

Circulating Biomarkers in Lower Extremity Artery Disease

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Abstract

Lower extremity artery disease (LEAD), a chronic condition with disturbed lower extremity circulation due to narrowing of the arteries, is predominantly caused by atherosclerosis and is associated with the presence of cardiovascular risk factors and an increased risk of cardiovascular events. LEAD is prevalent among older individuals and predicted to rise with the ageing population. In progressive disease, the patient experiences symptoms of ischaemia when walking and, in advanced critical limb-threatening ischaemia, even at rest. However, LEAD is asymptomatic in most patients, delaying diagnosis and treatment. In this setting, circulating biomarkers may facilitate earlier diagnosis in selected individuals. This review provides a broad overview of the circulating biomarkers investigated to date in relation to LEAD and discusses their usefulness in clinical practice.

Keywords

Peripheral artery disease, lower extremity artery disease, biomarkers

Disclosure: The authors have no conflicts of interest to declare.

Received: 20 December 2021 **Accepted:** 31 January 2022 **Citation:** *European Cardiology Review* 2022;17:e09. **DOI:** <https://doi.org/10.15420/ecr.2021.58>

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Lower Extremity Artery Disease: Definition, Epidemiology and Clinical Presentation

Lower extremity artery disease (LEAD), primarily an atherosclerosis -driven disease, causes varying degrees of disturbances in blood perfusion to the lower limb.^{1,2}

The prevalence of LEAD in the general population has been reported to be 3–10%, although prevalence varies depending on the age of the population, reaching around 18% for those aged ≥ 65 years.^{2,3}

In early stages, LEAD is usually asymptomatic, and the impaired limb circulation is only evident as an ankle–brachial index (ABI) ≤ 0.90 . With progression of the disease, intermittent claudication (IC) may occur, characterised by muscle fatigue and lower extremity pain triggered by physical activity and directly relieved when resting.⁴ In the more severe critical limb-threatening ischaemia (CLTI), the patient experiences pain at rest and/or presents with ischaemic ulceration or gangrene of the foot.⁴

The mere presence of LEAD, either asymptomatic or symptomatic, is a significant predictor of increased risk of cardiovascular events (CVE) and cardiovascular mortality.^{5–7} Among individuals with IC, there is a low risk of progression to CLTI, with only 2% requiring lower extremity amputation within 10 years from diagnosis.⁸ Conversely, in the CLTI population, first-year rates of amputation in most studies are greater than 15–20% and 1-year mortality rates increase markedly from a few per cent in IC to 20–30% in CLTI.^{9,10}

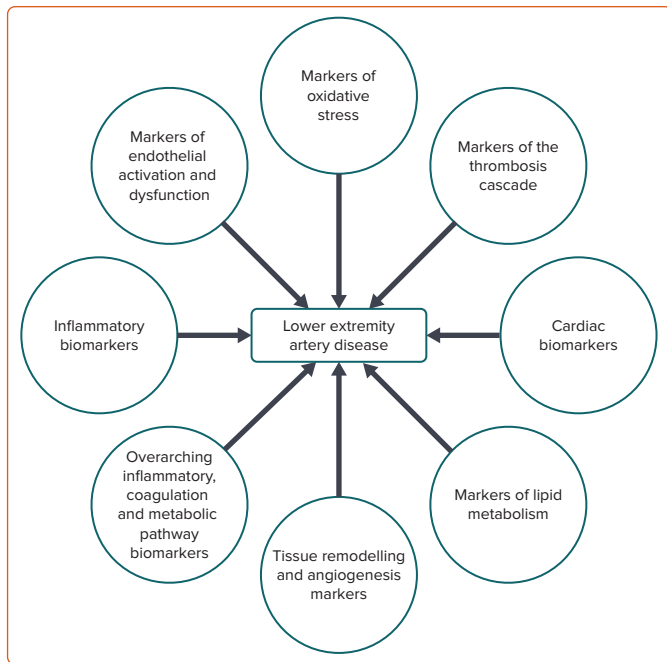
Compensatory development of new capillary networks (angiogenesis) and expansion of collateral arteries (arteriogenesis) are physiological responses to progressing limb ischaemia also encompassing pathological processes such as inflammation, apoptosis and vascular remodelling.¹¹ In the constantly hypoperfused tissue in CLTI, chronic inflammation induces endothelial dysfunction and oxidative stress, with subsequent mitochondrial damage, generation of free radicals, muscle degeneration, connective tissue damage, fibrosis and eventually risk of gangrene.^{12–14} These events are possible targets for treatment and potential sources of diagnostic and prognostic biomarkers.

Methods

The aim of this review is to provide a comprehensive overview of the most promising established and emerging circulating biomarkers for LEAD and to discuss the usefulness of these biomarkers in screening, risk stratification and monitoring of therapeutic effect in line with personalised medicine. The biomarkers are grouped according to their mechanism in atherothrombotic disease, as shown in *Figure 1*. In addition, major biomarkers with data regarding their discriminative performances are summarised in *Table 1*.

The search terms used in this review were: ‘lower extremity artery disease’ or ‘peripheral artery disease’ or ‘intermittent claudication’ or ‘critical limb-threatening ischemia’ or ‘critical limb ischaemia’ and ‘biomarker’. In additional searches, the term ‘biomarker’ was changed to the names of specific biomarkers.

Figure 1: Lower Extremity Artery Disease Biomarkers Grouped by Origin or Mechanism



The biomarkers presented in this review are grouped by their biological origin, pathophysiological mechanism or prior clinical application.

Biomarkers: Introduction and Definitions

The definition of a biomarker used in this review is the one presented by the Food and Drug Administration and the National Institutes of Health, specifically: “A defined characteristic that is measured as an indicator of normal biological processes, pathogenic processes or responses to an exposure or intervention.”¹⁵ The biomarker concept should be distinguished from clinical outcome assessments (i.e. evaluations of mental or physical status). To be of clinical value, a biomarker must be stable in the pre-analytical context, measurable without too much effort or cost and provide information on diagnosis and/or prognosis specific to the condition in question. In addition, a novel biomarker should add information incremental to the diagnostic tools and markers already used in clinical practice.

In this review, we focus on circulating biomarkers evaluated for diagnosis or assessment of prognosis in LEAD in combination with existing clinical biomarkers.

Circulating Biomarkers

As an adjunct to the clinical assessment tools, circulating biomarkers may contribute to the diagnostics of LEAD, prediction of future deterioration, including cardiovascular disease (CVD) progression in general, and evaluation of treatment effects. Despite demonstrated associations and biological relevance, most biomarkers are insufficient to reclassify individuals correctly above and beyond existing clinical and circulating biomarkers, such as ABI, which carries strong predictive power for CVD mortality.¹⁶ Moreover, due to large intraindividual variations in concentration, single-biomarker assessments may be of limited value.¹⁷ The biomarkers presented in this review are grouped by pathophysiological origin and mechanism (Figure 1).

Inflammatory Biomarkers

LEAD is primarily a consequence of progressive atherosclerosis, with or without atherothrombosis, in the lower extremity vasculature, with

resulting ischaemia and subsequent tissue damage. Given the established contribution of low-grade chronic inflammation in all stages of atherosclerosis, it is conceivable that inflammatory processes, such as those reflecting endothelial cell activation, synthesis and secretion of proinflammatory factors, the expression of adhesion molecules for inflammatory cells and prothrombotic activity, have been targeted in the search for disease biomarkers. These early processes, including monocyte recruitment, are driven by the innate immune system, and inflammatory cells or tissue cells activated during inflammation have been associated with a poor prognosis in LEAD.^{18,19}

Interleukin-6

Interleukin (IL)-6, central in atherosclerosis development and progression, is part of the IL-1 β /IL-6/C-reactive protein (CRP) pathway. IL-6 signals through two different pathways, with the classical IL-6 signalling pathway being the main mediator of the acute phase reaction with subsequent expression of adhesion molecules and the proliferation and transformation of vascular smooth muscle cells into foam cells.²⁰ The proinflammatory IL-6 trans-signalling pathway has not been analysed in LEAD.

In several case-control studies, IL-6 concentrations were higher in individuals with LEAD.^{17,21–23} This relationship was true also before and after treadmill exercise.²⁴ In the Edinburgh Artery study of middle-aged men and women, IL-6 was associated with deterioration in ABI after 5 and 12 years follow-up independent of cardiovascular risk factors, baseline CVD and ABI.²⁵ In line with the Edinburgh cohort, later studies have demonstrated that walking endurance in LEAD patients was inversely associated with high IL-6 concentrations.^{21,26,27} Moreover, persistently elevated IL-6 concentrations were associated with a more rapid functional decline compared with subjects with fluctuating or persistently low IL-6 concentrations.²⁸ In addition, decreased size of the calf muscle as a surrogate marker of impaired leg function was associated with IL-6.²⁹ In CLTI patients with diabetes, IL-6 was correlated with negative vascular outcome after endovascular revascularisation and was an independent predictor of in-stent restenosis 6 months after stenting in the femoropopliteal artery.^{30,31}

C-Reactive Protein

CRP is an acute-phase reactant and product of classical IL-6 signalling. Several studies have shown that CRP is associated with the risk of CVE, although causality has not been demonstrated. Several case-control studies reported higher CRP concentrations in subjects with LEAD than in healthy controls.^{17,21,32–34} The Physicians’ Health study and the population-based prospective ARIC cohort study demonstrated that CRP independently predicted incident LEAD.^{35,36} In the Rotterdam cohort study, CRP predicted progression of atherosclerosis, including LEAD, independently of traditional cardiovascular risk factors and in the Edinburgh Artery study and a prospective study by Aboyans et al., the same was shown specifically for LEAD.^{25,37,38} However, not all studies did demonstrate an association between CRP and LEAD.³⁹

