

Hydroxysafflor yellow A protects against angiotensin II-induced hypertrophy

BIN NI*, DONGLAI ZHOU*, YUNYAN JING and SHANXIN LIU

Department of Cardiology, The Affiliated Hospital of Hangzhou Normal University, Hangzhou, Zhejiang 310015, P.R. China

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Abstract. Myocardial infarction (MI) is life-threatening and is generally accompanied by myocardial hypertrophy. Notably, Hydroxysafflor yellow A (HSYA) can prevent tissue injuries. The objective of this study was to investigate the effect of HSYA on hypertrophy after MI. Hematoxylin and eosin (H&E) staining assays were performed to measure cell area. The protein synthesis rate was assessed using the ³H Leucine incorporation assay. Reverse transcription-quantitative polymerase chain reaction (RT-qPCR), western blot analysis and the immunohistochemical assay were used to detect the expression of target genes. The activity of superoxide dismutase (SOD), malondialdehyde (MDA) and the reactive oxygen species (ROS) generation were examined using commercial kits. Decreased myocardial hypertrophy was observed in animals treated with HSYA. Furthermore, the expression of nuclear factor (erythroid-derived 2)-like 2 (Nrf2) was higher in HSYA administration groups compared with that in the MI model group. In H9c2 cardiomyocytes, the pretreatment with HSYA increased the cell viability, however, it reduced protein synthesis rate, mitigated cell surface area and decreased the expression of Brain natriuretic factor (BNP) and β -myosin heavy chain (β -MHC). By contrast, the downregulation of Nrf2 deteriorated and reversed the effect of Ang II and HSYA. Furthermore, oxidative stress was alleviated by HSYA via inhibiting ROS generation, modulating the activities of SOD and MDA. In addition, the expression of NAD(P)H:quinone oxidoreductase 1 (NQO1) and heme oxygenase-1 (HO-1) were recovered by the pretreatment of HSYA that was combated by siNrf2. In conclusion, HSYA exerted anti-hypertrophic effects, which was pertinent with the activation of Nrf2/NQO-1/HO-1

signaling pathway. The findings of this study may inspire a novel strategy to combat MI.

Introduction

With heart failure becoming increasingly prevalent, it threatens the health of many individuals (1,2). As a common health problem, myocardial infarction (MI) is one of the primary causes of heart failure. The fibers of the myocardial tissue will be elongated under normal conditions. However, if the myocardial tissue works overload, the fibers may not properly shorten, therefore increasing the size of the myocardial fibers. This eventually leads to the fact that the blood pumping cannot be effectively exerted and the oxygen cannot be effectively delivered to the working tissues in the body (3). The risk factors for myocardial hypertrophy may be coronary artery disease or high blood pressure (4). As a compensatory response of the heart to pathological stimuli, myocardial hypertrophy is often accompanied with MI that is characterized by an enlarged heart (5).

From the molecular perspective, the myocardial hypertrophy is a consequence of the reprogramming of gene expression and the alteration of signaling pathways (6,7). Brain natriuretic factor (BNP) and β -myosin heavy chain (β -MHC) are two typical markers to indicate the myocardial hypertrophy (8). In addition, the activity of intracellular signaling pathway may produce even more profound effect on the biological events. Moreover, emerging evidences have demonstrated that reactive oxygen species (ROS) are critical in triggering the hypertrophic responses to various stimulus, for example, stretch (9) or Ang II (10-12). In addition, nuclear factor (erythroid-derived 2)-like 2 (Nrf2) is a transcription factor that regulates the expressions of target genes, for instance, NAD(P)H:quinone oxidoreductase 1 (NQO1) and heme oxygenase-1 (HO-1) (13), through antioxidant response element (ARE). The protective role of Nrf2 has been discovered in tissue injuries (14). However, the role of Nrf2 remains less known in hypertrophy after MI. Hence, investigating the effects of Nrf2 is of significance for the purpose of understanding the molecular event in heart failure.

The antihypertensive therapy has been proved to be increasingly effective (15); however, the prognosis of the patients suffering from acute MI remains unsatisfactory. Therefore, it is necessary to develop new options that are helpful to the treatment of myocardial hypertrophy induced

Correspondence to: Dr Shanxin Liu, Department of Cardiology, The Affiliated Hospital of Hangzhou Normal University, 126 Wenzhou Road, Hangzhou, Zhejiang 310015, P.R. China
E-mail: liushanxin_xslidr@163.com

*Contributed equally

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by MI. Increasing attention has recently been paid to the traditional Chinese medicine, which has been demonstrated to have valuable therapeutic effects by modern medicines (16). Hydroxysafflor yellow A (HSYA), a compound isolated from *Carthamus tinctorius* L., has its main activity in the treatment of cardiovascular diseases (17). HSYA is able to promote the blood circulation. Previous studies have proved its role in myocardial I/R injury (18). However, the effect of HSYA and the association between HSYA and Nrf2 in myocardial hypertrophy after MI remain unclear.

