ICU lower respiratory isolates. Significant declines in FEP and TZP susceptibility were observed in institutions that demonstrated higher CRPA frequency (p< 0.00001) (Table 1) with >25% of isolates showing resistance in CR2 and CR3. In contrast, C/T susceptibility remained above 90% for all categories. I/R susceptibility remained above 90% for CR1 and CR2.

Table 1. P. aeruginosa susceptibility among ICU lower respiratory tract isolates stratified by unit specific frequency of carbapenem resistance

Antimicrobial	CR Group 1 (N = 37) (n = 264, %)	CR Group 2 (N = 25) (n = 363, %)	CR Group 3 (N = 18) (n = 244, %)
Cefepime	83.7	74.9	63.1
Piperacillin-tazobactam	79.6	68.9	52.9
Meropenem	91.3	73.6	47.5
Levofloxacin	68.6	66.1	48.0
Ceftolozane-tazobactam	96.6	94.2	90.6
Imipenem-relebactam	98.1	91.7	81.6
N = number of institutions n = number of isolates CR Group 1: institutions with ≤ 20% CR Group 2: institutions with 21 – 40			
CR Group 2: Institutions with 21 – 40 CR Group 3: institutions with ≥ 41%			

Conclusion. C/T and I/R provide robust activity against PA isolates obtained from the lower respiratory tract of critically ill patients, regardless of CR frequency. Assessing CRPA frequency may be useful for identifying inflection points in which novel agents could be considered. In settings where CRPA frequency exceeded 20%, additional testing or early switch to C/T or I/R may be warranted. **Disclosures.** Kenneth Klinker, PharmD, Merck & Co, Inc (Employee) Daryl

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1451. Genetic Analysis of Antibiotic Resistance Profiles of *Acinetobacter baumannii* Using Whole Genome Sequencing

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Session: P-66. Resistance Mechanisms

Background. Acinetobacter is known to quickly develop resistance to commonly used antibiotics. Previously we performed whole genome sequencing (WGS) and whole genome multilocus sequence typing (wgMLST) analysis in clinical Acinetobacter isolates to determine sequence types (ST) of these isolates and mapped their distribution. In this study, we sought to characterize the genetic antibiotic resistance patterns in these isolates.

Methods. Sixty-two clinical Acinetobacter isolates collected in two distinct large tertiary care hospitals in Detroit were analyzed. The samples were subjected to WGS using the NextSeq instrument (Illumina). The contigs were *de novo* assembled using SPAdes (v3.7.1) and wgMLST analysis was performed using BioNumerics software v7.6. The genomic sequence for each isolate was uploaded in ResFinder 3.2 and known antibiotic resistance genes were analyzed.

Results. The most common resistance gene found is blaADC-25 conferring resistance to beta-lactams across all STs. Sulfonamide (sul1) and macrolide resistance (mphE/msrE) among STs were also common. ST2 (52%) was predominant for both hospitals (H1 and H2). ST2 in H2 exhibited the presence of the maximum number of resistance genes including resistance to aminoglycosides, macrolides (2), tetracyclines (tetB), beta-lactams, fluoroquinolones (aac(6')-Ib-cr), sulphonamides (sul1, sul2). ST2 had a slightly different resistance profile of beta-lactams in H1 when compared to H2. ST406 and ST15 exhibited similar antibiotic resistant.

Table 1. Antimicrobial Drug Resistance Profiles of all Acinetobacter Sequence Types (STs) in H1 and H2.

		Hospital 1
Strain Type (ST)	Total Number of Isolates	Resistance to Antibiotic Drugs
ST2	11	β-lactam, aminoglycoside, sulphonamide, phenicol, macrolide, tetracycline
ST15	2	β-lactam, aminoglycoside, sulphonamide
ST36	2	β-lactam
ST49	1	β-lactam
ST93	5	β-lactam
ST119	2	β-lactam
ST212	1	β-lactam
ST406	4	β-lactam, aminoglycoside, sulphonamide
ST667	1	β-lactam
		Hospital 2
Strain Type (ST)	Total Number of Isolates	Resistance to Antibiotic Drugs
ST2	20	β -lactam, aminoglycoside, sulphonamide, phenicol, macrolide, tetracycline
ST15	1	β-lactam, aminoglycoside, sulphonamide
ST20	1	β -lactam, aminoglycoside, sulphonamide, phenicol, macrolide, tetracycline
ST214	1	β-lactam
ST268	1	β-lactam
ST395	1	β-lactam
ST406	1	β-lactam, aminoglycoside, sulphonamide
ST427	2	β-lactam

Table 2. Antibiotic Resistance Gene Profiles of Sequence Type 2 (ST2) isolates in H1 and H2.