In patients with symptomatic LEAD, CRP was associated with an increased risk of future atherothrombotic events both in the lower extremities and in other vascular beds.^{32,40–42} Combining high-sensitivity C-reactive protein (hsCRP) measurement with the assessment of ABI improved cardiovascular risk stratification in patients with LEAD.^{32,43} However, hsCRP alone has been shown to be inferior to ABI in the prediction of disease severity.⁴⁴ These results were confirmed by a systematic meta-analysis of prospective studies investigating CRP as a predictor of CVE in LEAD patients.⁴⁵ Moreover, several studies have shown an inverse association between

Table 1: Summary of the Major Circulating Biomarkers in Lower Extremity Artery Disease Presented in This Review

Major Circulating Biomarkers Assessed	Pathophysiological Pathway/Mechanism	Type of Biomarker	Discriminative Performance
Inflammation			
IL-6 ^{17,21-31}	IL-1/IL-6/CRP pathway	Diagnostic, prognostic	IL-6, CRP, TNF- α and risk factors: AUC 1.00 for MALE and 0.91 for MACE ³⁰
CRP ^{17,21,32-48}	IL-1/IL-6/CRP pathway	Diagnostic, prognostic	Diagnosing LEAD: AUC 0.81 ³⁴ Predicting composite of amputation, revascularisation, all-cause death: AUC 0.57 ⁴⁰ CRP, IL-6, TNF- α and risk factors: AUC 1.00 for MALE and 0.91 for MACE ³⁰
TNF- α ^{21,23,24,27,30,49,50}	Adipokine and cytokine	Prognostic	TNF- α , IL-6, CRP and risk factors: AUC 1.00 for MALE and 0.91 for MACE ³⁰
GDF-15 ⁵⁸⁻⁶¹	Member of the TGF- β superfamily; regulation of inflammatory processes	Prognostic	Predicts all-cause mortality in LEAD with 90.0% sensitivity, 52.6% specificity ⁶⁰ Predicts all-cause mortality together with TRAIL-R2: AUC 0.74 ⁶¹
Endothelial Activation and Dysfunction			
VCAM-1, ICAM-1 ^{17,21,23-26,28,53-56}	Mediate leucocyte endothelial adhesion and transmigration	Diagnostic, prognostic	Diagnosing LEAD (VCAM-1): AUC 0.76 ⁵³
Oxidative Stress			
ADMA ^{63,67-71}	Inhibitor of NO production	Diagnostic, prognostic	Predict cardiovascular death: AUC 0.649 ⁷⁰
Coagulation			
Fibrinogen ^{33,35,43,73,77-79,81-84}	IL-1/IL-6/CRP pathway	Diagnostic, prognostic	Diagnosing LEAD: AUC 0.870 ³⁴
Cardiac			
NT-proBNP ^{43,88-92}	Precursor to BNP secreted by cardiomyocytes during ventricular stretch	Diagnostic, prognostic	Predicting all-cause mortality in LEAD: AUC 0.74 ⁹¹
Lipid Metabolism			
Lp(a) ⁹⁵⁻⁹⁷	LDL-like particle bound to apoB100	Diagnostic	Risk factors, OxPL/apoB100, Lp(a) diagnosing LEAD: AUC 0.759 in women and 0.736 in men ⁹⁷
Lp-PLA ₂ ⁹⁸⁻¹⁰²	Modifies oxidised LDL	Diagnostic	Diagnosing LEAD: AUC 0.807 ¹⁰⁰
Auxiliary			
Circulating miRNAs ¹²³⁻¹²⁵	Non-coding single-stranded RNAs from EVs, regulate gene expression	Diagnostic	Diagnosing LEAD: AUC >0.93 for miR-15b, miR-16 and miR-363 ¹²³

ADMA = asymmetric dimethylarginine; apoB100 = apolipoprotein B100; AUC = area under the curve; BNP = B-type natriuretic peptide; CRP = C-reactive protein; EVs = extracellular vesicles; GDF-15 = growth differentiation factor-15; IL = interleukin; LEAD = lower extremity artery disease; Lp(a) = lipoprotein (a); Lp-PLA₂ = lipoprotein-associated phospholipase A₂; MACE = major adverse cardiovascular events; MALE = major adverse limb events; miRNA = microRNA; NO = nitric oxide; NT-proBNP = N-terminal pro B-type natriuretic peptide; OxPL = oxidised phospholipids; TGF- β = transforming growth factor- β ; TNF- α = tumour necrosis factor- α ; TRAIL-R2 = tumour necrosis factor-related apoptosis-inducing ligand receptor 2; VCAM-1 = vascular cell adhesion molecule-1.

CRP and ABI, severe clinical LEAD and functional deterioration.^{19,25,33,34,46,47} High CRP concentrations also correlated negatively with walking duration.^{21,48} In addition, high CRP was associated with an enhanced risk of postoperative vascular events and failure of endovascular revascularisation in LEAD patients.³⁰

Tumour Necrosis Factor- α

Tumour necrosis factor (TNF)- α is a central mediator of inflammatory reactions inducing matrix metalloproteinase synthesis and contributing to atherosclerotic plaque instability. In case-control studies, TNF- α and circulating TNF- α receptor levels were higher in patients with LEAD than in healthy subjects.^{21,23,49} In a pre- and post-exercise analysis, TNF- α concentrations were still higher in subjects with LEAD, regardless of treadmill training.²⁴ Moreover, TNF- α expression and circulating levels of TNF- α were inversely associated with walking duration.^{21,50} Conversely, in a study of patients with verified LEAD, TNF- α concentrations were significantly correlated with an angiographic score, but an association with treadmill performance could not be demonstrated.²⁷ TNF- α concentrations were associated with increased vascular risk in CLTI patients with diabetes after endovascular intervention.³⁰

Markers of Endothelial Activation and Dysfunction Adhesion Molecules

Cell adhesion molecules (CAMs) are transmembrane glycoproteins that create binding sites for cell–cell and cell–extracellular matrix adhesion. CAMs, expressed on vascular endothelium and leucocytes subsequent to proinflammatory stimuli, mediate the tethering, rolling, adhesion to the endothelium and transmigration of recruited leucocytes into the subendothelial layer. In atherosclerosis, the three CAM families of importance are selectins, the immunoglobulin superfamily and integrins.

Selectins

Selectins are involved in inflammatory responses, such as atherosclerosis, and consist of three different types: E-selectin, expressed on endothelial cells; L-selectin, expressed on leucocytes; and P-selectin, expressed on platelets and endothelial cells.

Significantly higher concentrations of E-, L- and P-selectins have been reported in LEAD patients compared with controls.^{17,22,23} In addition, higher plasma P-selectin was associated with increased LEAD risk in MESA.⁵¹ However, the results regarding selectins in LEAD are not consistent.⁵²

Intercellular Adhesion Molecule-1 and Vascular Cell Adhesion Molecule-1

Soluble intercellular adhesion molecule (sICAM-1), part of the immunoglobulin superfamily, is mainly expressed on endothelial cells and leucocytes, whereas soluble vascular cell adhesion molecule-1 (sVCAM-1) is restricted to vascular endothelial cells, where it mediates leucocyte–endothelium adhesion and promotes signal transduction between adhered cells.

Several studies have demonstrated higher sICAM-1 and/or sVCAM-1 concentrations in LEAD patients compared with healthy controls, and these adhesion molecules have been proposed as suitable for the detection of LEAD.^{17,21,23,24,53} When investigating biomarker levels in relation to exercise, VCAM-1 and ICAM-1 concentrations were higher in LEAD patients both before and after treadmill exercise.²⁴ ICAM-1, but not VCAM-1, was independently associated with an increased risk of progressing to symptomatic LEAD in the prospective Physician's Health Study that included apparently healthy middle-aged men.⁵⁴ These results in men were later reproduced in both women and men in the Edinburgh Artery Study and in women exclusively in the Women's Health Study.^{25,55} In addition, higher sICAM-1 and sVCAM-1 concentrations were associated with reduced walking ability in some studies, whereas there was no association between sVCAM-1 and walking capacity in another study.^{21,26,28} However, sVCAM-1 was associated with reduced calf muscle area and strength in LEAD.²⁹ Regarding cardiovascular risk, high sVCAM-1 concentrations predicted increased risk in individuals with LEAD and improved the prognostic value of ABI.⁵⁶

Other Circulating Inflammatory Biomarkers

Neopterin, synthesised by activated macrophages upon interferon- γ stimulation, possesses pro-oxidant properties and is a marker of macrophage activity and inflammation in atherosclerosis. Neopterin concentrations have been demonstrated to be higher in asymptomatic and symptomatic LEAD patients than in healthy controls and to be negatively correlated with ABI.^{23,34,57} In addition, neopterin was found to be an effective predictor of LEAD.³⁴

Growth differentiation factor (GDF)-15 is involved in the regulation of cell growth, repair and apoptosis. GDF-15 is constitutively expressed in the reproductive organs, although its expression can be swiftly induced in other cell types by proinflammatory cytokines, such as IL-1 β and TNF- α . Several studies have demonstrated GDF-15 to be a biomarker of acute coronary syndrome and mortality in coronary artery disease (CAD).⁵⁸ In addition, GDF-15 concentrations are stable over time in CAD patients, suggesting that it is a marker of chronic disease.⁵⁸ In two cohorts with asymptomatic and symptomatic LEAD, including CLTI patients, strong correlations were demonstrated between circulating GDF-15 and future lower extremity amputation and all-cause mortality.⁵⁹ Moreover, GDF-15 predicted these outcomes equally efficiently as the combination of nine traditional vascular risk factors.⁵⁹ Other studies have confirmed that GDF-15 may be an effective predictor of all-cause mortality in LEAD patients.^{60,61} Thus, circulating GDF-15 could be of value in predicting which CLTI patients would benefit from intensified treatment and/or surgical intervention.