Thus, this study aimed to estimate the role of HSYA in myocardial hypertrophy induced by MI as well as its role in Nrf2 signaling pathway both *in vivo* and *in vitro*. Our results would not only provide a promising agent for the management of myocardial hypertrophy after MI, but also the possible molecular target in the prevention of heart failure.

Materials and methods

Animals and MI *in vivo* model. This study concerned animals trials and was approved by Animal Subjects Committee of the Affiliated Hospital of Hangzhou Normal University. All the clinical and surgical procedures were conducted under the Institutional Animal Care guidelines. Male Sprague-Dawley rats (275-300 g) were purchased from Guangdong Medical Laboratory Animal Center. The animals were randomly grouped into six groups: control group, sham surgery group (sham), model group, captopril (Ka) group, HSYA low concentration group (HSYA-L) and HSYA high concentration group (HSYA-H). The rats had access to standard rat chow and water *ad libitum* and they were fed in the Affiliated Hospital of Hangzhou Normal University. Permanent ligation of the left anterior descending coronary artery was performed to set up the MI model *in vivo*. The sham group was only performed with open-heart surgery. The treatment groups were intraperitoneally injected with captopril (10 mg/kg/day; Tianjin Lisheng Pharmaceutical Co., Ltd., Tianjin, China) or HSYA (2 or 5 mg/kg/day; Shanghai Yuanye Biotechnology Co. Ltd., Shanghai, China). The rats were anesthetized with 1.5% isoflurane at four weeks after MI. The heart were excised, rinsed in ice-cold saline and weighted. The doses of each agent were adopted as previously described (19,20).

Cell culture and grouping. H9c2, derived from the embryonic rat ventricle, was purchased from ATCC. Cells were maintained in Dulbecco's modified Eagle's medium (DMEM) supplemented with 1% penicillin/streptomycin, 10% fetal bovine serum (FBS) at 37°C with 5% CO₂ in a humidified incubator. The cell grouping was as follows: in control group: The normal cells; in Ang II group: The cells were treated with 1 μM Ang II for 12 h at 37°C; in Ang II/mock group (A/mock): The cells were transfected with si-negative control and treated with 1 μM Ang II; in Ang II/siNrf2 group (A/siNrf2): The cells were transfected with si-Nrf2 and treated with 1 μM Ang II; in Ang II/HSYA group (A/H): The cells were pretreated with 80 μM HSYA for 8 h and then incubated with 1 μM Ang II for 12 h; in Ang II/HSYA/mock group (A/H/mock): The cells were transfected with si-negative control and then treated with 1 μM Ang II for 12 h following 8 h incubation of 80 μM HSYA; in Ang II/HSYA/siNrf2

group (A/H/siNrf2): The cells were transfected with si-Nrf2 control and then treated with 1 μM Ang II for 12 h following 8 h incubation of 80 μM HSYA.

Hematoxylin and eosin (H&E) staining. The myocardial tissue was fixed 4% formaldehyde at 4°C for 72 h and was embedded in paraffin. The paraffin embedded tissue was sliced into 3-4 μm. Then, the slides were subject to the treatment as follows: Dewaxing with xylene for 15 min, incubation in gradient alcohol, hematoxylin staining for 15 min, incubation in 1% Hydrochloric acid alcohol for 15 sec, 1% eosin staining for 1 min. The images for H&E staining were captured under an inverted microscope (Olympus, Tokyo, Japan). The slides were sealed and the cell area was measured by Image-Pro Plus 6.0 software (Media Cybernetics, Inc., Rockville, MD, USA). Five random fields were selected.

Downregulation of Nrf2 by siRNA. Cells were transfected with siRNA using the Lipofectamine[®] 3000 reagent (Invitrogen; Thermo Fisher Scientific, Inc., Waltham, MA, USA). Gene specific siRNA for Nrf2 and negative control siRNA were purchased from GeneCopoeia, Inc. (Rockville, MD, USA). The transfection efficiency of RNA knockdown was estimated at 48 h after post-transfection.

Cell viability and ³H Leucine incorporation assay. Cells were seeded at a density of 3x10⁴ cells in 96-well plate. After being treated as designed for each group, the cells were prepared for conducting the cell viability assay. To explain further, first, the Cell Counting Kit-8 (CCK-8) solution (10 μl) was added into each well. After 4 h of incubation, the plate was removed from the incubator. The absorbance was read at 450 nm with a microplate reader (BioTek Instruments Inc., Winooski, VT, USA). For detecting protein synthesis rate, ³H Leucine incorporation assay was applied as previously described (21). To be more specific, the cells were pulsed with 1 μCi/ml [³H] leucine (PerkinElmer, Inc., Waltham, MA, USA) in PBS at 37°C for 2 h. After being washed by PBS buffer, the cells were incubated with 10% trichloroacetic acid. The glass micro-fiber filter under vacuum was used to collect the precipitate. The precipitate was then suspended in scintillation fluid. The incorporation of ³H-Leu [counts per minute (CPM)] was counted on the liquid scintillation counter (GE Healthcare Bio-Sciences, Pittsburgh, PA, USA).

