augmented levels of exosome-associated Tau have been reported in the cerebrospinal fluid and blood of individuals with AD and FTD (Saman et al., 2012; Fiandaca et al., 2015). These findings suggest that the exosomal process is involved in the cell to cell spread of Tau (Aguzzi and Rajendran, 2009; Rajendran et al., 2014; Polanco et al., 2016). Furthermore, the inhibition of exosome biosynthesis and depletion of microglia halt the spread of this protein in a mouse model of tauopathy (Asai et al., 2015). These studies highlight the involvement of microglia, through phagocytosis and the release of Tau-containing exosomes, in the propagation of Tau pathology.

MICROGLIA-KEY CELLS MAINTAINING BRAIN HOMEOSTASIS

Microglia are the resident immune cells of the CNS and represent approximately 15% of the cells in this area (Lawson et al., 1990; Perry, 1998; Alliot et al., 1999). They were termed by Pío del Río Hortega. He hypothesized that microglia have the ability to rapidly change their morphology, proliferate and migrate in response to changes in the microenvironment. He proposed that the basic function of these cells was phagocytosis. During early embryonic stages, microglia originate from myeloid progenitors in the yolk sac that migrate into the brain before the blood-brain barrier is formed (Alliot et al., 1999; Ginhoux et al., 2010). Under physiological conditions, microglia proliferate and self-renew constantly without a contribution from bone marrow-derived macrophages in order to maintain cell numbers. In contrast, in the presence of a disease, circulating peripheral monocytes contribute to the microglia population maintenance (Ginhoux et al., 2010; Bruttger et al., 2015).

Microglia are highly dynamic cells that regulate several processes during both development and adulthood in the CNS (Ransohoff and El Khoury, 2016). They perform a wide variety of functions, including the following: constant surveillance; removal of pathogens; phagocytosis of apoptotic cells and cellular debris; secretion of growth factors, pro-inflammatory and anti-inflammatory signals; synapse remodeling and elimination, among others (Sierra et al., 2010, 2014; Paolicelli et al., 2014; Michell-Robinson et al., 2015; Wolf et al., 2017). The main distinctive of microglia is their capability to quickly alter their morphology and function in response to variations in their microenvironment (Town et al., 2005; Karperien et al., 2013). In this regard, there are two main types of microglia, namely



FIGURE 1 | Microglia surrounding phospho-Tau in human brain tissue. Separate z-projection images of a z-stack image showing phospho-Tau surrounded by microglia published by our group previously (Bolós et al., 2015). The authors have the appropriate permissions from the copyright holders. Images show microglia in white (Iba-1), phospho-Tau in red (S396) and nuclei in blue (DAPI).

resting (ramified) and activated (ameboid; Town et al., 2005). Many studies have tried to explain the correlation between these two types of microglia and their roles in physiological and pathological conditions, but the exact function of the different morphological states remains unknown. Activated microglia are often classified into inflammatory (M1) and alternatively activated (M2) phenotypes (Boche et al., 2013; Tang and Le, 2016). M1 microglia produce and release pro-inflammatory cytokines, such as tumor necrosis factor (TNF)- α , interleukin (IL)-6, IL-12, IL-1 β , IL-23, nitric oxide (NO), among others (Hanisch, 2002). In addition, it has been described that M1 microglia predominate at the site of injury (Loane and Byrnes, 2010). In contrast, M2 microglia express extracellular matrix molecules and anti-inflammatory molecules, such as

transforming growth factor (TGF)- β and IL-10, and express phagocytic activity. M2 microglia act later, at a period more associated to repair processes (Michell-Robinson et al., 2015). However, this M1 and M2 classification is oversimplified since mixed activation phenotypes can co-occur (Ransohoff, 2016a).

Surveillance microglia participate in CNS homeostasis by actively contacting surrounding cells (Nimmerjahn et al., 2005). In this regard, soluble and membrane-bound cytokines mediate neuron-microglia communication. This cross-talk manages the balance between the neuroprotective and harmful actions of microglia on neurons (Sierra et al., 2014; Wolf et al., 2017). One example is how neurons modulate microglial function through the CX3CL1/CX3CR1 axis (Harrison et al., 1998; Biber et al., 2007). As CX3CL1 is produced almost exclusively by neurons



(Kim et al., 2011) and the receptor CX3CR1 is expressed almost exclusively on the surface of microglia (Zhan et al., 2014), this axis is a sensor of the neuronal signals. It has been recently shown that suitable cross-talk between microglia and neurons is essential for the accurate functional maturation of newborn granule neurons in the hippocampus and has an important role in the regulation of emotional behavior (Bolós et al., 2018). In this regard, it has been demonstrated that Tau binds to CX3CR1 increasing its own internalization by microglia, and that Tau competes with CX3CL1 to bind to this receptor (Bolós et al., 2017a). In addition, it has been shown that there is less internalization of phospho-Tau by microglial CX3CR1 than for the non-phosphorylated form of the protein. These observations thus suggest that phospho-Tau is internalized through other mechanisms. Furthermore, an increase in the expression of the CX3CL1/CX3CR1 axis and a greater number of activated microglia in the hippocampus of patients with an advanced stage of AD has been shown. However, the higher amount of phospho-Tau observed in brain tissue of AD patients should not be automatically interpreted to be the result of increased Tau phagocytosis, since this process may be compromised at some point of the disease development. Actually, a reduction in the phagocytic activity of microglia was proposed since a decrease in the number of phagocytic pouches in these cells was observed. These data support the idea that the disconnection of microglial activation and phagocytosis happens at advanced stages of AD.