Markers of Oxidative Stress

Oxidative stress is one of the pathogenetic mechanisms underpinning atherosclerosis, with the production of reactive oxygen species (ROS) and reduction in nitric oxide (NO) being associated with endothelial dysfunction. In LEAD, dysfunctional mitochondria mediate ROS production

in lower extremity muscles.⁶² The short ROS half-life prevents direct measurement, thus more stable molecular targets of ROS can function as indirect oxidative stress markers. Markers of oxidative stress, such as asymmetric dimethylarginine (ADMA), and 8-hydroxy-2-deoxyguanosine (8-OHdG), have been seen to be elevated in LEAD.^{63,64} Moreover, in a case-control study, men and women with LEAD had lower serum concentrations of NO metabolites and higher concentrations of the main producer of ROS, the NADPH oxidase 2-derived peptide (NOX2-dp) compared with controls.⁶⁵ In fact, NOX2-dp exhibited a negative correlation with ABI.⁶⁵ Levels of the oxidative stress marker 8-OHdG were also inversely correlated with markers of NO generation, indicating a connection between NO shortage and oxidative stress in LEAD.⁶⁴ Furthermore, 8-OHdG levels were correlated negatively with walking capacity.⁶⁴

Asymmetric Dimethylarginine

ADMA is the natural inhibitor of NO synthase, an enzyme catalysing NO production, by affecting the NO precursor arginine. Thus, ADMA is a potential marker of endothelial dysfunction, and several studies have demonstrated associations between elevated circulating ADMA concentrations, traditional cardiovascular risk factors and an increased risk of CVD.⁶⁶ Accordingly, plasma ADMA concentrations were higher in LEAD patients than in controls in a case-control study.⁶³ In another case-control study, ADMA concentrations did not differ between patients with CLTI, IC and healthy controls, whereas the ratio between arginine and ADMA was lower in CLTI patients than in IC patients and healthy controls.⁶⁷ Consistent data from prospective cohorts of asymptomatic or symptomatic LEAD patients have shown that increased plasma ADMA concentrations predict future CVE and both cardiovascular and all-cause mortality.^{68–71} Taken together, the available data suggest that ADMA may be a promising biomarker above all for detecting morbidity and mortality in LEAD patients.

Homocysteine

Circulating homocysteine (Hcy) mediates endothelial dysfunction when elevated. Moreover, there is evidence that Hcy potentiates the production of ROS.⁷² Approximately 30% of individuals with LEAD had increased circulating Hcy concentrations, compared with 1% in the general population.⁸ In addition, high Hcy concentrations were associated with worse functional outcome in LEAD.²⁶

Markers of the Coagulation Cascade

With endothelial dysfunction and subsequent inflammation and atherosclerosis, the local haemostatic balance is shifted towards a procoagulant state accompanied by resolving fibrinolysis, which is more distinct in LEAD than in CAD.⁷³

Procoagulant Markers

In both the COMPASS and VOYAGER trials, studying prophylactic treatment with the combination of aspirin and a low dose of the oral anticoagulant rivaroxaban in individuals with LEAD, a reduction in adverse lower limb events, cardiovascular death, MI and stroke was seen, corroborating the central role for a prothrombotic state in the pathology and complications of LEAD.^{74–76}

Plasminogen Activator Inhibitor-1

The serine protease inhibitor plasminogen activator inhibitor (PAI)-1 prevents fibrinolysis and heightens the hypercoagulable state by inhibiting tissue plasminogen activator (tPA). In case-control studies, PAI-1 concentrations were higher both at rest and after exercise in LEAD patients compared with healthy controls.^{17,77–79}

Thrombin Activation

Circulating concentrations of thrombin fragment 1+2 (F1+2) and thrombin–antithrombin III complex (TAT) are specific and sensitive markers of thrombin generation and have been seen to be higher in LEAD patients compared with controls.⁸⁰ In addition, CLTI patients have even higher TAT concentrations than IC patients.⁷⁹ In a case-control study of CLTI patients scheduled for revascularisation, CLTI patients preoperatively exhibited a prothrombotic condition together with fibrinolysis mirrored by increased TAT and fibrinogen and enhanced tPA and D-dimer compared with controls.⁷³ Immediately after reperfusion, F1+2 and TAT had further increased as a sign of thrombin generation, whereas fibrinogen concentrations were decreased.⁷³ This prothrombotic and fibrinolytic state persisted throughout the first postoperative month. In another study, subjects with LEAD exhibited increases in TAT and thrombin formation after exercise.⁷⁷

Platelet-Activating Factors

Tissue Factor and von Willebrand Factor

Tissue factor (TF) and von Willebrand factor (vWF) are part of the initial steps of the coagulation cascade, with the damaged integrity of the endothelium exposing TF to coagulation factors (F) VII and FVIIa and vWF promoting platelet adhesion.

Concentrations of both TF antigen and vWF were higher in individuals with LEAD compared with non-LEAD controls, and TF antigen concentrations were higher in CLTI than in other stages of LEAD.^{78,79} Moreover, subjects with LEAD exhibited a general platelet-activating state with elevated concentrations of platelet factor 4, sVCAM-1 and P-selectin.⁸⁰

Fibrinogen

Fibrinogen is an acute-phase reactant regulated by IL-6 and a marker of inflammation. In addition, fibrinogen stimulates platelet aggregation and is converted to fibrin by thrombin. Fibrinogen concentrations have been reported to be higher in individuals with LEAD than in healthy controls.^{33,77–79} In some studies, circulating fibrinogen predicted LEAD, including IC.^{35,81–83} Moreover, fibrinogen concentrations increased with the severity of LEAD.⁷⁹ High fibrinogen concentrations were also associated with the risk of fatal CVE and predicted mortality in LEAD.^{43,84} Fibrinogen was higher in CLTI patients undergoing infrainguinal bypass compared with controls.⁷³ In the same study, fibrinogen levels decreased immediately after reperfusion, possibly mirroring augmented thrombin-mediated conversion into fibrin.⁷³

Markers of Fibrinolysis

D-Dimer

D-dimer, a protein fragment arising from dissolving blood clots, is an indirect marker of fibrinolysis and is thus associated with the presence of venous and arterial thrombosis.

Circulating D-dimer concentrations were higher in subjects with LEAD than in healthy controls before and after treadmill exercise.^{17,77–79} Moreover, data are available regarding associations between increased D-dimer concentrations and both the presence and severity of LEAD.^{33,79} High D-dimer concentrations have been shown to be associated with poor calf muscle characteristics and inferior functional capacity.^{26,29,48} In addition, increased D-dimer concentrations in LEAD predicted CVE risk and mortality.^{43,85,86} In CLTI patients admitted for revascularisation, active fibrinolysis mirrored by enhanced D-dimer levels was seen before intervention and persisted in the first month after the intervention.⁷³ Despite plentiful research demonstrating the association between D-dimer and LEAD, there are conflicting data. In the prospective Edinburgh

Artery Study, elevated D-dimer was associated with LEAD progression, although the association was not independent of the IL-1/IL-6/CRP pathway, and it was demonstrated that individuals with concomitant increases in D-dimer and IL-6 experienced the largest deterioration.⁸⁷ Moreover, in a prospective study of individuals with LEAD followed for a median of 3 years, baseline D-dimer concentrations were neither associated with the risk of progression of LEAD nor with incident CVE, except for an increased risk of MI.³⁹

Tissue Plasminogen Activator

tPA is a protease present on vascular endothelial cells and is active in the conversion of plasminogen to plasmin, mediating the dissolution of blood clots. Circulating concentrations of tPA antigen were increased in LEAD compared with healthy controls, and higher in patients with more severe disease.^{77,79} Subjects with LEAD had higher tPA antigen plasma concentrations at rest and after treadmill exercise.⁷⁷ CLTI patients scheduled for revascularisation exhibited a prothrombotic state with high TAT and active fibrinolysis mirrored by enhanced tPA and D-dimer levels.⁷³

Cardiac Biomarkers

Increased concentrations of N-terminal pro B-type natriuretic peptide (NT-proBNP), a marker of cardiac failure and myocardial ischaemia, have been reported in subjects with LEAD and, together with copeptin, were associated with the incidence of LEAD during long-term follow-up.^{88,89} In the ARIC prospective cohort study, NT-proBNP and high-sensitivity troponin T (hsTnT), a marker of acute MI, were found to be predictive of incident LEAD.⁹⁰ Moreover, high NT-proBNP and hsTnT independently predicted increased all-cause mortality in LEAD.⁴³ In addition, hsTnT, but not carotid intima–media thickness or ABI, was predictive of reduced survival rate in a prospective LEAD cohort.⁹¹ Conversely, NT-proBNP was found to independently predict all-cause mortality after a 5-year follow-up in symptomatic LEAD patients,⁹² consistent with what has previously been demonstrated in heart failure.⁹³

Markers of Lipid Metabolism

Oxidised LDL (oxLDL) possesses proatherogenic properties and, compared with healthy controls, levels of total cholesterol, LDL, oxLDL and oxLDL antibodies were higher in subjects with LEAD.⁹⁴ In addition, oxLDL levels were positively correlated with total cholesterol and LDL.

Lipoprotein(a)

Lipoprotein(a), or Lp(a), is an LDL-like particle bound to an apolipoprotein B100 protein. Epidemiological studies and Mendelian randomisation (MR) analyses have demonstrated the association between Lp(a) and atherosclerotic CVD. Lp(a) concentrations and a single nucleotide polymorphism in its encoding gene *LPA* were also associated with LEAD, indicating causality between Lp(a) and LEAD.⁹⁵ Circulating Lp(a) was an independent predictor of LEAD.^{96,97} In addition, Lp(a) concentrations were positively correlated with disease severity, total cholesterol, LDL and apolipoprotein B.⁹⁶

Lipoprotein-Associated Phospholipase A₂

Lipoprotein-associated phospholipase A₂ (Lp-PLA₂) contributes to oxLDL modification, one of the earliest steps in the atherosclerosis process. Lp-PLA₂ binds to LDL particles in the circulation and is expressed by macrophages in atherosclerotic lesions. Lp-PLA₂ activity and mass were, either alone or together with CRP, predictors of LEAD risk in two population-based cohorts of middle-aged or elderly individuals.^{98,99} These results were reproduced in a prospective cohort study analysing the risk of future LEAD-related hospitalisation associated with high Lp-PLA₂ levels

in subjects free of LEAD at baseline.¹⁰⁰ In a hospital-based Chinese cross-sectional study, Lp-PLA₂ concentrations were associated with the prevalence of LEAD independent of inflammatory markers such as hsCRP, Hcy and fibrinogen.¹⁰¹ Conversely, in a population-based US multi-ethnic cohort study of 45–84 year olds, Lp-PLA₂ activity and mass were not associated with an increased risk of incident LEAD.¹⁰² Thus, results are conflicting regarding Lp-PLA₂ as a predictor of LEAD.