	Hospital Unit 1			Hospital Unit 2	
ST2			ST2		
ntibiotic Drugs	Antibiotic Resistance Genes	% of Isolates with Antibiotic Resistance Genes	Antibiotic Drugs	Antibiotic Resistance Genes	% of Isolates with Antibioti Resistance Genes
MINOGLYCOSIDE	and A1	72		aadA1	30
	andA2	9		aadA2	15
	armA	72		armA	70
	ant(2")-ia	9		ant(2")-la	15
		81		aph(3")-Ib	80
	aph(3")-Ib	9	AMINOGLYCOSIDE	aph(3')-Via	40
	aph(3')-Via	81		aph(3')-la	70
	aph(3')-1a			aph(6)-ld	80
	aph(6)-Id	81		aac(3)-IIa	5
	aac(3)-la	9		aac(6')-Ib	5
	aac(6')-1b3	63		aac(6')-Ib3	25
	aac(6')-Ib-cr	63		aac(6')-lb-cr	30
ICOL	catBB	63	PHENICOL	catB8	30
	mph(E)	72	MACROLIDE	mph(E)	80
ROLIDE	msr(E)	72		msr(E)	80
	sul1	90	SULPHONAMIDE	sul1	40
ONAMIDE	sul2	9	500 11010011100	sul2	10
CYCLINE		81	TETRACYCLINE	tet(B)	80
TCLINE	tet(B)	100		blaADC-25	100
BETALACTAM	blaADC-25			blaOXA-223	5
	blaOXA-23	27	BETALACTAM	blaOXA-23	80
	blaOXA-66	81	SCIADACTAM	blaOXA-66	20
	blaOXA-82	18		blaOXA-82	15
	blaTEM-1D	72		blaTEM-1D	35

Conclusion: This study provides us with a snapshot of antibiotic resistant genes among circulating *Acinetobacter isolates* prevalent in an area. All hospital isolates demonstrated resistance to beta-lactams. Multidrug resistant ST2 isolates from both hospitals demonstrate similar antibiotic resistance gene profiles suggesting a common circulating strain in the area. No colistin resistance genes were detected in any isolates. Because *Acinetobacter* infections are predominantly hospital acquired, it is important to continually monitor resistance profiles to determine the trends that may better serve both the patients and infection control practices.

Disclosures. Chetan Jinadatha, MD, MPH, AHRQ (Research Grant or Support)Department of Veterans Affairs (Other Financial or Material Support, Owner: Department of Veterans Affairs. Licensed to: Xenex Disinfection System, San Antonio, TX)Inventor (Other Financial or Material Support, Methods for organizing the disinfection of one or more items contaminated with biological agents)NiH/NINR (Research Grant or Support)NSF (Research Grant or Support)Xenex Healthcare Services (Research Grant or Support)

1452. Molecular Profile of β-Lactamase Genes and Siderophore-Dependent Iron Transporter Genes of Cefiderocol High MIC Isolates from SIDERO-WT Studies Yoshinori Yamano, PhD¹, Miki Takemura, MSc¹; Krystyna Kazmierczak, PhD²; Mark G G. Wise, PhD²; Meredith Hackel, MPH³; Daniel F. Sahm, PhD³; Roger Echols, MD⁴, ¹Shionogi & Co., Ltd., Osaka, Osaka, Japan; ²IHMA, Schaumburg, Illinois; ³IHMA, Inc., Schaumburg, Illinois; ⁴Infectious Disease Drug Development Consulting LLC, Easton, Connecticut

Session: P-66. Resistance Mechanisms

Background. Cefiderocol (CFDC) is a novel siderophore cephalosporin with efficacy against Gram-negative (GN) bacteria, including carbapenem-resistant Enterobacterales and non-glucose-fermenters such as *Pseudomonas aeruginosa and Acinetobacter baumannii*. In consecutive multinational surveillance (SIDERO-WT) studies (2014–2017), CFDC demonstrated activity with minimum inhibitory concentrations (MICs) of ≤ 4 mg/mL against 99.4% of 28,629 GN clinical isolates. We conducted molecular characterization of 161 isolates with CFDC MICs >4 mg/mL from the SIDERO-WT studies.