Other examples of dynamic communication between microglia and neurons that could be relevant in AD include the following neuron/microglia cross-talk: CD200/CD200R; ATP/P2Y; P2X; and CD22/CD45 (Mott et al., 2004; Minas and Liversidge, 2006; Surprenant and North, 2009; Verkhratsky et al., 2009; Eyo and Wu, 2013); among others.

In addition to performing active surveillance, microglia are tissue-resident macrophages, meaning that they are the phagocytic cells of the brain. Their function encompasses the clearance of cellular material and debris in order to maintain brain homeostasis. Phagocytosis is modulated in terms of activation or inhibition through various mechanisms (Parnaik et al., 2000; Peri and Nüsslein-Volhard, 2008; Sierra et al., 2010; Paolicelli et al., 2011; Schafer et al., 2012). One such mechanism is exerted by Toll-like receptors (TLRs), which recognize pathogens and induce a phagocytic response by microglia (Ribes et al., 2009). In addition, apoptotic neurons are phagocytosed after been recognized by various receptor systems like those described above. Other multiple factors regulate phagocytosis, such as the ciliary neurotrophic factor, CNTF, glia-derived neurotrophic factor, GDNF and macrophage colony-stimulating factor, M-CSF, the latter potentiating the phagocytic capacity of microglia (Mitrasinovic and Murphy, 2003; Chang et al., 2006; Lee et al., 2009). Substrate-bound complement component C1q enhances both FcR- and CR1-mediated phagocytosis (Webster et al., 2000), whereas the prostanoid receptor subtype 2, EP2,

downregulates phagocytosis (Shie et al., 2005). Therefore, the dysregulation of microglial phagocytosis mechanisms can impair the clearance of debris, thus inducing or exacerbating the inflammatory response and causing brain diseases such as AD.

MICROGLIA DYSFUNCTION CONTRIBUTES TO NEURODEGENERATION

Genome-wide association studies (GWAS) have recently identified several risk genes for AD. Specifically, the triggering receptor expressed on myeloid cells 2, TREM2 and cluster of differentiation 33 (CD33) can lead to a reduction in the activity of the complement system and decreased phagocytosis (Bradshaw et al., 2013; Jonsson et al., 2013). Several lines of evidence suggest that microglia prevent AD by stimulating the clearance of AB. Numerous microglial receptors appear to play a pivotal role in the clearance of this peptide (Yu and Ye, 2015), including microglial scavenger receptor 1 (Scara-1), cluster of differentiation 36 (CD36), the receptor for the Fc region of IgG IIb (FcyRIIb) and RAGE, the receptor of advanced glycation end products. For instance, during the progression of AD, the expression of these receptors in microglia decreases, and these cells lose their ability to clear A β (Derecki et al., 2014). The increase in the number of studies addressing the clearance of AB by microglia in recent years has brought about a deeper understanding of the contribution of microglia to clearance of this peptide. However, the clearance of extracellular Tau by microglia has received less attention. Nevertheless, several studies coincide on the crucial role played by microglial phagocytosis in the clearance of Tau and, consequently, in avoiding the spread of Tau and the progression of AD (Luo et al., 2015; Bolós et al., 2017c; Leyns and Holtzman, 2017; Figure 1). Therefore, it is now necessary to channel our efforts into elucidating the mechanisms by which pathological forms of Tau are cleared by glial cells to maintain brain homeostasis.

Abnormal and continuous activation of microglia has been reported in AD (Town et al., 2005; Ransohoff and El Khoury, 2016; Bolós et al., 2017c). In this regard, our recent data support the notion that although microglia are more activated in advanced stages of AD, their capacity to phagocytose Tau is impaired, thereby suggesting that there is a disconnection of microglial activation and phagocytosis at later stages of the disease (Bolós et al., 2017a). This effect has already been described for A β clearance by microglia in AD (Xiang et al., 2016) and in other neurodegenerative diseases (Sierra et al., 2013). Indeed, it has been proposed that neuron/microglial crosstalk is compromised in chronic neurodegenerative conditions (Sheridan and Murphy, 2013; Simon et al., 2018), and this impairment may explain the chronic maintenance of a pro-inflammatory state in AD.