Adiponectin

Adiponectin is an adipokine active in glucose and fatty acid metabolism that enhances insulin sensitivity and has anti-inflammatory and antioxidative properties. Lower adiponectin concentrations have been reported in men than in women, although adiponectin concentrations are lower in women with than without metabolic syndrome.¹⁰³ These sex differences in anti-inflammatory adiponectin may contribute, in part, to higher CVD risk in men and women with metabolic syndrome. In relation to LEAD, adiponectin concentrations were lower in women who developed LEAD than in those that did not.¹⁰⁴ In line with this finding in women, high adiponectin concentrations were associated with a decreased risk of developing symptomatic LEAD in men.¹⁰⁵ Moreover, in men with symptomatic LEAD, but not in their female counterparts, low adiponectin concentrations were associated with a higher risk of non-fatal CVE.¹⁰⁶

Tissue Remodelling and Angiogenesis Markers

Matrix Metalloproteinases

Matrix metalloproteinases (MMPs) are proteases with the ability to degrade extracellular matrix. MMPs are secreted by inflammatory cells and are active in vascular remodelling during the development and progression of atherosclerosis, including plaque rupture. MMP2 and MMP9 are also involved in the activation and regulation of platelet aggregation. In LEAD, concentrations of MMP2 and MMP9, and to some extent MMP3, were increased compared with healthy controls, and the progression and severity of LEAD have been associated with high concentrations of MMP2 and MMP9.^{17,22,23,107}

Galectin-3: Marker of Fibrosis and Calcification

Galectin-3 is induced by oxidative stress and is involved in inflammation, angiogenesis and fibrosis via mediation of cell–cell and cell–matrix interactions. Galectin-3 is also involved in macrophage maturation and has been associated with atherosclerotic CVD and LEAD.¹⁰⁸ Levels of galectin-3 predicted incident LEAD and were higher in subjects with pathological ABI than in individuals without LEAD.^{36,109} Moreover, in LEAD patients, high galectin-3 concentrations were associated with an increased risk of cardiovascular mortality.¹⁰⁸

Markers of Angiogenesis

Because angiogenesis is a physiological response to tissue ischaemia, circulating angiogenic factors may be relevant surrogates for disease severity via the increased production of angiogenic mitogens such as vascular endothelial growth factor (VEGF)-A and angiopoietin-2.

Vascular Endothelial Growth Factor-A

Increased circulating VEGF-A concentrations mirror pronounced angiogenesis. VEGF-A concentrations were higher in LEAD patients than in non-LEAD controls, and concentrations increased with the severity of the disease.^{110–112} Contrary to these findings, VEGF-A concentrations were lower in LEAD cases in a case-control study using controls with other cardiovascular risk factors and comorbidities other than LEAD.⁴⁹ This finding highlights the difficulties with biomarkers mirroring processes active in systemic diseases. In relation to exercise-dependent changes in

symptomatic LEAD, VEGF-A increased after home-based or supervised exercise.^{113,114}

Angiopoietin-2 and the Tie2 Receptor

Together with VEGF-A, angiopoietin-2 stimulates neovascularisation. The cognate angiopoietin-2 receptor is the Tie2 receptor expressed mainly on the vascular endothelium.

In LEAD patients, circulating angiopoietin-2 and the soluble Tie2 receptor were increased compared with healthy controls.¹¹¹ Moreover, no difference was seen in plasma angiopoietin-2 concentrations between IC and CLTI patients, whereas concentrations of the soluble Tie2 receptor were higher in CLTI than IC patients.¹¹¹ In a prospective cohort of symptomatic LEAD, increased angiopoietin-2 was found to be independently associated with an enhanced risk of major adverse CVE and all-cause mortality.¹¹⁵

Overarching Inflammatory, Coagulation and Metabolic Pathway Biomarkers

Extracellular Vesicles

Circulating extracellular vesicles (EVs), including exosomes, microparticles and apoptotic bodies, are secreted by different types of cells upon stimulation, and carry nucleic acids, proteins, lipids and metabolites from the host cell, thereby mediating vascular homeostasis and intercellular communication. The concentrations and types of circulating EVs in the blood and the effects mediated by EVs differ depending on the stimulus causing their secretion. In LEAD, platelet-derived EVs are by far the most common type, followed by EVs from endothelial cells, erythrocytes and leucocytes.¹¹⁶ Case-control studies have demonstrated higher levels of platelet-derived EVs in individuals with LEAD than in healthy controls, although one study did not show any difference between the groups.^{116–118} In addition, levels of platelet-derived EVs were correlated with LEAD severity.¹¹⁹ Circulating levels of endothelial cell-derived EVs, especially when containing proinflammatory monomeric CRP, were higher in LEAD.¹²⁰

Circulating MicroRNAs

MicroRNAs (miRNAs) are small, non-coding and single-stranded RNAs originating from EVs.¹²¹ MiRNAs are able to control gene expression at the post-transcriptional level, thereby inhibiting protein synthesis. MiRNAs are highly stable and expressed in a disease-specific manner, and therefore suitable as diagnostic biomarkers for CVD.¹²² A small case-control study performing transcriptomics on peripheral blood cells identified a group of miRNAs (miR-16, miR-363 and miR-15b) that had previously been associated with vascular pathophysiology as predictors of LEAD with outstanding diagnostic accuracy.¹²³ In a larger case-control study, circulating (serum) concentrations of miRNAs (miR-130a, miR-27b, miR-210) were associated with LEAD.¹²⁴ Moreover, in an all-male aortic aneurysm case-control study, four circulating miRNAs (let-7e, miR-15a, miR-196b and miR-411) were found to be associated both with aortic aneurysm and LEAD.¹²⁵

Perspectives

Neither diagnostic nor prognostic circulating biomarkers for LEAD are used clinically today. When suspicion is raised, a diagnosis of LEAD with ABI is cheap and easy with a trained operator. In primary care, where sometimes both equipment and skills to measure ABI are lacking, circulating biomarkers would add value to screening for LEAD. Notwithstanding, disease-specific biomarkers predictive of incident LEAD are challenging to find because the organ-specific features of LEAD, primarily hypoperfusion with subsequent ischaemic tissue damage, are

not present until later stages of the disease. Conversely, biomarkers indicating general atherosclerosis could function as a first-line screening marker for LEAD accompanied by ABI when appropriate. Later in the course of the disease, and certainly in symptomatic patients, organ-specific prognostic biomarkers may be of value together with imaging to stratify risk and evaluate the benefit of preventive medical treatment and/or interventional procedures. In addition, the association between LEAD and increased cardiovascular risk defines an important role for assessment of LEAD in personalised medicine⁵. Today, preventive medication for patients with LEAD is limited to drugs targeting traditional cardiovascular risk factors. In the development of novel, specific therapeutic targets for patients with LEAD, biomarkers representing pathways discussed in this review may be used to identify individuals suitable for treatment and to monitor the treatment effects of pharmacotherapies.

The traditional deductive approach in biomarker research has been challenged in recent years by inductive strategies using unbiased, large-scale, high-throughput plasma proteomic profiling. Such analyses demand large cohorts with enough statistical power to demonstrate associations and extensive bioinformatics to process large amounts of data. Conversely, new mechanisms and possible molecular targets can be discovered in corners previously not scrutinised.

Recent developments also highlight the future need for LEAD biomarkers. Using MR analysis and randomised controlled trials, the causal association between procoagulant factors and LEAD has been strengthened, and followed by the introduction of low-dose anticoagulant drugs in addition to antiplatelet therapy.^{126,75,76,2} However, biomarkers guiding the selection of patients for treatment and monitoring treatment effects are still lacking. The association between LEAD and inflammatory markers from the IL-1 β /IL-6/CRP pathway displays consistent results in epidemiological studies

and, in a recent randomised controlled trial, inhibition of the IL-1 β /IL-6/CRP pathway with canakinumab dampened LEAD progression.¹²⁷ In addition, positive associations between Lp(a) and LEAD in cohort and case-control studies, together with a demonstrated causal association between apolipoprotein B and LEAD in MR analyses, point to lipids as potential causal targets.¹²⁸ With this in mind, the need for biomarkers in selecting patients for targeted treatment and the identification of individuals at risk of future hospitalisation for LEAD is pivotal. In such a setting, Lp-PLA₂ holds great promise.¹⁰⁰

Conclusion

The pathophysiology underpinning LEAD is multifactorial and, together with the effective diagnostic and predictive tools already at hand, will likely require a multiple-biomarker approach to provide incremental predictive value. Moreover, the impact of each causal pathway differs in individual patients, demanding biomarkers to evaluate the magnitude of the respective pathway in risk stratification assessments and to guide clinicians in which patients to treat and how to treat them. Many of the biomarkers presented in this review are associated with several inflammatory and/or atherosclerosis-related conditions, complicating interpretation. In this setting, the relatively new field of disease-specific circulating miRNAs as diagnostic and prognostic biomarkers is promising as analyses become cheaper and easier to use.¹²²

Due to the late onset of symptoms and lack of diagnostic circulating biomarkers, LEAD patients are often diagnosed in late stages of the disease and, when diagnosed, biomarkers to guide the selection of patients for treatment and the monitoring of treatment effects are missing. However, the field is expanding, with many promising biomarkers continuously being investigated, together with potential targets for pharmacological treatment. □