Determination of ROS, malondialdehyde (MDA) and superoxide dismutase (SOD). The method of ROS detection and the cell treatment with Ang II was referred to previous studies (22,23). The ROS generation was assessed by a fluorescent probe dihydrodichlorofluorescein diacetate (DCFH-DA; Sigma-Aldrich; Merck KGaA, Darmstadt, Germany). The cells were treated as described in cell grouping. Then, the cells were detached with 0.25% trypsin-EDTA and washed with PBS buffer. Then, cells in each group were incubated with 10 μM DCFH-DA for 30 min at 37°C. Next, the cells were washed with serum-free DMEM. And then centrifugated for 5 min at 1,000 x g at room temperature. The supernatants were collected and prepared for the fluorescence determination. With the excitation wavelength of 480 nm and the emission wavelength of 525 nm, the fluorescent signal was determined

using a FLUOstarOPTIMA fluorescence spectrophotometer (BMG Labtech, Ortenberg, Germany). The mean fluorescence intensity (MFI) was considered as the index for the production of ROS. MFIs from five random fields were detected using Image J 1.41 software. The assessment of the activity of SOD and MDA was conducted following the protocols provided by each commercial kit (Beyotime Institute of Biotechnology, Haimen, China). The measurement was read adopting spectrophotometric methods with a plate reader (Bio-Rad Laboratories, Inc., Hercules, CA, USA). Absorbance at 412 nm was estimated for SOD. The result was expressed as SOD U/ml. The MDA level was expressed as MDA nmol/mg protein.

Reverse transcription-quantitative polymerase chain reaction (RT-qPCR). The total RNA was lysed with TRIzol reagent (Thermo Fisher Scientific, Inc., Waltham, MA, USA). The transparent RNA phase was taken by RNase free chloroform and transferred into a new RNase free tube containing 70% alcohol. The integrity of RNA was determined by gel electrophoresis. The QIAGEN RNA Cleanup kit was used to purify RNA. Invitrogen cDNA synthesis kit was used to synthesis cDNA. SYBR-Green Real Time PCR detection system was adopted to detect the gene expression on ABI 7500 instrument. The amplification conditions was as follows: At 50°C, 2 min, at 95°C, 10 min, and then 32 cycles at 95°C for 15 sec and at 60°C for 1 min. The expression levels were normalized to β -actin expression. The specific primers were as follows: Nrf2 Forward (F): 5'-TTCCTCTGCTGCCATTAGTCAGTC-3', and reverse (R): 5'-GCTCTTCCATTTCCGAGTCACTG-3'; NQO-1 F: 5'-GCAGTTTCTAAGAGCAGAACG-3', and R: 5'-GTAGAT TAGTCCTCACTCAGCCG-3'; HO-1 F: 5'-CTGGAAGAG GAGATAGAGCGAA-3', and R: 5'-TCTTAGCCTCTT CTGTCACCCT-3'; BNP F: 5'-TCGGCGCAGTCAGTC GCTTG-3', and R: 5'-CGCAGGCAGAGTCAGAAGCCG-3'; β -MHC F: 5'-TGCAAAGGCTCCAGGTCTGAT-3', and R: 5'-GCCAACACCAACCTGTCAAG-3'; β -actin F: 5'-GCC ATGTACGTAGCCATCCA-3', and R: 5'-GAACCGCTCATT GCCGATAG-3'.

Western blot analysis. Protein was isolated from cultured cells using lysis buffer that contained protease/phosphatase inhibitors. The quantification was determined by the BCA Protein Assay kit (Bio-Rad Laboratories, Inc., Hercules, CA, USA). Subsequently, after 5 min of denaturation at 95°C, samples in each group were separated by sodium dodecyl sulfate polyacrylamide (SDS-PAGE) gel. The PVDF membranes (EMD Millipore, Billerica, MA, USA) were used for the tank blot system. Next, the membrane was incubated with 5% skimmed milk containing 0.05% Tween-20 (TBST) at room temperature for 2 h. The primary antibodies were incubated at 4°C overnight: anti-Nrf2 (1:1,000, ab62352; Abcam, Cambridge, UK), anti-NQO-1 (1:800, ab34173), anti-HO-1 (1:1,000, ab137749), anti-BNP (1:500, ab19645), anti- β -MHC (1:500, ab23990), anti-GAPDH (ab9385, 1:5,000). The horseradish-peroxidase-coupled secondary IgG antibodies were incubated at room temperature for 1 h. ECL Western Blotting Substrate (Amersham; GE Healthcare, Chicago, IL, USA) was adopted to develop chemiluminescence signals using the ChemiDoc XRS Imaging System (Bio-Rad Laboratories, Inc.). Finally, the expression level

was calculated by the densitometric analysis (Quantity One; Bio-Rad Laboratories, Inc.).