REFERENCES

Aguzzi, A., and Rajendran, L. (2009). The transcellular spread of cytosolic amyloids, prions, and prionoids. *Neuron* 64, 783–790. doi: 10.1016/j.neuron. 2009.12.016

The transcriptome of microglia has been studied by several groups (Chiu et al., 2013; Hickman et al., 2013; Srinivasan et al., 2016; Keren-Shaul et al., 2017; Mathys et al., 2017). The overall gene expression profile of microglia revealed the upregulation of genes involved in neuroprotection and host defense during adulthood and aging in healthy brains (Hickman et al., 2013). However, using single-cell analysis in AD and other models of neurodegenerative disease, such as lateral amyotrophic sclerosis, the authors observed changes in several genes involved in microglia functions as the disease progressed (Chiu et al., 2013; Mathys et al., 2017). In a very interesting study using single-cell RNA-seq in an AD model, Keren-Shaul et al. (2017) identified a novel microglia subtype associated with neurodegenerative diseases (DAM) that is activated in a two-step process. In early stages, the expression of some genes, such as Cx3cr1, is upregulated, while other genes, such as Trem2, are downregulated. In late stages of AD, this ratio changes inversely, meaning that Trem2 is activated and Cx3cr1, among other genes, is downregulated. The observation that the expression of this set of genes is associated with phagocytic activity provides additional evidence of impaired microglia elementary functions in neurodegenerative diseases.

In summary, microglia are key cells in the maintenance of CNS homeostasis. They are essential for the clearance of proteins, such as the extracellular pathological Tau that characterizes tauopathies. As previously mentioned, the amount of phosphorylated Tau (phospho-Tau) at late stages of AD run in parallel with the number of activated microglia. However, a reduction in the phagocytic action of these cells has been shown in such stages. These findings support the notion that during the progression of the disease, there is an uncoupling of microglia activation and phagocytosis that specifically occurs at later stages of AD. Therefore, we propose that this disconnection is one of the mechanisms that drives neurodegeneration in AD and other tauopathies (**Figure 2**).

AUTHOR CONTRIBUTIONS

JP and ML-M contributed equally to this work. MB, JP, ML-M and JÁ drafted the manuscript. All authors read and approved the final manuscript.

FUNDING

This study was supported by the Spanish Ministry of Economy and Competitiveness (SAF-2014-53040-P (JÁ); SAF-2017-82185-R (ML-M)); the Centro de Investigación Biomédica en Red sobre Enfermedades Neurodegenerativas (CIBERNED, Spain, JÁ); the Alzheimer's Association AARG-17-528125 (ML-M); Institutional grants from the Fundación Ramón Areces and Banco de Santander to the CBMSO are also acknowledged.

Alliot, F., Godin, I., and Pessac, B. (1999). Microglia derive from progenitors, originating from the yolk sac, and which proliferate in the brain. *Dev. Brain Res.* 117, 145–152. doi: 10.1016/s0165-3806(99)00113-3

Andreadis, A. (2005). Tau gene alternative splicing: expression patterns, regulation and modulation of function in normal brain and neurodegenerative

diseases. Biochim. Biophys. Acta 1739, 91-103. doi: 10.1016/j.bbadis.2004. 08.010