- Aboyans V, Ricco JB, Bartelink MEL, et al. 2017 ESC guidelines on the diagnosis and treatment of peripheral arterial diseases, in collaboration with the European Society for Vascular Surgery (ESVS). *Eur Heart J* 2018;39:763–816. <https://doi.org/10.1093/eurheartj/ehx095>; PMID: 28886620.
- Frank U, Nikol S, Belch J, et al. ESVM guideline on peripheral arterial disease. *Vasa* 2019;48(Suppl 102):1–79. <https://doi.org/10.1024/0301-1526/a000834>; PMID: 31789115.
- Fowkes FGR, Rudan D, Rudan I, et al. Comparison of global estimates of prevalence and risk factors for peripheral artery disease in 2000 and 2010: a systematic review and analysis. *Lancet* 2013;382:1329–40. [https://doi.org/10.1016/S0140-6736\(13\)61249-0](https://doi.org/10.1016/S0140-6736(13)61249-0); PMID: 23915883.
- McDermott MM, Greenland P, Liu K, et al. Leg symptoms in peripheral arterial disease: associated clinical characteristics and functional impairment. *JAMA* 2001;286:1599–606. <https://doi.org/10.1001/jama.286.13.1599>; PMID: 11585483.
- Subherwal S, Patel MR, Kober L, et al. Peripheral artery disease is a coronary heart disease risk equivalent among both men and women: results from a nationwide study. *Eur J Prev Cardiol* 2015;22:317–25. <https://doi.org/10.1177/2047487313519344>; PMID: 24398369.
- Criqui MH, Langer RD, Fronek A, et al. Mortality over a period of 10 years in patients with peripheral arterial disease. *N Engl J Med* 1992;326:381–6. <https://doi.org/10.1056/NEJM199202063260605>; PMID: 1729621.
- Sigvant B, Lundin F, Wahlberg E. The risk of disease progression in peripheral arterial disease is higher than expected: a meta-analysis of mortality and disease progression in peripheral arterial disease. *Eur J Vasc Endovasc Surg* 2016;51:395–403. <https://doi.org/10.1016/j.ejvs.2015.10.022>; PMID: 26777541.
- Norgren L, Hiatt WR, Dormandy JA, et al. Inter-society consensus for the management of peripheral arterial disease (TASC II). *J Vasc Surg* 2007;45(Suppl S):S5–67. <https://doi.org/10.1016/j.jvs.2006.12.037>; PMID: 17223489.
- Duff S, Mafliios MS, Bhonsule P, Hasegawa JT. The burden of critical limb ischemia: a review of recent literature. *Vasc Health Risk Manag* 2019;15:187–208. <https://doi.org/10.2147/VHRM.S209241>; PMID: 31308682.
- Cambou JP, Aboyans V, Constans J, et al. Characteristics and outcome of patients hospitalised for lower extremity peripheral artery disease in France: the COPART registry. *Eur J Vasc Endovasc Surg* 2010;39:577–85. <https://doi.org/10.1016/j.ejvs.2010.02.009>; PMID: 20303804.
- Scholz D, Cai WJ, Schaper W. Arteriogenesis, a new concept of vascular adaptation in occlusive disease. *Angiogenesis* 2001;4:247–57. <https://doi.org/10.1023/A:1016094004084>; PMID: 12197469.
- Pipinos II, Judge AR, Selsby JT, et al. The myopathy of peripheral arterial occlusive disease: part 1. Functional and histomorphological changes and evidence for mitochondrial dysfunction. *Vasc Endovasc Surg* 2007;41:481–9. <https://doi.org/10.1177/1538574407311106>; PMID: 18166628.
- Pipinos II, Judge AR, Selsby JT, et al. The myopathy of peripheral arterial occlusive disease: part 2. Oxidative stress, neuropathy, and shift in muscle fiber type. *Vasc Endovasc Surg* 2008;42:101–12. <https://doi.org/10.1177/1538574408315995>; PMID: 18390972.
- Bhat HK, Hiatt WR, Hoppel CL, Brass EP. Skeletal muscle mitochondrial DNA injury in patients with unilateral peripheral arterial disease. *Circulation* 1999;99:807–12. <https://doi.org/10.1161/01.CIR.99.6.807>; PMID: 9989967.
- FDA-NIH Biomarker Working Group. *BEST (Biomarkers, EndpointS, and other Tools) Resource*. Silver Spring, MD, Bethesda, MD: Food and Drug Administration, National Institutes of Health, 2016. <https://www.ncbi.nlm.nih.gov/books/NBK326791> (accessed 1 March 2022).
- Fowkes FG, Aboyans V, Fowkes FJ, et al. Peripheral artery disease: epidemiology and global perspectives. *Nat Rev Cardiol* 2017;14:156–70. <https://doi.org/10.1038/nrcardio.2016.179>; PMID: 27853158.
- Engelberger RP, Limacher A, Kucher N, et al. Biological variation of established and novel biomarkers for atherosclerosis: results from a prospective, parallel-group cohort study. *Clin Chim Acta* 2015;447:16–22. <https://doi.org/10.1016/j.cca.2015.05.003>; PMID: 25979692.
- Libby P, Ridker PM, Hansson GK, Leducq Transatlantic Network on Atherothrombosis. Inflammation in atherosclerosis: from pathophysiology to practice. *J Am Coll Cardiol* 2009;54:2129–38. <https://doi.org/10.1016/j.jacc.2009.09.009>; PMID: 19942084.
- Murabito JM, Keyes MJ, Guo CY, et al. Cross-sectional relations of multiple inflammatory biomarkers to peripheral arterial disease: the Framingham offspring study. *Atherosclerosis* 2009;203:509–14. <https://doi.org/10.1016/j.atherosclerosis.2008.06.031>; PMID: 18701106.
- von der Thüsen JH, Kuiper J, van Berkel TJ, Biessen EA. Interleukins in atherosclerosis: molecular pathways and therapeutic potential. *Pharmacol Rev* 2003;55:133–66. <https://doi.org/10.1124/pr.55.1.5>; PMID: 12615956.
- Pande RL, Brown J, Buck S, et al. Association of monocyte tumor necrosis factor α expression and serum inflammatory biomarkers with walking impairment in peripheral artery disease. *J Vasc Surg* 2015;61:155–61. <https://doi.org/10.1016/j.jvs.2014.06.116>; PMID: 25095746.
- Signorelli SS, Anzaldi M, Fiore V, et al. Patients with unrecognized peripheral arterial disease (PAD) assessed by ankle-brachial index (ABI) present a defined profile of proinflammatory markers compared to healthy subjects. *Cytokine* 2012;59:294–8. <https://doi.org/10.1016/j.cyt.2012.04.038>; PMID: 22595645.
- Signorelli SS, Anzaldi M, Libra M, et al. Plasma levels of inflammatory biomarkers in peripheral arterial disease: results of a cohort study. *Angiology* 2016;67:870–4. <https://doi.org/10.1177/0003319716633339>; PMID: 26888895.
- Signorelli SS, Mazarino MC, Di Pino L, et al. High circulating levels of cytokines (IL-6 and TN α), adhesion molecules (VCAM-1 and ICAM-1) and selectins in patients with peripheral arterial disease at rest and after a treadmill test. *Vasc Med* 2003;8:15–9. <https://doi.org/10.1191/1358863x03vm4660a>; PMID: 12866607.
- Tzoulaki I, Murray GD, Lee AJ, et al. C-reactive protein, interleukin-6, and soluble adhesion molecules as predictors of progressive peripheral atherosclerosis in the general population: Edinburgh Artery study. *Circulation* 2005;112:976–83. <https://doi.org/10.1161/CIRCULATIONAHA.104.513085>; PMID: 16087797.
- McDermott MM, Liu K, Ferrucci L, et al. Circulating blood markers and functional impairment in peripheral arterial disease. *J Am Geriatr Soc* 2008;56:1504–10. <https://doi.org/10.1111/j.1532-5415.2008.01797.x>; PMID: 18662216.