Statistics. GraphPad Prism 6.0 (GraphPad Software, Inc., La Jolla, CA, USA) was used to perform statistical analyses. The data were shown as mean \pm standard deviation. One-way ANOVA with Dunnett's multiple comparison test was used to compare the difference the multiple groups (>2 groups). $P < 0.05$ was considered to indicate a statistically significant difference.

Results

HSYA reduced MI-induced hypertrophy in rats. As shown in Fig. 1A, animals in model group showed an obvious increase in the heart size, compared to sham and control group. By contrast, the treatment with captopril (Ka) or HSYA alleviated the cardiac hypertrophy. Moreover, there was a blunted (in HSYA-L group) or significant decrease (in Ka and HSYA-H group) in the heart weight/body weight ratio compared to model group (Fig. 1B). Furthermore, the H&E staining showed that the enlarged cell area of myocytes induced by MI was attenuated by the treatment with Ka or HSYA. And this reduction was more obvious in HSYA-H group than in HSYA-L group (Fig. 1C).

HSYA rescued the expression of Nrf2 in rats. Research has reported that the abrogation of Nrf2 may enhance atrial hypertrophy in response to exercise stress (24). Thus, we detected the expression of Nrf2 in myocardial tissue. The results showed that the Nrf2 expression was depressed sharply in model group both in mRNA and protein levels. The treatment with Ka or HSYA recovered its expression and the recovery effect was most obvious in HSYA-H group (Fig. 2A-C). There was no significant difference observed in Nrf2 protein levels in both HSYA-H and HSYA-L group.

HSYA rescued the expression of Nrf2 in H9c2 treated with Ang II. To further confirm the effect of HSYA on hypertrophy after MI and the role of Nrf2 in hypertrophy, we performed subsequent experiments in H9c2 cardiomyocytes. The transfection efficiency of Nrf2 was presented in Fig. 3A-C. Ang II was used to set up the hypertrophy model in the study. We observed that the Nrf2 expression was mitigated by about 50% in Ang II group both in terms of the transcriptional and translational levels. Noticeably, the treatment with HSYA inhibited this decrease in A/H group. By contrast, the combination use of Ang II and siNrf2 aggravated the reduced expression of Nrf2, however, the effect of HSYA was reversed by the siNrf2 treatment (Fig. 3D and E).

Downregulation of Nrf2 reversed the protective effect of HSYA in H9c2 treated with Ang II. CCK-8 results showed that the Ang II-induced decreased cell viability was deteriorated in A/siNrf2 group but improved in A/H group. And the cell viability was smaller in A/H/siNrf2 group than that in A/H group (Fig. 4A). Moreover, while the leucine incorporation was induced by Ang II, it was reduced by the treatment of HSYA. The treatment with siNrf2 enhanced the leucine incorporation both in A/siNrf2 group and A/H/siNrf2 group, compared to Ang II and A/H group (Fig. 4B). In addition, the expression of hypertrophy-related genes was examined. As shown in