- Arendt, T., Stieler, J. T., and Holzer, M. (2016). Tau and tauopathies. *Brain Res. Bull.* 126, 238–292. doi: 10.1016/j.brainresbull.2016.08.018
- Aronov, S., Aranda, G., Behar, L., and Ginzburg, I. (2001). Axonal tau mRNA localization coincides with tau protein in living neuronal cells and depends on axonal targeting signal. *J. Neurosci.* 21, 6577–6587. doi: 10.1523/JNEUROSCI. 21-17-06577.2001
- Asai, H., Ikezu, S., Tsunoda, S., Medalla, M., Luebke, J., Haydar, T., et al. (2015). Depletion of microglia and inhibition of exosome synthesis halt tau propagation. *Nat. Neurosci.* 18, 1584–1593. doi: 10.1038/nn.4132
- Avila, J., Jiménez, J. S., Sayas, C. L., Bolós, M., Zabala, J. C., Rivas, G., et al. (2016). Tau structures. *Front. Aging Neurosci.* 8:262. doi: 10.2514/6.1997-1232
- Ballatore, C., Lee, V. M., and Trojanowski, J. Q. (2007). Tau-mediated neurodegeneration in Alzheimer's disease and related disorders. *Nat. Rev. Neurosci.* 8, 663–672. doi: 10.1038/nrn2194
- Biber, K., Neumann, H., Inoue, K., and Boddeke, H. W. (2007). Neuronal 'On' and 'Off' signals control microglia. *Trends Neurosci.* 30, 596–602. doi: 10.1016/j.tins. 2007.08.007
- Boche, D., Perry, V. H., and Nicoll, J. A. (2013). Review: activation patterns of microglia and their identification in the human brain. *Neuropathol. Appl. Neurobiol.* 39, 3–18. doi: 10.1111/nan.12011
- Bolós, M., Llorens-Martín, M., Jurado-Arjona, J., Hernández, F., Rábano, A., and Avila, J. (2015). Direct evidence of internalization of tau by microglia *in vitro* and *in vivo. J. Alzheimers Dis.* 50, 77–87. doi: 10.3233/JAD-150704
- Bolós, M., Llorens-Martín, M., Perea, J. R., Jurado-Arjona, J., Rábano, A., Hernández, F., et al. (2017a). Absence of CX3CR1 impairs the internalization of Tau by microglia. *Mol. Neurodegener.* 12:59. doi: 10.1186/s13024-017-0200-1
- Bolós, M., Pallas-Bazarra, N., Terreros-Roncal, J., Perea, J. R., Jurado-Arjona, J., Ávila, J., et al. (2017b). Soluble Tau has devastating effects on the structural plasticity of hippocampal granule neurons. *Transl. Psychiatry* 7:1267. doi: 10.1038/s41398-017-0013-6
- Bolós, M., Perea, J. R., and Avila, J. (2017c). Alzheimer's disease as an inflammatory disease. *Biomol. Concepts* 8, 37–43. doi: 10.1515/bmc-2016-0029
- Bolós, M., Perea, J. R., Terreros-Roncal, J., Pallas-Bazarra, N., Jurado-Arjona, J., Avila, J., et al. (2018). Absence of microglial CX3CR1 impairs the synaptic integration of adult-born hippocampal granule neurons. *Brain Behav. Immun.* 68, 76–89. doi: 10.1016/j.bbi.2017.10.002
- Bradshaw, E. M., Chibnik, L. B., Keenan, B. T., Ottoboni, L., Raj, T., Tang, A., et al. (2013). CD33 Alzheimer's disease locus: altered monocyte function and amyloid biology. *Nat. Neurosci.* 16, 848–850. doi: 10.1038/nn.3435
- Brettschneider, J., Del Tredici, K., Lee, V. M., and Trojanowski, J. Q. (2015). Spreading of pathology in neurodegenerative diseases: a focus on human studies. *Nat. Rev. Neurosci.* 16, 109–120. doi: 10.1038/nrn3887
- Brion, J. P., Couck, A. M., Passareiro, E., and Flament-Durand, J. (1985). Neurofibrillary tangles of Alzheimer's disease: an immunohistochemical study. J. Submicrosc. Cytol. 17, 89–96.
- Bruttger, J., Karram, K., Wörtge, S., Regen, T., Marini, F., Hoppmann, N., et al. (2015). Genetic cell ablation reveals clusters of local self-renewing microglia in the mammalian central nervous system. *Immunity* 43, 92–106. doi: 10.1016/j. immuni.2015.06.012
- Calafate, S., Buist, A., Miskiewicz, K., Vijayan, V., Daneels, G., de Strooper, B., et al. (2015). Synaptic contacts enhance cell-to-cell tau pathology propagation. *Cell Rep.* 11, 1176–1183. doi: 10.1016/j.celrep.2015.04.043
- Chang, Y. P., Fang, K. M., Lee, T. I., and Tzeng, S. F. (2006). Regulation of microglial activities by glial cell line derived neurotrophic factor. J. Cell. Biochem. 97, 501–511. doi: 10.1002/jcb.20646
- Chiu, I. M., Morimoto, E. T., Goodarzi, H., Liao, J. T., O'Keeffe, S., Phatnani, H. P., et al. (2013). A neurodegeneration-specific gene-expression signature of acutely isolated microglia from an amyotrophic lateral sclerosis mouse model. *Cell Rep.* 4, 385–401. doi: 10.1016/j.celrep.2013.06.018
- de Calignon, A., Polydoro, M., Suarez-Calvet, M., William, C., Adamowicz, D. H., Kopeikina, K. J., et al. (2012). Propagation of tau pathology in a model of early Alzheimer's disease. *Neuron* 73, 685–697. doi: 10.1016/j.neuron.2011.11.033
- Derecki, N. C., Katzmarski, N., Kipnis, J., and Meyer-Luehmann, M. (2014). Microglia as a critical player in both developmental and late-life CNS pathologies. Acta Neuropathol. 128, 333–345. doi: 10.1007/s00401-014-1321-z