27. Nylaende M, Kroese A, Stranden E, et al. Markers of vascular inflammation are associated with the extent of atherosclerosis assessed as angiographic score and treadmill walking distances in patients with peripheral arterial occlusive disease. *Vasc Med* 2006;11:21–8. <https://doi.org/10.1191/1358863x06vm662oa>; PMID: 16669409.
28. McDermott MM, Liu K, Ferrucci L, et al. Relation of interleukin-6 and vascular cellular adhesion molecule-1 levels to functional decline in patients with lower extremity peripheral arterial disease. *Am J Cardiol* 2011;107:1392–8. <https://doi.org/10.1016/j.amjcard.2011.01.007>; PMID: 21371679.
29. McDermott MM, Ferrucci L, Guralnik JM, et al. Elevated levels of inflammation, D-dimer, and homocysteine are associated with adverse calf muscle characteristics and reduced calf strength in peripheral arterial disease. *J Am Coll Cardiol* 2007;50:897–905. <https://doi.org/10.1016/j.jacc.2007.05.017>; PMID: 17719478.
30. Biscetti F, Ferraro PM, Hiatt WR, et al. Inflammatory cytokines associated with failure of lower-extremity endovascular revascularization (LER): a prospective study of a population with diabetes. *Diabetes Care* 2019;42:1939–45. <https://doi.org/10.2337/dci19-0408>; PMID: 31371431.
31. Guo S, Zhang Z, Wang L, et al. Six-month results of stenting of the femoropopliteal artery and predictive value of interleukin-6: comparison with high-sensitivity C-reactive protein. *Vascular* 2020;28:715–21. <https://doi.org/10.1177/1708538120921005>; PMID: 32408853.
32. Beckman JA, Preis O, Ridker PM, Gerhard-Herman M. Comparison of usefulness of inflammatory markers in patients with versus without peripheral arterial disease in predicting adverse cardiovascular outcomes (myocardial infarction, stroke, and death). *Am J Cardiol* 2005;96:1374–8. <https://doi.org/10.1016/j.amjcard.2005.07.041>; PMID: 16275181.
33. Unlü Y, Karapolat S, Karaca Y, Kizilunç A. Comparison of levels of inflammatory markers and hemostatic factors in the patients with and without peripheral arterial disease. *Thromb Res* 2006;117:357–64. <https://doi.org/10.1016/j.thromres.2005.03.019>; PMID: 15890391.
34. Ceasosvich A, Sorodoc V, Onofrei Aursulesi V, et al. Biomarker utility for peripheral artery disease diagnosis in real clinical practice: a prospective study. *Diagnostics (Basel)* 2020;10:723. <https://doi.org/10.3390/diagnostics10090723>; PMID: 32962217.
35. Ridker PM, Stampfer MJ, Rifai N. Novel risk factors for systemic atherosclerosis: a comparison of C-reactive protein, fibrinogen, homocysteine, lipoprotein(a), and standard cholesterol screening as predictors of peripheral arterial disease. *JAMA* 2001;285:2481–5. <https://doi.org/10.1001/jama.285.19.2481>; PMID: 11368701.
36. Ding N, Yang C, Ballew SH, et al. Fibrosis and inflammatory markers and long-term risk of peripheral artery disease: the ARIC study. *Arterioscler Thromb Vasc Biol* 2020;40:2322–31. <https://doi.org/10.1161/ATVBAHA.120.314824>; PMID: 32698688.
37. Van Der Meer IM, De Maat MP, Hak AE, et al. C-reactive protein predicts progression of atherosclerosis measured at various sites in the arterial tree: the Rotterdam study. *Stroke* 2002;33:2750–5. <https://doi.org/10.1161/01.STR.0000044168.00485.02>; PMID: 12468765.
38. Aboyans V, Criqui MH, Denenberg JO, et al. Risk factors for progression of peripheral arterial disease in large and small vessels. *Circulation* 2006;113:2623–9. <https://doi.org/10.1161/CIRCULATIONAHA.105.608679>; PMID: 16735675.
39. Musicant SE, Taylor LM Jr, Peters D, et al. Prospective evaluation of the relationship between C-reactive protein, D-dimer and progression of peripheral arterial disease. *J Vasc Surg* 2006;43:772–80. <https://doi.org/10.1016/j.jvs.2005.12.051>; PMID: 16616235.
40. Høgh AL, Joensen J, Lindholt JS, et al. C-Reactive protein predicts future arterial and cardiovascular events in patients with symptomatic peripheral arterial disease. *Vasc Endovasc Surg* 2008;42:341–7. <https://doi.org/10.1177/1538574408316138>; PMID: 18458051.
41. Rossi E, Biasucci LM, Citterio F, et al. Risk of myocardial infarction and angina in patients with severe peripheral vascular disease: predictive role of C-reactive protein. *Circulation* 2002;105:800–3. <https://doi.org/10.1161/hc0702.104126>; PMID: 11854118.
42. Vainas T, Stassen FR, de Graaf R, et al. C-Reactive protein in peripheral arterial disease: relation to severity of the disease and to future cardiovascular events. *J Vasc Surg* 2005;42:243–51. <https://doi.org/10.1016/j.jvs.2005.03.060>; PMID: 16102622.
43. Kremers B, Wübbke L, Mees B, et al. Plasma biomarkers to predict cardiovascular outcome in patients with peripheral artery disease: a systematic review and meta-analysis. *Arterioscler Thromb Vasc Biol* 2020;40:2018–32. <https://doi.org/10.1161/ATVBAHA.120.314774>; PMID: 32640905.
44. Eldrup N, Sillesen H, Prescott E, Nordestgaard BG. Ankle brachial index, C-reactive protein, and central augmentation index to identify individuals with severe atherosclerosis. *Eur Heart J* 2006;27:316–22. <https://doi.org/10.1093/eurheartj/ehi644>; PMID: 16278227.
45. Singh TP, Morris DR, Smith S, et al. Systematic review and meta-analysis of the association between C-reactive protein and major cardiovascular events in patients with peripheral artery disease. *Eur J Vasc Endovasc Surg* 2017;54:220–33. <https://doi.org/10.1016/j.ejvs.2017.05.009>; PMID: 28666785.
46. Maksimovic M, Vlainjac H, Radak D, et al. Relationship between high-sensitivity C-reactive protein and risk factors in patients with peripheral arterial disease – a cross-sectional study. *Angiology* 2013;64:230–6. <https://doi.org/10.1177/0003319712440303>; PMID: 22499952.
47. Urbonaviciene G, Frystyk J, Flyvbjerg A, et al. Markers of inflammation in relation to long-term cardiovascular mortality in patients with lower-extremity peripheral arterial disease. *Int J Cardiol* 2012;160:89–94. <https://doi.org/10.1016/j.ijcard.2011.03.030>; PMID: 21463908.
48. McDermott MM, Greenland P, Green D, et al. D-Dimer, inflammatory markers, and lower extremity functioning in patients with and without peripheral arterial disease. *Circulation* 2003;107:3191–8. <https://doi.org/10.1161/01.CIR.0000074227.53616.CC>; PMID: 12810614.
49. Gardner AW, Parker DE, Montgomery PS, et al. Impaired vascular endothelial growth factor A and inflammation in patients with peripheral artery disease. *Angiology* 2014;65:683–90. <https://doi.org/10.1177/0003319713501376>; PMID: 24006146.
50. DePalma RG, Hayes VW, May PE, et al. Statins and biomarkers in claudicants with peripheral arterial disease: cross-sectional study. *Vascular* 2006;14:193–200. <https://doi.org/10.2310/6670.2006.00039>; PMID: 17026909.
51. Wassel CL, Berardi C, Pankow JS, et al. Soluble P-selectin predicts lower extremity peripheral artery disease incidence and change in the ankle brachial index: the Multi-Ethnic Study of Atherosclerosis (MESA). *Atherosclerosis* 2015;239:405–11. <https://doi.org/10.1016/j.atherosclerosis.2015.01.022>; PMID: 25682040.
52. Brevetti G, Schiano V, Chiariello M. Cellular adhesion molecules and peripheral arterial disease. *Vasc Med* 2006;11:39–47. <https://doi.org/10.1191/1358863x06vm645ra>; PMID: 16669412.