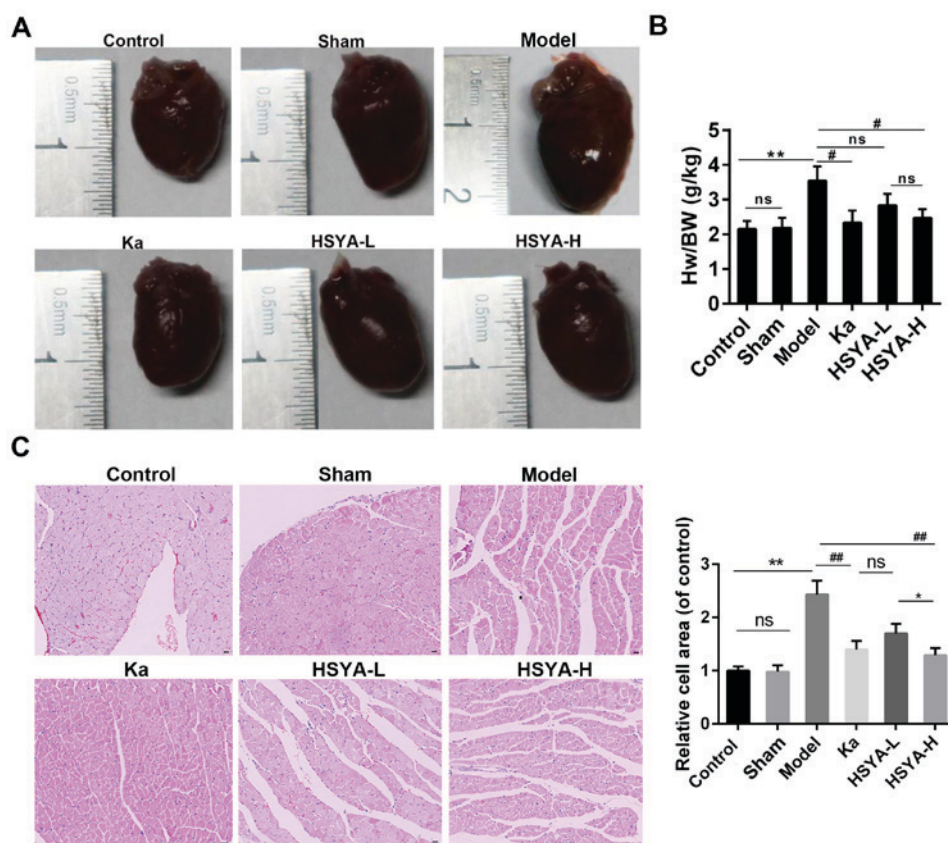


Figure 1. HSYA reduced hypertrophy after MI *in vivo*. (A) The heart size in each group; (B) The heart weight/body weight ratio; ** $P < 0.01$, # $P < 0.05$ as indicated. ns, no significant difference; (C) The H&E staining for myocardial tissue, scale bar, 20 μm . HSYA, Hydroxysafflor yellow A; MI, myocardial infarction; H&E, hematoxylin and eosin.

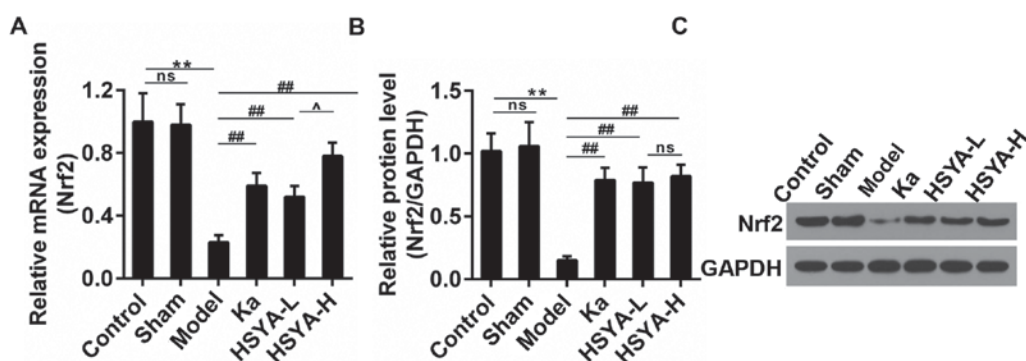


Figure 2. HSYA rescued the declined expression of Nrf2 caused by MI. The expression of Nrf2 was measured by (A) RT-qPCR and (B and C) western blot analysis. ** $P < 0.01$, ## $P < 0.01$, ^ $P < 0.05$ as indicated. ns, no significant difference. HSYA, Hydroxysafflor yellow A; MI, myocardial infarction; RT-qPCR, reverse transcription-quantitative polymerase chain reaction.

Fig. 5A and B, the increased mRNA expressions of BNP and β -MHC caused by Ang II was depressed by HSYA, and the effect of HSYA was mitigated by the downregulation of Nrf2. Consistently, the protein level of BNP and β -MHC showed a similar trend to mRNA level (Fig. 5C and D), suggesting that the protective effect delivered by HSYA was reversed by the downregulation of Nrf2.

HSYA altered the oxidative stress and the activation of NQO-1/HO-1 signaling pathway. As shown in Fig. 6A-C, the treatment of Ang II caused the oxidative stress in

H9c2, and this was achieved by triggering ROS accumulation, increasing the MDA content and inhibiting the SOD activity. The oxidative stress was increased and decreased by the down-regulation of Nrf2 and the treatment of HSYA, respectively. By contrast, the treatment of siNrf2 partly reversed the effect of HSYA. NQO-1/HO-1 are the typical downstream targets of Nrf2 through ARE. Thus, we detected the expressions of these two genes. Our results revealed that the treatment of Ang II inhibited the expressions of NQO-1 and HO-1, and combined treatment with Ang II and siNrf2 presented even more obvious effects than treatment with