- Drubin, D. G., and Kirschner, M. W. (1986). Tau protein function in living cells. *J. Cell Biol.* 103, 2739–2746. doi: 10.1083/jcb.103.6.2739
- Eyo, U. B., and Wu, L. J. (2013). Bidirectional microglia-neuron communication in the healthy brain. *Neural Plast.* 2013:456857. doi: 10.1155/2013/456857
- Fiandaca, M. S., Kapogiannis, D., Mapstone, M., Boxer, A., Eitan, E., Schwartz, J. B., et al. (2015). Identification of preclinical Alzheimer's disease by a profile of pathogenic proteins in neurally derived blood exosomes: a case-control study. *Alzheimers Dement*. 11, 600.e1–700.e1. doi: 10.1016/j.jalz. 2014.06.008
- Ginhoux, F., Greter, M., Leboeuf, M., Nandi, S., See, P., Gokhan, S., et al. (2010). Fate mapping analysis reveals that adult microglia derive from primitive macrophages. *Science* 330, 841–845. doi: 10.1126/science.1194637
- Goedert, M., and Spillantini, M. G. (2011). Pathogenesis of the tauopathies. J. Mol. Neurosci. 45, 425–431. doi: 10.1007/s12031-011-9593-4
- Gómez-Ramos, A., Díaz-Hernández, M., Cuadros, R., Hernández, F., and Avila, J. (2006). Extracellular tau is toxic to neuronal cells. *FEBS Lett.* 580, 4842–4850. doi: 10.1016/j.febslet.2006.07.078
- Hanisch, U. K. (2002). Microglia as a source and target of cytokines. *Glia* 40, 140–155. doi: 10.1002/glia.10161
- Harrison, J. K., Jiang, Y., Chen, S., Xia, Y., Maciejewski, D., McNamara, R. K., et al. (1998). Role for neuronally derived fractalkine in mediating interactions between neurons and CX3CR1-expressing microglia. *Proc. Natl. Acad. Sci.* U S A 95, 10896–10901. doi: 10.1073/pnas.95.18.10896
- Hickman, S. E., Kingery, N. D., Ohsumi, T. K., Borowsky, M. L., Wang, L. C., Means, T. K., et al. (2013). The microglial sensome revealed by direct RNA sequencing. *Nat. Neurosci.* 16, 1896–1905. doi: 10.1038/nn.3554
- Holmes, B. B., and Diamond, M. I. (2014). Prion-like properties of Tau protein: the importance of extracellular Tau as a therapeutic target. J. Biol. Chem. 289, 19855–19861. doi: 10.1074/jbc.R114.549295
- Holtzman, D. M., Morris, J. C., and Goate, A. M. (2011). Alzheimer's disease: the challenge of the second century. *Sci. Transl. Med.* 3:77sr1. doi:10.1126/scitranslmed.3002369
- Hu, W., Zhang, X., Tung, Y. C., Xie, S., Liu, F., and Iqbal, K. (2016). Hyperphosphorylation determines both the spread and the morphology of tau pathology. *Alzheimers Dement.* 12, 1066–1077. doi: 10.1016/j.jalz.2016. 01.014
- Hyman, B. T. (2014). Tau propagation, different tau phenotypes, and prion-like properties of tau. *Neuron* 82, 1189–1190. doi: 10.1016/j.neuron.2014. 06.004
- Ittner, L. M., and Götz, J. (2011). Amyloid-β and tau—a toxic pas de deux in Alzheimer's disease. *Nat. Rev. Neurosci.* 12, 65–72. doi: 10.1038/nrn2967
- Jonsson, T., Stefansson, H., Steinberg, S., Jonsdottir, I., Jonsson, P. V., Snaedal, J., et al. (2013). Variant of TREM2 associated with the risk of Alzheimer's disease. *N. Engl. J. Med.* 368, 107–116. doi: 10.1056/NEJMoa1211103
- Kanmert, D., Cantlon, A., Muratore, C. R., Jin, M., O'Malley, T. T., Lee, G., et al. (2015). C-terminally truncated forms of tau, but not full-length tau or its C-terminal fragments, are released from neurons independently of cell death. *J. Neurosci.* 35, 10851–10865. doi: 10.1523/JNEUROSCI.0387-15.2015
- Karch, C. M., Jeng, A. T., and Goate, A. M. (2012). Extracellular Tau levels are influenced by variability in Tau that is associated with tauopathies. J. Biol. Chem. 287, 42751–42762. doi: 10.1074/jbc.M112.380642
- Karperien, A., Ahammer, H., and Jelinek, H. F. (2013). Quantitating the subtleties of microglial morphology with fractal analysis. *Front. Cell. Neurosci.* 7:3. doi: 10.3389/fncel.2013.00003
- Keren-Shaul, H., Spinrad, A., Weiner, A., Matcovitch-Natan, O., Dvir-Szternfeld, R., Ulland, T. K., et al. (2017). A unique microglia type associated with restricting development of Alzheimer's disease. *Cell* 169, 1276.e17–1290.e17. doi: 10.1016/j.cell.2017.05.018
- Kim, K. W., Vallon-Eberhard, A., Zigmond, E., Farache, J., Shezen, E., Shakhar, G., et al. (2011). *In vivo* structure/function and expression analysis of the CX3C chemokine fractalkine. *Blood* 118, e156–e167. doi: 10.1182/blood-2011-04-348946
- Lawson, L. J., Perry, V. H., Dri, P., and Gordon, S. (1990). Heterogeneity in the distribution and morphology of microglia in the normal adult mouse brain. *Neuroscience* 39, 151–170. doi: 10.1016/0306-4522(90)90229-w
- Lee, T. I., Yang, C. S., Fang, K. M., and Tzeng, S. F. (2009). Role of ciliary neurotrophic factor in microglial phagocytosis. *Neurochem. Res.* 34, 109–117. doi: 10.1007/s11064-008-9682-0