Methods. A total of 161 isolates underwent whole genome sequencing by Illumina Hiseq. Analyses were done using the CLC genomics workbench (Qiagen) for possible resistance-related genes (e.g. *β*-lactamases, porin channels or penicillin-binding protein genes) and some TonB-dependent siderophore uptake receptor genes (*fu*, *cir*, *piu*, *pir*). Fiu and Cir in *Escherichia coli* and Piu in *P. aeruginosa* are the iron transporters involved in CFDC transport.

Results. Of 161 isolates with CFDC MIC >4 mg/mL, 128 were *A. baumannii*, 22 Enterobacterales, 7 *Burkholderia multivorans*, 2 *P. aeruginosa*, and 2 *Stenotrophomonas maltophilia*. Genes encoding PER/VEB extended-spectrum β -lactamases and NDMtype metallo- β -lactamases were detected in some isolates, but other β -lactamase genes (*bla*) were not shown to be linked to high CFDC MICs. *bla_{PER}/bla_{VEB}* were found only in *A. baumannii* and *bla_{NDM}* was found in *A. baumannii* and *Klebsiella pneumoniae*. In 128 *A. baumannii* isolates, 103 harbored PER or VEB, including PER positive isolates from Russia (n=87) and Turkey (n=6) and 4 VEB positive isolates from USA. Nine NDMpositive isolates (7 *K. pneumoniae*, 2 *A. baumannii*) were found. Disruption of iron transport genes was also detected in some isolates, including *piuA* (11 *A. baumannii*, 1 *P. aeruginosa*), *pirA* (2 *A. baumannii*), and *fiuA* (4 *B. multivorans*, 1 *Proteus mirabilis*). No *cir* homologs were found in 2 *B. multivorans*.

Conclusion. PER and NDM could reduce susceptibility to CFDC, as such isolates have been seen in some countries. Iron transporter disruption was also observed in some isolates with high CFDC MICs; the contribution of these deficiencies in *A. baumannii* and *B. multivorans* requires further study.

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1453. PBP2, PBP2a and PBP4 Clone-specific Polymorphisms are not Associated to Ceftaroline (CPT) Susceptibility in Chilean Clinical Isolates of Methicillin-Resistant *Staphylococcus aureus* (MRSA)

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Session: P-66. Resistance Mechanisms

Background. CPT is a last-generation cephalosporin active against MRSA due to its affinity for PBP2a. CPT-resistance (CPT-R) is well-described, with mutations in the active transpeptidase domain of PBP2a associated to high-level resistance. The accumulation of changes in the non-penicillin-binding domain of PBP2a has been linked to elevations of the minimal inhibitory concentration (MIC) to CPT to levels around 2-4ug/mL. PBP4 and PBP2 have also been implicated as potentially relevant *mecA*-independent mechanisms of CPT-R. We recently reported high rates of CPT-non-susceptibility in clinical MRSA strains from Chile. However, the mutational landscape of PBPs in clinical MRSA isolates from Chile and its relation to CPT susceptibility has not been assessed.

Methods. We analyzed 180 MRSA isolates collected from 2000-2018 in Santiago, Chile. Identification was confirmed by MALDI-TOF and methicillin resistance with cefoxitin disk-diffusion. CPT susceptibility was performed by BMD

following CLSI-2019 guidance. Whole-genome sequencing was performed for all isolates; the mutational profile of PBPs was determined using reference sequences for PBP2 (AGY89563.1), PBP2a (NG_047938.1) and PBP4 (X91786.1).

Results. All isolates were phenotypically-confirmed MRSA and harbored *mecA*. The MIC_{50}/MIC_{50} by BMD was 2/2µg/dL; only 71 (39%) isolates were CPT-susceptible (MIC <1µg/mL). Most isolates belonged to ST5/SCCmecI (70%,126/180), ST105/SCCmecII (10%,18/180) and ST8/SCCmecIV (5%, 9/180). All ST5/SCCmecI isolates carried the mutations in PBP2 (Y156D), PBP2a (M122I and E150K), and PBP4 (T189S, L234H, and T409A); CPT-susceptibility among ST5/SCCmecI was only 22%. On the other hand, all ST105/SCCmecII isolates had mutations in PBP2 (S707L) and PBP4 (T189S, L234H, and T409A) and exhibited a higher CPT-susceptibility rate (67%). All 9 isolates belonging to the ST8/SCCmecIV lineage harbored a non-coding mutation in PBP2a (g-6t) and the previously observed L234H change in PBP4. Importantly, no association between specific polymorphisms and MIC to CPT was found.