53. Edlinger C, Lichtenauer M, Wernly B, et al. Disease-specific characteristics of vascular cell adhesion molecule-1 levels in patients with peripheral artery disease. *Heart Vessels* 2019;34:976–83. <https://doi.org/10.1007/s00380-018-1315-1>; PMID: 30535754.
54. Pradhan AD, Rifai N, Ridker PM. Soluble intercellular adhesion molecule-1, soluble vascular adhesion molecule-1, and the development of symptomatic peripheral arterial disease in men. *Circulation* 2002;106:820–5. <https://doi.org/10.1161/01.CIR.0000025636.03561.EE>; PMID: 12176954.
55. Pradhan AD, Shrivastava S, Cook NR, et al. Symptomatic peripheral arterial disease in women: nontraditional biomarkers of elevated risk. *Circulation* 2008;117:823–31. <https://doi.org/10.1161/CIRCULATIONAHA.107.719369>; PMID: 18227386.
56. Silvestro A, Brevetti G, Schiano V, et al. Adhesion molecules and cardiovascular risk in peripheral arterial disease. Soluble vascular cell adhesion molecule-1 improves risk stratification. *Thromb Haemostasis* 2005;93:559–63. <https://doi.org/10.1160/TH04-07-0440>; PMID: 15735810.
57. Signorelli SS, Anzaldi M, Fiore V, et al. Neopterin: a potential marker in chronic peripheral arterial disease. *Mol Med Rep* 2013;7:1855–8. <https://doi.org/10.3892/mmr.2013.1407>; PMID: 23563241.
58. Wollert KC, Kempf T, Wallentin L. Growth differentiation factor 15 as a biomarker in cardiovascular disease. *Clin Chem* 2017;63:140–51. <https://doi.org/10.1373/clinchem.2016.255174>; PMID: 28062617.
59. De Haan JJ, Haitjema S, den Ruijter HM, et al. Growth differentiation factor 15 is associated with major amputation and mortality in patients with peripheral artery disease. *J Am Heart Assoc* 2017;6:e006225. <https://doi.org/10.1161/JAHA.117.006225>; PMID: 28855167.
60. Hsu LA, Wu S, Juang JJ, et al. Growth differentiation factor 15 may predict mortality of peripheral and coronary artery diseases and correlate with their risk factors. *Mediators Inflamm* 2017;2017:9398401. <https://doi.org/10.1155/2017/9398401>; PMID: 28798540.
61. Skau E, Henriksen E, Leppert J, et al. Targeted multiplex proteomics for prediction of all-cause mortality during long-term follow-up in outpatients with peripheral arterial disease. *Atherosclerosis* 2020;311:143–9. <https://doi.org/10.1016/j.atherosclerosis.2020.06.015>; PMID: 32711845.
62. Pipinos II, Judge AR, Zhu Z, et al. Mitochondrial defects and oxidative damage in patients with peripheral arterial disease. *Free Radic Biol Med* 2006;41:262–9. <https://doi.org/10.1016/j.freeradbiomed.2006.04.003>; PMID: 16814106.
63. Böger RH, Bode-Böger SM, Thiele W, et al. Biochemical evidence for impaired nitric oxide synthesis in patients with peripheral arterial occlusive disease. *Circulation* 1997;95:2068–74. <https://doi.org/10.1161/01.CIR.95.8.2068>; PMID: 9133517.
64. Loffredo L, Pignatelli P, Cangemi R, et al. Imbalance between nitric oxide generation and oxidative stress in patients with peripheral arterial disease: effect of an antioxidant treatment. *J Vasc Surg* 2006;44:525–30. <https://doi.org/10.1016/j.jvs.2006.05.023>; PMID: 16950429.
65. Loffredo L, Carnevale R, Cangemi R, et al. NOX2 up-regulation is associated with artery dysfunction in patients with peripheral artery disease. *Int J Cardiol* 2013;165:184–92. <https://doi.org/10.1016/j.ijcard.2012.01.069>; PMID: 22336250.
66. Dowsett L, Higgins E, Alanazi S, et al. ADMA: a key player in the relationship between vascular dysfunction and inflammation in atherosclerosis. *J Clin Med* 2020;9:3026. <https://doi.org/10.3390/jcm9093026>; PMID: 32962225.
67. Ismaeel A, Papoutsis E, Miserlis D, et al. The nitric oxide system in peripheral artery disease: connection with oxidative stress and biopterins. *Antioxidants (Basel)* 2020;9:590. <https://doi.org/10.3390/antiox9070590>; PMID: 32640613.
68. Mittermayer F, Krzyzanowska K, Exner M, et al. Asymmetric dimethylarginine predicts major adverse cardiovascular events in patients with advanced peripheral artery disease. *Arterioscler Thromb Vasc Biol* 2006;26:2536–40. <https://doi.org/10.1161/01.ATV.0000242801.38419.48>; PMID: 16931791.
69. Böger RH, Endres HG, Schwedhelm E, et al. Asymmetric dimethylarginine as an independent risk marker for mortality in ambulatory patients with peripheral arterial disease. *J Intern Med* 2011;269:349–61. <https://doi.org/10.1111/j.1365-2796.2010.02322.x>; PMID: 21175900.
70. Jud P, Hafner F, Verheyen N, et al. Homocysteine/ADMA ratio and homocysteine/SDMA ratio as independent predictors of cardiovascular mortality and cardiovascular events in lower extremity arterial disease. *Sci Rep* 2018;8:14197. <https://doi.org/10.1038/s41598-018-32607-8>; PMID: 30242192.
71. Staniszewska A, Rajagopalan S, Al-Shaheen A, et al. Increased levels of symmetric dimethylarginine are associated with all-cause mortality in patients with symptomatic peripheral arterial disease. *J Vasc Surg* 2015;61:1292–8. <https://doi.org/10.1016/j.jvs.2015.01.002>; PMID: 25776186.
72. Tyagi N, Sedoris KC, Steed M, et al. Mechanisms of homocysteine-induced oxidative stress. *Am J Physiol Heart Circ Physiol* 2005;289:H2649–56. <https://doi.org/10.1152/ajpheart.00548.2005>; PMID: 16085680.
73. Pärsson H, Holmberg A, Siegbahn A, Bergqvist D. Activation of coagulation and fibrinolytic systems in patients with CLI is not normalized after surgical revascularisation. *Eur J Vasc Endovasc Surg* 2004;27:186–92. <https://doi.org/10.1016/j.ejvs.2003.10.015>; PMID: 14718902.
74. Eikelboom JW, Connolly SJ, Bosch J, et al. Rivaroxaban with or without aspirin in stable cardiovascular disease. *N Engl J Med* 2017;377:1319–30. <https://doi.org/10.1056/NEJMoa1709118>; PMID: 28844192.
75. Anand SS, Bosch J, Eikelboom JW, et al. Rivaroxaban with or without aspirin in patients with stable peripheral or carotid artery disease: an international, randomised, double-blind, placebo-controlled trial. *Lancet* 2018;391:219–29. [https://doi.org/10.1016/S0140-6736\(17\)32409-1](https://doi.org/10.1016/S0140-6736(17)32409-1); PMID: 29132880.
76. Bonaca MP, Bauersachs RM, Anand SS, et al. Rivaroxaban in peripheral artery disease after revascularization. *N Engl J Med* 2020;382:1994–2004. <https://doi.org/10.1056/NEJMoa2000052>; PMID: 32222135.
77. Mustonen P, Lepäntalo M, Lassila R. Physical exertion induces thrombin formation and fibrin degradation in patients with peripheral atherosclerosis. *Arterioscler Thromb Vasc Biol* 1998;18:244–9. <https://doi.org/10.1161/01.ATV.18.2.244>; PMID: 9484989.
78. Lee AJ, MacGregor AS, Hau CM, et al. The role of haematological factors in diabetic peripheral arterial disease: the Edinburgh Artery study. *Br J Haematol* 1999;105:648–54. <https://doi.org/10.1046/j.1365-2141.1999.01382.x>; PMID: 10354125.
79. Wiecek R, Kulwas A, Rośc D. Implications of hemostasis disorders in patients with critical limb ischemia – an in-depth comparison of selected factors. *J Clin Med* 2020;9:659. <https://doi.org/10.3390/jcm9030659>; PMID: 32121363.
80. Zamzam A, Syed MH, Rand ML, et al. Altered coagulation profile in peripheral artery disease patients. *Vascular* 2020;28:368–77. <https://doi.org/10.1177/1708538120915997>;