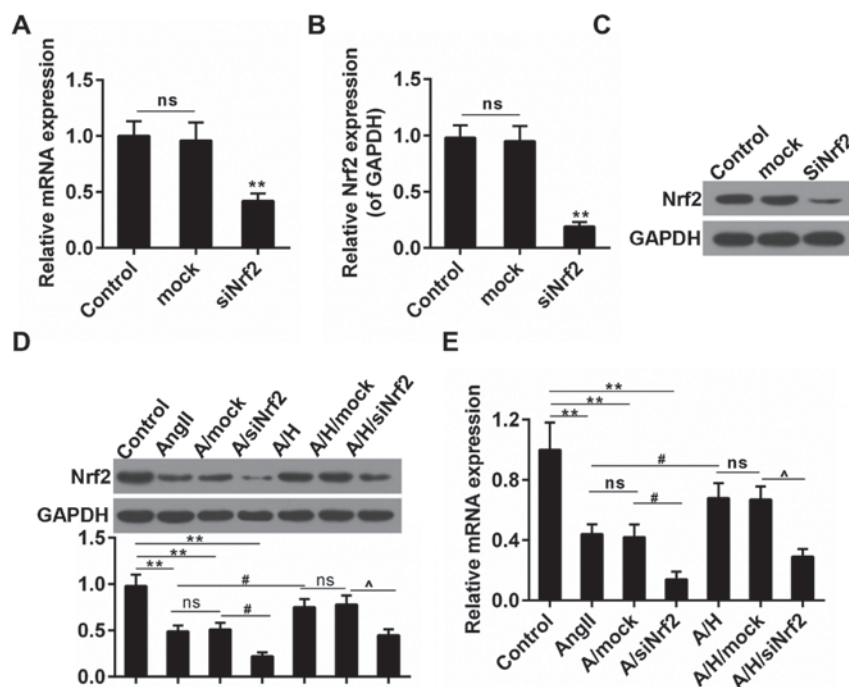


Figure 3. The effect of HSYA on Nrf2 expression *in vitro*. (A-C) The transfection efficiency of siNrf2 was examined by (A) RT-qPCR and (B and C) western blot analysis; (D and E) The effect of HSYA on the expression of Nrf2 through (D) western blotting and (E) RT-qPCR; #P<0.05 and ^P<0.05, **P<0.01 as indicated. ns, no significant difference; HSYA, Hydroxysafflor yellow A; Nrf2, nuclear factor (erythroid-derived 2)-like 2; RT-qPCR, reverse transcription-quantitative polymerase chain reaction.

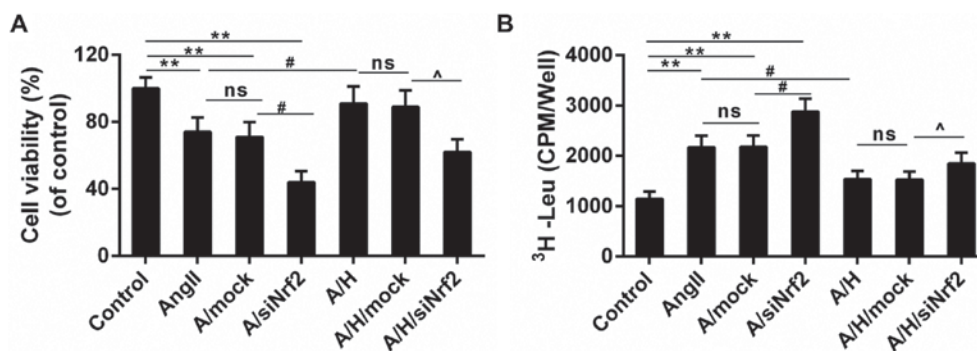


Figure 4. HSYA improved the cell viability and reduced the cardiac hypertrophy. (A) CCK-8 test for cell viability; **P<0.01, #P<0.05 and ^P<0.05, no significant difference (ns); (B) ³H Leucine incorporation assay to detect the protein synthesis rate; **P<0.01, #P<0.05 and ^P<0.05 as indicated. ns, no significant difference. HSYA, Hydroxysafflor yellow A; CCK-8, Cell Counting Kit-8.

Ang II alone. Therefore, while HSYA apparently abolished the effects of Ang II on the expressions of NQO-1 and HO-1, the down-regulation of Nrf2 partly blocked the effect produced by HSYA (Fig. 6D-G).

Discussion

The onset of MI would happen when the coronary artery occlude, and this could induce cardiac hypertrophy (25,26). This pathological hypertrophy in the heart will cause many ailments, leading to morbidity and mortality (27,28). This study uncovered a protective effect of HSYA on cardiac hypertrophy after MI *in vivo* and *in vitro*. Animals and/or H9c2 cells treated with HSYA exhibited reduced heart size, declined hypertrophic area and oxidative stress, which were closely related to Nrf2 signaling pathway.

In this work, HSYA alleviated the heart size and heart weight/body weight ratio induced by MI in rats. The cross-sectional area of myocardial cells, an indicator of hypertrophy, was reduced by the administration of HSYA in rats. Importantly, the expression of Nrf2 was depressed in hypertrophic cardiomyocyte *in vivo*, which was recovered by HSYA. These results pointed to the protective role of HSYA, in which the activation of Nrf2 may be involved.

