DOI: 10.14814/phy2.14836

### EDITORIAL



# Probing the early upper respiratory responses to SARS-CoV-2

Since the emergence of the novel respiratory coronavirus, designated SARS-CoV-2 in 2019 there have been more than 2.5 million deaths worldwide and the numbers continue to rise. Much of what we know about the disease comes from data from hospitalized patients, who have most likely been infected for a long time and often have severe symptoms. Understanding how SARS-CoV-2 infection influences the immune response early in the course of infection is of upmost importance to fully understand how the infection progresses. Since the emergence of the novel respiratory coronavirus, designated SARS-CoV-2 in 2019, there have been more than 2.5 million deaths worldwide, and the numbers continue to rise (Zhu et al., 2020). Symptoms from infection by SARS-CoV-2, range from entirely asymptomatic to severe acute respiratory distress syndrome (ARDS) (Wiersinga et al., 2020) (Yuki et al., 2020). Severe cases of COVID-19, the disease caused by SARS-CoV-2, are complicated by ARDS and hyper-inflammation, with many patients requiring mechanical ventilation and ICU admission due to hypoxia (Wiersinga et al., 2020) (Edler et al., 2020). Much of what we know about the disease comes from data from hospitalized patients, who have most likely been infected for a long time and often have severe symptoms. Understanding how SARS-CoV-2 infection influences the immune response early in the course of infection is of upmost importance to fully understand how the infection progresses.

The manuscript by Liou et al. (2020) uses an innovative approach to examine the impact of early SARS-CoV-2 infection on the innate immune response in the upper respiratory system. By using nasal pharyngeal swab biospecimens from both SARS-CoV-2 positive patients and SARS-CoV-2 negative patients, the authors are able to determine the impact of a probable early infection on several factors that are important in the early response. In addition, by using both transcriptional and proteomic analysis, this study demonstrates how SARS-CoV-2 infection may influence both transcriptional and post-transcriptional regulation of inflammatory mediators.

One of the most striking findings of the study is that there is a clear correlation of IP-10 (CXCL10) expression with viral load not only at the transcript level but also at the protein level.

CXCL10 expression in the upper respiratory tract has been shown in previous studies to be a hallmark of a respiratory infection (Landry & Foxman, 2018). This chemokine attracts several different immune cell types to the site of infection including macrophages, Natural Killer cells, and activated T cells (Liu et al., 2011). CXCL10 is well known as a factor in ARDS progression, and it has been shown to be elevated in the lungs of patients infected with highly pathogenic avian influenza (HPAI) and SARS-CoV-1 (Jiang et al., 2005) (Peiris et al., 2004). Interestingly, it not only has chemotactic activity, but also controls apoptosis, cell growth and proliferation, and angiogenesis (Liu et al., 2011). CXCL10 has been shown to contribute to oxidative burst as well as chemotaxis of activated neutrophils in models of lung injury (Ichikawa et al., 2013). In COVID-19 patients a biomarker of disease severity is elevated levels of CXCL10 in the plasma (Yang et al., 2020) (Chen et al., 2020). Several previous studies using different models have also shown that nasal epithelial cells express CXCL10 (Gamage et al., 2020) (Lieberman et al., 2020).

Another finding is that in many cases the protein levels do not correspond to what is seen at the mRNA level for the various inflammatory mediators looked at. However, proteins for CXCL10 and several interferon stimulated genes (ISGs) did correlate with transcript level. Further studies are necessary to determine mechanisms behind this interesting observation. It is unclear is this is due to a lack of translation or an active degradation process. Also, it is unknown if this a specific feature of SARS-CoV-2 infection where the virus is actively involved in the regulation of host inflammatory mediators, or if this is part of the host process in the tight regulation of the innate immune response during early infection.

This study gives important insight into the early immune and anti-viral responses to initial SARS-CoV-2 infection of the upper respiratory tract. Understanding these early stages of infection will allow us to more fully understand the pathology of COVID-19.

#### Amanda M. Jamieson 回

Division of Biology and Medicine, Department of Molecular Microbiology and Immunology, Brown University, Providence, RI, USA

This is an open access article under the terms of the Creative Commons Attribution License, which permits use, distribution and reproduction in any medium, provided the original work is properly cited.

<sup>© 2021</sup> The Authors. Physiological Reports published by Wiley Periodicals LLC on behalf of The Physiological Society and the American Physiological Society



#### Correspondence

Amanda M. Jamieson, Division of Biology and Medicine, Department of Molecular Microbiology and Immunology, Brown University, Providence, RI, USA. Email: Amanda Jamieson@brown.edu