- PMID: 32252612.
81. Wattanakit K, Folsom AR, Selvin E, et al. Risk factors for peripheral arterial disease incidence in persons with diabetes: the Atherosclerosis Risk In Communities (ARIC) study. *Atherosclerosis* 2005;180:389–97. <https://doi.org/10.1016/j.atherosclerosis.2004.11.024>; PMID: 15910867.
 82. Smith FB, Lee AJ, Hau CM, et al. Plasma fibrinogen, haemostatic factors and prediction of peripheral arterial disease in the Edinburgh Artery study. *Blood Coagul Fibrinolysis* 2000;11:43–50. <https://doi.org/10.1097/00001721-200011010-00005>; PMID: 10691098.
 83. Murabito JM, D'Agostino RB, Silbershatz H, Wilson WF. Intermittent claudication. A risk profile from the Framingham heart study. *Circulation* 1997;96:44–9. <https://doi.org/10.1161/01.CIR.96.1.44>; PMID: 9236415.
 84. Doweik L, Maca T, Schilling M, et al. Fibrinogen predicts mortality in high risk patients with peripheral artery disease. *Eur J Vasc Endovasc Surg* 2003;26:381–6. [https://doi.org/10.1016/S1078-5884\(03\)00340-X](https://doi.org/10.1016/S1078-5884(03)00340-X); PMID: 1451999.
 85. Vidula H, Tian L, Liu K, et al. Biomarkers of inflammation and thrombosis as predictors of near-term mortality in patients with peripheral arterial disease: a cohort study. *Ann Intern Med* 2008;148:85–93. <https://doi.org/10.7326/0003-4819-148-2-200801150-00003>; PMID: 18195333.
 86. McDermott MM, Liu K, Green D, et al. Changes in D-dimer and inflammatory biomarkers before ischemic events in patients with peripheral artery disease: the BRAVO study. *Vasc Med* 2016;21:12–20. <https://doi.org/10.1177/1358863X15617541>; PMID: 26647446.
 87. Tzoulaki I, Murray GD, Price JF, et al. Hemostatic factors, inflammatory markers, and progressive peripheral atherosclerosis: the Edinburgh Artery study. *Am J Epidemiol* 2006;163:334–41. <https://doi.org/10.1093/aje/kwj051>; PMID: 16357107.
 88. Falkensammer J, Frech A, Duschek N, et al. Prognostic relevance of ischemia-modified albumin and NT-proBNP in patients with peripheral arterial occlusive disease. *Clin Chim Acta* 2015;438:255–60. <https://doi.org/10.1016/j.cca.2014.08.031>; PMID: 25195005.
 89. Fatemi S, Acosta S, Gottsäter A, et al. Copeptin, B-type natriuretic peptide and cystatin C are associated with incident symptomatic PAD. *Biomarkers* 2019;24:615–21. <https://doi.org/10.1080/1354750X.2019.1631886>; PMID: 31215249.
 90. Matsushita K, Kwak L, Yang C, et al. High-sensitivity cardiac troponin and natriuretic peptide with risk of lower-extremity peripheral artery disease: the Atherosclerosis Risk In Communities (ARIC) study. *Eur Heart J* 2018;39:2412–9. <https://doi.org/10.1093/eurheartj/ehy106>; PMID: 29579246.
 91. Clemens RK, Annema W, Baumann F, et al. Cardiac biomarkers but not measures of vascular atherosclerosis predict mortality in patients with peripheral artery disease. *Clin Chim Acta* 2019;495:215–20. <https://doi.org/10.1016/j.cca.2019.04.061>; PMID: 30981846.
 92. Mueller T, Dieplinger B, Poelz W, et al. Amino-terminal pro-B-type natriuretic peptide as predictor of mortality in patients with symptomatic peripheral arterial disease: 5-year follow-up data from the Linz Peripheral Arterial Disease study. *Clin Chem* 2009;55:68–77. <https://doi.org/10.1373/clinchem.2008.108753>; PMID: 18988753.
 93. Di Angelantonio E, Chowdhury R, Sarwar N, et al. B-Type natriuretic peptides and cardiovascular risk: systematic review and meta-analysis of 40 prospective studies. *Circulation* 2009;120:2177–87. <https://doi.org/10.1161/CIRCULATIONAHA.109.884866>; PMID: 19917883.
 94. Andican G, Seven A, Uncu M, et al. Oxidized LDL and anti-oxLDL antibody levels in peripheral atherosclerotic disease. *Scand J Clin Lab Invest* 2008;68:473–8. <https://doi.org/10.1080/00365510701842996>; PMID: 18609113.
 95. Laschkolnig A, Kollerits B, Lamina C, et al. Lipoprotein(a) concentrations, apolipoprotein(a) phenotypes, and peripheral arterial disease in three independent cohorts. *Cardiovasc Res* 2014;103:28–36. <https://doi.org/10.1093/cvr/cvu107>; PMID: 24760552.
 96. Cheng SW, Ting AC, Wong J. Lipoprotein(a) and its relationship to risk factors and severity of atherosclerotic peripheral vascular disease. *Eur J Vasc Endovasc Surg* 1997;14:17–23. [https://doi.org/10.1016/S1078-5884\(97\)80220-1](https://doi.org/10.1016/S1078-5884(97)80220-1); PMID: 9290555.
 97. Bertoia ML, Pai JK, Lee JH, et al. Oxidation-specific biomarkers and risk of peripheral artery disease. *J Am Coll Cardiol* 2013;61:2169–79. <https://doi.org/10.1016/j.jacc.2013.02.047>; PMID: 23541965.
 98. Fatemi S, Gottsäter A, Zarrouk M, et al. Lp-PLA₂ activity and mass and CRP are associated with incident symptomatic peripheral arterial disease. *Sci Rep* 2019;9:5609. <https://doi.org/10.1038/s41598-019-42154-5>; PMID: 30948779.
 99. Garg PK, Arnold AM, Hinckley Stukovsky KD, et al. Lipoprotein-associated phospholipase A₂ and incident peripheral arterial disease in older adults: the cardiovascular health study. *Arterioscler Thromb Vasc Biol* 2016;36:750–6. <https://doi.org/10.1161/ATVBAHA.115.306647>; PMID: 26848158.
 100. Garg PK, Norby FL, Polfus LM, et al. Lipoprotein-associated phospholipase A₂ and risk of incident peripheral arterial disease: findings from the Atherosclerosis Risk In Communities study (ARIC). *Atherosclerosis* 2018;268:12–8. <https://doi.org/10.1016/j.atherosclerosis.2017.11.007>; PMID: 29169030.
 101. Li SB, Yang F, Jing L, et al. Correlation between plasma lipoprotein-associated phospholipase A₂ and peripheral arterial disease. *Exp Ther Med* 2013;5:1451–5. <https://doi.org/10.3892/etm.2013.1005>; PMID: 23737897.
 102. Garg PK, Jorgensen NW, McClelland RL, et al. Lipoprotein-associated phospholipase A₂ and risk of incident peripheral arterial disease in a multi-ethnic cohort: the Multi-Ethnic Study of Atherosclerosis. *Vasc Med* 2017;22:5–12. <https://doi.org/10.1177/1358863X16671424>; PMID: 28215109.
 103. Ter Horst R, van den Munckhof ICL, Schraa K, et al. Sex-specific regulation of inflammation and metabolic syndrome in obesity. *Arterioscler Thromb Vasc Biol* 2020;40:1787–800. <https://doi.org/10.1161/ATVBAHA.120.314508>; PMID: 32460579.
 104. Ho DY, Cook NR, Britton KA, et al. High-molecular-weight and total adiponectin levels and incident symptomatic peripheral artery disease in women: a prospective investigation. *Circulation* 2011;124:2303–11. <https://doi.org/10.1161/CIRCULATIONAHA.111.045187>; PMID: 22025604.
 105. Joosten MM, Josphura KJ, Pai JK, et al. Total adiponectin and risk of symptomatic lower extremity peripheral artery disease in men. *Arterioscler Thromb Vasc Biol* 2013;33:1092–7. <https://doi.org/10.1161/ATVBAHA.112.301089>; PMID: 23448969.
 106. Urbonaviciene G, Frystyk J, Flyvbjerg A, et al. Association of serum adiponectin with risk for cardiovascular events in patients with peripheral arterial disease. *Atherosclerosis* 2010;210:619–24. <https://doi.org/10.1016/j.atherosclerosis.2009.12.030>; PMID: 20096841.
 107. Tayebjee MH, Tan KT, MacFadyen RJ, Lip GY. Abnormal circulating levels of metalloproteinase 9 and its tissue inhibitor 1 in angiographically proven peripheral arterial disease: relationship to disease severity. *J Intern Med* 2005;257:110–6. <https://doi.org/10.1111/j.1365-2796.2004.01431.x>; PMID: 15606382.
 108. Madrigal-Matute J, Lindholt JS, Fernandez-Garcia CE, et al. Galectin-3, a biomarker linking oxidative stress and inflammation with the clinical outcomes of patients with atherothrombosis. *J Am Heart Assoc* 2014;3:e000785. <https://doi.org/10.1161/JAHA.114.000785>; PMID: 25095870.
 109. Casanegra AI, Stoner JA, Tafur AJ, et al. Differences in galectin-3, a biomarker of fibrosis, between participants with peripheral artery disease and participants with normal ankle-brachial index. *Vasc Med* 2016;21:437–44. <https://doi.org/10.1177/1358863X16644059>; PMID: 27155290.
 110. Wiecek R, Rośc D, Wiecek AM, Kulwas A. VASCULAR-1 and VASCULAR-2 as a new potential angiogenesis and endothelial dysfunction markers in peripheral arterial disease. *Clin Appl Thromb Hemost* 2019;25:1076029619877440. <https://doi.org/10.1177/1076029619877440>; PMID: 31564130.
 111. Findley CM, Mitchell RG, Duscha BD, et al. Plasma levels of soluble Tie2 and vascular endothelial growth factor distinguish critical limb ischemia from intermittent claudication in patients with peripheral arterial disease. *J Am Coll Cardiol* 2008;52:387–93. <https://doi.org/10.1016/j.jacc.2008.02.045>; PMID: 18652948.
 112. Stehr A, Töpel I, Müller S, et al. VEGF: a surrogate marker for peripheral vascular disease. *Eur J Vasc Endovasc Surg* 2010;39:330–2. <https://doi.org/10.1016/j.ejvs.2009.09.025>; PMID: 19889554.
 113. Gardner AW, Parker DE, Montgomery PS. Changes in vascular and inflammatory biomarkers after exercise rehabilitation in patients with symptomatic peripheral artery disease. *J Vasc Surg* 2019;70:1280–90. <https://doi.org/10.1016/j.jvs.2018.12.056>; PMID: 30922751.
 114. Dopheide JF, Geissler P, Rubrech J, et al. Influence of exercise training on proangiogenic Tie-2 monocytes and circulating angiogenic cells in patients with peripheral arterial disease. *Clin Res Cardiol* 2016;105:666–76. <https://doi.org/10.1007/s00392-016-0966-0>; PMID: 26830098.
 115. Höbaus C, Pesau G, Herz CT, et al. Angioprotein-2 and survival in peripheral artery disease patients. *Thromb Haemost* 2018;118:791–7. <https://doi.org/10.1055/s-0038-1636543>; PMID: 29618157.
 116. Saenz-Piñon G, San Martín P, Planell N, et al. Functional and transcriptomic analysis of extracellular vesicles identifies calprotectin as a new prognostic marker in peripheral arterial disease (PAD). *J Extracell Vesicles* 2020;9:1729646. <https://doi.org/10.1080/20013078.2020.1729646>; PMID: 32158521.
 117. Zeiger F, Stephan S, Hoheisel G, et al. P-Selectin expression, platelet aggregates, and platelet-derived microparticle formation are increased in peripheral arterial disease. *Blood Coagul Fibrinolysis* 2000;11:723–8. <https://doi.org/10.1097/00001721-200012000-00005>; PMID: 11132650.
 118. van der Zee PM, Biró E, Ko Y, et al. P-Selectin- and CD63-exposing platelet microparticles reflect platelet activation in peripheral arterial disease and myocardial infarction. *Clin Chem* 2006;52:657–64. <https://doi.org/10.1373/clinchem.2005.057414>; PMID: 16439610.
 119. Tan KT, Tayebjee MH, Lynd C, et al. Platelet microparticles and soluble P selectin in peripheral artery disease: relationship to extent of disease and platelet activation markers. *Ann Med* 2005;37:61–6. <https://doi.org/10.1080/07853890410018943>; PMID: 15902848.
 120. Crawford JR, Trial J, Nambi V, et al. Plasma levels of endothelial microparticles bearing monomeric C-reactive protein are increased in peripheral artery disease. *J Cardiovasc Transl Res* 2016;9:184–93. <https://doi.org/10.1007/s12265-016-9678-0>; PMID: 26891844.
 121. Mitchell PS, Parkin RK, Kroh EM, et al. Circulating microRNAs as stable blood-based markers for cancer detection. *Proc Natl Acad Sci USA* 2008;105:10513–8. <https://doi.org/10.1073/pnas.0804549105>; PMID: 18663219.
 122. Zhou SS, Jin JP, Wang JQ, et al. miRNAs in cardiovascular diseases: potential biomarkers, therapeutic targets and challenges. *Acta Pharmacol Sin* 2018;39:1073–84. <https://doi.org/10.1038/aps.2018.30>; PMID: 29877320.
 123. Stather PW, Sylvius N, Wild JB, et al. Differential microRNA expression profiles in peripheral arterial disease. *Circ Cardiovasc Genet* 2013;6:490–7. <https://doi.org/10.1161/CIRCGENETICS.111.000053>; PMID: 24129592.
 124. Li T, Cao H, Zhuang J, et al. Identification of miR-130a, miR-27b and miR-210 as serum biomarkers for atherosclerosis obliterans. *Clin Chim* 2011;412:66–70. <https://doi.org/10.1016/j.cca.2010.09.029>; PMID: 20888330.
 125. Stather PW, Sylvius N, Sidloff DA, et al. Identification of microRNAs associated with abdominal aortic aneurysms and peripheral arterial disease. *Br J Surg* 2015;102:755–66. <https://doi.org/10.1002/bjs.9802>; PMID: 25832031.
 126. Small AM, Huffman JE, Klarin D, et al. Mendelian randomization analysis of hemostatic factors and their contribution to peripheral artery disease – brief report. *Arterioscler Thromb Vasc Biol* 2021;41:380–6. <https://doi.org/10.1161/ATVBAHA.119.313847>; PMID: 32847391.
 127. Russell KS, Yates DP, Kramer CM, et al. A randomized, placebo-controlled trial of canakinumab in patients with peripheral artery disease. *Vasc Med* 2019;24:414–21. <https://doi.org/10.1177/1358863X19859072>; PMID: 31277561.
 128. Levin MG, Zuber V, Walker VM, et al. Prioritizing the role of major lipoproteins and subfractions as risk factors for peripheral artery disease. *Circulation* 2021;144:353–64. <https://doi.org/10.1161/CIRCULATIONAHA.121.053797>; PMID: 34139859.