To confirm the role of HSYA and Nrf2 in hypertrophy caused by MI, we conducted experiments using H9c2 *in vitro*. Consistently, the expression of Nrf2 was decreased in H9c2 under the stimuli of Ang II that was combated by the treatment of HSYA. Although in HSYA-L group, the expression of Nrf2 was significantly rescued, the Hw/BW was not decreased significantly. This discrepancy may be caused by the complexity of the intracellular regulation network. The

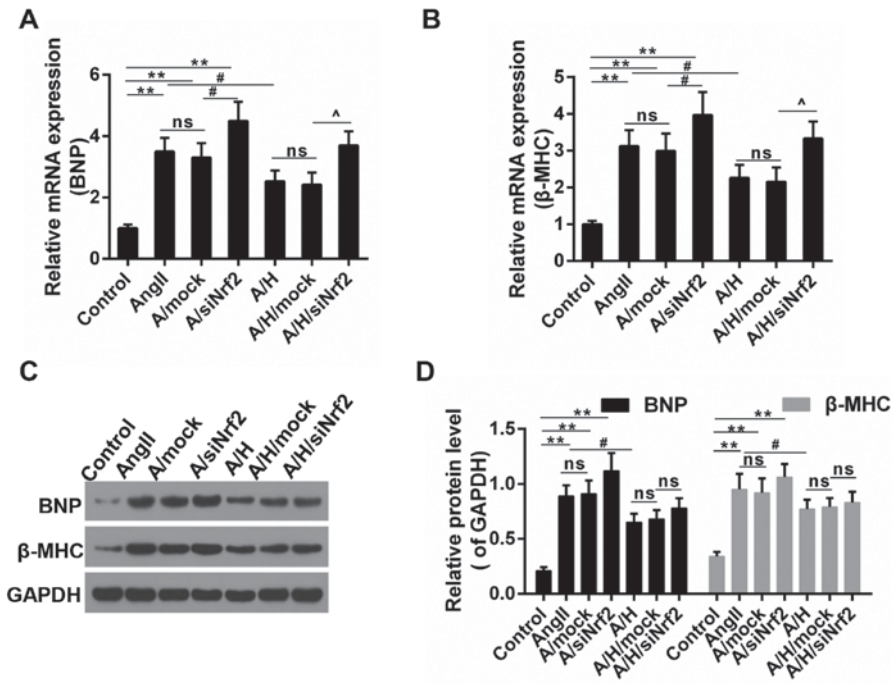


Figure 5. HSYA decreased the expression of hypertrophic factors. RT-qPCR [(A) for BNP, and (B) for β-MHC] and western blot analysis [(C-D) for BNP and β-MHC]. **P<0.01, #P<0.05 and ^P<0.05 as indicated. ns, no significant difference. HSYA, Hydroxysafflor yellow A; RT-qPCR, reverse transcription-quantitative polymerase chain reaction; BNP, brain natriuretic factor; β-MHC, β-myosin heavy chain.

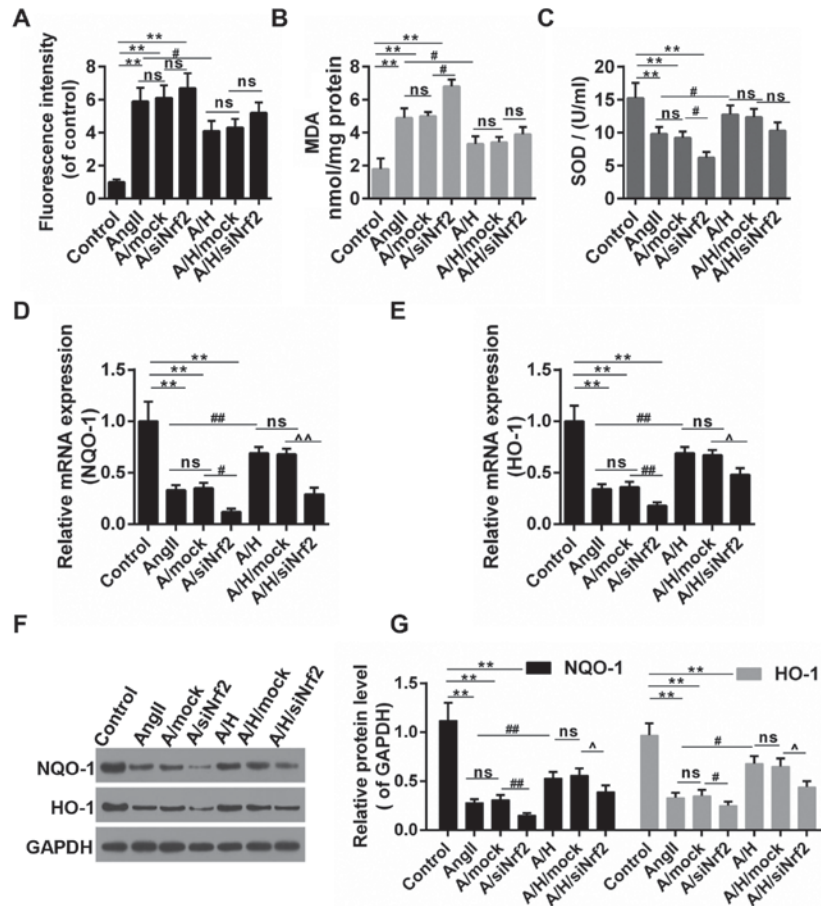


Figure 6. HSYA ameliorated oxidative stress and increased the expression of NQO-1 and HO-1. (A) ROS production was examined by DCHF-DA; (B) The content of MDA; (C) The activity of SOD; (D-G) The expression NQO-1 and HO-1 by RT-qPCR [(D) for NQO-1, (E) for HO-1] and (F and G) Western blot assays. **P<0.01, #P<0.05, ##P<0.01, ^P<0.05 and ^^P<0.01 as indicated. ns, no significant difference. HSYA, Hydroxysafflor yellow A; RT-qPCR, reverse transcription-quantitative polymerase chain reaction; NQO-1, NAD(P)H:quinone oxidoreductase 1 (NQO1) HO-1 heme oxygenase-1; MDA, malondialdehyde; SOD, superoxide dismutase; ROS, reactive oxygen species.