## ORCID

Amanda M. Jamieson D https://orcid. org/0000-0002-5781-9231

## REFERENCES

- Chen, Y., Wang, J., Liu, C., Su, L., Zhang, D., Fan, J., Yang, Y., Xiao, M., Xie, J., Xu, Y., Li, Y., & Zhang, S. (2020). IP-10 and MCP-1 as biomarkers associated with disease severity of COVID-19. Molecular Medicine, 26, 97. https://doi.org/10.1186/s10020-020-00230-x
- Edler, C., Schröder, A. S., Aepfelbacher, M., Fitzek, A., Heinemann, A., Heinrich, F., Klein, A., Langenwalder, F., Lütgehetmann, M., Meißner, K., Püschel, K., Schädler, J., Steurer, S., Mushumba, H., & Sperhake, J.-P. (2020). Dying with SARS-CoV-2 infection-an autopsy study of the first consecutive 80 cases in Hamburg, Germany. International Journal of Legal Medicine, 134, 1275-1284. https://doi.org/10.1007/s00414-020-02317-w
- Gamage, A. M., Tan, K. S., Chan, W. O. Y., Liu, J., Tan, C. W., Ong, Y. K., Thong, M., Andiappan, A. K., Anderson, D. E., Wang, D. Y., & Wang, L.-F. (2020). Infection of human Nasal Epithelial Cells with SARS-CoV-2 and a 382-nt deletion isolate lacking ORF8 reveals similar viral kinetics and host transcriptional profiles. PLoS Path, 16, e1009130. https://doi.org/10.1371/journal.ppat.1009130
- Ichikawa, A., Kuba, K., Morita, M., Chida, S., Tezuka, H., Hara, H., Sasaki, T., Ohteki, T., Ranieri, V. M., dos Santos, C. C., Kawaoka, Y., Akira, S., Luster, A. D., Lu, B., Penninger, J. M., Uhlig, S., Slutsky, A. S., & Imai, Y. (2013). CXCL10-CXCR3 enhances the development of neutrophil-mediated fulminant lung injury of viral and nonviral origin. American Journal of Respiratory and Critical Care Medicine, 187, 65-77. https://doi.org/10.1164/rccm.20120 3-0508OC
- Jiang, Y., Xu, J., Zhou, C., Wu, Z., Zhong, S., Liu, J., Luo, W., Chen, T., Qin, Q., & Deng, P. (2005). Characterization of Cytokine/ Chemokine profiles of severe acute respiratory syndrome. American Journal of Respiratory and Critical Care Medicine, 171, 850-857. https://doi.org/10.1164/rccm.200407-857OC
- Landry, M. L., & Foxman, E. F. (2018). Antiviral response in the nasopharynx identifies patients with respiratory virus infection.

Lieberman, N. A. P., Peddu, V., Xie, H., Shrestha, L., Huang, M.-L., Mears, M. C., Cajimat, M. N., Bente, D. A., Shi, P.-Y., Bovier, F., Roychoudhury, P., Jerome, K. R., Moscona, A., Porotto, M., & Greninger, A. L. (2020). In vivo antiviral host transcriptional response to SARS-CoV-2 by viral load, sex, and age. PLoS Biology, 18, e3000849. https://doi.org/10.1371/journal.pbio.3000849

org/10.1093/infdis/jix648

- Liou, T. G., Adler, F. R., Cahill, B. C., Cox, D. R., Cox, J. E., Grant, G. J., Hanson, K. E., Hartsell, S. C., Hatton, N. D., Helms, M. N., Jensen, J. L., Kartsonaki, C., Li, Y., Leung, D. T., Marvin, J. E., Middleton, E. A., Osburn-Staker, S. M., Packer, K. A., Shakir, S. M., ... Sturrock, A. B. (2020). SARS-CoV-2 innate effector associations and viral load in early nasopharyngeal infection. medRxiv, https://doi.org/10.1101/2020.10.30.20223545
- Liu, M., Guo, S., Hibbert, J. M., Jain, V., Singh, N., Wilson, N. O., & Stiles, J. K. (2011). CXCL10/IP-10 in infectious diseases pathogenesis and potential therapeutic implications. Cytokine & Growth Factor Reviews, 22(3):121-130. https://doi.org/10.1016/j.cytog fr.2011.06.001
- Peiris, J., Yu, W., Leung, C., Cheung, C., Ng, W., Nicholls, J., Ng, T., Chan, K., Lai, S., Lim, W., Yuen, K., & Guan, Y. (2004). Re-emergence of fatal human influenza A subtype H5N1 disease. The Lancet, 363, 617-619. https://doi.org/10.1016/S0140 -6736(04)15595-5
- Wiersinga, W. J., Rhodes, A., Cheng, A. C., Peacock, S. J., & Prescott, H. C. (2020). Pathophysiology, transmission, diagnosis, and treatment of coronavirus disease 2019 (COVID-19): A review. JAMA, 324, 782. https://doi.org/10.1001/jama.2020.12839
- Yang, Y., Shen, C., Li, J., Yuan, J., Wei, J., Huang, F., Wang, F., Li, G., Li, Y., Xing, L., Peng, L., Yang, M., Cao, M., Zheng, H., Wu, W., Zou, R., Li, D., Xu, Z., Wang, H., ... Liu, Y. (2020). Plasma IP-10 and MCP-3 levels are highly associated with disease severity and predict the progression of COVID-19. Journal of Allergy and Clinical Immunology, 146, 119-127.e4. https://doi.org/10.1016/j. jaci.2020.04.027
- Yuki, K., Fujiogi, M., & Koutsogiannaki, S. (2020). COVID-19 pathophysiology: A review. Clinical Immunology, 215, 108427. https:// doi.org/10.1016/j.clim.2020.108427
- Zhu, N., Zhang, D., Wang, W., Li, X., Yang, B., Song, J., Zhao, X., Huang, B., Shi, W., Lu, R., Niu, P., Zhan, F., Ma, X., Wang, D., Xu, W., Wu, G., Gao, G. F., & Tan, W. (2020). A Novel coronavirus from patients with pneumonia in China, 2019. New England Journal of Medicine, 382, 727-733. https://doi.org/10.1056/ NEJMoa2001017