- Leyns, C. E. G., and Holtzman, D. M. (2017). Glial contributions to neurodegeneration in tauopathies. *Mol. Neurodegener.* 12:50. doi: 10.1186/ s13024-017-0192-x
- Liu, F., and Gong, C. X. (2008). Tau exon 10 alternative splicing and tauopathies. Mol. Neurodegener. 3:8. doi: 10.1186/1750-1326-3-8
- Loane, D. J., and Byrnes, K. R. (2010). Role of microglia in neurotrauma. Neurotherapeutics 7, 366–377. doi: 10.1016/j.nurt.2010.07.002
- Luo, W., Liu, W., Hu, X., Hanna, M., Caravaca, A., and Paul, S. M. (2015). Microglial internalization and degradation of pathological tau is enhanced by an anti-tau monoclonal antibody. *Sci. Rep.* 5:11161. doi: 10.1038/ srep11161
- Maphis, N., Xu, G., Kokiko-Cochran, O. N., Jiang, S., Cardona, A., Ransohoff, R. M., et al. (2015). Reactive microglia drive tau pathology and contribute to the spreading of pathological tau in the brain. *Brain* 138, 1738–1755. doi: 10.1093/brain/awv081
- Mathys, H., Adaikkan, C., Gao, F., Young, J. Z., Manet, E., Hemberg, M., et al. (2017). Temporal tracking of microglia activation in neurodegeneration at single-cell resolution. *Cell Rep.* 21, 366–380. doi: 10.1016/j.celrep.2017.09.039
- Medina, M., and Avila, J. (2014a). Is tau a prion-like protein? J. Alzheimers Dis. 40, S1–S3. doi: 10.3233/JAD-140755
- Medina, M., and Avila, J. (2014b). The role of extracellular Tau in the spreading of neurofibrillary pathology. *Front. Cell. Neurosci.* 8:113. doi: 10.3389/fncel.2014. 00113
- Medina, M., Hernández, F., and Avila, J. (2016). New features about tau function and dysfunction. *Biomolecules* 6:E21. doi: 10.3390/biom6020021
- Michell-Robinson, M. A., Touil, H., Healy, L. M., Owen, D. R., Durafourt, B. A., Bar-Or, A., et al. (2015). Roles of microglia in brain development, tissue maintenance and repair. *Brain* 138, 1138–1159. doi: 10.1093/brain/awv066
- Minas, K., and Liversidge, J. (2006). Is the CD200/CD200 receptor interaction more than just a myeloid cell inhibitory signal? *Crit. Rev. Immunol.* 26, 213–230. doi: 10.1615/critrevimmunol.v26.i3.20
- Mirbaha, H., Holmes, B. B., Sanders, D. W., Bieschke, J., and Diamond, M. I. (2015). Tau trimers are the minimal propagation unit spontaneously internalized to seed intracellular aggregation. J. Biol. Chem. 290, 14893–14903. doi: 10.1074/jbc.M115.652693
- Mitrasinovic, O. M., and Murphy, G. M. Jr. (2003). Microglial overexpression of the M-CSF receptor augments phagocytosis of opsonized Aβ. *Neurobiol. Aging* 24, 807–815. doi: 10.1016/s0197-4580(02)00237-3
- Morris, M., Knudsen, G. M., Maeda, S., Trinidad, J. C., Ioanoviciu, A., Burlingame, A. L., et al. (2015). Tau post-translational modifications in wild-type and human amyloid precursor protein transgenic mice. *Nat. Neurosci.* 18, 1183–1189. doi: 10.1038/nn.4067
- Mott, R. T., Ait-Ghezala, G., Town, T., Mori, T., Vendrame, M., Zeng, J., et al. (2004). Neuronal expression of CD22: novel mechanism for inhibiting microglial proinflammatory cytokine production. *Glia* 46, 369–379. doi: 10.1002/glia.20009
- Nimmerjahn, A., Kirchhoff, F., and Helmchen, F. (2005). Resting microglial cells are highly dynamic surveillants of brain parenchyma *in vivo*. *Science* 308, 1314–1318. doi: 10.1126/science.1110647
- Paolicelli, R. C., Bisht, K., and Tremblay, M. E. (2014). Fractalkine regulation of microglial physiology and consequences on the brain and behavior. *Front. Cell. Neurosci.* 8:129. doi: 10.3389/fncel.2014.00129
- Paolicelli, R. C., Bolasco, G., Pagani, F., Maggi, L., Scianni, M., Panzanelli, P., et al. (2011). Synaptic pruning by microglia is necessary for normal brain development. *Science* 333, 1456–1458. doi: 10.1126/science.1202529
- Parnaik, R., Raff, M. C., and Scholes, J. (2000). Differences between the clearance of apoptotic cells by professional and non-professional phagocytes. *Curr. Biol.* 10, 857–860. doi: 10.1016/s0960-9822(00)00598-4
- Pérez, M., Cuadros, R., Hernández, F., and Avila, J. (2016). Secretion of full-length tau or tau fragments in a cell culture model. *Neurosci. Lett.* 634, 63–69. doi: 10.1016/j.neulet.2016.09.026
- Peri, F., and Nüsslein-Volhard, C. (2008). Live imaging of neuronal degradation by microglia reveals a role for v0-ATPase a1 in phagosomal fusion *in vivo*. *Cell* 133, 916–927. doi: 10.1016/j.cell.2008.04.037
- Perry, V. H. (1998). A revised view of the central nervous system microenvironment and major histocompatibility complex class II antigen presentation. J. Neuroimmunol. 90, 113–121. doi: 10.1016/s0165-5728(98)00145-3