crosstalk between signals is very common in the living cells. Some other signals (that was not identified in this study) may also affect the Hw/BW.

The downregulation of Nrf2 blunted the effect of HSYA. Accompanied by the reduced cell viability, Ang II increased the [³H] leucine incorporation of cardiomyocytes. This was in line with the results obtained in a previous study (29). Moreover, the expressions of hypertrophic markers, BNP and β -MHC, was significantly induced by Ang II. The actions of Ang II were deteriorated by the decreased Nrf2. By contrast, the treatment of HSYA attenuated the effect of Ang II. Nevertheless, the effect of HSYA was partly blocked by the down-regulation of Nrf2. These results suggested that the anti-hypertrophic effect of HSYA was largely dependent on the activation of Nrf2. Consistently, in a previous study, the over-expression of Nrf2 could significantly inhibit the expression of the hypertrophic factor (30). These listed results confirmed that the activation of Nrf2 was helpful in combating the cardiac hypertrophy.

Researchers proposed that ROS contributed to cardiomyocyte hypertrophy (11,31). Moreover, the inhibition of ROS accumulation is helpful to the amelioration of hypertrophy (32). In our work, the ROS burst caused by Ang II was alleviated by HSYA. The intracellular ROS accumulation will lead to the disruption of redox system, for example, the activity of MDA and SOD (33,34). In this study, the content of MDA and the activity of SOD were dysregulated by Ang II, however, the treatment of HSYA recovered their balance. And the down-regulation of Nrf2 reversed the effect of HSYA. Taken together, we concluded that anti-hypertrophic effect produced by HSYA was closely associated with the anti-oxidative capacity. Coincidentally, Nrf2 is outstanding through its ARE in the protection against oxidative stress. NQO-1 and HO-1 are two downstream targets of Nrf2. In addition, the induction of NQO-1 and HO-1 was believed to have protective effects on oxidative injury (35-37). Therefore, we speculated that HSYA may prevent cardiac hypertrophy in relation with the activation of NQO-1/HO-1. Our results showed that the expressions of NQO-1 and HO-1 were higher in A/H group than those in Ang II and A/siNrf2 groups in terms of transcriptional and translational levels, suggesting that the activations of NQO-1 and HO-1 contributed to the protective role of HSYA. In addition, the results showed that the down-regulation of Nrf2 reversed the effect of HSYA on the expressions of NQO-1 and HO-1. These data suggested that Nrf2 was necessary for the protective effects of HSYA against cardiac hypertrophy caused by MI. The cardio-protective role HSYA as such was consistent with previous studies (38-40). Therefore, the results illustrated that HSYA delivered a positive effect on the activation of Nrf2/NQO-1/HO-1 pathway, which is a promising thread for increasing the antioxidant and anti-hypertrophic effect.

Collectively, we discovered that the protective effect of HSYA on cardiac hypertrophy after MI. The anti-oxidant capacity of HSYA may be associated with this work model. Moreover, some studies have pointed out that HSYA could exert anti-apoptotic effect on tissue injury (20,41,42). Thus, to assess other aspects of this protective effect could also lead to productive outcome. Furthermore, we found that Nrf2 signaling was necessary to the effect produced by HSYA. Nevertheless, due to the complexity of cellular signal cascades, we did not

exclude the possibility that other signaling pathways may play a part in the protective effect conferred by HSYA.

In summary, HSYA significantly mitigated cardiac hypertrophy *in vitro* and *in vivo*. This effect may depend on the antioxidant effects via Nrf2/NQO-1/HO-1 signaling pathway. Our study inspired insight into the mechanisms that is responsible for HSYA. Our results may provide effective regimen and strategies to relieve heart failure.

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Availability of data and materials

The analyzed data sets generated during the study are available from the corresponding author on reasonable request.

Authors' contributions

BN, SL and DZ provided substantial contributions to conception and design of the present study. YJ contributed to the data acquisition, data analysis and interpretation. SL drafted the article and critically revised it for important intellectual content. All authors read and approved the final manuscript.

Ethics approval and consent to participate

The present approved by Animal Subjects Committee of the Affiliated Hospital of Hangzhou Normal University.

Patient consent for publication

Not applicable.

Competing interests

The authors declare that they have no competing interests.

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