Table 1. PBPs mutations compared to CPT MICs by MLST and SCCmec

MLST SCCmec	Mutational profile			MIC						
	Sectinee	PBP2	PBP2a	PBP4	0.125	0.25	0.5	1	2	4
ST 5	SCCmec I	Y156D	M122I/E150K	T189S, L234H, T409A	1		5	22	86	12
ST 5	SCCmec IV		g-6t	T189S, L234H, T409A		1				
ST 105	SCCmec II	\$707L		T189S, L234H, T409A		1	8	3	3	3
ST 225	SCCmec II	S707L		T189S, L234H, T409A			1	1		
ST 125	SCCmec IV		g-6t	T1895, T409A			1	1		
ST 72	SCCmec IV		g-6t	T25A, T189S, P235S, Q383K, T409A			4		2	
ST 72	SCCmec VI		g-6t	T25A, T189S, P235S, Q383K, T409A			3		1	
ST 1472	SCCmec IV		g-6t	T25A, L234H, T409A			1			
ST 22	SCCmec IV		g-6t	T25A, D98E, E398A, T409A		1				
ST 2802	SCCmec IV		g-6t	P235S			2			
ST 2039	SCCmec III			P2358				1		
ST 923	SCCmec IV		g-6t	P235S		1	1			
ST 8	SCCmec IV		g-6t	L234H			8		1	
ST 239	SCCmec III		-	L234H				3	1	
ST UN	SCCmec IV		g-6t	1234H			1			

Conclusion: Changes in the studied PBPs were frequent among MRSA circulating in Chile and were conserved among different genetic backgrounds. However, these changes were not associated with the level of CPT MIC among these isolates.

Disclosures. Cesar A. Arias, MD, MSc, PhD, FIDSA, Entasis Therapeutics (Scientific Research Study Investigator)MeMed (Scientific Research Study Investigator)Merck (Grant/Research Support)

1454. Plazomicin Activity against *Enterobacterales* Isolates Producing Extended-Spectrum β-Lactamases (ESBLs), Carbapenemases, and Aminoglycoside-Modifying Enzymes (AMEs) from United States (US) Hospitals

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Session: P-66. Resistance Mechanisms

Background. Limited therapeutic options are available for the treatment of multidrug resistant (MDR) organisms. Plazomicin (PLZ) is an aminoglycoside developed to overcome common aminoglycoside-resistance mechanisms. We evaluated the activity of PLZ and comparators against *Enterobacterales* isolates collected in 2018-2019 carrying genes encoding ESBLs, carbapenemases, and AMEs.

Methods. Among 3,899 *Enterobacterales* isolates from US hospitals susceptibility (S) tested using reference broth microdilution method, 619 isolates from selected species displaying elevated MIC values for cephalosporins, carbapenems and/or resistance (CLSI criteria) to amikacin (AMK), gentamicin (GEN) and tobramycin (TOB) were submitted to whole genome sequencing for detection of resistance genes.

Results. Most isolates producing ESBLs (n= 418) carried bla_{CTX-M} (n= 386). The activity of PLZ (99.3% susceptible [S]) was comparable to that of collstin and higher than other comparators against ESBL isolates (Figure). AMK inhibited 96.4% of the isolates and GEN and TOB inhibited 57.9% and 43.5%, respectively. Only 34 isolates produced carbapenemases, including 19 KPC-2, 10 KPC-3, 1 each with VIM-1, OXA-181, NDM-5 and KPC-2-like plus 1 isolate carrying the genes encoding NDM-1 and OXA-232. These isolates displayed higher resistance rates to comparators and only PLZ, and tigecycline inhibited >90% of these isolates. AMK and GEN inhibited 67.6% and 55.9% of these isolates, respectively. PLZ was active against 97.7% of isolates carrying AME genes (n= 306) that carried *aac(6')-lb-cr* (n =177), *aac(3)-IIa* (n = 159) and *TOB* (only 10.8-14.1% S), but 92.8% were S to AMK. Three *K. pneumoniae* isolates carried 168 rRNA methyltransferases, 1 *armA* (also harboring NDM-1) and 2 *rmIB1*.

Conclusion. The activity of PLZ against *Enterobacteriaceae* isolates carrying AMEs, ESBLs, and carbapenemases was greater than the activity of other aminogly-cosides tested and comparable to those of tigecycline and colistin against carbapenemase-producing organisms. Isolates carrying genes encoding ESBLs, AMEs and carbapenemases are usually MDR and PLZ had activity against these organisms collected in the US.