- Pittman, A. M., Fung, H. C., and de Silva, R. (2006). Untangling the tau gene association with neurodegenerative disorders. *Hum. Mol. Genet.* 15, R188-R195. doi: 10.1093/hmg/ddl190
- Polanco, J. C., Scicluna, B. J., Hill, A. F., and Götz, J. (2016). Extracellular vesicles isolated from the brains of rTg4510 mice seed tau protein aggregation in a threshold-dependent manner. J. Biol. Chem. 291, 12445–12466. doi: 10.1074/jbc.M115.709485
- Pooler, A. M., Phillips, E. C., Lau, D. H., Noble, W., and Hanger, D. P. (2013). Physiological release of endogenous tau is stimulated by neuronal activity. *EMBO Rep.* 14, 389–394. doi: 10.1038/embor.2013.15
- Rajendran, L., Bali, J., Barr, M. M., Court, F. A., Krämer-Albers, E. M., Picou, F., et al. (2014). Emerging roles of extracellular vesicles in the nervous system. *J. Neurosci.* 34, 15482–15489. doi: 10.1523/JNEUROSCI.3258-14.2014
- Ransohoff, R. M. (2016a). A polarizing question: do M1 and M2 microglia exist? Nat. Neurosci. 19, 987–991. doi: 10.1038/nn.4338
- Ransohoff, R. M. (2016b). How neuroinflammation contributes to neurodegeneration. *Science* 353, 777–783. doi: 10.1126/science.aag2590
- Ransohoff, R. M., and El Khoury, J. (2016). Microglia in health and disease. Cold Spring Harb. Perspect. Biol. 8:a020560. doi: 10.1101/cshperspect. a020560
- Ribes, S., Ebert, S., Czesnik, D., Regen, T., Zeug, A., Bukowski, S., et al. (2009). Tolllike receptor prestimulation increases phagocytosis of *Escherichia coli* DH5α and *Escherichia coli* K1 strains by murine microglial cells. *Infect. Immun.* 77, 557–564. doi: 10.1128/IAI.00903-08
- Saman, S., Kim, W., Raya, M., Visnick, Y., Miro, S., Saman, S., et al. (2012). Exosome-associated tau is secreted in tauopathy models and is selectively phosphorylated in cerebrospinal fluid in early Alzheimer disease. *J. Biol. Chem.* 287, 3842–3849. doi: 10.1074/jbc.M111.277061
- Sanchez-Mejias, E., Navarro, V., Jimenez, S., Sanchez-Mico, M., Sanchez-Varo, R., Nunez-Diaz, C., et al. (2016). Soluble phospho-tau from Alzheimer's disease hippocampus drives microglial degeneration. *Acta Neuropathol.* 132, 897–916. doi: 10.1007/s00401-016-1630-5
- Schafer, D. P., Lehrman, E. K., Kautzman, A. G., Koyama, R., Mardinly, A. R., Yamasaki, R., et al. (2012). Microglia sculpt postnatal neural circuits in an activity and complement-dependent manner. *Neuron* 74, 691–705. doi: 10.1016/j.neuron.2012.03.026
- Sheridan, G. K., and Murphy, K. J. (2013). Neuron-glia crosstalk in health and disease: fractalkine and CX3CR1 take centre stage. *Open Biol.* 3:130181. doi: 10.1098/rsob.130181
- Shie, F. S., Breyer, R. M., and Montine, T. J. (2005). Microglia lacking E Prostanoid Receptor subtype 2 have enhanced Aβ phagocytosis yet lack Aβactivated neurotoxicity. Am. J. Pathol. 166, 1163–1172. doi: 10.1016/s0002-9440(10)62336-x
- Sierra, A., Abiega, O., Shahraz, A., and Neumann, H. (2013). Janus-faced microglia: beneficial and detrimental consequences of microglial phagocytosis. *Front. Cell. Neurosci.* 7:6. doi: 10.3389/fncel.2013.00006
- Sierra, A., Encinas, J. M., Deudero, J. J., Chancey, J. H., Enikolopov, G., Overstreet-Wadiche, L. S., et al. (2010). Microglia shape adult hippocampal neurogenesis through apoptosis-coupled phagocytosis. *Cell Stem Cell* 7, 483–495. doi: 10.1016/j.stem.2010.08.014
- Sierra, A., Tremblay, M. È., and Wake, H. (2014). Never-resting microglia: physiological roles in the healthy brain and pathological implications. *Front. Cell. Neurosci.* 8:240. doi: 10.3389/fncel.2014.00240
- Simón, D., García-García, E., Royo, F., Falcón-Pérez, J. M., and Avila, J. (2012). Proteostasis of tau. Tau overexpression results in its secretion via membrane vesicles. *FEBS Lett.* 586, 47–54. doi: 10.1016/j.febslet.2011. 11.022
- Simon, E., Obst, J., and Gomez-Nicola, D. (2018). The evolving dialogue of microglia and neurons in Alzheimer's disease: microglia as necessary transducers of pathology. *Neuroscience* doi: 10.1016/j.neuroscience.2018.01. 059 [Epub ahead of print].
- Spillantini, M. G., and Goedert, M. (2013). Tau pathology and neurodegeneration. Lancet Neurol. 12, 609–622. doi: 10.1016/S1474-4422(13)70090-5
- Srinivasan, K., Friedman, B. A., Larson, J. L., Lauffer, B. E., Goldstein, L. D., Appling, L. L., et al. (2016). Untangling the brain's neuroinflammatory and neurodegenerative transcriptional responses. *Nat. Commun.* 7:11295. doi: 10.1038/ncomms11295

- Surprenant, A., and North, R. A. (2009). Signaling at purinergic P2X receptors. Annu. Rev. Physiol. 71, 333–359. doi: 10.1146/annurev.physiol.70.113006. 100630
- Tang, Y., and Le, W. (2016). Differential roles of M1 and M2 microglia in neurodegenerative diseases. *Mol. Neurobiol.* 53, 1181–1194. doi: 10.1007/ s12035-014-9070-5
- Town, T., Nikolic, V., and Tan, J. (2005). The microglial "activation" continuum: from innate to adaptive responses. J. Neuroinflammation 2:24. doi: 10.1186/1742-2094-2-24
- Verkhratsky, A., Anderova, M., and Chvatal, A. (2009). Differential calcium signalling in neuronal-glial networks. *Front. Biosci.* 14, 2004–2016. doi: 10.2741/3359
- Webster, S. D., Yang, A. J., Margol, L., Garzon-Rodriguez, W., Glabe, C. G., and Tenner, A. J. (2000). Complement component C1q modulates the phagocytosis of Aβ by microglia. *Exp. Neurol.* 161, 127–138. doi: 10.1006/exnr. 1999.7260
- Weingarten, M. D., Lockwood, A. H., Hwo, S. Y., and Kirschner, M. W. (1975). A protein factor essential for microtubule assembly. *Proc. Natl. Acad. Sci. U S A* 72, 1858–1862. doi: 10.1073/pnas.72.5.1858
- Wischik, C. M., Novak, M., Edwards, P. C., Klug, A., Tichelaar, W., and Crowther, R. A. (1988). Structural characterization of the core of the paired helical filament of Alzheimer disease. *Proc. Natl. Acad. Sci. U S A* 85, 4884–4888. doi: 10.1073/pnas.85.13.4884
- Wolf, S. A., Boddeke, H. W., and Kettenmann, H. (2017). Microglia in physiology and disease. Annu. Rev. Physiol. 79, 619–643. doi: 10.1146/annurev-physiol-022516-034406
- Wood, J. G., Mirra, S. S., Pollock, N. J., and Binder, L. I. (1986). Neurofibrillary tangles of Alzheimer disease share antigenic determinants with the axonal microtubule-associated protein tau (tau). *Proc. Natl. Acad. Sci. U S A* 83, 4040–4043. doi: 10.1073/pnas.83.11.4040
- Wu, J. W., Herman, M., Liu, L., Simoes, S., Acker, C. M., Figueroa, H., et al. (2013). Small misfolded Tau species are internalized via bulk endocytosis and

anterogradely and retrogradely transported in neurons. J. Biol. Chem. 288, 1856–1870. doi: 10.1074/jbc.M112.394528

- Xiang, X., Werner, G., Bohrmann, B., Liesz, A., Mazaheri, F., Capell, A., et al. (2016). TREM2 deficiency reduces the efficacy of immunotherapeutic amyloid clearance. *EMBO Mol. Med.* 8, 992–1004. doi: 10.15252/emmm. 201606370
- Yamada, K., Holth, J. K., Liao, F., Stewart, F. R., Mahan, T. E., Jiang, H., et al. (2014). Neuronal activity regulates extracellular tau *in vivo*. J. Exp. Med. 211, 387–393. doi: 10.1084/jem.20131685
- Yu, Y., and Ye, R. D. (2015). Microglial Aβ receptors in Alzheimer's disease. *Cell. Mol. Neurobiol.* 35, 71–83. doi: 10.1007/s10571-014-0101-6
- Zhan, Y., Paolicelli, R. C., Sforazzini, F., Weinhard, L., Bolasco, G., Pagani, F., et al. (2014). Deficient neuron-microglia signaling results in impaired functional brain connectivity and social behavior. *Nat. Neurosci.* 17, 400–406. doi: 10.1038/nn.3641
- Zhang, Y., Tian, Q., Zhang, Q., Zhou, X., Liu, S., and Wang, J. Z. (2009). Hyperphosphorylation of microtubule-associated tau protein plays dual role in neurodegeneration and neuroprotection. *Pathophysiology* 16, 311–316. doi: 10.1016/j.pathophys.2009.02.003

Conflict of Interest Statement: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Copyright © 2018 Perea, Llorens-Martín, Ávila and Bolós. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.